

## Review

# An Update Review of the Pathogenesis Hypothesis in Preeclampsia

Rui Lian<sup>1,2</sup>, Bao-Sheng Zhu<sup>3</sup>, Xi Zeng<sup>1,2,\*</sup><sup>1</sup>Department of Gynecology and Obstetrics, The West China Second University Hospital, Sichuan University, 610041 Chengdu, Sichuan, China<sup>2</sup>Key Laboratory of Birth Defects and Related Diseases of Women and Children, Sichuan University, Ministry of Education, 610041 Chengdu, Sichuan, China<sup>3</sup>Department of Genetic Diagnosis Center, First People's Hospital of Yunnan Province, 650032 Kunming, Yunnan, China\*Correspondence: [zengxi1003@scu.edu.cn](mailto:zengxi1003@scu.edu.cn) (Xi Zeng)

Academic Editor: Paolo Ivo Cavoretto

Submitted: 2 February 2022 Revised: 17 April 2022 Accepted: 18 April 2022 Published: 22 July 2022

## Abstract

**Objectives:** Hypertensive disorders occur in approximately 12% to 22% of pregnancies and cause substantial perinatal morbidity and mortality of both mother and fetus. Hypertensive disease is directly responsible for approximately 20% of maternal deaths and can be classified as chronic hypertension, gestational hypertension, preeclampsia-eclampsia, and chronic hypertension with superimposed preeclampsia. At present, the pathogenesis of preeclampsia is still unclear, we wrote this article to make a uptodate review of this disease.

**Mechanism:** A comprehensive search of several databases was conducted from inception up to March 2022. The searched databases were Web of Science, MEDLINE, Ovid, and Cochrane Database of Systematic Reviews. The search strategy included the combinations of the following medical terms: Hypertensive disorders; preeclampsia; mechanism; pathogenesis hypothesis. **Findings in Brief:** At present, the pathogenesis of preeclampsia is still unclear, the theory of Genetic, Inflammatory Response, Immune Imbalance in Maternal-Fetal Interface, Oxidative Stress, Vascular Endothelial Cell Damage are supposed involved in the progress of preeclampsia. **Conclusions:** Although there are various theories mentioned above, none of the hypothesis can fully explain preeclampsia. More research is needed on the mechanism of preeclampsia.

**Keywords:** preeclampsia; pathogenesis hypothesis; mechanism; review

## 1. Introduction

Hypertensive disorders occur in approximately 12% to 22% of pregnancies and cause substantial perinatal morbidity and mortality of both mother and fetus. Hypertensive disease is directly responsible for approximately 20% of maternal deaths and can be classified as chronic hypertension, gestational hypertension, preeclampsia-eclampsia, and chronic hypertension with superimposed preeclampsia [1,2].

Preeclampsia is the development of hypertension with proteinuria after 20 weeks of gestation, with or without proteinuria, in conjunction with fetal growth restriction (FGR), maternal endothelial dysfunction, and chronic immune activation [3]. The main risk factors for the development of preeclampsia (PE) are first pregnancy, previous or family history of PE, chronic hypertension, diabetes, antiphospholipid syndrome (APS), obesity and thrombophilia (Table 1, Ref. [4,5]). The basic pathophysiological changes are systemic arteriole spasm, vascular endothelial injury and ischemia, causing damage to multiple organs [3].

Preeclampsia progresses in 2 stages: in the first stage, uterine spiral artery remodeling disordered, resulting in abnormal trophoblastic infiltration and superficial placenta implantation in the maternal myometrium. In the second stage, some maternal syndrome appears, which is associated with the systemic inflammatory response caused by the placenta, leading to the signs of preeclampsia [3,6].

Delivery can resolve most signs and symptoms; however, preeclampsia can persist after delivery and recurrence in the postpartum period [3].

At present, the pathogenesis of preeclampsia is still unclear, large number of hypothesis have been proposed for the placental dysfunction, including oxidative stress, abnormal natural killer cells at the maternal-fetal interface, and genetic and environmental factors, though none have conclusive evidence in humans [6]. This article summarized the research status of pathogenesis in preeclampsia as follows (Fig. 1).

## 2. Genetic Theory

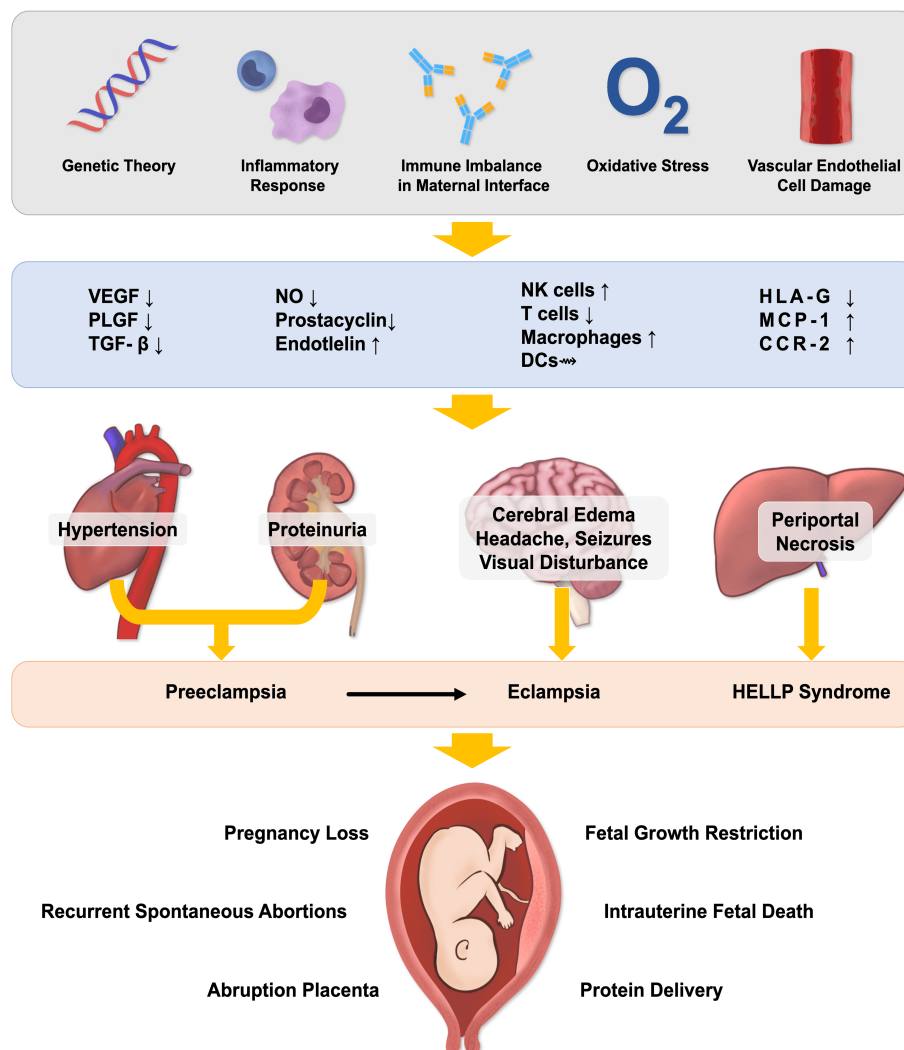
Family history is an independent risk factor of preeclampsia [7]. As early as the 1870s, the literature reported a family gathering phenomenon of preeclampsia [8]. The preeclampsia incidence of pregnant women can be increased by 2 to 5 times, whose immediate family members suffered with preeclampsia previously [9,10]. A family history of preeclampsia can significantly increase the risk of individual women suffering from preeclampsia by 24% to 163% [11]. A study from Norway showed that women born with preeclampsia has a 2.2-fold increased risk of preeclampsia during her pregnancy, indicating that preeclampsia was genetic predisposition [12].

For men whose mothers had preeclampsia, the men might also increase the incidence of preeclampsia in their



**Table 1. Major risk factors for the preeclampsia [4,5].**

Risk factors	Odds ratio (OR) or Relative risk (RR) (95% CI)
Antiphospholipid antibody syndrome	9.7 (4.3–21.7)
Renal disease	7.8 (2.2–28.2)
Preeclampsia in previous pregnancy	7.2 (5.8–8.8)
Systemic lupus erythmatosis	5.7 (2.0–16.2)
Nulliparity	5.4 (2.8–10.3)
Maternal age >40 years	3–4 (4.7–9.6)
Chronic hypertension	3.8 (3.4–4.3)
Diabetes Mellitus	3.6 (2.5–5.0)
Sister with preeclampsia	3.3 (1.5–7.5)
Strong family history of cardiovascular disease	3.2 (1.4–7.7)
Twin pregnancy	3 (2–4.2)
COVID-19	2.84 (1.67–4.82)
Obesity	2.5 (1.7–3.7)
Body mass index >25.8	2.3–2.7 (2.8–4.4)
Previous partner had PE	1.8 (1.6–3.9)
Excessive weight gain	1.7–2.8 (1.9–4.4)
Artificial insemination	1.6–2.5 (1.7–2.9)
Use of contraceptive barrier method	1.6–2.1 (4.3–7.3)
Pregnant woman born with low birth weight	1.4 (1.6–3.0)
Bleeding in 1st trimester	1.4 (1.4–2.6)



**Fig. 1. Mechanism of preeclampsia.**

spouses [13]. Dekker [14] claimed that, for the husbands, if their original spouses had a history of preeclampsia, when they had a next spouses without preeclampsia, the probability of preeclampsia in the new one would also increase significantly. Nearly half of preeclampsia family aggregation is caused by maternal genetic factors; and environmental, fetal or paternal factors account for the rest [15]. Nowadays, a variety of genetic models have been proposed to explain the inheritance of preeclampsia, including classic Mendelian Inheritance models [16], mitochondrial genes [17]. However, no hypothesis can fully explain preeclampsia. Therefore, preeclampsia is thought to be the result of complex interactions between maternal and fetal genotypes in addition to the environment [18].

### 3. Theory of Inflammatory Response

Compared with healthy pregnant women, the placenta of preeclampsia patients often present vascular abnormalities and inflammation, leading the reduction of placental spiral artery remodeling and increasing vasoconstriction [19]. Macrophages, natural killer cells (NK), dendritic cells (DCs), and T cells infiltrate in the decidua around the trophoblast cells, controlling the remove of inherent cells in the spiral artery and the invasion of trophoblasts [20–22]. Any imbalance of these local inflammatory reactions may lead to placental malformations, impairing placental blood supply, resulting in preeclampsia, urine protein and edema, and even pregnancy loss [23].

#### 3.1 Natural Killer (NK) Cells

Uterine Natural Killer (uNK) cells are the main lymphocytes infiltrating the decidua, accounting for about 70% of the immune cells at the maternal-fetal interface and regulating the depth of trophoblast cell infiltration and angiogenesis, playing a role in the abnormal placentation observed in preeclampsia [18,24]. Unlike peripheral NKs, uNK is not cytotoxic [18]. The uNK cells in the preeclampsia were significantly higher than those in the normal pregnancy group between 12 and 20 weeks of pregnancy [25].

There are specific receptors, such as killer cell immunoglobulin-like receptors (KIRs) on the surface of uNK cells in the maternal-fetal interface, which are related to the onset of preeclampsia [26]. The HLA-G of trophoblasts can contact with KIRs to regulate the invasion ability of trophoblasts [24,27]. However, when the uNK cells express the KIR inhibitory, the trophoblasts will tend to express HLA-C2, which will inhibit the function of NK cells in the maternal-fetal interface, and the resistance to spiral artery remodeling will increase significantly [28]. Uterine NK cells also mediate immune tolerance by recognizing major histocompatibility complex (MHC) ligands of fetal trophoblast cells [24].

#### 3.2 T Cells

T lymphocytes play an important role in mediating the occurrence of preeclampsia [29]. The imbalance of T cells, appearing in the blood circulation of women with low placental function, leads to a chronic inflammatory state, which is believed to be one of the causes of the disease [30].

During a normal pregnancy, maternal Th1 and Th2 establish a new balance, when the immune state drifts to Th2, the Th1 cytokines are down-regulated, promoting the normal placenta development and avoiding immune rejection of embryos in utero [31,32]. Th1 cells can secrete interleukin-2 (IL-2), tumor necrosis factor (TNF)- $\alpha$ , etc., and these inflammatory factors is related to preeclampsia. Excessive immune inflammatory response can be suppressed by Treg cells. Treg is essential during pregnancy and promotes immune tolerance by regulating Th1 from primitive T cells [32,33].

Both Th1 and Th2 cells were differentiated from Th0 cells [32]. This differentiation of Th0 cells is regulated by regulatory T cells (Treg cells) [31,34]. Treg cells play an anti-inflammatory effect and the main effector cytokine is IL-10, helping form immune tolerance and hinder the differentiation [31]. During this differentiation process, specific transcription factors GATA-3 promotes Th0 cells to differentiate into Th2 cells, and T-bet induces Th0 cells to differentiate into Th1 cells [35].

In preeclampsia patients, the levels of Treg and indoleamine 2,3-dioxygenase (IDO) are significantly lower than that in normal pregnant women [36]. IDO is an amino acid converting enzyme, which can effectively regulate Treg activity. When IDO decreases, Tregs activity will also decrease, promoting the transformation of Th0 cells to Th17 cells [37,38]. IL-17 is a pro-inflammatory factor (most of the core cytokines secreted by Th17), which can promote inflammation. Immune imbalance between Th17 cells and Treg cells is one of the important causes of preeclampsia [39].

#### 3.3 Macrophages

In the process of human embryo implantation, extravillous trophoblasts (EVT) derived from blastocysts invades the decidua, interacting with natural killer cells, enhancing the expression of angiogenic factors and microangiogenesis [40], increasing blood flow in the uterus and promoting the development of the fetus and placenta [41].

In early pregnancy, uterine macrophages participate in the initiation apoptosis of vascular smooth muscle cells and endothelial cells during decidual vascular remodeling [42]. The macrophage polarization imbalance damages trophoblast and placental function, and leads to preeclampsia, FGR, miscarriage and even preterm delivery [43]. Colony stimulating factor (CSF) is an effective inducer of macrophage proliferation, differentiation and activation [44]. Serum CSF in pregnant women are elevated and quickly return to baseline levels after the delivery [44]. The

dynamic phenotypic changes of macrophages indicate that: (1) The macrophage expression is elevated in decidua during preeclampsia [45]. (2) preeclampsia significantly enhances the expression of CSF in early decidual cells [46].

In late pregnancy, the elevated macrophage and loss of trophoblast were observed under some abnormal pregnancy conditions, including early pregnancy loss, preeclampsia, FGR and gestational trophoblastic disease, indicating macrophages homeostasis is essential for maintaining decidual homeostasis in human pregnancy [47–49].

### 3.4 Dendritic Cells (DCs)

Th1 and Treg cells in the placenta and decidua are differentiated from naive CD4<sup>+</sup> T cells induced by antigen presenting cells (APCs), the most important of which are dendritic cells (DCs) [50]. Low-level stimulatory molecules expressed by immature DCs (such as MHC-II and CD80/CD86, etc.) can induce naive CD4<sup>+</sup> T cells to differentiate into Treg cells and form immune tolerance [51]. While mature DCs express high levels of stimulatory molecules would induce naive CD4<sup>+</sup> T cells to differentiate into activated Th1 cells and mediate inflammation [52]. During a normal pregnancy, the number of DCs at the maternal-fetal interface increases and maintain immature [52,53]. Between the patients with preeclampsia and the normal pregnant patients, the number of DCs in the decidua is no significant difference; however, the ability of DCs inducing Treg cell is significantly impaired in patients with preeclampsia, and the mechanism is still not sure [51].

## 4. Theory of Immune Imbalance in Maternal-Fetal Interface

Pregnancy can be considered as an immune-homeostasis between the pregnant woman's tolerance and the protection of the fetus [54]. Normal pregnancy is a state that immune system is activated and pregnancies affected by preeclampsia are regarded to involve more complex immunological processes. The boundary between fetal components and maternal tissues is the maternal-fetal interface. The balance of maternal-fetal immune regulation is the vital to successful pregnancy [55]. In normal pregnancy, there is an increase in systemic immune cell activation, especially monocyte (the macrophage precursor) [43]. When infiltrated into tissues, circulating monocytes will differentiate to steady state macrophages [23]. Both of monocytes and macrophages are related to trophoblast growth, spiral artery remodeling and angiogenesis [56]. Among the pregnant women with preeclampsia, the monocyte chemoattractant protein 1 (MCP-1), mRNA of MCP-1 and C-C chemokine receptors 2 (CCR2, the receptor of MCP-1) in serum and placental tissue are significantly increased [57]. It is believed MCP-1 may induce excessive local placental inflammation by binding to its receptor CCR2, disrupting the immune balance and causing preeclampsia [57].

For pregnant women with preeclampsia, after losing the immunity homeostasis, the immunity to tolerate fetal antigens would decrease, causing abnormal placental immune function, inducing inflammatory reactions and inhibiting blood vessel production [58]. In addition, DCs in the decidua also induce mother's immune tolerance to the fetus by promoting Th2 cells in uterus and placenta [36]. In addition, the appropriate concentration of human leukocyte antigen G (HLA-G) can protect the embryo from the attack by cytotoxic T lymphocytes (CTL) [59]. Johnsen showed that when the expression of HLA-G in trophoblast cells is reduced, it is vulnerable to maternal immune cells, which can lead to preeclampsia, miscarriage or FGR [60].

## 5. Theory of Oxidative Stress

Oxidative stress (OS) is defined as an imbalance between oxidants and antioxidants, leading to a disruption of redox signaling and control and/or molecular damage [61,62]. The placenta is hypoxic in the first trimester of pregnancy, and the blood flows through the villus space in the later period and releases a large amount of reactive oxygen species (ROS), with an increase of antioxidants in the placenta to protect pregnant women and fetus from oxidative stress outbreaks [63].

### 5.1 Abnormal Placental Vascular Remodeling

Preeclampsia is associated with insufficient trophoblast infiltration and impaired vascular remodeling in spiral arteries [64]. Impaired vascular remodeling in preeclampsia reduces placental perfusion and oxygenation, and increases oxidative stress with ROS and toxic lipid peroxides [65]. Free radicals activate monocytes and neutrophils to produce pro-inflammatory cytokines, and further produce ROS through the action of various enzymes and the large amount of ROS produced by oxidative stress can damage the trophoblast cell mitochondrial membrane, permeable transport pore, mitochondrial DNA, and mitochondrial enzymes, which in turn triggers preeclampsia, premature rupture of membrane, etc. [63]. The expression of ROS damaging the vascular endothelium is significantly increased in the blood of women with preeclampsia; while the concentration of antioxidants weaken the activity of ROS in preeclampsia is significantly reduced [66,67].

### 5.2 Immune Dysregulation and Oxidative Stress

In preeclampsia women, the maternal immune response, against the fetus, interferes with trophoblast invasion and angiogenesis, resulting in oxidative stress and a maternal systemic inflammatory response [68]. Immunity imbalance of Th1/Th2 cells, Th17/Treg cells and abnormal regulation of decidual NK cells will lead to the failure of trophoblast invasion and the reduction of placental perfusion, which would generate of reactive oxygen species, and the aggravate of oxidative stress Autoreactive antibodies to the angiotensin receptor AT1 are elevated in preeclamptic



mothers and inhibit trophoblast invasion, thereby promoting oxidative stress [69]. Complement system also plays a crucial role in maintaining a healthy pregnancy, and abnormal activation of the complement system would cause maternal systemic inflammatory response, leading to the occurrence of preeclampsia.

Malonyldialdehyde (MDA) is a lipid peroxide, which is an important monitoring indicator of oxidative stress [70]. When the placental ischemia and hypoxia occur, the oxidative stress is intensified and releases a large amount of oxygen free radicals, which penetrate the cell membrane and attack the polyunsaturated fatty acids in the cell, converting them into lipid peroxides, and MDA as the product of lipid peroxidation reaction will increase significantly [71,72].

8-isoprostane is another final product of lipid peroxidation, catalyzed by arachidonic acid through oxygen free radicals [73]. Its level is consistent with the change trend of lipid peroxidation, which could be an indicator of lipid peroxidation reaction [74]. The more severe preeclampsia, the significantly higher 8-isoprostane in serum; which indicated the oxidative stress in preeclampsia is correlated with the severity of the disease [75].

### 5.3 Mitochondrial Oxidative Stress

Patients with preeclampsia got mitochondrial dysfunction, increased mitochondrial lipid peroxidation, and susceptibility to oxides [76]. In mitochondria of women with preeclampsia, those genes responsible for energy production and oxidative electron exchange, such as cytochrome C oxidase, are abnormally expressed [77]. Compared with normal pregnancy, endothelial cells treated with plasma from women with preeclampsia had significantly higher intramitochondrial ROS, inhibiting cell death ROS-induced [78].

## 6. Theory of Vascular Endothelial Cell Damage

The pathological mechanisms of preeclampsia include insufficient placental bed vascular remodeling, placental ischemia and hypoxia, secondary to systemic endothelial injury [16]. It is currently believed that the increase in the adhesion of leukocytes to endothelial cells and the endothelial cell permeability mediated by oxygen free radicals are critical processes in the development of preeclampsia [24,79].

### 6.1 Vascular Endothelial Growth Factor

Vascular endothelial growth factor (VEGF) can enhance vascular permeability, increase the ability of neo-vascular endothelial division and proliferation, regulate the permeability of placental vascular endothelial cells and promote blood vessels formation through the mediation of nitric oxide (NO) [74]. When the vascular endothelium is damaged, VEGF levels and the function of placental vascular endothelial cells will decrease, involving the occur-

rence of preeclampsia [80]. The levels of VEGF protein and mRNA in placental tissues of pregnant women with preeclampsia were lower than those of normal pregnant women [81]. As pregnancy progressed, the density of micro-vessel in normal placental were increased, while decreased in placental with preeclampsia, indicating VEGF inhibited the occurrence of preeclampsia [82]. The VEGF level in placenta preeclampsia was related to the uterine spiral artery resistance index and systolic period of pregnant women with preeclampsia, indicating the lower the serum VEGF level in pregnant women with preeclampsia, the greater blood flow resistance, and the less in placental perfusion [83].

Also, vascular endothelial growth factor or placental growth factor (PlGF) and anti-angiogenic factors like soluble fms-like tyrosine kinase 1 (sFlt-1) are known to be related to the disease pathogenesis [84]. Recombinant sFlt-1 could block development of endothelial tubes and inhibit vasodilatory effects of VEGF and PlGF in vasculature [85]. In preeclampsia, circulating maternal serum levels of sFlt-1 are increased, and PlGF levels are decreased [86].

In addition, endothelial cell damage can promote the release of tissue factor (TF), plasminogen activator inhibitor (PAI), thrombomodulin, platelets, etc., and activate the coagulation system [87]. The expression of TF in pregnant patients with hypertension was significantly higher than that in normal pregnant women. It was believed that the change of TF may cause abnormal coagulation function in pregnant women, promoting the occurrence of preeclampsia consequently [88]. Also, PAI elevated in pregnant women with preeclampsia [89,90].

Thrombomodulin is a transmembrane glycoprotein existing in vascular endothelial cells and trophoblast cells. Thrombomodulin is the receptor for thrombin, which can bind to thrombin to form a thrombomodulin-thrombin complex [91]. This substance has an activating effect on protein C form and can activate it into protein C, which can protect endothelial cells, reduce inflammation, anti-thrombosis, improve microcirculation in turn [92,93].

When the expression of thrombomodulin in the placenta tissue is reduced, the inflammatory response is prone to over-activation, damaging vascular endothelial cells [93, 94]. A lower expression of thrombomodulin in the placental tissue of pregnant women may be related to the evolution of early-onset severe preeclampsia [94]. Platelet may promote of blood coagulation and mediate inflammation in the occurrence and development of preeclampsia [95].

### 6.2 Nitric Oxide

Nitric Oxide (NO) is one of the key endothelial release factors in the body and the primary neurotransmitter; playing a role in regulating vascular blood flow, arterial blood pressure, maternal organ perfusion and placental blood flow, maintaining the metabolism, and keeping dynamic balance of blood pressure in cardiovascular [96].

In vascular endothelium, nitric oxide synthase (NOS) is the rate-limiting enzyme in NO synthesis, and the expression of NOS depends on the regulation of nuclear factor kappa-B (NF- $\kappa$ B) [97]. NF- $\kappa$ B exists in almost all cells, and plays a key role in cellular inflammatory and immune responses, which can be activated in the placenta of preeclampsia [98,99]. In patients with preeclampsia, hypoxic trophoblast cells produce plenty of endogenous digitalis-like factors, including NO and endothelin, which affect vasomotor and cell proliferation.

In vascular smooth muscle cells, NO can effect on cyclic guanosine phosphate, causing calcium outflow, promoting the conversion of free calcium into bound calcium and vasodilation [100]. Blocking NO synthesis with NOS inhibitors in pregnant rats would cause preeclampsia-like symptoms, including hypertension, proteinuria, FGR, even stillbirths and malformations [100,101]. In pregnant mice with NOS knockout, the uterine blood flow were reduced and spiral arteries were shortened, leading placenta oxygen supply insufficient [102,103]. The reduction of NO may cause an imbalance of vascular factors, followed by arteriole spasm and increased peripheral resistance, especially in kidney, uterus and placenta, which promotes the development of hypertension and preeclampsia [104]. NO is the important link in vascular endothelial cell damage and oxidative stress, which can ensure the blood supply of the placenta and the nutrition and oxygen supply of the fetus [105].

## 7. Other Theory

### 7.1 Oocyte Donation and In-Vitro Fertilization (IVF)

IVF is associated with the onset and progression of PE. Defective placentation and placental insufficiency may predispose IVF patients to preeclampsia and may manifest as first-trimester bleeding [106]. In IVF protocols including oocyte donation, the absence of corpus luteum and subsequent deficiency of relaxin can disturb maternal circulation and precipitate the development of preeclampsia [107]. Oocyte donation is an independent risk factor for preeclampsia due to immunological maladaptation or intolerance [108].

### 7.2 Pregestational Diabetics

Among women with pregestational diabetes, preeclampsia complicates 10% to 20% of the pregnancies [109]. GDM and preeclampsia share many risk factors, including advanced maternal age, nulliparity, multifetal pregnancies, non-white race/ethnicity and pre-pregnancy obesity [110].

### 7.3 COVID-19

Coronavirus disease 2019 (COVID-19) has been shown to cause systemic complications such as high blood pressure, kidney disease, thrombocytopenia, and liver injury [111]. COVID-19 during pregnancy is strongly associated with preeclampsia. COVID-19 severity does not

seem to be a factor in this association [112]. Women with preeclampsia should be considered a particularly vulnerable group with regard to the risks posed by COVID-19 [111].

## 8. Conclusions

Preeclampsia is a serious complication during pregnancy, which greatly threatens the pregnant women and fetuses. The pathogenesis of preeclampsia is still unclear, the theory of Genetic, Inflammatory Response, Immune Imbalance in Maternal-Fetal Interface, Oxidative Stress, Vascular Endothelial Cell Damage are supposed involved in the progress of preeclampsia. Although there are various theories mentioned above, none of them can fully explain all the biological behaviors of preeclampsia. More research is needed on the mechanism of preeclampsia.

## Author Contributions

RL—Project development, Literature Collection, Manuscript writing. BZ—Literature Collection, Critical revision of the manuscript, Supervision. XZ—Project development, Literature Collection, Critical revision of the manuscript, Supervision. All authors read and approved the final manuscript.

## Ethics Approval and Consent to Participate

Not applicable.

## Acknowledgment

We would like to express our gratitude to all those who helped us during the writing of this manuscript. Thanks to all the peer reviewers for their opinions and suggestions.

## Funding

This research received no external funding.

## Conflict of Interest

The authors declare no conflict of interest.

## References

- [1] Folk DM. Hypertensive Disorders of Pregnancy: Overview and Current Recommendations. *Journal of Midwifery and Women's Health*. 2018; 63: 289–300.
- [2] Wilkerson RG, Ogunbodede AC. Hypertensive Disorders of Pregnancy. *Emergency Medicine Clinics of North America*. 2019; 37: 301–316.
- [3] Rana S, Lemoine E, Granger JP, Karumanchi SA. Preeclampsia: pathophysiology, challenges, and perspectives. *Circulation Research*. 2019; 124: 1094–1112.
- [4] Ramos J, Sass N, Costa S. Preeclampsia. *Revista Brasileira De Ginecologia E Obstetrícia*. 2017; 39: 496–512.
- [5] Armaly Z, Jadaon JE, Jabbour A, Abassi ZA. Preeclampsia: novel mechanisms and potential therapeutic approaches. *Frontiers in Physiology*. 2018; 9: 973.
- [6] El-Sayed AAF. Preeclampsia: a review of the pathogenesis and possible management strategies based on its pathophysiological derangements. *Taiwanese Journal of Obstetrics and Gynecology*. 2017; 56: 593–598.

- [7] Oudejans CBM, van Dijk M, Oosterkamp M, Lachmeijer A, Blankenstein MA. Genetics of preeclampsia: paradigm shifts. *Human Genetics*. 2007; 120: 607–612.
- [8] Chesley LC, Anntito JE, Cosgrove RA. The Familial Factor in Toxemia of Pregnancy. *Obstetrics and Gynecology*. 1968; 32: 303–311.
- [9] Timokhina EV, Strizhakov AN, Ignatko IV, Belousova VS, Ibragimova SM. Genetic Aspects of Preeclampsia: The Role of Polymorphisms in the Genes of the Renin–Angiotensin System. *Biochemistry*. 2019; 84: 181–186.
- [10] Young BC, Levine RJ, Karumanchi SA. Pathogenesis of preeclampsia. *Annual Review of Pathology: Mechanisms of Disease*. 2010; 5: 173–192.
- [11] Boyd HA, Tahir H, Wohlfahrt J, Melbye M. Associations of Personal and Family Preeclampsia History with the Risk of Early-, Intermediate- and Late-Onset Preeclampsia. *American Journal of Epidemiology*. 2013; 178: 1611–1619.
- [12] Skjærven R, Vatten LJ, Wilcox AJ, Rønning T, Irgens LM, Lie RT. Recurrence of pre-eclampsia across generations: exploring fetal and maternal genetic components in a population based cohort. *British Medical Journal*. 2005; 331: 877.
- [13] Dawson LM, Parfrey PS, Hefferton D, Dicks EL, Cooper MJ, Young D, *et al.* Familial Risk of Preeclampsia in Newfoundland: a Population-Based Study. *Journal of the American Society of Nephrology*. 2002; 13: 1901–1906.
- [14] Dekker G. The partner's role in the etiology of preeclampsia. *Journal of Reproductive Immunology*. 2002; 57: 203–215.
- [15] Cnattingius S, Reilly M, Pawitan Y, Lichtenstein P. Maternal and fetal genetic factors account for most of familial aggregation of preeclampsia: a population-based Swedish cohort study. *American Journal of Medical Genetics Part A*. 2004; 130: 365–371.
- [16] Gregg AR. Preeclampsia. In Emery and Rimoin's Principles and Practice of Medical Genetics and Genomics. 2022; 4: 217–234.
- [17] Vishnyakova PA, Volodina MA, Tarasova NV, Marey MV, Tsvirkun DV, Vavina OV, *et al.* Mitochondrial role in adaptive response to stress conditions in preeclampsia. *Scientific Reports*. 2016; 6: 32410.
- [18] Moffett A, Colucci F. Uterine NK cells: active regulators at the maternal-fetal interface. *Journal of Clinical Investigation*. 2014; 124: 1872–1879.
- [19] Cornelius DC. Preeclampsia: from Inflammation to Immunoregulation. *Clinical Medicine Insights: Blood Disorders*. 2018; 11: 1179545X17752325.
- [20] Skog A, Lagnefeldt L, Conner P, Wahren-Herlenius M, Sonesson S. Outcome in 212 anti-Ro/SSA-positive pregnancies and population-based incidence of congenital heart block. *Acta Obstetrica Et Gynecologica Scandinavica*. 2016; 95: 98–105.
- [21] Murthi P, Pinar AA, Dimitriadis E, Samuel CS. Inflammasomes—A Molecular Link for Altered Immunoregulation and Inflammation Mediated Vascular Dysfunction in Preeclampsia. *International Journal of Molecular Sciences*. 2020; 21: 1406.
- [22] Steegers EA, von Dadelszen P, Duvekot JJ, Pijnenborg R. Preeclampsia. *The Lancet*. 2010; 376: 631–644.
- [23] Michalczyk M, Celewicz A, Celewicz M, Woźniakowska-Gondek P, Rzepka R. The Role of Inflammation in the Pathogenesis of Preeclampsia. *Mediators of Inflammation*. 2020; 2020: 3864941.
- [24] Yang X, Yang Y, Yuan Y, Liu L, Meng T. The Roles of Uterine Natural Killer (NK) Cells and KIR/HLA-C Combination in the Development of Preeclampsia: A Systematic Review. *BioMed Research International*. 2020; 2020: 4808072.
- [25] Du M, Wang W, Huang L, Guan X, Lin W, Yao J, *et al.* Natural killer cells in the pathogenesis of preeclampsia: a double-edged sword. *The Journal of Maternal-Fetal & Neonatal Medicine*. 2022; 35: 1028–1035.
- [26] Larsen TG, Hackmon R, Geraghty DE, Hviid TVF. Fetal human leukocyte antigen-C and maternal killer-cell immunoglobulin-like receptors in cases of severe preeclampsia. *Placenta*. 2019; 75: 27–33.
- [27] Xu X, Zhou Y, Wei H. Roles of HLA-G in the Maternal-Fetal Immune Microenvironment. *Frontiers in Immunology*. 2020; 11: 2767.
- [28] Johnsen GM, Fjeldstad HES, Drabbels JJM, Haasnoot GW, Eikmans M, Størvold GL, *et al.* A possible role for HLA-G in development of uteroplacental acute atherosclerosis in preeclampsia. *Journal of Reproductive Immunology*. 2021; 144: 103284.
- [29] Rahimzadeh M, Norouzian M, Arabpour F, Naderi N. Regulatory T-cells and preeclampsia: an overview of literature. *Expert Review of Clinical Immunology*. 2016; 12: 209–227.
- [30] Robertson SA, Green ES, Care AS, Moldenhauer LM, Prins JR, Hull ML, *et al.* Therapeutic potential of regulatory T cells in preeclampsia—opportunities and challenges. *Frontiers in Immunology*. 2019; 10: 478.
- [31] Vargas-Rojas MI, Solleiro-Villavicencio H, Soto-Vega E. Th1, Th2, Th17 and Treg levels in umbilical cord blood in preeclampsia. *The Journal of Maternal-Fetal & Neonatal Medicine*. 2016; 29: 1642–1645.
- [32] Dong M, He J, Wang Z, Xie X, Wang H. Placental imbalance of Th1- and Th2-type cytokines in preeclampsia. *Acta Obstetrica Et Gynecologica Scandinavica*. 2005; 84: 788–793.
- [33] Redman CWG, Sargent IL, Taylor RN. Immunology of Normal Pregnancy and Preeclampsia. *Chesley's Hypertensive Disorders in Pregnancy*. 2015; 60: 161–179.
- [34] Han X, Ghaemi MS, Ando K, Peterson LS, Ganio EA, Tsai AS, *et al.* Differential dynamics of the maternal immune system in healthy pregnancy and preeclampsia. *Frontiers in Immunology*. 2019; 10: 1305.
- [35] Ribeiro VR, Romao-Veiga M, Romagnoli GG, Matias ML, Nunes PR, Borges VTM, *et al.* Association between cytokine profile and transcription factors produced by T-cell subsets in early- and late-onset pre-eclampsia. *Immunology*. 2017; 152: 163–173.
- [36] Hosseini A, Dolati S, Hashemi V, Abdollahpour-Alitappeh M, Yousefi M. Regulatory T and T helper 17 cells: their roles in preeclampsia. *Journal of Cellular Physiology*. 2018; 233: 6561–6573.
- [37] Robertson SA, Care AS, Moldenhauer LM. Regulatory T cells in embryo implantation and the immune response to pregnancy. *The Journal of Clinical Investigation*. 2018; 128: 4224–4235.
- [38] Chang R, Li D, Li M. The role of indoleamine-2,3-dioxygenase in normal and pathological pregnancies. *American Journal of Reproductive Immunology*. 2018; 79: e12786.
- [39] Eghbal-Fard S, Yousefi M, Heydarlou H, Ahmadi M, Taghavi S, Movasaghpour A, *et al.* The imbalance of Th17/Treg axis involved in the pathogenesis of preeclampsia. *Journal of Cellular Physiology*. 2019; 234: 5106–5116.
- [40] Pollheimer J, Vondra S, Baltayeva J, Beristain AG, Knöfler M. Regulation of placental extravillous trophoblasts by the maternal uterine environment. *Frontiers in Immunology*. 2018; 9: 2597.
- [41] Qi Y, Ning F, Lash GE. Uterine macrophages: Essential roles for a successful human pregnancy. *Reproductive Immunology*. 2021; 262: 39–53.
- [42] Qi D, Wei M, Jiao S, Song Y, Wang X, Xie G, *et al.* Hypoxia inducible factor 1 $\alpha$  in vascular smooth muscle cells promotes angiotensin II-induced vascular remodeling via activation of CCL7-mediated macrophage recruitment. *Cell Death & Disease*. 2019; 10: 544.
- [43] Ma Y, Ye Y, Zhang J, Ruan C, Gao P. Immune imbalance is associated with the development of preeclampsia. *Medicine*. 2019; 98: e15080.

- [44] Pan L, Fu Z, Yin P, Chen D. Pre-existing medical disorders as risk factors for preeclampsia: an exploratory case-control study. *Hypertension in Pregnancy*. 2019; 38: 245–251.
- [45] Sun F, Wang S, Du M. Functional regulation of decidual macrophages during pregnancy. *Journal of Reproductive Immunology*. 2021; 143: 103264.
- [46] Krasnyi AM, Gracheva MI, Sadekova AA, Vtorushina VV, Balashov IS, Kan NE, *et al.* Complex Analysis of Total and Fetal DNA and Cytokines in Blood Plasma of Pregnant Women with Preeclampsia. *Bulletin of Experimental Biology and Medicine*. 2018; 164: 721–725.
- [47] Wheeler KC, Jena MK, Pradhan BS, Nayak N, Das S, Hsu CD, *et al.* VEGF may contribute to macrophage recruitment and M2 polarization in the decidua. *PLoS ONE*. 2018; 13: e0191040.
- [48] Faas MM, De Vos P. Innate immune cells in the placental bed in healthy pregnancy and preeclampsia. *Placenta*. 2018; 69: 125–133.
- [49] Vishnyakova P, Poltavets A, Nikitina M, Midiber K, Mikhaleva L, Muminova K, *et al.* Expression of Estrogen Receptor  $\alpha$  by Decidual Macrophages in Preeclampsia. *Biomedicine*. 2021; 9: 191.
- [50] Sahu MB, Deepak V, Gonzales SK, Rimawi B, Watkins KK, Smith AK, *et al.* Decidual cells from women with preeclampsia exhibit inadequate decidualization and reduced sFlt1 suppression. *Pregnancy Hypertension*. 2019; 15: 64–71.
- [51] Cornelius DC, Wallace K. Decidual natural killer cells: a critical pregnancy mediator altered in preeclampsia. *EBioMedicine*. 2019; 39: 31–32.
- [52] Cornelius DC, Cottrell J, Amaral LM, LaMarca B. Inflammatory mediators: a causal link to hypertension during preeclampsia. *British Journal of Pharmacology*. 2019; 176: 1914–1921.
- [53] Jung YJ, Park Y, Kim H, Lee HJ, Kim Y, Lee J, *et al.* Abnormal lymphatic vessel development is associated with decreased decidual regulatory T cells in severe preeclampsia. *American Journal of Reproductive Immunology*. 2018; 80: e12970.
- [54] Jin S, Wu C, Zhang Y. Complement in structure and immune homeostasis in placenta. 2021. (in press)
- [55] Yang F, Zheng Q, Jin L. Dynamic function and composition changes of immune cells during normal and pathological pregnancy at the maternal-fetal interface. *Frontiers in Immunology*. 2019; 10: 2317.
- [56] Lu H, Hu R. The role of immunity in the pathogenesis and development of pre-eclampsia. *Scandinavian Journal of Immunology*. 2019; 90: e12756.
- [57] Faas MM, Spaans F, De Vos P. Monocytes and macrophages in pregnancy and pre-eclampsia. *Frontiers in Immunology*. 2014; 5: 298.
- [58] Yang F, Zheng Q, Jin L. Dynamic Function and Composition Changes of Immune Cells During Normal and Pathological Pregnancy at the Maternal-Fetal Interface. *Frontiers in Immunology*. 2019; 10: 2317.
- [59] Sheridan MA, Yang Y, Jain A, Lyons AS, Yang P, Brahmasani SR, *et al.* Early onset preeclampsia in a model for human placental trophoblast. *Proceedings of the National Academy of Sciences*. 2019; 116: 4336–4345.
- [60] Johnsen GM, Størvold GL, Drabbe J, Haasnoot GW, Eikmans M, Spruyt-Gerritse MJ, *et al.* The combination of maternal KIR-B and fetal HLA-C2 is associated with decidua basalis acute atherosclerosis in pregnancies with preeclampsia. *Journal of Reproductive Immunology*. 2018; 129: 23–29.
- [61] Sies H. Oxidative stress: Concept and some practical aspects. *Antioxidants*. 2020; 9: 852.
- [62] Sies H, Berndt C, Jones DP. Oxidative Stress. *Annual Review of Biochemistry*. 2017; 86: 715–748.
- [63] Covarrubias AE, Lecarpentier E, Lo A, Salahuddin S, Gray KJ, Karumanchi SA, *et al.* AP39, a Modulator of Mitochondrial Bioenergetics, Reduces Antiangiogenic Response and Oxidative Stress in Hypoxia-Exposed Trophoblasts: relevance for preeclampsia pathogenesis. *The American Journal of Pathology*. 2019; 189: 104–114.
- [64] Mukherjee I, Dhar R, Singh S, Sharma JB, Nag TC, Mridha AR, *et al.* Oxidative stress-induced impairment of trophoblast function causes preeclampsia through the unfolded protein response pathway. *Scientific Reports*. 2021; 11: 18415.
- [65] Yang N, Wang Q, Ding B, Gong Y, Wu Y, Sun J, *et al.* Expression profiles and functions of ferroptosis-related genes in the placental tissue samples of early- and late-onset preeclampsia patients. *BMC Pregnancy and Childbirth*. 2022; 22: 87.
- [66] Tong W, Giussani DA. Preeclampsia link to gestational hypoxia. *Journal of Developmental Origins of Health and Disease*. 2019; 10: 322–333.
- [67] Matsubara K. Hypoxia in the pathogenesis of preeclampsia. *Hypertension Research in Pregnancy*. 2018; HRP2017-2014.
- [68] Guerby P, Tasta O, Swiader A, Pont F, Bujold E, Parant O, *et al.* Role of oxidative stress in the dysfunction of the placental endothelial nitric oxide synthase in preeclampsia. *Redox Biology*. 2021; 40: 101861.
- [69] Murray EJ, Gumusoglu SB, Santillan DA, Santillan MK. Manipulating CD4+ T Cell Pathways to Prevent Preeclampsia. *Frontiers in Bioengineering and Biotechnology*. 2021; 9: 811417.
- [70] Cherian D, Peter T, Narayanan A, Madhavan S, Achammada S, Vynat G. Malondialdehyde as a marker of oxidative stress in periodontitis patients. *Journal of Pharmacy and Bioallied Sciences*. 2019; 11: S297–S300.
- [71] Asiltas B, Surmen-Gur E, Uncu G. Prediction of first-trimester preeclampsia: Relevance of the oxidative stress marker MDA in a combination model with PP-13, PAPP-a and beta-HCG. *Pathophysiology*. 2018; 25: 131–135.
- [72] Mazloomi S, Alimohammadi S, Khodadadi I, Ghiasvand T, Shafiee G. Evaluation of methylenetetrahydrofolate reductase (MTHFR) activity and the levels of homocysteine and malondialdehyde (MDA) in the serum of women with preeclampsia. *Clinical and Experimental Hypertension*. 2020; 42: 590–594.
- [73] Rahman RA, Murthi P, Singh H, Gurungsinghe S, Leaw B, Mockler JC, *et al.* Hydroxychloroquine mitigates the production of 8-isoprostane and improves vascular dysfunction: implications for treating preeclampsia. *International Journal of Molecular Sciences*. 2020; 21: 2504.
- [74] Kawasaki K, Kondoh E, Chigusa Y, Kawamura Y, Mogami H, Takeda S, *et al.* Metabolomic Profiles of Placenta in Preeclampsia: Antioxidant Effect of Magnesium Sulfate on Trophoblasts in Early-Onset Preeclampsia. *Hypertension*. 2019; 73: 671–679.
- [75] Li F, Han N, Wang Y, Xu Q. Gadd45a knockdown alleviates oxidative stress through suppressing the p38 MAPK signaling pathway in the pathogenesis of preeclampsia. *Placenta*. 2018; 65: 20–28.
- [76] Marín R, Chiarello DI, Abad C, Rojas D, Toledo F, Sobrevia L. Oxidative stress and mitochondrial dysfunction in early-onset and late-onset preeclampsia. *Biochimica Et Biophysica Acta (BBA) - Molecular Basis of Disease*. 2020; 1866: 165961.
- [77] Vaka R, Deer E, LaMarca B. Is Mitochondrial Oxidative Stress a Viable Therapeutic Target in Preeclampsia? *Antioxidants*. 2022; 11: 210.
- [78] Hu X-Q, Zhang L. Hypoxia and mitochondrial dysfunction in pregnancy complications. *Antioxidants*. 2021; 10: 405.
- [79] Liberis A, Stanulov G, Ali EC, Hassan A, Pagalos A, Kontomanolis EN. Pre-eclampsia and the vascular endothelial growth factor: a new aspect. *Clinical and Experimental Obstetrics & Gynecology*. 2016; 43: 9–13.
- [80] Peng M, Yang M, Ding Y, Yu L, Deng Y, Lai W, *et al.* Mechanism of endogenous digitalis-like factor-induced vascular endothelial cell damage in patients with severe preeclampsia. *International*



Journal of Molecular Medicine. 2018; 41: 985–994.

- [81] Gong W, Wan J, Yuan Q, Man Q, Zhang X. Ferulic acid alleviates symptoms of preeclampsia in rats by upregulating vascular endothelial growth factor. *Clinical and Experimental Pharmacology and Physiology*. 2017; 44: 1026–1031.
- [82] Miller EC. Preeclampsia and Cerebrovascular Disease: the maternal brain at risk. *Hypertension*. 2019; 74: 5–13.
- [83] Sahay AS, Jadhav AT, Sundrani DP, Wagh GN, Mehendale SS, Chavan-Gautam P, *et al.* VEGF and VEGFR1 levels in different regions of the normal and preeclampsia placentae. *Molecular and Cellular Biochemistry*. 2018; 438: 141–152.
- [84] Nikuei P, Rajaei M, Roozbeh N, Mohseni F, Poordarvishi F, Azad M, *et al.* Diagnostic accuracy of sFlt1/PIGF ratio as a marker for preeclampsia. *BMC Pregnancy and Childbirth*. 2020; 20: 80.
- [85] Lecarpentier E, Zsengellér ZK, Salahuddin S, Covarrubias AE, Lo A, Haddad B, *et al.* Total Versus Free Placental Growth Factor Levels in the Pathogenesis of Preeclampsia. *Hypertension*. 2020; 76: 875–883.
- [86] Zeisler H, Llorba E, Chantraine F, Vatish M, Staff AC, Sennström M, *et al.* Predictive Value of the sFlt-1: PIGF Ratio in Women with Suspected Preeclampsia. *New England Journal of Medicine*. 2016; 374: 13–22.
- [87] Xu C, Li Y, Zhang W, Wang Q. Analysis of perinatal coagulation function in preeclampsia. *Medicine*. 2021; 100: e26482.
- [88] Liu R, Ma Q, Wen A, Tian G, Li M, Wang W, *et al.* Increased tissue factor expression and promoter hypomethylation in preeclampsia placentas in a Chinese population. *Pregnancy Hypertension*. 2017; 10: 90–95.
- [89] Tuten A, Gungor Z, Ekmekci H, Ekmekci OB, Kucur M, Yilmaz N, *et al.* Relationship between LPA SNPs and inflammatory burden in patients with preeclampsia to address future cardiovascular risk. *The Journal of Maternal-Fetal & Neonatal Medicine*. 2021; 34: 898–906.
- [90] Brien M, Larose J, Greffard K, Julien P, Bilodeau JF. Increased placental phospholipase A2 gene expression and free F2-isoprostane levels in response to oxidative stress in preeclampsia. *Placenta*. 2017; 55: 54–62.
- [91] Alpoim PN, Perucci LO, Godoi LC, Goulart COL, Dusse LMS. Oxidative stress markers and thrombomodulin plasma levels in women with early and late severe preeclampsia. *Clinica Chimica Acta*. 2018; 483: 234–238.
- [92] Lindoff C, Ingemarsson I, Martinsson G, Segelmark M, Thysell H, Åstedt B. Preeclampsia is associated with a reduced response to activated protein C. *American Journal of Obstetrics and Gynecology*. 1997; 176: 457–460.
- [93] Taleb YMA, Mohammed HF, El-Khazragy NN, Ahmed SK. Evaluation of Serum level of thrombomodulin in cases with preeclampsia. *QJM: An International Journal of Medicine*. 2021; 114: hcab115. 036.
- [94] Turner RJ. Endothelial pathology in preeclampsia. Leiden University: Netherlands. 2018.
- [95] Wang ZM, Zhu QY, Zhang JF, Wu JL, Yang R, Wang DM. Changes of platelet parameters in early severe preeclampsia. *Clinical and Experimental Obstetrics & Gynecology*. 2017; 44: 259–263.
- [96] Farah C, Michel LYM, Balligand J. Nitric oxide signalling in cardiovascular health and disease. *Nature Reviews Cardiology*. 2018; 15: 292–316.
- [97] Jiang Y, Liu J, Zhou Z, Liu K, Liu C. Diosmetin attenuates Akt signaling pathway by modulating nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB)/inducible nitric oxide synthase (iNOS) in streptozotocin (STZ)-induced diabetic nephropathy mice. *Medical Science Monitor*. 2018; 24: 7007–7014.
- [98] Liu T, Zhang L, Joo D, Sun S. NF-κB signaling in inflammation. *Signal Transduction and Targeted Therapy*. 2017; 2: 17023.
- [99] Lingappan K. NF-κB in oxidative stress. *Current Opinion in Toxicology*. 2018; 7: 81–86.
- [100] Sarkar O, Li Y, Anand-Srivastava MB. Nitric oxide attenuates overexpression of Giα proteins in vascular smooth muscle cells from SHR: Role of ROS and ROS-mediated signaling. *PLoS ONE*. 2017; 12: e0179301.
- [101] Shi J, Yang Y, Cheng A, Xu G, He F. Metabolism of vascular smooth muscle cells in vascular diseases. *American Journal of Physiology-Heart and Circulatory Physiology*. 2020; 319: H613–H631.
- [102] Zölner J, Lambden S, Nasri NM, Johnson MR, Leiper J. Inhibition of Dimethylarginine Dimethylaminohydrolase 1 Improves the Outcome of Sepsis in Pregnant Mice. *Shock*. 2020; 54: 498–506.
- [103] Hu X-Q, Song R, Zhang L. Effect of oxidative stress on the estrogen-NOS-NO-KCa channel pathway in uteroplacental dysfunction: Its implication in pregnancy complications. *Oxidative Medicine and Cellular Longevity*. 2019; 2019: 9194269.
- [104] Ishkaraeva-Yakovleva VV, Fedorova OV, Solodovnikova NG, Frolova EV, Bzhelyansky AM, Emelyanov IV, *et al.* DigiFab Interacts with Endogenous Cardiotonic Steroids and Reverses Preeclampsia-Induced Na/K-ATPase Inhibition. *Reproductive Sciences*. 2012; 19: 1260–1267.
- [105] Erez O, Romero R, Vaisbuch E, Than NG, Kusanovic JP, Mazaki-Tovi S, *et al.* Tissue factor activity in women with preeclampsia or SGA: a potential explanation for the excessive thrombin generation in these syndromes. *The Journal of Maternal-Fetal & Neonatal Medicine*. 2018; 31: 1568–1577.
- [106] Chen XK, Wen SW, Bottomley J, Smith GN, Leader A, Walker MC. In vitro fertilization is associated with an increased risk for preeclampsia. *Hypertension in Pregnancy*. 2009; 28: 1–12.
- [107] Blazquez A, García D, Vassena R, Figueras F, Rodriguez A. Risk of preeclampsia in pregnancies resulting from double gamete donation and from oocyte donation alone. *Pregnancy Hypertension*. 2018; 13: 133–137.
- [108] Gui J, Ling Z, Hou X, Fan Y, Xie K, Shen R. In vitro fertilization is associated with the onset and progression of preeclampsia. *Placenta*. 2020; 89: 50–57.
- [109] Castiglioni MT, Valsecchi L, Cavoretto P, Pirola S, Di Piazza L, Maggio L, *et al.* The risk of preeclampsia beyond the first pregnancy among women with type 1 diabetes parity and preeclampsia in type 1 diabetes. *Pregnancy Hypertension: An International Journal of Women's Cardiovascular Health*. 2014; 4: 34–40.
- [110] Weissgerber TL, Mudd LM. Preeclampsia and Diabetes. *Current Diabetes Reports*. 2015; 15: 9.
- [111] Papageorgiou AT, Deruelle P, Gunier RB, Rauch S, García-May PK, Mhatre M, *et al.* Preeclampsia and COVID-19: results from the INTERCOVID prospective longitudinal study. *American Journal of Obstetrics and Gynecology*. 2021; 225: 289.e281–289.e17.
- [112] Jayaram A, Buhimschi IA, Aldasqi H, Hartwig J, Owens T, Elam GL, *et al.* Who said differentiating preeclampsia from COVID-19 infection was easy? *Pregnancy Hypertension*. 2021; 26: 8–10.