

Review

# Epigenetic Modifications in Genome Help Remembering the Stress Tolerance Strategy Adopted by the Plant

Suresh Kumar<sup>1,\*</sup>, Trilochan Mohapatra<sup>2,\*</sup><sup>1</sup>Division of Biochemistry, ICAR-Indian Agricultural Research Institute, 110012 New Delhi, India<sup>2</sup>Protection of Plant Varieties and Farmers' Rights Authority, 110012 New Delhi, India\*Correspondence: [sureshkumar3\\_in@yahoo.co.uk](mailto:sureshkumar3_in@yahoo.co.uk); [sureshkumar@iari.res.in](mailto:sureshkumar@iari.res.in) (Suresh Kumar);  
[mohapatrat1962@gmail.com](mailto:mohapatrat1962@gmail.com); [tmnrpcb@gmail.com](mailto:tmnrpcb@gmail.com) (Trilochan Mohapatra)

Academic Editor: Changsoo Kim

Submitted: 18 November 2023 Revised: 15 January 2024 Accepted: 26 January 2024 Published: 22 March 2024

## Abstract

Genetic information in eukaryotic organisms is stored, replicated, transcribed, and inherited through the nucleus of a cell. Epigenetic modifications in the genetic material, including DNA methylation, histone modification, changes in non-coding RNA (ncRNA) biogenesis, and chromatin architecture play important roles in determining the genomic landscape and regulating gene expression. Genome architecture (structural features of chromatin, affected by epigenetic modifications) is a major driver of genomic functions/activities. Segregation of euchromatin (transcriptionally active) from heterochromatin (transcriptionally repressed chromosome) and positioning of genes in specific nuclear space in eukaryotic cells emphasise non-randomness in the organization of the genetic information. Not only does the base sequence of a gene carry the genetic information but the covalent modifications of bases, three-dimensional positioning of the genome, and chromatin loops are vital for switching on/off the gene and regulating its expression during growth/environmental stress. The epigenetic dynamics depend on the activities of writers and erasers under changing environmental conditions. The discovery of non-coding RNAs (one of the players in *de novo* methylation of DNA), increased DNA methylation protein (guide for the DNA demethylase), and methylation monitoring sequence (that helps keep a balance between DNA demethylation and methylation) have been some of the new developments in the era of epigenomics. To respond to environmental stimuli, plants depend on modulating gene expression through different mechanisms including biochemical, molecular, genetic, and epigenetic alterations. Studies on plants might provide better insights into epigenetic stress memory and molecular bases of adaptability to enable (epi)genome editing of crops for climate resilience and sustainable agriculture in the present era of multifaceted climate change.

**Keywords:** epigenetic modification; 5-methylcytosine; histone modification; genome architecture; abiotic stress; epigenetic memory

## 1. Introduction

Unlike animals, plants are continuously exposed to a variety of environmental stresses. Hence, several strategies (morphological, physiological, biochemical, molecular, genetic, and epigenetic) have been evolved by plants to cope with environmental stresses [1,2]. Our understanding of the stress sensing, signaling, and responses of plants has considerably increased over the past few decades. Growing evidence suggests that chromatin remodeling, histone modifications, DNA methylation, and non-coding RNAs play important roles in epigenetic regulations of gene expression under abiotic stresses [3,4]. Evidence also suggests that DNA not only carries the genetic information required for the expression of a trait but epigenetic modifications on nitrogenous bases of DNA modulate chromatin architecture to control the expression of gene/trait [5]. The genome is a high-order organization of genetic information [6], which is modulated by epigenetic modifications. During the last two decades, the genome and the epigenome of eukaryotic organisms have been extensively studied to better understand the gene regulatory mechanisms. It is becoming clear that the genetic information and functions de-

termined by a genome can be modulated under changing environmental conditions through epigenetic modulations leading to alterations in 3D chromatin organization in the nucleus [7,8]. Since the epigenetic state of the genetic material (chromatin) varies with the changing environmental conditions (depending on the activity of the epigenetic machinery), the transfer of a gene/trait from one species to another would not only require the transfer of the gene but also the epigenetic milieu for the gene to express appropriately [9]. Therefore, understanding the epigenetic setup of a gene of interest in the donor plant becomes necessary to ensure the effective transfer of the gene/trait to the recipient.

Recent advances in high-throughput genome and epigenome analyses have provided unprecedented opportunities to generate genomic and epigenomic maps in unraveling the (epi)genomic landscape at single-base resolution. However, there are certain challenges in presenting the whole epigenome, particularly under changing environmental conditions in different tissues of an organism. The task becomes further complicated in compiling the unified information for methylome, histone modifications, and ncRNA biogenesis, as these epigenomic com-



ponents vary differently with the changing environmental conditions in different cells/tissues. Generally, the epigenetic changes observed in a regular analysis present the average changes of a population of cells in the sample tissue; hence, epigenomic analysis at the single-cell level needs to be captured/performed [10,11]. Thus, challenges in epigenomics include the required technological advancements to generate reliable/consistent data necessary for understanding the stable, reversible, and heritable components of the epigenome.

Modifications in DNA bases (at the 5th carbon of cytosine residue and/or N<sup>6</sup> position of adenine residue) play important roles in the epigenetic regulation of genes in animals as well as plants [5]. Methylation of DNA bases is catalyzed by different methyltransferases, wherein the required methyl group is supplied by S-adenosyl-L-methionine. In contrast, active DNA demethylation is performed through the base excision repair (BER) pathway in mammals as well as plants [5,12]. Though the RNA-directed DNA methylation (RdDM) pathway has been reported to play vital roles in *de novo* DNA methylation in plants, it has not been reported in mammals [13]. While active DNA demethylation in mammals initiates with deamination (removal of the amino group) and/or oxidation of 5-methylcytosine (5-mC), the excision of 5-mC is performed by methylcytosine DNA glycosylase in plants [12,14]. Another important base modification in DNA is N<sup>6</sup>-methyladenine (6-mA) which plays regulatory roles in plants [5]. While *Arabidopsis* has been a model plant species to understand the epigenetic machinery in plants, they are being tested/validated in rice and other crop plants. The presence of alkylating agents in the environment was reported to generate 1-mA in *Escherichia coli*, which prohibits regular Watson–Crick base pairing and blocks DNA replication [15]. An *AlkB* gene in *E. coli* is induced by the alkylating agents, which is responsible for the adaptive response of the organism. A human *AlkB* homolog was reported to perform similar functions [16]. Similarly, the presence of an alkylating agent caused the generation of N<sup>7</sup>-methylguanine (7-mG). A significantly higher level of 6-mA in *C. elegans* and *Drosophila melanogaster* has been reported, but a negligible amount of 5-mC or 5-hmC was recorded [17,18]. While DNA 6-mA-demethylase (DMAD, a TET-homolog) serves as an eraser for 6-mA, SeqA protein was reported to act as a reader of 6-mA.

Accessibility of a genomic region to transcriptional machinery is also modulated by post-translational modifications in the histone proteins and chromatin-remodeling complex that regulates nucleosome assembly and spacing [19]. Many amino acid residues, mostly at the N-terminal tail protruding out of the core histone, are subjected to covalent post-translational modifications. The rate of transcription of a gene is affected by different sites/degrees of histone modifications (e.g., H3K27ac, H3K9me2, H3K9me3, and H3K27me3), which influence the association between

DNA and histone proteins, thus affecting the chromatin accessibility [20–22]. In addition, histone modifications are recognized by proteins like ATP-dependent chromatin remodelers which affect local chromatin status and regulate the expression of the gene [19,23]. H3K9me2, established by SUVH4/5/6, is required for maintaining DNA methylation at heterochromatic regions in *Arabidopsis* [24,25]. Ubiquitination of histone, when occurs at the C-terminal tail of H2A, causes transcriptional repression; whereas ubiquitination at the C-terminal tail of H2B causes transcriptional activation [26]. Moreover, histone variants have also been reported to influence nucleosome stability and interaction with the mRNA processing factors [27]. Several studies report that such epigenetic marks are involved in flowering, seed development, nitrogen fixation, and abiotic/biotic stress responses [26,28,29]. Roles of the important components like ncRNA (triggering *de novo* DNA methylation), Increased DNA Methylation (IDM) protein (involved targeted DNA demethylation), and methylation-sensing genetic element (maintains DNA methylation/demethylation homeostasis) in epigenomic regulation of genes/traits are being deciphered [25,30].

## 2. Genomics of Environmental Adaptation

Genome in the eukaryotes is not randomly placed in the nucleus, but it is packed into a high-order chromatin structure, which plays important roles in its functions. Understanding nuclear genome organization is attracting significant attention because processes like DNA replication/transcription, genome stability/integrity, etc. (necessary for appropriate growth/development, and stress tolerance) are regulated through chromatin structures. Eukaryotic genome organization can be studied at three levels namely (1) linear genome (nucleotide sequence), (2) epigenomic alterations (modified DNA bases, histone modifications), and (3) three-dimensional (3D) structure of the genome (depicting the arrangements of chromatins/chromosomes in the eukaryotic nucleus) [11,31]. Chromatins are dynamic during cyclic compaction/decompaction through cell division, cellular differentiation/developmental processes, as well as defense responses, which regulate/fine-tune the expression of genes [32,33]. Environmental fluctuations necessitate modulations in cellular processes by switching on/off the gene in response to stress [11,34]. Transient chromatin compaction (e.g., loosely-packed euchromatin or more compact heterochromatin) and modulation in chromatin architecture under environmental stress have been demonstrated in animals as well as plants [34–37]. Environmental stresses are reported to cause alterations in chromatin architecture, improving accessibility of the stress-associated genes to transcriptional machinery through chromatin remodeling (shifting/removal of histones) [38], post-translational modifications of histones [19,39] or replacing the histone variants [40].

In addition to the genetic factors, gene expression is also controlled by the combined actions of other regulatory mechanisms that alter chromatin architecture including DNA base modifications [5,11], histone modifications [41,42], chromatin remodeling [43,44], etc. Different chromatin remodelers like CHD, ISWI, INO80, and Switch/Sucrose non-fermenting (SWI/SNF) act on chromatin architecture under different environmental conditions to alter the transcriptionally inactive to transcriptionally active chromatin state. Alterations in chromatin architecture at the promoter region are more crucial in affecting the expression of the gene [45,46]. Developmental/environmental stimuli alter the epigenetic landscape/chromatin architecture that modulates the expression of the gene(s) necessary for proper growth and to cope with stress [11]. Some of these epigenetic marks/transcriptional repressors cooperate with chromatin remodelers to alter chromatin structure and fine-tune the expression of the gene(s). Some of these epigenetic modifications may get transmitted through the cell division/reproductive cycle and help to cope with the reoccurring stress [47]. However, identification/validation of the stress-induced heritable epigenetic mark would be necessary for utilizing it in stress tolerance improvement programs.

### 3. Epigenetics of Modified DNA Bases

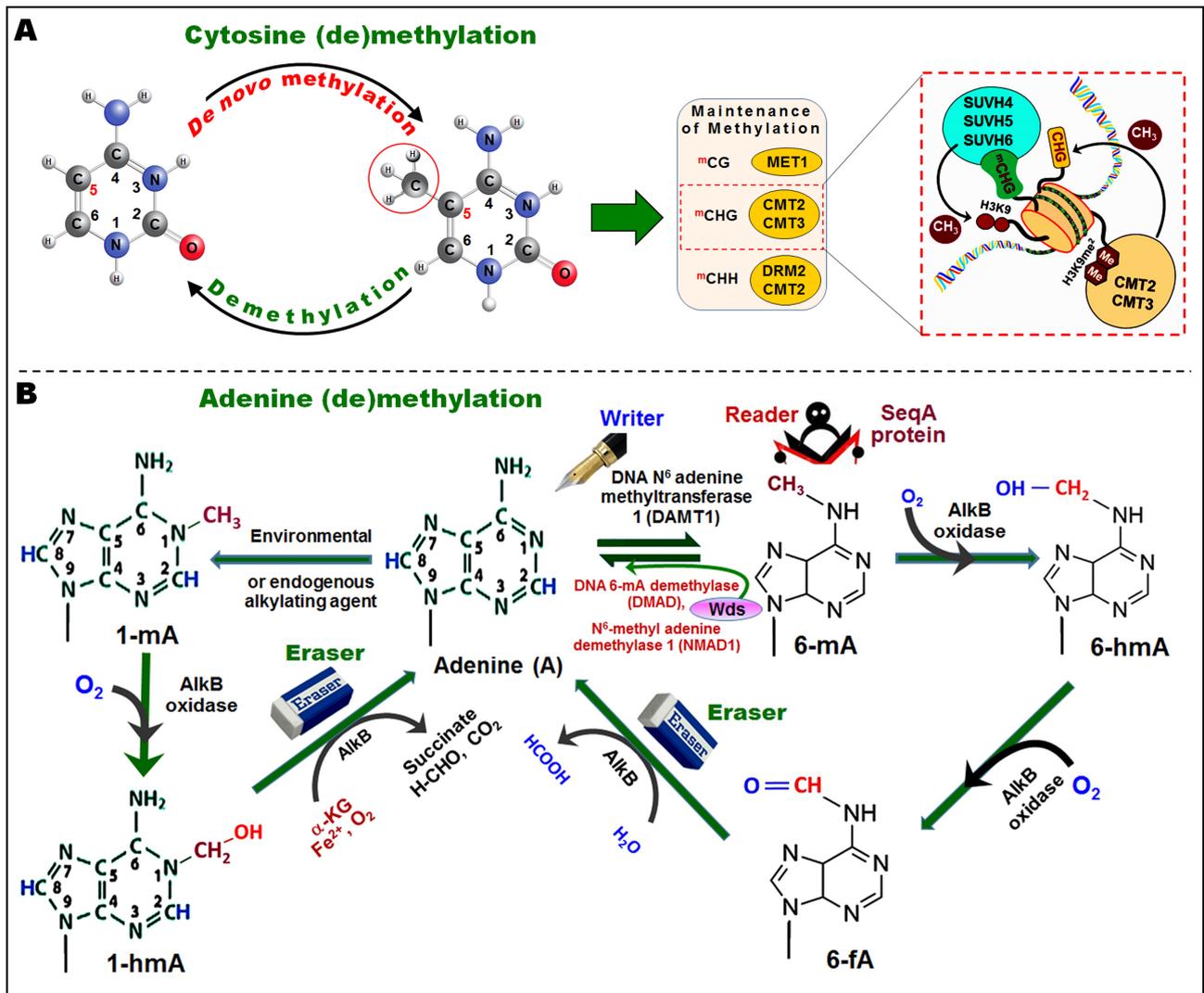
Initially considered to be a host defensive mechanism in prokaryotes, DNA base modifications, mainly cytosine methylation, have now been recognized to play vital roles in the regulation of gene expression in eukaryotes. In the last two decades, several epigenetic changes occurring during the developmental processes/abiotic stresses in animals and plant genomes have been reported [2,48]. In most cases, methylation of promoter has been reported to repress the rate of transcription of the gene, mainly because of the formation of a repressive-chromatin structure affecting the binding of transcription factor on acquiring its bonding site by a methylated-DNA binding protein [49]. However, hypermethylation of promoter not causing transcriptional repression has also been reported [50], indicating that transcriptional responses of promoter methylation are context-specific. Reversal of the epigenetic modification is required for restating the gene activity, particularly in responses to environmental stress.

Only some of the genes have been reported to be silenced due to methylation of promoter in Arabidopsis, which indicates that DNA base modification is not the only epigenetic regulatory mechanism. About one-third of the genes in Arabidopsis were reported to be methylated at gene-body [51]. Gene-body methylation appears to be a common phenomenon in plants, as it can affect up to 60% of the genes in some species [52]. Though the function of gene-body methylation in plants is still controversial, recent findings suggest that it occurs under environmental stresses and is correlated with the fitness/adaptability of the

plant [52,53]. Generally, transposable/repetitive elements are heavily methylated in all the cytosine (CG, CHG, and CHH) contexts (Fig. 1A), but gene-body methylation meagrely occurs in non-CG contexts [51,54,55]. Gene-body methylation (occurring in the coding region between transcription start and termination sites) [56] and methylation in different cytosine contexts of transposable element (TE) or repetitive elements present in the introns of a gene might control mRNA processing. Epigenetic modulation of retrotransposon in the homeotic gene was reported to affect alternative splicing resulting in premature termination of transcription [57].

Similar to cytosine methylation in DNA, an adenine can also get methylated due to the addition of a CH<sub>3</sub> group at the N<sup>6</sup> or N<sup>1</sup> position [5,58]. Methylation of exocyclic NH<sub>2</sub> at the sixth carbon (C6) of the purine ring of adenine forms N<sup>6</sup>-methyladenine (6-mA), while methylation of cyclic N (N<sup>1</sup>) results in N<sup>1</sup>-methyladenine (1-mA) in the presence of endogenous/environmental alkylating agent [15]. While an AlkB homolog was reported to perform adaptive responses to environmental stresses in humans [16], a similar factor is also expected to be present in plants. Although the role of methylated DNA-binding proteins has been reported to be conserved in many organisms, several questions regarding the role of adenine (de)methylation homeostasis in plants remain unanswered. Generally, the detection of 6-mA is difficult in higher eukaryotes mainly because of its low abundance or presence for a shorter period. Recent advances in high-throughput and more sensitive techniques have resulted in a better ability to detect 6-mA, which helps understand the epigenetic functions in animals and plants [59,60]. The enzymes involved in the conversion of adenine to 6-mA in animals have been well known [61], but little is known about adenine methyltransferase in plants [62]. The occurrence of 6-mA in the genome of Arabidopsis was reported adjacent to the transcription start site (TSS), which positively correlates with the expression of the gene. Only little is known about the proteins (readers/erasers) interacting with 6-mA in eukaryotes. The occurrence of 6-mA in the GAGG context was reported to be 20% in genes while 14% in TEs in rice [63]. Decreased 6-mA content in *HsfA1* (a repressor of HSP70) in a heat-tolerant genotype was reported to improve stress tolerance in rice [64]. Thus, the possible action of 6-mA includes activation as well as silencing of genes, affecting transgenerational chromatin functions under stress [65].

However, oxidation of methyl group (by demethylase, e.g., AlkB dioxygenase) of 6-mA results in its conversion to N<sup>6</sup>-hydroxymethyladenosine (6-hmA), which further converted to N<sup>6</sup>-formyladenosine (6-fA) and then gets demethylated to adenine [5] (Fig. 1B). Similarly, 1-mA might also get demethylated by the action of AlkB oxidase/AlkB enzyme via N<sup>1</sup>-hydroxymethyladenine (1-hmA). The 6-mA attracts the binding of an effector molecule (reader) that changes chromatin conformation



**Fig. 1. DNA base methylation in plant.** (A) *De novo* methylation of DNA in CG, CHG, and CHH contexts. After DNA replication, methylation in CG context is performed by methyltransferase 1 (MET1). Methylation in the CHG context is performed by chromomethylase 2 (CMT2) or CMT3, while methylation in the CHH context is performed by CMT2 or DRM2 through the RdDM pathway. Methylation in the CHG context attracts binding of H3K9-specific suppressor of variegation 3-9 homolog 4 (SUVH4), SUVH5, SUVH6 that generate dimethylated-H3K9 (H3K9me2), which promotes CMT2 and CMT3. (B) Adenine gets methylated by the addition of the CH<sub>3</sub> group at the N<sup>6</sup> position by the enzyme DNA adenine methyltransferase 1 which converts it to N<sup>6</sup>-methyladenine (6-mA). Moreover, hemimethylated 6-mA DNA attracts the binding of SeqA protein. The 6-mA might get hydroxylated to N<sup>6</sup>-hydroxymethylcytosine (6-hmA) by AlkB oxidase. Subsequently, an eraser like DNA 6-mA demethylase or N<sup>6</sup>-methyladenine demethylase-1 deaminates 6-mA to adenine. Environmental/endogenous alkylating agents can methylate adenine to N<sup>1</sup>-methyladenine (1-mA), which might get demethylated by the AlkB oxidase back to adenine via N<sup>1</sup>-hydroxymethyl adenine (1-hmA).

and/or transcription of the gene. A reader, SeqA protein, specifically binds to the hemimethylated (6-mA) DNA. The presence of 6-mA in the promoter region was reported to silence the gene, while it activates the expression of the gene when present in the coding region. These suggest that cytosine and adenine (de)methylation are context-specific, dynamic, and coordinated events. Although the functions of 6-mA in abiotic stress tolerance are still elusive, studies show dynamic changes in 6-mA level under heat, cold, and salt stress in rice [64]. Comprehensive analysis of coordi-

nated/combined actions of more than one epigenetic mark (e.g., 5-mC and 6-mA) in stress responses needs to be experimentally elucidated. Therefore, it would be quite interesting to investigate the interactions among the epigenetic marks to understand the complex epigenetic regulatory mechanisms and the epigenetic codons to unravel the biological enigmas [5,66].

Changes in DNA methylation have been detected in several prokaryotic and eukaryotic organisms. Modifications in DNA bases in context- and region-specific manner are catalyzed by specific enzymes. In plants, heterochromatic regions are generally enriched with 5-mCs, mainly in the repetitive sequences and TEs. The dynamics of DNA base modification depend on the revocability of the processes, depending on the activity of enzymes involved in the processes that switch on/off the gene. The complex epigenetic changes are being discovered in different organisms and possible combinations/interactions among the epigenetic marks (epimarks) indicate that epigenetic codons are more complex than they are being thought [5]. Four bifunctional DNA glycosylases including Demeter (DME), Demeter-like protein 2 (DML2), DML3, and Repressor of silencing 1 (Ros1) have been known in Arabidopsis [67] that can remove methylation in any sequence context [68–70]. An AT-rich TE is favourably demethylated by DME, leading to changes in the expression of nearby genes [71–73]. Demethylation of TE by ROS1 affects transposon activity and silences the nearby genes [74]. ROS1 is also involved in demethylation of the RdDM-independent regions [75,76]. The genomic regions demethylated by ROS1 are characterized by lowered H3K27me and/or H3K9me2 but increased H3K18Ac and/or H3K27me3 marks [74]. Some of the target sites (methylated DNA sequences) of ROS1 are created by the binding of a histone acetyltransferase “increased DNA methylation 1” (IDM1), which acetylates H3 at the site deprived of H3K4me2 and H3K4me3 [77]. Interestingly, a 39 bp “methylation monitoring sequence” (MEMS) is present in the promoter of *ROS1* that functions as a sensor of MET1 and RdDM activities [5]. Moreover, the promoter of *ROS1* also contains a transposon (Helitron) upstream of MEMS, which binds to cytosine methylation factors and thus makes the promoter responsive to the methylation level. Thus, like a thermostat of a machine, MEMS acts as a “methylstat” that senses and maintains the ROS1-dependent methylation level in a genomic region [78,79]. The presence of such “methylstat” is essential for cytosine methylation dynamics and it has been reported not only in plants but also in animal systems [80,81].

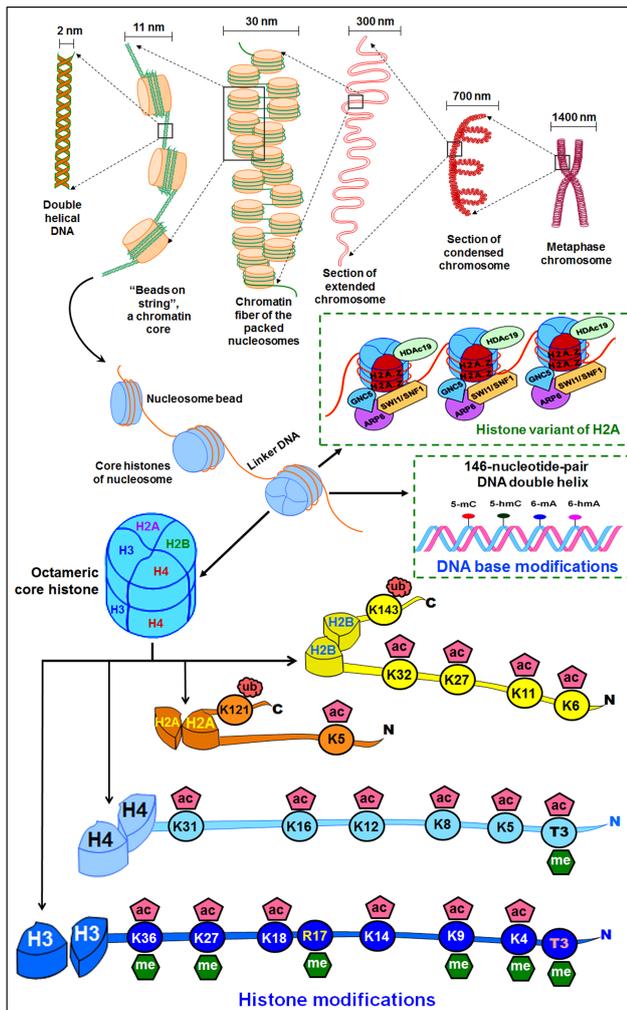
#### 4. Epigenetics of Histone Modifications

A histone octamer (two copies of each of the H2A, H2B, H3, and H4) wrapped around by a stretch of DNA (146 bp) forms a nucleosome (Fig. 2). The amino acid residues mostly at the protruding N-terminal tail, mainly of histone H3, are subjected to covalent post-translational modifications including acetylation, methylation, phosphorylation, ubiquitination, and SUMOylation, which play important roles in regulating gene expression during growth, development, and exposure to environmental stresses [82]. The post-translational modification of histone proteins, generally methylation and acetylation of histone 3 (H3)

at lysine-4 (i.e., H3K4), lysine-9 (H3K9), and lysine-27 (H3K27) are some of the most common histone marks associated with controlling gene expression [83]. Acetylation of H3, such as lys-9 (H3K9ac) and lys-27 (H3K27ac), correlates with transcriptional activation of a gene [84]. Similarly, trimethylation of H3 at lys-9 (H3K9me3) and lys-27 (H3K27me3) have varying effects based on the site of methylation. While H3K9me3 and H3K27me3 have been known to cause transcriptional suppression, H3K4me3 is associated with the transcriptional activation of a gene [85, 86]. The involvements of H3K4me2, H3K4me3, H3K9ac, H4K5ac, H4K8ac, and H4K12ac in the environmental stress tolerance of plants are becoming evident now [87].

Acetylation of a lysine residue in histone is dynamically controlled by the action of enzymes histone acetyltransferase (HAT) and histone deacetylase (HDAC). Acetyl-CoA is a cofactor of HAT in catalysing the transfer of an acetyl group to the  $\epsilon$ -amino group of a lysine residue [82]. This leads to the neutralization of the positive charge on lysine residue resulting in the weakening of the electrostatic interactions between histones and DNA; thereby, relaxation of the chromatin to improve accessibility of transcription machinery to the gene for transcription to take place [88]. Phosphorylation of histone may take place on serine, threonine and tyrosine residues at the N-terminal tail. Its dynamics are controlled by the action of kinase in adding a phosphate from ATP to the hydroxyl group of the amino acid side chain or by phosphatase in removing Pi. Phosphorylation adds a negative charge to the histone which affects the compactness of chromatin. Methylation of a histone mainly occurs at the lysine and arginine residues. However, unlike acetylation and phosphorylation, methylation does not cause any alteration in charge of the histone. Moreover, an additional level of complexity can be seen in the case of methylation; as lysine can be mono-, di- or tri-methylated, while arginine can be di-methylated (symmetrically or asymmetrically) or mono-methylated [82]. Interestingly, ubiquitination of histone is a bigger molecular change through covalent attachment of ubiquitin (a 76-amino acid polypeptide) to a lysine residue by a sequential action of activation (E1), conjugation (E2), and ligation (E3) enzymes. SUMOylation is another modification of histone that involves the covalent addition of a ubiquitin-like modifier to lysine by the actions of E1, E2, and E3 enzymes. SUMOylation can occur on all the core histones which antagonizes the effects of acetylation and ubiquitination, if it occurs on the same lysine residue [89,90].

Though modification of DNA base(s) and amino acid(s) at the tail of histones alter the chromatin architecture, the complexity of regulation of gene expression increases due to the cross-talk between different epigenetic modifications, which might help to fine-tune the gene expression (Fig. 2). The cross-talk may occur between the modified DNA base and the histone modification or be-



**Fig. 2. Organization of the genome and epigenetic modifications.** Packaging of chromosomal DNA in the nuclei through wrapping of DNA double helix on a histone octamer. The positively-charged histone proteins adhere with the negatively-charged DNA, forming a nucleosome. The nucleosomes fold into 30 nm chromatin fiber, forming loops of 300 nm, which coils to form 700 nm condensed chromatid of ~1400 nm Metaphase chromosome. DNA base modifications as well as the histone variants affect the genome functions and regulate gene expression. Modifications at the tail of core histone (H2A, H2B, H3, H4) proteins cause chromatin remodeling that affects the expression of the gene under abiotic stress.

tween the histone modifications. The effects of cross-talk between histone modifications might be (i) competitive antagonism, particularly when more than one modifications target the same site (as in the case of lysine where acetylation, methylation, and/or ubiquitination might occur), (ii) cooperative, or (iii) disruptive [82]. Similarly, there might be cooperation between DNA methylation and histone modification as it was reported for the binding of UHRF1 protein to methylated (H3K9me3) nucleosome in HeLa cell, which is significantly enhanced when the nu-

cleosome is methylated at CG site [91]. Moreover, DNA methylation can also inhibit protein binding to modified histone as it has been reported for the binding of KDM2A, which binds only to the nucleosome having H3K9me3 but not methylated at the DNA level in HeLa cell.

In addition to the modifications of canonical histone proteins, the incorporation of a histone variant (at different genomic regions, affecting the nucleosome stability and interaction with mRNA processing factors) as well as the chromatin-remodeling complex (regulating nucleosome assembly and spacing) also affect the nucleosome functions [19,27,92–94]. Histone variants are characterized by changes in a few amino acids which are encoded by separate genes. They not only affect nucleosome packaging but also modify the chromatin properties [95]. Evidence suggests that histone variants have effects on epigenetic state/transcription, chromosomal segregation, and repair of DNA damage [96]. All the core histones (except for the histone H4) and linker H1 show variants in plants. However, the focus has been on variants of H2A and H3. Among these, H2A.Z and H3.3 are evolutionarily conserved in the eukaryotes. Moreover, some tissue-specific and lineage-specific variants like H2A.W (functioning in flowering plants) and H3.10 (exists in the sperm cells of Arabidopsis only) are known [95]. Variant histone proteins exhibit distinct patterns of chromatin deposition. For example, the variants H2A.X and H2A.Z are concentrated in the euchromatin, while H2A.W is located mainly in the heterochromatic region. Furthermore, combinations of histone variants form thousands of different types of nucleosomes conferring extended potential for epigenetic codons in regulating various biological processes in plants, particularly under environmental stress. Variants of H2B are relatively less understood. One of the H2B variants (H2B.S) has been reported in sperm cells as well as in mature embryonic cells [95]. Three main variants of H3 have been known in plants including H3.1, H3.3, and centromeric H3 (CenH3) variants. The H3 variants (H3.1 and H3.3) are distinguished mainly by the amino acids present at four different locations namely 31st (Ala vs. Thr), 41st (Phe vs. Tyr), 87th (Ser vs. His), and 90th (Ala vs. Leu). These four different amino acids in H3.1 facilitate the recruitment of PRC2, which ensures the silencing of some of the development-related genes. Although H3.3 is predominant at the transcription end sites and reported to be associated with activation histone modification marks like H3K4me3, H3K9me3, and H3K36me3, it specifically affects transcription of genes associated with abiotic and biotic stresses. CenH3 is located mainly in the centromere for its formation and kinetochore assembly, particularly in the mitosis G2 phase. In recent years, manipulation of CenH3 has been exploited in haploid induction (HI) which can create true-breeding lines in a shorter period to accelerate the pace of plant breeding [97]. In addition to the above-mentioned variants of H3, some other variants of H3 like H3.10 and H3.15 are known in

Arabidopsis. While H3.10 plays important roles in epigenetic reprogramming in sperm/spermatocytes, H3.15 plays essential roles in callus formation in plants [98,99].

Research findings indicate that H2A.Z plays important roles in photo-morphogenesis and thermal morphogenesis in plants. While the accumulation of H2A.Z is essential for photo-morphogenesis, it plays an inhibitory role in thermal-morphogenesis. In rice, phosphate deficiency was reported to reduce H2A.Z accumulation on gene bodies of stress-responsive genes to facilitate the expression of the genes [100]. Transcription of drought-responsive genes was reported to be negatively correlated with the accumulation of H2A.Z in the gene body in Arabidopsis [101]. Similarly, heat stress affects flowering time in plants by altering the deposition of H2A.Z with species-specific effects [102]. H3.3 is involved in regulating flowering and seed germination as it is essential for transcriptional regulation of genes during germination [103].

Modification of histones (like H3K4me3, H3K9ac, H4K5ac, H4K8ac, and H4K12ac) in the promoter region of transcription factor (e.g., *WRKY*) gene has been implied to activate the expression of defensive gene(s) [104], suggesting their roles in epigenetic (histone modification) stress memory in plants. Application of acibenzolar S-methyl (a salicylic acid analog) was associated with histone modifications like H3K4me2, H3K4me3, H3K9ac H4K5ac, H4K8ac, and H4K12ac at the promoter of some of the defense associated genes [105]. Exogenous application of acetic acid was reported to promote jasmonic acid (JA) synthesis and enrichment of histone H4 acetylation, which primes the JA signaling pathway and enhanced drought tolerance in plants (Arabidopsis, rice, rapeseed, maize, and wheat) [106]. Methyl jasmonate (MeJA) was reported to increase the expression of defense-associated genes (*OsBBPI* and *OsPOX*) and modulated histone modification as well as DNA methylation. Thus, covalent (yet reversible) modification of histone interacts with DNA base modification and plays important roles in regulating gene expression. Nucleosome remodeling was reported to be implicated with heat stress memory in Arabidopsis [107]. FORGETTER1 (*FGT1*), encoding for a helicase, interacts with chromatin remodeling complexes (like SWI/SNF chromatin remodeler BRM) and functions as a factor required for heat stress memory. Studies also suggest that cold stress causes histone modifications (methylation and acetylation) at cold-responsive genes that can be manipulated for enhanced cold tolerance in plants [44,108].

## 5. Epigenomics of ncRNA Biogenesis

Non-coding RNAs (ncRNAs) are the regulatory RNAs that have significant impacts on the expression of coding genes. Based on the size, the regulatory RNAs are grouped into two types: (i) small non-coding RNAs (sncRNAs, <200 bases), which include miRNAs as well as siRNAs, and (ii) long non-coding RNAs (lncRNAs),

which are several hundred to several thousand nucleotides long [109]. The small interfering RNAs (siRNAs) are non-coding RNAs of 21-25 nt [110]. For rice, siRNAs have been grouped into two types, (i) heterochromatic siRNA (hc-siRNA) and (ii) trans-acting siRNA (tasiRNA) depending on their origin, processing, and sequence diversity [111]. While the hc-siRNAs (24 nt in size) are involved in DNA methylation/histone modification to silence TEs for genome integrity [112], tasiRNAs (21 nt long) derived from a double-stranded precursor transcript [113] are involved in post-transcriptional silencing of genes in plants [114].

Long non-coding RNA (lncRNA) is another important group of ncRNAs having >200 nt long transcripts in eukaryotes [115]. Like mRNAs, the lncRNAs are also transcribed by RNA polymerase II, followed by capping, splicing, and polyadenylation [116,117]. The lncRNAs are categorized into subgroups: (i) long intergenic non-coding RNAs (lincRNAs), (ii) enhancer RNAs (eRNAs), (iii) circular RNAs (circRNAs), and (iv) natural antisense transcripts (NATs) based on their structure and mode of action [118]. They have features like exon-intron splicing, 5' capping, polyA tail, and a lack of open reading frame (ORF) [119]. Many lncRNAs are involved in regulatory functions like chromosome dosage compensation, transcription, splicing, translation, imprinting, nuclear and cytoplasmic trafficking, cell cycle control, cell differentiation as well as epigenetic regulations [120–122]. Reports show that many lncRNAs play important roles as *cis* and/or *trans* regulators of biological processes like chromosomal dynamics, gene silencing, and embryonic stem cell differentiation in plants [123]. An eRNA is generated/transcribed from an enhancer domain in the proximity of protein-coding genes [124]. The eRNA directly affects the expression of nearby genes through transcription factors (TFs), chromatin-looping, and lysine-specific demethylase-2. An eRNA can recruit ATP-dependent nucleosome remodeller, which helps the formation of a histone-modification complex [125].

The circRNAs are evolutionarily conserved abundant ncRNAs in plants having a loop-like (covalently closed) structure [126]. It has also been reported to be produced as an intermediate/by-product of mRNA splicing [127] or from other ncRNAs by RNA editing enzymes [128]. In rice, the majority of circRNAs are flanked by non-GT/AG splicing signals, which suggests that plants also have an alternative mechanism for their biogenesis [129]. The circRNA interacts with miRNA as a sponge and it can regulate the gene expression negatively or positively [130]. Although circRNAs are derived from protein-coding genes, they are not translated into proteins; hence, classified as non-coding RNAs [131]. As a competitor, the lncRNA can inhibit the attachment of DNA-binding protein (e.g., TF) to the target site. The lncRNA inhibits binding of DNMT1 to the target DNA sequence and thus affects DNA methylation [132]. Moreover, lncRNA can reinforce the DNA methylation process by recruiting epigenetic modifiers to the target

site [133]. In mammals, lncRNA has also been reported to act as a precursor of miRNA by the action of Dicers [134].

## 6. Epigenetic Stress Memory and Adaptability

In animals, the inheritance of epigenetic changes over the generations requires their transit through germline cells without being erased by methylation surveillance mechanisms [135]. Like animals, plants do not own germline cells, instead, they produce gametes during sexual reproduction. Therefore, some of the epigenetic modifications attained during the vegetative phase of plant growth might be inherited through mitotic and meiotic cell divisions [136,137]. Hence, plants can remember the strategies used to fight against stressful conditions and it might be used by the plant to respond quickly to the stress on reoccurrence of the stress [138]. Plants have been reported to memorize defensive strategies against stress and trigger responses quickly on the recurrence of stress [139]. A part of the epigenetic changes induced by abiotic stress may be mitotically heritable (within the generation), which can last for several days (short-term memory) or even for the rest of plant life (long-term memory). However, abiotic stress can also induce certain epigenetic changes in plants that may show transgenerational heritability, at least to one non-stressed progeny (transgenerational memory). Although the basics of stress memories in plants have not been clear to date, reports on biochemical/metabolic, transcriptomic and/or epigenetic alterations indicate their role in short- as well as long-term memories [140–142].

A transcription activation mark (H3K4me3) was reported to play an important role in transcriptional memory [143]. To identify the heritable epigenetic marks/components associated with a trait of interest continuous efforts are being made so that they can be utilized in epigenome editing for crop improvement towards the development of climate-resilient crop varieties. However, the genome of many crop plants is considerably large and complex mainly because of their polyploid nature and occurrence of repetitive elements. How epigenetic changes superimpose the multiple copies of a gene in conferring genetic plasticity may provide some clues in developing tailor-made crop varieties to cope with multiple stresses being faced due to global climate change. Epigenetic engineering might be a potential way to achieve the desired adaptive plasticity without altering the underlying DNA sequence. However, stable inheritance of such epiallele would be essential to provide adaptive fitness/adaptability to the plant.

Under cold stress, acetylation of histone H3 by histone acetyltransferase (HAC1) was reported to promote the activation of *COR* genes which is involved in cold-stress memory [144]. Similarly, sRNA was reported to play a role in drought stress memory in Arabidopsis [136]. Up to 70% of the stress-induced epigenetic alterations have been reported

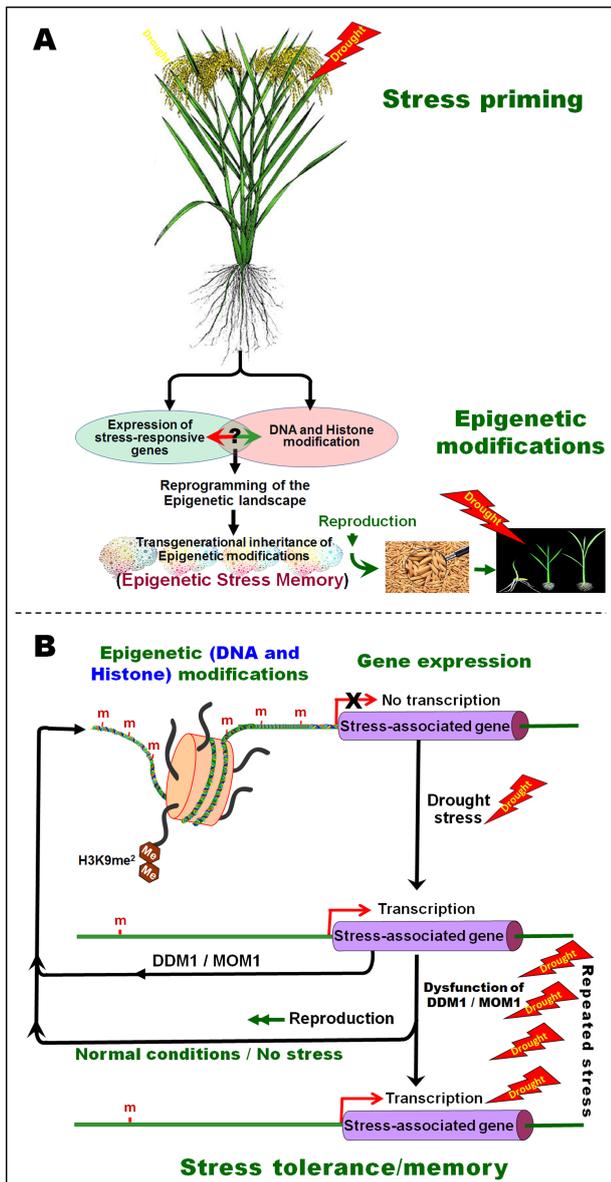
to revert to the original state after withdrawal of the stress, but one-third of the changes might get inherited as epigenetic stress memory [9,140,145]. Some of the salt stress-induced modulations in the methylation of DNA were reported to be transmitted over the generations in *Arabidopsis thaliana*, particularly through female gametes [146]. *SDC* gene, coding for the suppressor of *DRM1*, *DRM2*, and *CMT3*, was reported to get silenced by methylation at the promoter of the gene; however, the reoccurring heat stress was reported to activate *SDC* [147].

Assessment of the contribution of a heritable epimark in the inheritance of a phenotypic alteration has been a challenging task because many of the epigenetic changes and DNA sequence polymorphisms co-segregate with the altered gene expression. However, evidence indicates that some of the heritable epiallelic changes in plants can be associated with a trait of interest which can be utilized in crop improvement programs. Molecular analyses indicate that variations in methylation in an epiRIL are stable across the generations. Since epiRILs are identical for DNA sequences but differ for the epigenetic marks, the phenotypic differences among them can be attributed to epigenetic variations. Thus, epiRILs can be valuable assets for epigenetic studies in plants. Epigenetic changes occurring in response to environmental stress accumulate in the first generation, but only stable/heritable components might get inherited that might be involved in the adaptability of the plant.

Repeated exposure of rice to drought stress for several generations was reported to improve adaptability through epi-mutations and transmission of the altered DNA methylation to the progenies [140] (Fig. 3A). In Arabidopsis, Morpheus Molecule 1 (MOM1) and DDM1 were reported to be involved in the removal of stress-induced epigenetic marks under stress-free environment. In a double mutant for *ddm1-mom1*, the stress-induced epigenetic marks were reported to be inherited by the progeny. However, a single mutant (either *ddm1* or *mom1*) failed to inherit the stress-induced epigenetic mark [148]. These indicate that DDM1 and MOM1 block the inheritance of stress-induced epimark (Fig. 3B). Remembering an episode of stress and responding more efficiently to subsequent occurrences of stress constitute the way plants adapt to environmental changes. However, transgenerational epigenetic stress memory needs to be meiotically stable and heritable to at least two stress-free generations [149].

## 7. Future Perspectives

Although considerable progress has been made in epigenomics of gene regulation in animals, the enzymes involved in DNA methylation and histone modification in plants are still being characterized. Dynamic chromatin architecture affects the accessibility of genes to transcription machinery, thus modulating the interpretation of the genetic information encoded in the DNA sequence. To date, only a little is known about the epigenetic modulators and



**Fig. 3. Possible role of epigenetic changes in stress tolerance/memory in plants.** (A) Abiotic stress alters the expression of stress-responsive genes through different epigenetic modifications. In the stress-primed plants, the epigenetic landscape is altered and a part of the epigenetic modifications gets inherited, which serves as epigenetic stress memory. (B) Stress-induced epigenetic modifications cause differential expression of the stress-responsive genes. Under normal conditions (after removal of stress), Morpheus Molecule 1 (MOM1) and DDM1 play important roles in the removal of the stress-induced epigenetic marks. However, dysfunction of MOM1 and DDM1 results in transmission of the stress-induced epigenetic marks over the generation, particularly on repeated exposure to stress.

their functions during developmental processes and environmental stress in plants. Moreover, little is known about the possible interactions between epigenetic marks. Dif-

ferent types of epigenetic modifications, including DNA methylation, histone modification, histone variants, and nucleosome occupancy, are being reported to be implicated in stress memory [150,151]. Recent technological advancements like super-resolution technology and single-cell *omics* approaches might help in understanding the phenomenon of stress memory [11]. In addition, an assay for transposase-accessible chromatin sequencing (ATAC-seq) [152] would help decipher the interactions between DNA base methylation and histone modifications and their role in stress memory.

Furthermore, epitranscriptomic modifications [RNA base modifications, like 5-methylcytosine ( $m^5C$ ), 5-hydroxymethylcytosine ( $hm^5C$ ),  $N^6$ -methyladenosine ( $m^6A$ ),  $N^1$ -methyladenosine ( $m^1A$ ),  $N^7$ -methylguanosine ( $m^7G$ ), Pseudouridine,  $N^6$ -2'-O-dimethyladenosine ( $m^6Am$ )] [153] playing important roles in plant growth, development, and stress responses, might also be involved in keeping of memory. Once the heritable components of epigenome associated with stress memory are identified/established, they can be utilized in epigenome editing [154,155] to enhance the plasticity of crop plants toward the development of climate-resilient varieties for sustainable agriculture.

## 8. Conclusions

Sequencing, assembly, annotation, and functional validation of the biological function(s) of protein-coding as well as non-coding/regulatory genes have been some of the revolutionary research during the last four decades. On understanding the biological functions of genes/proteins, it is obvious now that the DNA sequence information alone is not sufficient enough to depict/decide the expression of a gene/trait. Efforts are also being made to explore the epigenetic and 3D/4D genomic mechanisms involved in the plasticity of an organism. Interestingly, the nucleotide sequence variation and rate of meiotic recombination are correlated with 3D genomic structures. Topologically associating domains (TADs) were reported to show more single-nucleotide polymorphism, increased rate of recombination, and structural variations (SVs) compared to that observed in the inter-TAD regions [156]. Accumulating evidence suggests the involvement of epigenetic modifications such as DNA base (6-mA) modification [65] and histone modification ( $H3K4me3$ ) mark [157] in stress memory and adaptation of plants. In many cases, stress memory is reported to be reset just after one stress-free generation. Though it is suggested that transgenerational stress memory over several generations might be disadvantageous to the plant [148], it might be beneficial for the adaptability of plants, particularly under fluctuating environmental conditions. 3D genomic study at the single-cell level using live-cell imaging techniques paves the way to discover the gene regulatory mechanisms needed for the development of climate-smart crops. Combined studies on genome, epigenome, epi-

transcriptome, and epiproteome (multi-omics analyses) and their effects/contributions to stress memory would greatly improve our understanding of keeping stress memory in plants.

### Author Contributions

SK and TM conceived the review. SK prepared the initial draft. SK and TM revised the manuscript and approved the final draft. Both authors contributed to the article and approved the final version. Both authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

### Ethics Approval and Consent to Participate

Not applicable.

### Acknowledgment

Not applicable.

### Funding

Epigenomics research on abiotic stress tolerance is being carried out with financial support from the National Agricultural Science Fund (NASF/ABP-70161/2018-19) and Extramural Research Grant [18(3)/2018-O&P, NASF/EMR/(OG)/06/2021] from the Indian Council of Agricultural Research, Government of India, New Delhi, India.

### Conflict of Interest

The authors declare no conflict of interest.

### References

- [1] Kumar S, Beena AS, Awana M, Singh A. Physiological, Biochemical, Epigenetic and Molecular Analyses of Wheat (*Triticum aestivum*) Genotypes with Contrasting Salt Tolerance. *Frontiers in Plant Science*. 2017; 8: 1151.
- [2] Kumar S, Mohapatra T. Dynamics of DNA Methylation and Its Functions in Plant Growth and Development. *Frontiers in Plant Science*. 2021; 12: 596236.
- [3] Kumar S. Epigenomics of plant responses to environmental stress. *Epigenomes*. 2018; 2: 6.
- [4] Kumar S, Singh AK, Mohapatra T. Epigenetics: History, present status and future perspective. *Indian Journal of Genetics and Plant Breeding*. 2017; 77: 445–463.
- [5] Kumar S, Chinnusamy V, Mohapatra T. Epigenetics of Modified DNA Bases: 5-Methylcytosine and Beyond. *Frontiers in Genetics*. 2018; 9: 640.
- [6] Rowley MJ, Corces VG. Organizational principles of 3D genome architecture. *Nature Reviews. Genetics*. 2018; 19: 789–800.
- [7] Doğan ES, Liu C. Three-dimensional chromatin packing and positioning of plant genomes. *Nature Plants*. 2018; 4: 521–529.
- [8] Grob S. Three-dimensional chromosome organization in flowering plants. *Briefings in Functional Genomics*. 2020; 19: 83–91.
- [9] Kumar S. Epigenetic memory of stress responses in plants. *Journal of Phytochemistry and Biochemistry*. 2018; 2: e102.
- [10] Nagano T, Lubling Y, Várnai C, Dudley C, Leung W, Baran Y, *et al.* Cell-cycle dynamics of chromosomal organization at single-cell resolution. *Nature*. 2017; 547: 61–67.
- [11] Kumar S, Kaur S, Seem K, Kumar S, Mohapatra T. Understanding 3D Genome Organization and Its Effect on Transcriptional Gene Regulation Under Environmental Stress in Plant: A Chromatin Perspective. *Frontiers in Cell and Developmental Biology*. 2021; 9: 774719.
- [12] Li Y, Kumar S, Qian W. Active DNA demethylation: mechanism and role in plant development. *Plant Cell Reports*. 2018; 37: 77–85.
- [13] Matzke MA, Moshier RA. RNA-directed DNA methylation: an epigenetic pathway of increasing complexity. *Nature Reviews. Genetics*. 2014; 15: 394–408.
- [14] Law JA, Jacobsen SE. Establishing, maintaining and modifying DNA methylation patterns in plants and animals. *Nature Reviews. Genetics*. 2010; 11: 204–220.
- [15] Sedgwick B, Bates PA, Paik J, Jacobs SC, Lindahl T. Repair of alkylated DNA: recent advances. *DNA Repair*. 2007; 6: 429–442.
- [16] Westbye MP, Feyzi E, Aas PA, Vågbo CB, Talstad VA, Kavli B, *et al.* Human AlkB homolog 1 is a mitochondrial protein that demethylates 3-methylcytosine in DNA and RNA. *The Journal of Biological Chemistry*. 2008; 283: 25046–25056.
- [17] Greer EL, Blanco MA, Gu L, Sendinc E, Liu J, Aristizábal-Corrales D, *et al.* DNA Methylation on N6-Adenine in *C. elegans*. *Cell*. 2015; 161: 868–878.
- [18] Zhang G, Huang H, Liu D, Cheng Y, Liu X, Zhang W, *et al.* N6-methyladenine DNA modification in *Drosophila*. *Cell*. 2015; 161: 893–906.
- [19] Clapier CR, Iwasa J, Cairns BR, Peterson CL. Mechanisms of action and regulation of ATP-dependent chromatin-remodelling complexes. *Nature Reviews. Molecular Cell Biology*. 2017; 18: 407–422.
- [20] Miller JL, Grant PA. The role of DNA methylation and histone modifications in transcriptional regulation in humans. *Subcellular Biochemistry*. 2013; 61: 289–317.
- [21] Padeken J, Methot SP, Gasser SM. Establishment of H3K9-methylated heterochromatin and its functions in tissue differentiation and maintenance. *Nature Reviews. Molecular Cell Biology*. 2022; 23: 623–640.
- [22] Chang W, Zhao Y, Rayê D, Xie Q, Suzuki M, Zheng D, *et al.* Dynamic changes in whole genome DNA methylation, chromatin and gene expression during mouse lens differentiation. *Epigenetics & Chromatin*. 2023; 16: 4.
- [23] Oliver SS, Denu JM. Dynamic interplay between histone H3 modifications and protein interpreters: emerging evidence for a “histone language”. *Chembiochem: a European Journal of Chemical Biology*. 2011; 12: 299–307.
- [24] Du J, Zhong X, Bernatavichute YV, Stroud H, Feng S, Caro E, *et al.* Dual binding of chromomethylase domains to H3K9me2-containing nucleosomes directs DNA methylation in plants. *Cell*. 2012; 151: 167–180.
- [25] Zhang H, Lang Z, Zhu JK. Dynamics and function of DNA methylation in plants. *Nature Reviews. Molecular Cell Biology*. 2018; 19: 489–506.
- [26] Zhou Y, Romero-Campero FJ, Gómez-Zambrano Á, Turck F, Calonje M. H2A monoubiquitination in *Arabidopsis thaliana* is generally independent of LHP1 and PRC2 activity. *Genome Biology*. 2017; 18: 69.
- [27] Martire S, Banaszynski LA. The roles of histone variants in fine-tuning chromatin organization and function. *Nature Reviews. Molecular Cell Biology*. 2020; 21: 522–541.
- [28] Cao Y, Dai Y, Cui S, Ma L. Histone H2B monoubiquitination in the chromatin of FLOWERING LOCUS C regulates flowering time in *Arabidopsis*. *The Plant Cell*. 2008; 20: 2586–2602.
- [29] Huang X, Lu Z, Wang X, Ouyang Y, Chen W, Xie K, *et al.* Imprinted gene OsFIE1 modulates rice seed development by influencing nutrient metabolism and modifying genome H3K27me3.

The Plant Journal: for Cell and Molecular Biology. 2016; 87: 305–317.

- [30] Liu J, He Z. Small DNA Methylation, Big Player in Plant Abiotic Stress Responses and Memory. *Frontiers in Plant Science*. 2020; 11: 595603.
- [31] Bonev B, Cavalli G. Organization and function of the 3D genome. *Nature Reviews. Genetics*. 2016; 17: 661–678.
- [32] Dixon JR, Jung I, Selvaraj S, Shen Y, Antosiewicz-Bourget JE, Lee AY, *et al.* Chromatin architecture reorganization during stem cell differentiation. *Nature*. 2015; 518: 331–336.
- [33] Boltsis I, Grosveld F, Giraud G, Kolovos P. Chromatin Conformation in Development and Disease. *Frontiers in Cell and Developmental Biology*. 2021; 9: 723859.
- [34] Sun L, Jing Y, Liu X, Li Q, Xue Z, Cheng Z, *et al.* Heat stress-induced transposon activation correlates with 3D chromatin organization rearrangement in Arabidopsis. *Nature Communications*. 2020; 11: 1886.
- [35] Lupiáñez DG, Kraft K, Heinrich V, Krawitz P, Brancati F, Kłopotki E, *et al.* Disruptions of topological chromatin domains cause pathogenic rewiring of gene-enhancer interactions. *Cell*. 2015; 161: 1012–1025.
- [36] Fei T, Li W, Peng J, Xiao T, Chen CH, Wu A, *et al.* Deciphering essential cistromes using genome-wide CRISPR screens. *Proceedings of the National Academy of Sciences of the United States of America*. 2019; 116: 25186–25195.
- [37] Li X, Xu B, Li X, Wang D, An Z, Jia Y, *et al.* Widespread transcriptional responses to the thermal stresses are prewired in human 3D genome. *BioRxiv*. 2019. (preprint)
- [38] Perrella G, Zioutopoulou A, Headland LR, Kaiserli E. The impact of light and temperature on chromatin organization and plant adaptation. *Journal of Experimental Botany*. 2020; 71: 5247–5255.
- [39] Eberharter A, Becker PB. Histone acetylation: a switch between repressive and permissive chromatin. Second in review series on chromatin dynamics. *EMBO Reports*. 2002; 3: 224–229.
- [40] Sokolova V, Sarkar S, Tan D. Histone variants and chromatin structure, update of advances. *Computational and Structural Biotechnology Journal*. 2022; 21: 299–311.
- [41] Rowley MJ, Rothi MH, Böhmendorfer G, Kuciński J, Wierzbicki AT. Long-range control of gene expression via RNA-directed DNA methylation. *PLoS Genetics*. 2017; 13: e1006749.
- [42] Park J, Lim CJ, Shen M, Park HJ, Cha JY, Iniesto E, *et al.* Epigenetic switch from repressive to permissive chromatin in response to cold stress. *Proceedings of the National Academy of Sciences of the United States of America*. 2018; 115: E5400–E5409.
- [43] Peterson CL, Workman JL. Promoter targeting and chromatin remodeling by the SWI/SNF complex. *Current Opinion in Genetics & Development*. 2000; 10: 187–192.
- [44] Bhadouriya SL, Mehrotra S, Basantani MK, Loake GJ, Mehrotra R. Role of Chromatin Architecture in Plant Stress Responses: An Update. *Frontiers in Plant Science*. 2021; 11: 603380.
- [45] Tannenbaum M, Sarusi-Portuguez A, Krispil R, Schwartz M, Loza O, Benichou JIC, *et al.* Regulatory chromatin landscape in *Arabidopsis thaliana* roots uncovered by coupling INTACT and ATAC-seq. *Plant Methods*. 2018; 14: 113.
- [46] Barragán-Rosillo AC, Peralta-Alvarez CA, Ojeda-Rivera JO, Arzate-Mejia RG, Recillas-Targa F, Herrera-Estrella L. Genome accessibility dynamics in response to phosphate limitation is controlled by the PHR1 family of transcription factors in *Arabidopsis*. *Proceedings of the National Academy of Sciences of the United States of America*. 2021; 118: e2107558118.
- [47] Gallusci P, Dai Z, Génard M, Gauffretau A, Leblanc-Fournier N, Richard-Molard C, *et al.* Epigenetics for Plant Improvement: Current Knowledge and Modeling Avenues. *Trends in Plant Science*. 2017; 22: 610–623.
- [48] Bartels A, Han Q, Nair P, Stacey L, Gaynier H, Mosley M, *et al.* Dynamic DNA Methylation in Plant Growth and Development. *International Journal of Molecular Sciences*. 2018; 19: 2144.
- [49] Bird A. DNA methylation patterns and epigenetic memory. *Genes & Development*. 2002; 16: 6–21.
- [50] de Mendoza A, Nguyen TV, Ford E, Poppe D, Buckberry S, Pflueger J, *et al.* Large-scale manipulation of promoter DNA methylation reveals context-specific transcriptional responses and stability. *Genome Biology*. 2022; 23: 163.
- [51] Zhang X, Yazaki J, Sundaresan A, Cokus S, Chan SWL, Chen H, *et al.* Genome-wide high-resolution mapping and functional analysis of DNA methylation in arabidopsis. *Cell*. 2006; 126: 1189–1201.
- [52] Muyle AM, Seymour DK, Lv Y, Huettel B, Gaut BS. Gene Body Methylation in Plants: Mechanisms, Functions, and Important Implications for Understanding Evolutionary Processes. *Genome Biology and Evolution*. 2022; 14: evac038.
- [53] Shahzad Z, Moore JD, Zilberman D. Gene body methylation mediates epigenetic inheritance of plant traits. *BioRxiv*. 2021. (preprint)
- [54] Cokus SJ, Feng S, Zhang X, Chen Z, Merriman B, Haudenschild CD, *et al.* Shotgun bisulphite sequencing of the Arabidopsis genome reveals DNA methylation patterning. *Nature*. 2008; 452: 215–219.
- [55] Takuno S, Gaut BS. Gene body methylation is conserved between plant orthologs and is of evolutionary consequence. *Proceedings of the National Academy of Sciences of the United States of America*. 2013; 110: 1797–1802.
- [56] Bewick AJ, Schmitz RJ. Gene body DNA methylation in plants. *Current Opinion in Plant Biology*. 2017; 36: 103–110.
- [57] Ong-Abdullah M, Ordway JM, Jiang N, Ooi SE, Kok SY, Sarpan N, *et al.* Loss of Karma transposon methylation underlies the mantled somaclonal variant of oil palm. *Nature*. 2015; 525: 533–537.
- [58] Ratel D, Ravanat JL, Berger F, Wion D. N<sup>6</sup>-methyladenine: the other methylated base of DNA. *BioEssays: News and Reviews in Molecular, Cellular and Developmental Biology*. 2006; 28: 309–315.
- [59] Huang W, Xiong J, Yang Y, Liu SM, Yuan BF, Feng YQ. Determination of DNA adenine methylation in genomes of mammals and plants by liquid chromatography/mass spectrometry. *RSC Advances*. 2015; 5: 64046–64054.
- [60] Liang D, Wang H, Song W, Xiong X, Zhang X, Hu Z, *et al.* The decreased N<sup>6</sup>-methyladenine DNA modification in cancer cells. *Biochemical and Biophysical Research Communications*. 2016; 480: 120–125.
- [61] Vanyushin BF. Adenine methylation in eukaryotic DNA. *Molecular Biology*. 2005; 39: 473–481.
- [62] Li Z, Zhao P, Xia Q. Epigenetic Methylations on N<sup>6</sup>-Adenine and N<sup>6</sup>-Adenosine with the same Input but Different Output. *International Journal of Molecular Sciences*. 2019; 20: 2931.
- [63] Zhou C, Wang C, Liu H, Zhou Q, Liu Q, Guo Y, *et al.* Identification and analysis of adenine N<sup>6</sup>-methylation sites in the rice genome. *Nature Plants*. 2018; 4: 554–563.
- [64] Zhang Q, Liang Z, Cui X, Ji C, Li Y, Zhang P, *et al.* N<sup>6</sup>-Methyladenine DNA Methylation in Japonica and Indica Rice Genomes and Its Association with Gene Expression, Plant Development, and Stress Responses. *Molecular Plant*. 2018; 11: 1492–1508.
- [65] Liang Z, Riaz A, Chachar S, Ding Y, Du H, Gu X. Epigenetic Modifications of mRNA and DNA in Plants. *Molecular Plant*. 2020; 13: 14–30.
- [66] Kumar S. Epigenetic control of apomixis: A new perspective of an old enigma. *Advances in Plants and Agriculture Research*. 2017; 7: 00243.
- [67] Ortega-Galisteo AP, Morales-Ruiz T, Ariza RR, Roldán-Arjona T. Arabidopsis DEMETER-LIKE proteins DML2 and DML3

- are required for appropriate distribution of DNA methylation marks. *Plant Molecular Biology*. 2008; 67: 671–681.
- [68] Morales-Ruiz T, Ortega-Galisteo AP, Ponferrada-Marín MI, Martínez-Macias MI, Ariza RR, Roldán-Arjona T. DEMETER and REPRESSOR OF SILENCING 1 encode 5-methylcytosine DNA glycosylases. *Proceedings of the National Academy of Sciences of the United States of America*. 2006; 103: 6853–6858.
- [69] Penterman J, Zilberman D, Huh JH, Ballinger T, Henikoff S, Fischer RL. DNA demethylation in the Arabidopsis genome. *Proceedings of the National Academy of Sciences of the United States of America*. 2007; 104: 6752–6757.
- [70] Zhu JH, Kapoor A, Sridhar VV, Agius F, Zhu JK. The DNA glycosylase/lyase ROS1 functions in pruning DNA methylation patterns in Arabidopsis. *Current Biology*. 2007; 17: 54–59.
- [71] Gehring M, Bubb KL, Henikoff S. Extensive demethylation of repetitive elements during seed development underlies gene imprinting. *Science (New York, N.Y.)*. 2009; 324: 1447–1451.
- [72] Hsieh TF, Ibarra CA, Silva P, Zemach A, Eshed-Williams L, Fischer RL, *et al.* Genome-wide demethylation of Arabidopsis endosperm. *Science (New York, N.Y.)*. 2009; 324: 1451–1454.
- [73] Ibarra CA, Feng X, Schoft VK, Hsieh TF, Uzawa R, Rodrigues JA, *et al.* Active DNA demethylation in plant companion cells reinforces transposon methylation in gametes. *Science (New York, N.Y.)*. 2012; 337: 1360–1364.
- [74] Tang K, Lang Z, Zhang H, Zhu JK. The DNA demethylase ROS1 targets genomic regions with distinct chromatin modifications. *Nature Plants*. 2016; 2: 16169.
- [75] He XJ, Hsu YF, Zhu S, Wierzbicki AT, Pontes O, Pikaard CS, *et al.* An effector of RNA-directed DNA methylation in Arabidopsis is an ARGONAUTE 4- and RNA-binding protein. *Cell*. 2009; 137: 498–508.
- [76] Gao Z, Liu HL, Daxinger L, Pontes O, He X, Qian W, *et al.* An RNA polymerase II- and AGO4-associated protein acts in RNA-directed DNA methylation. *Nature*. 2010; 465: 106–109.
- [77] Qian W, Miki D, Zhang H, Liu Y, Zhang X, Tang K, *et al.* A histone acetyltransferase regulates active DNA demethylation in Arabidopsis. *Science (New York, N.Y.)*. 2012; 336: 1445–1448.
- [78] Lei M, Zhang H, Julian R, Tang K, Xie S, Zhu JK. Regulatory link between DNA methylation and active demethylation in Arabidopsis. *Proceedings of the National Academy of Sciences of the United States of America*. 2015; 112: 3553–3557.
- [79] Williams BP, Pignatta D, Henikoff S, Gehring M. Methylation-sensitive expression of a DNA demethylase gene serves as an epigenetic rheostat. *PLoS Genetics*. 2015; 11: e1005142.
- [80] Jones MJ, Goodman SJ, Kobor MS. DNA methylation and healthy human aging. *Aging Cell*. 2015; 14: 924–932.
- [81] Baylin SB, Jones PA. Epigenetic Determinants of Cancer. *Cold Spring Harbor Perspectives in Biology*. 2016; 8: a019505.
- [82] Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. *Cell Research*. 2011; 21: 381–395.
- [83] Ramakrishnan M, Zhang Z, Mullasserri S, Kalendar R, Ahmad Z, Sharma A, *et al.* Epigenetic stress memory: A new approach to study cold and heat stress responses in plants. *Frontiers in Plant Science*. 2022; 13: 1075279.
- [84] Shahbazian MD, Grunstein M. Functions of site-specific histone acetylation and deacetylation. *Annual Review of Biochemistry*. 2007; 76: 75–100.
- [85] Kouzarides T. Chromatin modifications and their function. *Cell*. 2007; 128: 693–705.
- [86] Black JC, Van Rechem C, Whetstone JR. Histone lysine methylation dynamics: establishment, regulation, and biological impact. *Molecular Cell*. 2012; 48: 491–507.
- [87] Turgut-Kara N, Arikan B, Celik H. Epigenetic memory and priming in plants. *Genetica*. 2020; 148: 47–54.
- [88] Allis CD, Jenuwein T. The molecular hallmarks of epigenetic control. *Nature Reviews. Genetics*. 2016; 17: 487–500.
- [89] Shiio Y, Eisenman RN. Histone sumoylation is associated with transcriptional repression. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100: 13225–13230.
- [90] Nathan D, Ingvarsdottir K, Sterner DE, Bylebyl GR, Dokmanovic M, Dorsey JA, *et al.* Histone sumoylation is a negative regulator in *Saccharomyces cerevisiae* and shows dynamic interplay with positive-acting histone modifications. *Genes & Development*. 2006; 20: 966–976.
- [91] Bartke T, Vermeulen M, Xhemalce B, Robson SC, Mann M, Kouzarides T. Nucleosome-interacting proteins regulated by DNA and histone methylation. *Cell*. 2010; 143: 470–484.
- [92] Lusser A, Kadonaga JT. Chromatin remodeling by ATP-dependent molecular machines. *BioEssays: News and Reviews in Molecular, Cellular and Developmental Biology*. 2003; 25: 1192–1200.
- [93] Hargreaves DC, Crabtree GR. ATP-dependent chromatin remodeling: genetics, genomics and mechanisms. *Cell Research*. 2011; 21: 396–420.
- [94] Armeev GA, Gribkova AK, Pospelova I, Komarova GA, Shaytan AK. Linking chromatin composition and structural dynamics at the nucleosome level. *Current Opinion in Structural Biology*. 2019; 56: 46–55.
- [95] Wu X, Zhang X, Huang B, Han J, Fang H. Advances in biological functions and mechanisms of histone variants in plants. *Frontiers in Genetics*. 2023; 14: 1229782.
- [96] Jiang D, Berger F. Variation is important: Warranting chromatin function and dynamics by histone variants. *Current Opinion in Plant Biology*. 2023; 75: 102408.
- [97] Aboobucker SI, Zhou L, Lübberstedt T. Haploid male fertility is restored by parallel spindle genes in *Arabidopsis thaliana*. *Nature Plants*. 2023; 9: 214–218.
- [98] Borg M, Jacob Y, Susaki D, LeBlanc C, Buendía D, Axelsson E, *et al.* Targeted reprogramming of H3K27me3 resets epigenetic memory in plant paternal chromatin. *Nature Cell Biology*. 2020; 22: 621–629.
- [99] Yan A, Borg M, Berger F, Chen Z. The atypical histone variant H3.15 promotes callus formation in *Arabidopsis thaliana*. *Development (Cambridge, England)*. 2020; 147: dev184895.
- [100] Zahraeifard S, Foroozani M, Sepehri A, Oh DH, Wang G, Mangu V, *et al.* Rice H2A.Z negatively regulates genes responsive to nutrient starvation but promotes expression of key housekeeping genes. *Journal of Experimental Botany*. 2018; 69: 4907–4919.
- [101] Sura W, Kabza M, Karlowski WM, Bieluszewski T, Kus-Słowińska M, Pawełszek Ł, *et al.* Dual Role of the Histone Variant H2A.Z in Transcriptional Regulation of Stress-Response Genes. *The Plant Cell*. 2017; 29: 791–807.
- [102] Abelenda JA, Trabanco N, Del Olmo I, Pozas J, Martín-Trillo MDM, Gómez-Garrido J, *et al.* High ambient temperature impacts on flowering time in *Brassica napus* through both H2A.Z-dependent and independent mechanisms. *Plant, Cell & Environment*. 2023; 46: 1427–1441.
- [103] Zhao T, Lu J, Zhang H, Xue M, Pan J, Ma L, *et al.* Histone H3.3 deposition in seed is essential for the post-embryonic developmental competence in Arabidopsis. *Nature Communications*. 2022; 13: 7728.
- [104] Fabrizio P, Garvis S, Palladino F. Histone Methylation and Memory of Environmental Stress. *Cells*. 2019; 8: 339.
- [105] Laura B, Silvia P, Francesca F, Benedetta S, Carla C. Epigenetic control of defense genes following MeJA-induced priming in rice (*O. sativa*). *Journal of Plant Physiology*. 2018; 228: 166–177.
- [106] Kim JM, To TK, Matsui A, Tanoi K, Kobayashi NI, Matsuda F, *et al.* Acetate-mediated novel survival strategy against drought

- in plants. *Nature Plants*. 2017; 3: 17097.
- [107] Brzezinka K, Altmann S, Czesnick H, Nicolas P, Gorka M, Benke E, *et al.* *Arabidopsis* FORGETTER1 mediates stress-induced chromatin memory through nucleosome remodeling. *eLife*. 2016; 5: e17061.
- [108] Kidokoro S, Shinozaki K, Yamaguchi-Shinozaki K. Transcriptional regulatory network of plant cold-stress responses. *Trends in Plant Science*. 2022; 27: 922–935.
- [109] Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. *Nature Reviews. Genetics*. 2016; 17: 47–62.
- [110] Chi JT, Chang HY, Wang NN, Chang DS, Dunphy N, Brown PO. Genomewide view of gene silencing by small interfering RNAs. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100: 6343–6346.
- [111] Hua X, Berkowitz ND, Willmann MR, Yu X, Lyons E, Gregory BD. Global Analysis of RNA-Dependent RNA Polymerase-Dependent Small RNAs Reveals New Substrates and Functions for These Proteins and SGS3 in *Arabidopsis*. *Non-coding RNA*. 2021; 7: 28.
- [112] Roessler K, Bousios A, Meca E, Gaut BS. Modeling Interactions between Transposable Elements and the Plant Epigenetic Response: A Surprising Reliance on Element Retention. *Genome Biology and Evolution*. 2018; 10: 803–815.
- [113] Axtell MJ, Jan C, Rajagopalan R, Bartel DP. A two-hit trigger for siRNA biogenesis in plants. *Cell*. 2006; 127: 565–577.
- [114] Axtell MJ. Classification and comparison of small RNAs from plants. *Annual Review of Plant Biology*. 2013; 64: 137–159.
- [115] Charles Richard JL, Eichhorn PJA. Platforms for Investigating LncRNA Functions. *SLAS Technology*. 2018; 23: 493–506.
- [116] Schulz D, Schwalb B, Kiesel A, Baejen C, Torkler P, Gagneur J, *et al.* Transcriptome surveillance by selective termination of noncoding RNA synthesis. *Cell*. 2013; 155: 1075–1087.
- [117] St Laurent G, Wahlstedt C, Kapranov P. The Landscape of long noncoding RNA classification. *Trends in Genetics: TIG*. 2015; 31: 239–251.
- [118] Fernandes JCR, Acuña SM, Aoki JI, Floeter-Winter LM, Muxel SM. Long Non-Coding RNAs in the Regulation of Gene Expression: Physiology and Disease. *Non-coding RNA*. 2019; 5: 17.
- [119] Wang J, Meng X, Dobrovolskaya OB, Orlov YL, Chen M. Non-coding RNAs and Their Roles in Stress Response in Plants. *Genomics, Proteomics & Bioinformatics*. 2017; 15: 301–312.
- [120] Kazmierczyk M, Kasprovicz MK, Kasprzyk ME, Wrzesinski J. Human Long Noncoding RNA Interactome: Detection, Characterization and Function. *International Journal of Molecular Sciences*. 2020; 21: 1027.
- [121] Rinn JL, Chang HY. Long Noncoding RNAs: Molecular Modalities to Organismal Functions. *Annual Review of Biochemistry*. 2020; 89: 283–308.
- [122] Statello L, Guo CJ, Chen LL, Huarte M. Gene regulation by long non-coding RNAs and its biological functions. *Nature Reviews. Molecular Cell Biology*. 2021; 22: 96–118.
- [123] Liao P, Li S, Cui X, Zheng Y. A comprehensive review of web-based resources of non-coding RNAs for plant science research. *International Journal of Biological Sciences*. 2018; 14: 819–832.
- [124] Ding M, Liu Y, Liao X, Zhan H, Liu Y, Huang W. Enhancer RNAs (eRNAs): New Insights into Gene Transcription and Disease Treatment. *Journal of Cancer*. 2018; 9: 2334–2340.
- [125] Gil N, Ulitsky I. Production of Spliced Long Noncoding RNAs Specifies Regions with Increased Enhancer Activity. *Cell Systems*. 2018; 7: 537–547.e3.
- [126] Wang M, Yu F, Wu W, Zhang Y, Chang W, Ponnusamy M, *et al.* Circular RNAs: A novel type of non-coding RNA and their potential implications in antiviral immunity. *International Journal of Biological Sciences*. 2017; 13: 1497–1506.
- [127] Quan G, Li J. Circular RNAs: biogenesis, expression and their potential roles in reproduction. *Journal of Ovarian Research*. 2018; 11: 9.
- [128] Weirick T, Militello G, Uchida S. Long Non-coding RNAs in Endothelial Biology. *Frontiers in Physiology*. 2018; 9: 522.
- [129] Chu Q, Shen E, Ye CY, Fan L, Zhu QH. Emerging roles of plant circular RNAs. *Journal of Plant Cell Development*. 2018; 1: 1–14.
- [130] Tang B, Hao Z, Zhu Y, Zhang H, Li G. Genome-wide identification and functional analysis of circRNAs in *Zea mays*. *PLoS One*. 2018; 13: e0202375.
- [131] Ragan C, Goodall GJ, Shirokikh NE, Preiss T. Insights into the biogenesis and potential functions of exonic circular RNA. *Scientific Reports*. 2019; 9: 2048.
- [132] Hung T, Wang Y, Lin MF, Koegel AK, Kotake Y, Grant GD, *et al.* Extensive and coordinated transcription of noncoding RNAs within cell-cycle promoters. *Nature Genetics*. 2011; 43: 621–629.
- [133] Yap KL, Li S, Muñoz-Cabello AM, Raguz S, Zeng L, Mujtaba S, *et al.* Molecular interplay of the noncoding RNA ANRIL and methylated histone H3 lysine 27 by polycomb CBX7 in transcriptional silencing of *INK4a*. *Molecular Cell*. 2010; 38: 662–674.
- [134] Keniry A, Oxley D, Monnier P, Kyba M, Dandolo L, Smits G, *et al.* The H19 lincRNA is a developmental reservoir of miR-675 that suppresses growth and *Igf1r*. *Nature Cell Biology*. 2012; 14: 659–665.
- [135] Lange UC, Schneider R. What an epigenome remembers. *BioEssays: News and Reviews in Molecular, Cellular and Developmental Biology*. 2010; 32: 659–668.
- [136] Ding Y, Fromm M, Avramova Z. Multiple exposures to drought ‘train’ transcriptional responses in *Arabidopsis*. *Nature Communications*. 2012; 3: 740.
- [137] Kinoshita T, Seki M. Epigenetic memory for stress response and adaptation in plants. *Plant & Cell Physiology*. 2014; 55: 1859–1863.
- [138] Hilker M, Schmölling T. Stress priming, memory, and signalling in plants. *Plant, Cell & Environment*. 2019; 42: 753–761.
- [139] Thiebaut F, Hemerly AS, Ferreira PCG. A Role for Epigenetic Regulation in the Adaptation and Stress Responses of Non-model Plants. *Frontiers in Plant Science*. 2019; 10: 246.
- [140] Zheng X, Chen L, Xia H, Wei H, Lou Q, Li M, *et al.* Trans-generational epimutations induced by multi-generation drought imposition mediate rice plant’s adaptation to drought condition. *Scientific Reports*. 2017; 7: 39843.
- [141] Schulze WX, Altenbuchinger M, He M, Kränzlein M, Zörb C. Proteome profiling of repeated drought stress reveals genotype-specific responses and memory effects in maize. *Plant Physiology and Biochemistry: PPB*. 2021; 159: 67–79.
- [142] Kumar S, Seem K, Mohapatra T. Biochemical and Epigenetic Modulations under Drought: Remembering the Stress Tolerance Mechanism in Rice. *Life (Basel, Switzerland)*. 2023; 13: 1156.
- [143] Kim JM, To TK, Ishida J, Matsui A, Kimura H, Seki M. Transition of chromatin status during the process of recovery from drought stress in *Arabidopsis thaliana*. *Plant & Cell Physiology*. 2012; 53: 847–856.
- [144] Friedrich T, Faivre L, Bäurle I, Schubert D. Chromatin-based mechanisms of temperature memory in plants. *Plant, Cell & Environment*. 2019; 42: 762–770.
- [145] Crisp PA, Ganguly D, Eichten SR, Borevitz JO, Pogson BJ. Reconsidering plant memory: Intersections between stress recovery, RNA turnover, and epigenetics. *Science Advances*. 2016; 2: e1501340.
- [146] Wibowo A, Becker C, Marconi G, Durr J, Price J, Hagemann J, *et al.* Hyperosmotic stress memory in *Arabidopsis* is mediated

- by distinct epigenetically labile sites in the genome and is restricted in the male germline by DNA glycosylase activity. *eLife*. 2016; 5: e13546.
- [147] Sanchez DH, Paszkowski J. Heat-induced release of epigenetic silencing reveals the concealed role of an imprinted plant gene. *PLoS Genetics*. 2014; 10: e1004806.
- [148] Iwasaki M, Paszkowski J. Identification of genes preventing transgenerational transmission of stress-induced epigenetic states. *Proceedings of the National Academy of Sciences of the United States of America*. 2014; 111: 8547–8552.
- [149] Lämke J, Bäurle I. Epigenetic and chromatin-based mechanisms in environmental stress adaptation and stress memory in plants. *Genome Biology*. 2017; 18: 124.
- [150] Borg M, Jiang D, Berger F. Histone variants take center stage in shaping the epigenome. *Current Opinion in Plant Biology*. 2021; 61: 101991.
- [151] Oberkofler V, Pratz L, Bäurle I. Epigenetic regulation of abiotic stress memory: maintaining the good things while they last. *Current Opinion in Plant Biology*. 2021; 61: 102007.
- [152] Grandi FC, Modi H, Kampman L, Corces MR. Chromatin accessibility profiling by ATAC-seq. *Nature Protocols*. 2022; 17: 1518–1552.
- [153] Kumar S, Mohapatra T. Deciphering epitranscriptome: Modification of mRNA bases provides a new perspective for post-transcriptional regulation of gene expression. *Frontiers in Cell and Developmental Biology*. 2021; 9: 628415.
- [154] Kumar S. Genome editing to epigenome editing: towards unravelling the enigmas in developmental biology. *Trends in Developmental Biology*. 2019; 12: 1–8.
- [155] Seem K, Kaur S, Kumar S, Mohapatra T. Epigenome editing for targeted DNA (de)methylation: a new perspective in modulating gene expression. *Critical Reviews in Biochemistry and Molecular Biology*. 2024; 1–30. (online ahead of print)
- [156] Liao Y, Zhang X, Chakraborty M, Emerson JJ. Topologically associating domains and their role in the evolution of genome structure and function in *Drosophila*. *Genome Research*. 2021; 31: 397–410.
- [157] Nguyen NH, Vu NT, Cheong JJ. Transcriptional Stress Memory and Transgenerational Inheritance of Drought Tolerance in Plants. *International Journal of Molecular Sciences*. 2022; 23: 12918.