# Catalase activity, serum trace element and heavy metal concentrations, vitamin A, vitamin D and vitamin E levels in hydatidiform mole

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### **Summary**

Purpose of investigation: In this study we aimed to measure the activity of catalase, which is an antioxidant enzyme, the concentrations of some trace elements and heavy metals, and vitamin A, D and E levels in serum samples of patients with hydatidiform mole, normal pregnancies and healthy non pregnant women. Methods: Seventy-two women were enrolled in this study. Of these, 24 were healthy women in the first trimester of pregnancy (HP), 24 were healthy non-pregnant women (NP) and 24 were patients with complete hydatidiform mole (CHM). Results: Serum levels of catalase, Zn, Co, vitamin A, D and E were significantly lower in the CHM group when compared with the HP and NP groups (p < 0.001). Serum levels of Cu, Fe, and Cd were significantly higher in the CHM group when compared with the HP and NP groups (p < 0.001). Conclusion: The assessment of oxidant/antioxidant imbalance in pregnant women could be useful in the early determination of molar pregnancy and supplementation with antioxidants may be useful in the treatment of CHM, and may prevent recurrent molar pregnancy.

Key words: Catalase activity; Heavy metal; Trace element; Vitamin levels; Hydatidiform mole.

# Introduction

Hydatidiform mole (HM) is a disorder of fertilization. It is characterized by a conceptus of hyperplastic trophoblastic tissue attached to the placenta [1]. Broad variations in the distribution of gestational trophoblastic disease (GTD) exist worldwide, with higher frequencies in some parts of Asia, the Middle East and Africa, with Turkey ranking in the middle of these countries [2]. The etiology of molar pregnancy is not completely understood [3]. Preeclamptic patients are exposed to increased oxidative stress [4, 5]. Placental hypersecretion of lipid peroxides or decreased placental antioxidant enzyme production may lead to endothelial dysfunction [6]. Placental abnormality and associated metabolic changes cause increased oxidative stress [7]. Similar metabolic changes are present in molar pregnancy [8]. The antioxidant enzymes such as superoxide dismutase, catalase, and glutathione peroxidase (GPx) are synthesized in the body. Catalase is an effective scavenger of aqueous peroxide radicals [9]. The maternal blood levels of vitamin C, vitamin E, vitamin A, and  $\beta$ -carotene which has a strong provitamin A activity are decreased [10, 11]. Vitamin D might exert subtle oxidative stress, which could stimulate the detoxification mechanisms to protect cells from the subsequent stress challenges [12]. The studies on the roles of trace elements in health and disease over the past 50 years have led to a good understanding of their mode of action and why they are essential for life [13]. Some metals are essential to maintain the metabolism of the human body, whereas they can lead to poisoning because they tend to bioaccumulate [14]. Lead (Pb) and cadmium (Cd) increase oxidative stress [15].

In this study, we aimed to measure the activity of catalase which is an antioxidant enzyme; the concentrations of some trace elements and heavy metals (Cu, Zn, Cd, Co and Fe); vitamin A (retinol), vitamin D (cholecalciferol) and vitamin E ( $\alpha$ -tocopherol) levels in serum samples of patients with hydatidiform mole, with normal pregnancies and healthy non-pregnant women.

### **Material and Methods**

Seventy-two women were enrolled in this study. Of these, 24 were healthy women in the first trimester of pregnancy with a single viable fetus (HP). Twenty-four healthy non-pregnant women also participated as controls (NP). The remaining 24 subjects had complete hydatidiform mole (CHM). Written informed consent was obtained from all subjects.

None of these patients were using any drugs at the time of collection of the blood samples. Venous blood samples were drawn from each patient during the 10<sup>th</sup>-19<sup>th</sup> week of gestation after a fasting overnight period. Serum was separated by centrifugation and the samples were processed immediately. The serum samples were placed in deionized polyethylene tubes and kept in a deep-freeze at -20°C (without thawing) until the study day.

Biochemical analysis of catalase activity in erythrocytes was determined according to the method defined by Aebi, in short, the supernatant (0.1 ml) was added to a quartz cuvette containing 2.95 ml of 19 mmol l<sup>1</sup> H<sub>2</sub>O<sub>2</sub> solution prepared in potassium phosphate buffer (0.05M, pH 7.00) [16]. The change in absorbance was monitored at 240 nm over a 5-min period using a spectrophotometer (Shimadzu UV-1201, Japan). Determination of serum concentrations of Cu, Zn, Co, Cd, and Fe was performed by atomic absorption measurements, in which a

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UNICAM-929 spectrophotometer (Unicam Ltd, York Street, Cambridge, UK) was used. In addition, vitamin E, vitamin D and vitamin A levels were determined by a high-performance liquid chromatography (HPLC) method [17]. Data were expressed as mean ± standard error (SE) and analyzed using one-way repeated measures of analysis of variance (ANOVA). Duncan's test was used to test for differences among means for which one-way ANOVA indicated a significant (p < 0.01) F ratio.

### Results

Demographic data of the subjects are shown in Table 1. There were no differences in mean age, gestational age, gravidity and parity between patients with CHM and HP (p > 0.05).

Table 1. — Characteristics of complete hydatidiform mole (CHM), healthy pregnant (HP) and healthy non pregnant (NP)

	CHM group (n = 24) Mean (SE)	HP group (n = 24) Mean (SE)	NP group (n = 24) Mean (SE)	p value
Age (years)	28.88 ± 1.71	29.17 ± 1.11	$28.83 \pm 0.87$	NS
Gestational age	e			
(weeks)	$15.71 \pm 040$	$15.33 \pm 0.56$	_	NS
Gravidity	$4.46 \pm 0.74$	$4.00 \pm 0.49$	_	NS
Parity	$3.13 \pm 0.67$	$2.67 \pm 0.50$	_	NS

NS: nonsignificant.

Serum levels of catalase, vitamin A, D and E were significantly lower in the CHM group when compared with the HP and NP groups (p < 0.001). The mean values of serum levels of catalase, vitamin A, vitamin D, and vitamin E are given in Table 2.

Table 2. — Serum levels of catalase and vitamins (A, D, E) of complete hydatidiform mole (CHM), healthy pregnant (HP), and non-pregnant (NP) groups.

	CHM group (n = 24) Mean (SE)	HP group (n = 24) Mean (SE)	NP group (n = 24) Mean (SE)
Catalase EU/(gHb)-1	$3.01 \pm 0.33^{a}$	$10.22 \pm 1.12^{\text{b}}$	11.86 ± 1.61 <sup>b</sup>
Vitamin A (mmol/l)	$0.50 \pm 0.04^{a}$	$0.72 \pm 0.02^{b}$	$0.74 \pm 0.03^{b}$
Vitamin D (mmol/l)	$0.02 \pm 0.00^{a}$	$0.03 \pm 0.00^{b}$	$0.03 \pm 0.00^{b}$
Vitamin E (mmol/l)	$4.72 \pm 0.34^{a}$	$6.26 \pm 0.32^{b}$	$6.36 \pm 0.20^{b}$

Values are means  $\pm$  SE; different letters represent significant differences (p < 0.01).

Serum levels of Cu, Fe, and Cd was significantly higher in the CHM group when compared with the HP and NP groups (p < 0.001). Serum levels of Zn and Co were statistically lower in the CHM group compared with the HP and NP groups (p < 0.001). Mean values of serum levels of Cu, Zn, Fe, Co and Cd are given in Table 3.

Table 3. — Serum levels of Cu, Zn, Fe, Cd and Co of complete hydatidiform mole (CHM), healthy pregnant (HP), and nonpregnant (NP) groups.

	CHM group (n = 24) Mean (SE)	HP group (n = 24) Mean (SE)	NP group (n = 24) Mean (SE)
Cu (µg/dl)	$3.49 \pm 0.16^{a}$	$2.82 \pm 0.09^{b}$	1.81 ± 0.08°
Zn (µg/dl)	$1.25 \pm 0.08^{a}$	$1.71 \pm 0.07^{\text{b}}$	$1.88 \pm 0.09^{\circ}$
Fe (µg/dl)	$4.48 \pm 0.71^{a}$	$1.79 \pm 0.32^{\text{b}}$	$1.30 \pm 0.07^{\circ}$
Cd (µg/dl)	$0.26 \pm 0.03^{a}$	$0.02 \pm 0.00^{b}$	$0.03 \pm 0.00^{\circ}$
Co (µg/dl)	$0.03 \pm 0.00^{a}$	$0.26 \pm 0.03^{b}$	$0.24 \pm 0.04^{\circ}$

Values are means  $\pm$  SE; different letters represent significant differences (p < 0.01).

### Discussion

Reactive oxygen species (ROS) are continuously produced during normal physiologic events, and removed by antioxidant defence mechanisms. In pathological conditions, ROS are over produced and result in lipid peroxidation and oxidative damage [18, 19]. There is increasing evidence that oxidative stress is an important contributing factor in the pathogenesis of preeclampsia [9]. Similar increasing oxidative stress is present in molar pregnancy [8]. We found that catalase was significantly lower in the CHM group when compared with the HP and NP groups. The importance of vitamin E and β-carotene in preventing free radical damage is well known and their levels have been reported to be significantly decreased in preeclampsia [11]. Maternal vitamin D deficiency may be an independent risk factor for preeclampsia. Vitamin D supplementation in early pregnancy should be explored for preventing preeclampsia and promoting neonatal well-being [12]. There is no study evaluating vitamin A, D and E levels in hydatidiform moles in the English literature. In the present study, we observed significantly lower levels of vitamin A, vitamin D and vitamin E in CHM patients when compared with the HP and NP groups. Significant changes in serum trace metal concentrations, particularly zinc and copper, have been documented during normal pregnancies. Harma et al. reported that levels of zinc in serum were found to be significantly higher and levels of copper in serum were significantly lower in hydatidiform mole patients than controls [20]. On the contrary, Wang et al. reported that the contents of Cu in erythrocytes were found to be significantly higher and the contents of Zn significantly lower in patients with hydatidiform mole than those in normal females [21]. In our study, we found that zinc was significantly lower and copper was significantly higher in the CHM group when compared with the HP and NP groups.

Some metals are essential to maintain the metabolism of the human body, whereas they can lead to poisoning because they tend to bioaccumulate [14]. The Industrial Revolution, which increased opportunities of occupational and environmental exposure to metals among women, revealed their adverse effects on pregnancy [22]. Lead and cadmium are established toxic and carcinogenic metals [23]. Lead and cadmium increase oxidative stress [15]. Iron is a redox-active transition metal and can participate in single electron reactions and catalyse formation of free radicals, including the undesirable hydroxyl radicals [24, 25]. There is no study evaluating heavy metal levels in hydatidiform moles in the literature. Serdar et al. observed a significant increase in serum iron levels in mild and severe preeclampsia patients when compared to normotensive healthy women [26]. In our study we found that cadmium and iron were significantly higher and cobalt was significantly lower in the CHM group when compared with the HP and NP groups.

The assessment of oxidant/antioxidant imbalance in pregnant women could be useful in the early determination of molar pregnancy. Supplementation with antioxidants in pregnant women with low antioxidant status may be useful in the treatment of CHM and may prevent recurrent molar pregnancy. However, more studies are needed to verify and clarify the relationship between antioxidant levels and heavy metal levels and hydatidiform mole.

# References

- Di Cintio E., Parazzini F., Rosa C., Chatenoud L., Benzi G.: "The epidemiology of gestational trophoblastic disease". *Gen. Diagn.* Pathol., 1997, 143, 103.
- [2] Altieri A., Franceschi S., Ferlay J., Smith J., La Vecchia C.: "Epidemiology and aetiology of gestational trophoblastic diseases". *Lancet Oncol.*, 2003, 4, 670.
- [3] Berkowitz R.S., Bernstein M.R., Harlow B.L., Rice L.W., Lage J.M., Goldstein D.P. et al.: "Case-control study of risk factors for partial molar pregnancy". Am. J. Obstet. Gynecol., 1995, 173, 788.
- [4] Many A., Hubel C.A., Fisher S.J., Roberts J.M., Zhou Y.: "Invasive cytotrophoblasts manifest evidence of oxidative stress in preeclampsia". Am. J. Pathol., 2000, 156, 321.
- [5] Hubel C.A.: "Oxidative stress in the pathogenesis of preeclampsia". Proc. Soc. Exp. Biol. Med., 1999, 222, 222.
- [6] Maseki M., Nishigaki I., Hagihara M., Tomoda Y., Yagi K.: "Lipid peroxide levels and lipids content of serum lipoprotein fractions of pregnant subjects with or without pre-eclampsia". Clin. Chim. Acta, 1981, 115, 155.
- [7] Merabishvili N., Sanikidze T., Sioridze E.: "Contemporary understanding of etiopathogenesis of preeclampsia". Tbilisi State Medical University Annals of Biomedical Research and Education, 2002, 2, 95.
- [8] Harma M., Harma M., Erel O.: "Increased oxidative stress in patients with hydatidiform mole". Swiss Med. Wkly, 2003, 133, 563.
- [9] Derevianko A., D'Amico R., Graeber T., Keeping H., Simms H.H.: "Endogenous PMN-derived reactive oxygen intermediates provide feedback regulation on respiratory burst signal transduction". J. Leukocyte Biol., 1997, 62, 268.
- [10] Wang Y.P., Walsh S.W., Guo J.D., Zhang J.Y.: "The imbalance between thromboxane and prostacyclin in preeclampsia is associated with an imbalance between lipid peroxides and vitamin E in maternal blood". Am. J. Obstet. Gynecol., 1991, 165, 1695.
- [11] Mikhail M.S., Anyaegbunam A., Garfinkel D., Palan P.R., Basu J., Romney S.L.: "Preeclampsia and antioxidant nutrients: decreased plasma levels of reduced ascorbic acid, alpha-tocopherol, and beta-carotene in women with preeclampsia". *Am. J. Obstet. Gynecol.*, 1994, *171*, 150.

- [12] Bodnar L.M., Catov J.M., Simhan H.N., Holick M.F., Powers R.W., Roberts J.M.: "Maternal vitamin D deficiency increases the risk of preeclampsia". J. Clin. Endocrinol. Metab., 2007, 92, 3517.
- [13] Yaman M., Kaya G., Simsek M.: "Comparison of trace element concentrations in cancerous and noncancerous human endometrial and ovary tissues". *Int. J. Gynecol. Cancer*, 2007, 17, 220.
- [14] Rani A.U.: "Cadmium-induced bioaccumulation in the selected tissues of a freshwater teleost, Oreochromis mossambicus (Tilapia)". Ann. NY Acad. Sci., 2000, 919, 318.
- [15] Stohs S.J., Bagchi D.: "Oxidative mechanisms in the toxicity of metal ions". Free Radic. Biol. Med., 1995, 18, 321.
- [16] Aebi H.: "Catalase in vitro". Methods Enzymol., 1984, 105, 121.
- [17] Meral I., Mert H., Mert N., Deger Y., Yoruk I., Yetkin A. et al.: "Effects of 900-Mhz electromagnetic field emitted from cellular phone on brain oxidative stress and some vitamin levels of guinea pigs". Brain Res., 2007, 1169, 120.
- [18] Cho C.H., Pfeiffer C.J., Misra H.P.: "Ulcerogenic mechanism of ethanol and the activation of sulphanilyl on the rat stomach in vivo". J. Pharm. Pharmacol., 1991, 43, 495.
- [19] Kanter M., Demir H., Karakaya C., Ozbek H.: "Gastroprotective activity of Nigella Sativa L oil and its constituent, thymoquinone against acute alcohol induced gastric mucosal injury in rats". World J. Gastroenterol., 2005, 11, 6662.
- [20] Harma M., Harma M., Kocyigit A., Keles H.: "Serum levels of zinc and copper in hydatidiform mole". Arch. Gynecol. Obstet., 2005, 271, 304.
- [21] Wang C.Y., Qi X.M.: "Studies on some trace elements and cell-mediated immunity in patients with gestational trophoblastic disease". J. Tongji Med. Univ., 1993, 13, 51.
- [22] Jarup L.: "Hazards of heavy metal contamination". Br. Med. Bull., 2003, 68, 167.
- [23] Agency for Toxic Substances and Disease Registry. US Department of Health and Human Services, Public Health Service, 1999.
- [24] Casanueva E., Viteri F.E.: "Iron and oxidative stress in pregnancy". J. Nutr., 2003, 133, 1700.
- [25] Gurzau E.S., Neagu C., Gurzau A.E.: "Essential metals-case study on iron". *Ecotoxicol. Environ. Saf.*, 2003, 56, 190.
- [26] Serdar Z., Gur E., Develioglu O.: "Serum iron and copper status and oxidative stres in severe and mild preeclampsia". Cell. Biochem. Funct., 2006, 24, 209.

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