# Estradiol inhibits LDL oxidation: Do the progestins medroxyprogesterone acetate and norethisterone acetate influence this effect?

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## Summary

Estrogen replacement therapy in postmenopausal women must be combined with progestin to avoid endometrial cancer. However, progestin addition could antagonize cardioprotective effects of estradiol. Therefore we investigated the effect of the two most commonly used progestins-medroxyprogesterone acetate (progesterone-derivative) and norethisterone acetate (nortestosterone-derivate)-alone and in combination with 17 $\beta$ -estradiol on copper-mediated oxidation of low density lipoprotein (LDL). Whereas 17 $\beta$ -estradiol alone inhibited the onset of LDL oxidation at the concentrations 0.5, 1.0, 5 and 10  $\mu$ M, the progestins alone did not demonstrate any significant effect. In the estrogen-progestin combinations of 0.5 $\mu$ M 17 $\beta$ -estradiol with 0.5, 1.0, 5 and 10  $\mu$ M progestin, respectively, the estradiol effect was not changed. These results suggest that medroxyprogesterone acetate as well as norethisterone acetate do not counteract the beneficial effect of 17 $\beta$ -estradiol on LDL oxidation when used in hormone replacement therapy.

Key words: Estradiol; Medroxyprogesterone acetate; Norethisterone acetate; LDL oxidation.

#### Introduction

A disturbed lipid metabolism, especially the oxidation of LDL, is generally considered to be an important factor in the etiology of cardiovascular diseases. Estrogens not only influence the concentrations of lipid subfractions i.e. reduce LDL-cholesterol and increase HDL-cholesterol [1], but also inhibit the oxidation of LDL [2, 3]. This effect may contribute to the well-established cardioprotective action of estradiol replacement therapy in postmenopausal women [4].

Treating women with unopposed estrogen enhances the risk of endometrial carcinomas. In combination with progestins, however, this can be avoided [5, 6, 7]. The progestins differ in their pharmacodynamic properties. The main criterion for the choice of the progestins is their efficacy with regards to endometrial action and tolerability. Two types of progestins are in common use: "C21 progestins", which are derived from natural progesterone and are known to be particularly tolerable but less effective [8, 9], and "C19 progestins", derived from nortestosterone, which are especially efficacious on the endometrium but less tolerable [10, 11]. For hormonal replacement the most frequently used progestins are medroxyprogesterone acetate (MPA), a C21 progestin, and norethisterone acetate (NETA), a C19 progestin.

The present study was designed to investigate the effect on LDL oxidation by the progestins MPA and NETA. Of particular interest was whether the beneficial estradiolinduced inhibition of LDL oxidation would be impaired by MPA and NETA. This could have consequences for hormonal replacement therapy in postmenopausal women notably in the case of patients with cardiovascular disease.

#### **Materials and Methods**

17β-estradiol (E2) and medroxyprogesterone acetate (MPA) were purchased from Sigma (Deisenhofen, Germany) and norethisterone acetate (NETA) from Ciba-Geigy (Basel, Switzerland).

LDL oxidation was investigated following the method described by Esterbauer et al. [12] using pooled blood serum from healthy premenopausal women containing the antioxidants EDTA (1 mg/ml) and butylated hydroxytoluene (BHT) (4.4 µg/ml). LDL was obtained by ultracentrifugation within 4 hours after the collection of blood samples. In brief: 1.638 g sodium bromide was added to 3 ml serum which was overlayed with saline. After ultracentrifugation at 105,000 g for 9 h (fixed angle rotor), the LDLlayer (density 1.02-1.05 g/ml) was aspirated by a syringe. Oxidation of LDL was started after ultracentrifugation as follows: LDL-solution was separated from BHT and EDTA by gelfiltration (Sephadex G-25, column 10x1.5 cm, eluens: saline). Samples of 150 µl of LDL (adjusted to 300 µg protein/ml) were each mixed with 850 μl saline containing 10 μM CuCl<sub>2</sub> and the test substances. The tested steroids, dissolved in ethanol, were added to this mixture to give final concentrations of 0.5, 1, 5 and 10 µM. E2, MPA and NETA alone were tested in each of these concentrations. In the estrogen/progestin-combinations estradiol was in each instance 0.5 µM, whereas MPA and NETA were added in the concentrations 0.5, 1, 5, 10 µM, respectively. Control values were obtained by the addition of alcohol alone in the same concentrations as in the test substances (final ethanol concentration in all samples = 1%).

The rise of conjugated diene formation, characteristic for the oxidation of LDL, was monitored spectrometrically at 234 nm. Tangents were drawn to the segments of the absorption curve corresponding to the lag phase and propagation phase of LDL oxi-

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dation. The length of the lag phase = lag time was determined as the intercept of the two tangents.

The antioxidative effect of the substances tested is expressed as the elongation of the lag time after the ox-LDL formation of the control. The increase in lag time was measured in minutes up to a maximum of 300 min. Each test substance, alone or in combination, was tested in duplicate in nine different LDL-pools.

Protein content of LDL was determined by a colorimetric protein assay (Bio-Rad, Müchen, Germany). Statistical analysis was performed using the Student's t-test.

#### Results

The control values for ox-LDL formation showed an average lag time of 98.5±11.2 min.

Table 1 shows the effect of 17ß-estradiol (E2), medroxyprogesterone acetate (MPA) and norethisterone acetate (NETA) on the onset of LDL oxidation. E2 elongated the onset at the concentrations of 0.5 and 1 µM for 24.5±3.6 min and 73.6±6.3 min, respectively and exceeded the observation time of 300 min at 5 and 10 µM. Neither MPA nor NETA exhibited any significant effect on the onset of LDL oxidation in the concentration range tested.

The progestins combined with 0.5 µM E2, i.e. the lowest effective concentration of E2, did not antagonize the estradiol-induced elongation of the lag time over the entire concentration range tested (Table 2).

Table 1. — Increase of the lag time of LDL transformed into oxidated LDL after addition of 17β-estradiol, medroxyprogesterone acetate and norethisterone acetate, respectively. The values in minutes are expressed as means  $\pm$  SD, n=9.

	0.5 μΜ	1 μΜ	5μΜ	10μΜ
Estradiol	24.5 (3.6)*	73.6 (6.3)**	>300**	>300**
Medroxy- progesterone acetate	<5	<5	<5	<5
Norethisterone acetate	<5	<5	<5	<5

<sup>\*</sup> p<0.05, \*\* p<0.01 compared to control value.

Table 2. — *Increase of the lag time of LDL transformed into oxi*dated LDL after addition of the combinations of 0.5 µM 17\betaestradiol (E2) with 0.5, 1, 5 and 10 µM medroxyprogesterone acetate (MPA) and norethisterone acetate (NETA), respectively. The values in minutes are expressed as means  $\pm$  SD, n=9.

E2	0.5 μM E2 + 0.5 μM MPA	0.5 μM E2 + 1 μM MPA	0.5 μM E2 5 μM MPA	0.5 μM E2 + 10 μM MPA
24.5 (3.6)*	24.1 (3.7)*	23.8 (3.5)*	25.9 (3.8)*	26.1 (3.2)*
E2	0.5 μM E2 + 0.5 μM NETA	0.5 μM E2 + 1 μM NETA	0.5 μM E2 5 μM NETA	0.5 μM E2 + 10 μM NETA
24.5 (3.6)*	23.6 (3.4)*	25.9 (2.9)*	26.8 (3.3)*	24.3 (3.2)*

<sup>\*</sup> p<0.05, \*\* p<0.01 compared to control value.

#### Discussion

The inhibition of oxidation of LDL is known to be an important step in preventing cardiovascular disease. Oxidated LDL is internalized by macrophages more efficiently than native LDL [13]. Lipid-laden macrophages accumulate in the intima of vascular vessels and form foam cells which are believed to be precursors of atherosclerotic lesions [13]. Furthermore, oxidated LDL is toxic to endothelial and smooth muscle cells of vascular vessels [14] and can impair the cardiovascular system by reducing nitric oxide synthesis [15], enhancing endothelin production [16] and increasing intracellular calcium in vascular cells [17].

The present study demonstrates that 17ß-estradiol is able to delay the onset of LDL oxidation in vitro thus confirming the results of our previous study [18] and also of other groups [2, 3].

Whereas estradiol replacement therapy has been shown to have a beneficial effect on cardiovascular disease the effect of progestins is so far not clear. Progestins can counteract estrogens in many instances [19]. For example NETA decreases HDL-cholesterol [10, 20] and thereby antagonizes the HDL-increasing effect of estradiol. Nevertheless NETA is one of the most commonly used progestins, as it potentially transforms a proliferative endometrium into a secretory one and thus protects from the development of endometrial hyperplasia and cancer [6, 11, 21]. MPA does not counterbalance the HDLincreasing effect of estrogen [9] but has been shown to be less efficient concerning the secretory transformation of the endometrium [8].

### Conclusion

Although the two progestins MPA and NETA show differences in their pharmacodynamic actions, e.g. concerning the influence on HDL-cholesterol and on the endometrium, the present results indicate that estradiol-induced inhibition of LDL oxidation is not influenced by either of the progestins. Even in high dosages they do not eliminate the beneficial estradiol effect. This is of clinical relevance for hormonal replacement therapy in postmenopausal women especially for prophylaxis and therapy of cardiovascular diseases.

# References

- Krauss R.M.: "Lipids and lipoproteins and effects of hormone replacement therapy". In: Lobo R.A. (ed.): "Treatment of postmenopausal woman: basic and clinical aspects". First edition. New York, Raven Press Ltd., 1994, 235.
- Marziere C., Auclair M., Ronveaux M.-F., Salmon S., Santus R., Maziere J.-C.: "Estrogens inhibit copper and cell-mediated modification of low density lipoprotein". Atherosclerosis, 1991, 89, 175.
- Rifici V.A., Chong L., Khachadurian A.K.: "Inhibition of low density lipoprotein oxidation by 17 beta estradiol". Clin. Res., 1992, 40, 208A.

- [4] Bush T.L., Barrett-Connor E., Cowan L.D.: "Cardiovascular mortality and noncontraceptive use of estrogen in women: results from the Lipid Research Clinics Program Follow-up Study". Circulation, 1987, 75, 1102.
- [5] Gambrell R.D.: "Prevention of endometrial cancer with progestogens". *Maturitas*, 1986, 8, 159.
- [6] Whitehead M.I., Fraser D.: "The effect of estrogens and progestogens on the endometrium". Obstet. Gynecol. Clin. North. Am., 1987, 14, 299.
- [7] Writing group for the PEPI trial: "Effects of hormone replacement therapy on endometrial histology in postmenopausal women". *JAMA*, 1996, 275, 370.
- [8] Lane G., Siddle N.C., Ryder T.A., Pryse-Davies J., King R.J.B., Whitehead M.I.: "Is Provera the ideal progestogen for addition to postmenopausal estrogen therapy?". Fertil. Steril., 1986, 45, 345.
- [9] Writing group for the PEPI trial: "Effects of estrogen or estrogen/progestin regimens on heart disease risk factors in postmenopausal women". *JAMA*, 1995, 273, 199.
- [10] Mueck A.O., Salbach B., Rabe T., von Holst T., Runnebaum B.: "Serumlipide unter Behandlung mit transdermalem Östradiol und oralem Norethisteronacetat". *Geburtsh. u. Frauenheilk*, 1995, 55, 393.
- [11] Whitehead M.I., Towsend P.T., Pryse-Davies J., Ryder T. A., King R.J.B.: "Effects or estrogens and progestins on the biochemistry and morphology of the postmenopausal endometrium". *N. Engl. J. Med.*, 1981, *305*, 1599.
- [12] Esterbauer H., Striegl G., Puhl H., Rotheneder M.: "Continuous monitoring of in vitro oxidation of human low density lipoprotein". Free Rad. Res. Comms., 1989, 6, 67.
- [13] Parthasarathy S., Steinberg D., Witzum J.L.: "The role of oxidized low-density lipoprotein in the pathogenesis of atherosclerosis". *Annu. Rev. Med.*, 1992, 43, 219.
- [14] Morel D.W., DiCorleto P., Chisholm G.M.: "Endothelial and smooth muscle cells alter low density lipoprotein in vitro by free radical oxidation". *Arteriosclerosis*, 1994, 4, 357.

- [15] Galle J., Mülsch A., Busse R., Bassenge E.: "Effects of native and oxidized low density lipoproteins on formation and inactivation of endothelium-derived relaxing factor". *Arterioscler. Thromb.*, 1991, 11, 198.
- [16] Boulanger C.M., Tanner F.C., Bea M.L., Hahn A.W.A., Werner W., Lüscher T.F.: "Oxidized low density lipoproteins induce mRNA expression and release endothelin from human and procine endothelium". Circ. Res., 1992, 70, 1191.
- [17] Galle J., Bassenge E., Busse R.: "Oxidized low density lipoproteins potentiate vasoconstrictions to various agonists by direct interaction with vascular smooth muscle". Circ. Res., 1990, 676, 1287.
- [18] Seeger H., Mueck A.O., Lippert T.H.: "Effect of estradiol metabolites on the susceptibility of low density lipoprotein to oxidation". *Life Sciences*, 1997, *61*, 865.
  [19] Kuhl H.: "Oral contraception and replacement therapy: The
- [19] Kuhl H.: "Oral contraception and replacement therapy: The significance of the progestogen for cardiovascular diseases". *Geburtsh. u. Frauenheilk.*, 1992, 52, 653.
- [20] Munk-Jensen N., Ulrich L.G., Obel E.B., Nielsen S.P., Edwards D., Meinertz H.: "Continuous combined and sequential estradiol and norethindrone acetate treatment of postmenopausal women: effect on plasma lipoproteins in a two-year placebo-controlled trial". Am. J. Obstet. Gynecol., 1994, 171, 132.
- [21] Fraser D. I., Parsons A., Whitehead M. I;, Wordsworth J., Stuart G., Pryse-Davies J.: "The optimal dose of oral norethisterone acetate for addition to estradiol". *Fertil. Steril.*, 1990, 53, 460.

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