

Biomarkers in lung cancer

Alessandra Bearz¹, Massimiliano Berretta¹, Alessandro Cappellani², Arben Lleshi¹, Eleonora Berto, Lucia Fratino¹, Umberto Tirelli¹

¹Department of Medical Oncology, CRO-IRCCS, Aviano (PN), Italy, ²Department of Surgery, University of Catania, Italy

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Circulating markers
 - 3.1. Small cell lung cancer
 - 3.1.1. Neuron-specific enolase
 - 3.1.2. Chromogranin A (CgA)
 - 3.2. Non small cell lung cancer
 - 3.2.1. Cyfra 21-1
 - 3.2.2. Ca 15-3
 - 3.2.3. Carcinoembryonic Antigen
 - 3.2.4. Ca 19-9
 - 3.2.5. Squamous cell carcinoma
4. Tissutal markers
 - 4.1. Excision repair cross-complementing-1
 - 4.2. RRM1
 - 4.3. BRAC1
 - 4.4. Epidermal growth factor receptor
 - 4.5. k-RAS
 - 4.6. Calcitonin
5. Acknowledgments
6. References

1. ABSTRACT

Here we review the role of tissutal and circulating biomarkers in the management of lung cancer. In the past they were considerate quite ineffective tools as regards prognosis and prediction of treatment activity, nowadays instead, they are becoming a crucial key point as potential predictive issues in driving therapy, with possibly prognostic values as well.

2. INTRODUCTION

Lung cancer is the major cause of cancer-related deaths in the world (1). Two main pathological entities of lung cancer are recognized depending on cell type: small cell (SCLC) and non-small cell lung cancer (NSCLC), overall 25% and 75%, respectively (2). Lung cancer is generally diagnosed at advanced stage, and is associated with poor prognosis, with a median survival of 8-10 months (3). If for SCLC few improvement has been achieved in the last decade, for NSCLC overall survival has been significantly prolonged due to the discovery of new therapeutic agents. NSCLC is further divided in three major histological subtypes: squamous cell carcinoma, adenocarcinoma and large cell carcinoma (4).

Adenocarcinoma is a malignant epithelial tumor with glandular differentiation or mucin production, showing acinar, papillary, bronchoalveolar, or solid with mucin growth patterns or a mixture of these patterns. Common molecular characteristics of NSCLC adenocarcinoma are a higher thymidylate synthase expression than in squamous cell carcinoma (5), frequent point mutations of dominant oncogenes, such as k-RAS genes and p53 and p16Ink4, higher expression of TTF-1 (up to 75% of adenocarcinomas) (4).

Squamous cell carcinoma is a malignant epithelial tumor showing keratinization and/or intercellular bridges that arise from bronchial epithelium. These features vary with degree of differentiation, being prominent in well-differentiated tumors and focal in poorly differentiated tumors and focal in poorly differentiated tumors. Usually k-RAS mutations are rare in squamous cell carcinoma, as well as HER2/Neu expression; most squamous cell carcinomas demonstrate large 3p segments of allelic loss, whereas most adenocarcinomas have smaller chromosome areas of 3p allelic loss (4)

Large cell carcinoma is an undifferentiated NSCLC that lacks the cytologic and architectural features

Markers in lung cancer

of small cell carcinoma and glandular or squamous differentiation. K-RAS mutations, p-53 mutations and Rb pathway alterations occur with the same frequency as in other NSCLC subtypes (4).

Tumor markers have been extensively studied in lung cancer, but none is specific for this malignancy. Neuron-specific enolase (NSE) is the tumor marker of choice in SCLC (6), whereas there is no specific tumor marker for NSCLC; nevertheless most of the circulating markers in patients affected by lung cancer have no prognostic or predictive value. On the contrary, tissutal markers may give a hint about tumor genotype that together with host characteristics could influence the treatment algorithm. Summarizing, we divided markers for SCLC and NSCLC, and circulating markers and tissue markers.

3. CIRCULATING MARKERS

The role of most of serum markers for lung cancer remains undefined. Several markers, such as carcinoembryonic antigen (CEA), small cell carcinoma, as well as cytokeratins including CYFRA 21-1, tissue polypeptide antigen (TPA), and Ca 15-3 have been proposed as useful contributions to the diagnosis of lung cancer, although their role has not been demonstrated clearly (7). NSE is specifically used in SCLC only (8).

Up to now, the most accepted clinical use of tumor circulating markers in lung cancer is in the follow-up, with contradictory results in relation to their possible prognostic or predictive value.

3.1. Small cell lung cancer

SCLC usually arises as a rapidly growing tumor of the major airways and it is considered the anaplastic variety of this spectrum of neuroendocrine (NE) tumors. TTF-1 is positive in the majority of both large and small cell NE carcinomas and is therefore useful to differentiate those lesions from poorly differentiated NE carcinomas of other sites. High molecular weight cytokeratins are not expressed in SCLC, in contrast with non-NE carcinomas (9).

3.1.1. Neuron-specific enolase

SCLC yields neuroendocrine properties that are considered to be part of its aggressive clinical behavior (10). However, the serum NSE level is not absolutely specific to the NE differentiation. Clinical experience has shown low levels of NSE in limited SCLC and high levels in NSCLC, mostly large cell, therefore there is concern about the sensitivity of this marker (11). Its role is for follow-up of the disease during treatment.

3.1.2. Chromogranin A

Chromogranin A (CgA) is a 49 KD acidic soluble protein, initially recognized in the core of the adrenal medullary catecholamine storage vesicles (12). This protein is ubiquitously present in neuroendocrine cells and often found in the serum of patients affected by SCLC (13). Its role is in the follow-up of the disease during treatment and for prediction of recurrence.

3.2. Non small cell lung cancer

3.2.1. Cyfra 21-1

Cyfra 21-1 is a fragment of cytokeratin subunit 19 and can be measured in serum by an immunoradiometric assay. The prognostic information provided by serum Cyfra 21-1 seems to be higher than the one produced by the cytokeratin 18 marker, or tissue polypeptide specific antigen (TPS) (14).

Other authors have demonstrated that CEA and Cyfra 21-1 are the most sensitive tumor markers in NSCLC (15-17). According to several authors the serum Cyfra 21-1 distribution is highest in squamous cell carcinoma, metastatic stage and poor performance status (18); however not all the authors agree with correlation between histology and serum levels of Cyfra 21-1 (19).

3.2.2. Ca 15-3

Episialin or MUC-1 is a transmembrane glycoprotein expressed at the apical side of normal glandular cells (20). It is expressed by most "wet" epithelia such as bladder, breast, stomach, pancreas, ovary and respiratory tract. In a tumor, the cell polarization is lost and the normal tissue architecture is disrupted by the growing neoplastic tissue, allowing MUC-1 to be shed into the circulation where it can be measured by immunoassays (21). MUC-1 mucin as detected by a Ca 15-3 sandwich capture assay had been the first marker to correlate with treatment response in breast cancer (22-25). As a matter of fact, Ca 15-3 is elevated in 54-80% of breast cancer patients, but it may rise even in other malignancies, e.g., lung, ovarian, endometrial, bladder and gastrointestinal carcinomas. *In vitro*, MUC-1 inhibits the E-cadherin mediated cell-cell adhesion system to extra cellular matrix (ECM), which stimulates the metastatic process (26). It has been suggested that MUC-1 and epidermal growth factor receptor (EGFR) expression are likely to be activated within different pathogenic pathways. In lung cancer, MUC-1 is highly correlated with adenocarcinoma, poor prognosis and tumor spreading (27, 28).

So far, the incidence and role of pathological Ca 15-3 serum levels in patients with lung cancer, especially adenocarcinoma, are still unknown.

We already suggested the possible correlation between low levels of Ca 15-3 and response to EGFR tyrosine kinase inhibitors (TKI) in patients affected by adenocarcinoma of the lung with bronchioloalveolar carcinoma features (29), nevertheless the possibly predictive role of Ca 15-3 needs further confirmation.

3.2.3. Carcinoembryonic Antigen

CEA is mainly found in adenocarcinomas, but it is unable to distinguish between NSCLC and SCLC (15, 16).

It has been proposed that, in case of SCLC with normal serum NSE levels, CEA and squamous cell carcinoma (SCC) should be normal as well (19). It is unclear if CEA could have a prognostic value, in particular for recurrence risk (30, 31).

Markers in lung cancer

3.2.4. Ca 19.9

The sensitivity of the mucins and among them, Ca 19.9., is lower than that of other tumor markers in NSCLC, however their highest concentration is found in adenocarcinomas. Some authors have reported the prognostic utility of Ca 19.9. in this disease, but those observations need confirmation (32).

3.2.5. Squamous cell carcinoma

SCC is a tumor marker with low specificity for lung cancer, but high relationship with NSCLC, mainly squamous cell histotype. It is used for monitoring NSCLC, although some reports discourage its routine use because of low sensitivity (33).

4. TISSUTAL MARKERS

Tissutal biomarkers are important for their potential use in customizing a therapy for NSCLC patients together other host factors.

The host factors include age, race, weight loss, performance status, gender, comorbidities, social factors; they all together concur to a personalized-driven therapy. The tumor characteristics are potentially as important as host factors; in particular genes and proteins with activity in peculiar biochemical pathways may be important in the identification of different risks for survival and other clinical relevant outcomes.

Up to now, their potential role involves predominantly NSCLC; for SCLC no putative biomarker has been recognized with predictive or prognostic value.

4.1. Excision repair cross-complementing-1

The nucleotide excision repair cross-complementing-1 (ERCC1) gene plays a key role in DNA repair after cisplatin damage. It is responsible for the 5' incision required for the removal of DNA adducts that are the basis for platinum cytotoxicity (34). The balance of DNA damage to DNA repair dictates tumor cell death or survival after cisplatin therapy. ERCC1 mRNA levels are prognostic after surgical resection of NSCLC (35). ERCC1 mRNA levels are predictive of improved response and survival from cisplatin-based therapy (36).

Immunohistochemical positivity for ERCC1 is predictive of failure for adjuvant therapy platin-based in NSCLC according to analysis of 761 tissue samples of patients recruited in the International Adjuvant Lung Cancer trial (37).

It is still controversial if analysis of ERCC1 should be through immunohistochemistry or mRNA levels, or whether both the systems are equally significative and reliable.

4.2. RRM1

RRM1 is the molecular target of gemcitabine and a component of ribonucleotide reductase, which is required for deoxynucleotide production (38). RRM1 protein levels in tumor specimens are predictive of disease response in patients with advanced NSCLC and treated with gemcitabine (39).

4.3. BRAC1

A growing body of evidence suggests that the BRAC1 confers sensitivity to apoptosis induced by antimicrotubule drugs (paclitaxel and vincristine) but induces resistance to DNA-damaging agents (cisplatin and etoposide) (40-42). Gene expression signatures have been reported to predict survival outcome in resected stage I (43). In particular, Rosell *et al* reported that BRCA1 mRNA expression is closely related to ERCC1 and RRM1, but it is the most significant prognostic marker of relapse in patients affected by NSCLC stage I and resected (44).

4.4. Epidermal growth factor receptor

The EGFR family of genes encodes transmembrane molecules that have been implicated in the development and progression of cancer (45, 46). After ligand binding, the transmembrane receptor forms homodimers or heterodimers, internalizes, and autophosphorylates tyrosine residues in its cytoplasmic domain, thereby triggering a cascade that leads to cellular proliferation, angiogenesis, metastasis, and inhibition of apoptosis. The EGFR gene is frequently expressed in solid tumors, and NSCLC as well- Non-small-cell lung cancer frequently expresses EGFR (47). At present, somatic activating mutations in EGFR tyrosin kinase (TK) domains (exons 18-21) represent the major molecular determinant of the clinical response to treatment with TKI, Gefitinib or Erlotinib (48, 49). Increasing clinical evidence suggests that patients with EGFR mutations may receive more benefit from EGFR TKI therapies than chemotherapy (50, 51); on the other hand, KRAS mutations, which have been found to be mutually exclusive from EGFR ones, are associated with lack of response (52). Recently, mutations in the kinase domain of ERBB2 have been detected in a small fraction (2-4%) of NSCLC (53). The presence of such mutation has been implicated in TKI sensitivity (53, 54). NSCLC tumors that over express both EGFR and HER2 are more sensitive to EGFR TKI than are tumors that over express EGFR but are HER2 negative (55).

4.5. k-RAS

Somatic mutations of the k-RAS oncogene have been assessed as a mechanism of de-novo resistance to EGFR TKI in patients with NSCLC, and to anti-EGFR monoclonal antibodies in patients with metastatic colorectal cancer (mCRC). A systematically review of 252 manuscripts about expression of k-RAS in NSCLC showed that k-RAS mutations are highly specific negative predictors of response to TKIs in advanced NSCLC; and similarly to anti-EGFR monoclonal antibodies alone or in combination with chemotherapy in patients with mCRC (52).

The authors in the meta analysis suggested that the low sensitivity of k-RAS mutations for determining non-responsiveness to TKI clearly shows that additional mechanisms of resistance to EGFR inhibitors exist.

5. ACKNOWLEDGEMENTS

The authors thank Mrs Paola Favetta for her skilled and precious help in revising the manuscript.

6. REFERENCES

1. F. Kamangar, G.M. Dores and W.F. Anderson: Patterns of cancer incidence, mortality, and prevalence across five continents: defining priorities to reduce cancer disparities in different geographic regions of the world. *J Clin Oncol* 24, 2137-2150 (2006)
2. G. D'Addario and E.E. Felip, ESMO Guidelines Working Group: Non-small-cell lung cancer: ESMO clinical recommendations for diagnosis, treatment and follow-up. *Ann Oncol* 20 (Suppl 4), 68-70 (2009)
3. D.M. Parkin, F. Bray, J. Ferlay and P. Pisani: Global cancer statistics, 2002. *CA Cancer J Clin* 55, 74-108 (2005)
4. Pathology and genetics of tumors of the lung, pleura, thymus and heart. IARC Press 2004
5. P. Ceppi, M. Volante, S. Saviozzi, I. Rapa, S. Novello, A. Cambieri, M. Lo Iacono, S. Cappia, M. Papotti and G.V. Scagliotti: Squamous cell carcinoma of the lung compared with other histotypes shows higher messenger RNA and protein levels for thymidylate synthase. *Cancer* 107, 1589-1596 (2006)
6. L.G. Jørgensen, K. Osterlind, J. Genollá, S.A. Gomm, J.R. Hernández, P.W. Johnson, J. Løber, T.A. Splinter and M. Szturmowicz: Serum neuron-specific enolase (S-NSE) and the prognosis in small cell lung cancer (SCLC): a combined multivariate analysis on data from nine centers. *Br J Cancer* 74, 463-467 (1996)
7. V. Macchia, A. Mariano, M. Cavalcanti, A. Coppa, C. Cecere, G. Fraioli, S. Elia and G. Ferrante: Tumor markers and lung cancer: correlation between serum and bronchial secretion levels of CEA, TPA, Can Ag Ca-50, NSE and ferritin. *Int J Biol Markers* 2, 151-156 (1987)
8. J.L. Pujol, X. Quantin, M. Jacot, J.M. Boher, J. Grenier and P.J. Lamy: Neuroendocrine and cytokeratin serum markers as prognostic determinants of small cell lung cancer. *Lung Cancer* 39, 131-138 (2003)
9. L. Viberti, M. Bongiovanni, S. Croce and G. Bussolati: 34bE12 Cytokeratin immunodetection in the differential diagnosis of small cell tumors of lung. *Int J Surg Pathol* 8, 317-322 (2000)
10. D.N. Carney, P.J. Marangos, D.C. Ihde, P.A. Bunn Jr, M.H. Cohen, J.D. Minna and A.F. Gazdar: Serum neuron-specific enolase a marker for disease extent and response to therapy for small-cell lung cancer. *Lancet* 1, 583-585 (1982)
11. T. Shibayama, T. Ohnoshi, H. Ueoka, T. Horiguchi, T. Kodani, Y. Segawa, T. Maeda, K. Miyatake, N. Takigawa and I. Kimura: Serum neuron specific enolase (NSE) in patients with non small cell lung cancer. *Nihon Kyobu Shikkan Gakkai Zasshi* 30, 1097-1102 (1992)
12. F.R. Nobels, D.J. Kwekkeboom, W. Coopmans, C.H. Schoenmakers, J. Lindemans, W.W. De Herder, E.P. Krenning, R. Bouillon and S.W. Lamberts: Chromogranin A as serum marker for neuroendocrine neoplasia: comparison with neuron-specific enolase and the alpha-subunit of glycoprotein hormones. *J Clin Endocrinol Metab* 82, 2622-2628 (1997)
13. L. Drivsholm, L.I. Paloheimo and K. Osterlind: Chromogranin A, a significant prognostic factor in small cell lung cancer. *Br J Cancer* 81, 667-671 (1999)
14. J.L. Pujol, J. Grenier, E. Parrat, M. Lehmann, T. Lafontaine, X. Quantin and F.B. Michel: Cytokeratins as serum markers in lung cancer: a comparison of Cyfra 21-1 and TPS. *Am J Resp Crit Care Med* 154, 725-733 (1996)
15. European Group on Tumor Markers: Consensus recommendations. *Anticancer Res* 19, 2785-2820 (1999)
16. J. Schneider, N. Bitterlich, H.G. Velcovsky, H. Morr, N. Katz and E. Eigenbrodt: Fuzzy logic-based tumor marker profiles improved sensitivity in the diagnosis of lung cancer. *Int J Clin Oncol* 7, 145-151 (2002)
17. R. Molina, X. Filella, J.M. Augé, R. Fuentes, I. Bover, J. Rifa, V. Moreno, E. Canals, N. Vinals, A. Marquez, E. Barreiro, J. Borrás and P. Viladiu. Tumor markers (CEA, Ca 125, Cyfra 21-1, SCC and NSE) in NSCLC patients as aid in histological diagnosis and prognostic: comparison with the main clinical, pathological and anthropomorphic prognostic factors. *Tumor Biol* 24, 209-218 (2003)
18. J.L. Pujol, J.M. Boher, J. Grenier and X. Quantin: Cyfra 21-1, neuron specific enolase and prognosis of non-small cell lung cancer: prospective study in 621 patients. *Lung Cancer* 31, 221-231 (2001)
19. R. Molina, J.M. Auge, J.M. Escudero, R. Marrades, N. Vinolas, E. Carcereny, J. Ramirez, X. Filella: Mucins Ca 125, Ca 19.9., Ca 15-3 and TAG 72.3. as tumor markers in patients with lung cancer: comparison with Cyfra 21.1., CEA, Scc and NSE. *Tumor Biol* 29, 371-380 (2008)
20. A. López-Ferrer, V. Curull, C. Barranco, M. Garrido, J. Lloreta, F.X. Real and C. de Bolós: Mucins as differentiation markers in bronchial epithelium. Squamous cell carcinoma and adenocarcinoma display similar expression patterns. *Am J Respir Cell Mol Biol* 24, 22-29 (2001)
21. D.F. Hayes, V.R. Zurawski and D.W. Kufe: Comparison of circulating Ca 15-3 and carcinoembryonic antigen levels in patients with breast cancer. *J Clin Oncol* 10, 1542-1550 (1986)
22. D. Hayes, H. Sekine, T. Ohao, M. Abe, K. Keefe and D.W. Kufe: Use of murine monoclonal antibody for detection of circulating plasma DF3 antigen levels in breast cancer patients. *J Clin Invest* 75, 1671-1678 (1985)

Markers in lung cancer

23. D.F. Hayes, V.R. Zurawski and D.W. Kufe: Comparison of circulating Ca15-3 and carcinoembryonic antigen levels in patients with breast cancer. *J Clin Oncol* 10, 1542-1550 (1986)
24. D.M.F. Pons-Anicet, B.P. Krebs and M. Namer: Value of Ca15-3 in the follow-up of breast cancer patients. *Br J Surg* 55, 567-569 (1987)
25. C. Todini, D.F. Hayes, R. Gelman, I.C. Henderson and D. Kufe: Comparison of Ca15-3 and carcinoembryonic antigen in monitoring of the clinical course of patients with metastatic breast cancer. *Cancer Res* 48, 41107-41112 (1988)
26. J. Wesseling, S.W. van der Valk and J. Hilkens: A mechanism for inhibition of E-cadherin-mediated cell-cell adhesion by the membrane-associated mucin episialin/MUC1. *Mol Biol Cell* 7, 565-577 (1996)
27. F. Guddo, A. Giatromanolaki, M.I. Koukourakis, C. Reina, A.M. Vignola, G. Chlouverakis, J. Hilkens, K.C. Gatter, A.L. Harris and G. Bonsignore: MUC-1 (episialin) expression in non-small cell lung cancer is independent of EGFR and c-erbB-2 expression and correlates with poor survival in node positive patients. *J Clin Pathol* 51, 667-671 (1998)
28. J.A. Jarrard, R.I. Linnoila, H. Lee, S.M. Steinberg, H. Witschki and E. Szabo: MUC-1 is a novel marker for the type II pneumocyte lineage during lung carcinogenesis. *Cancer Res* 58, 5582-5589 (1998)
29. A. Bearz, R. Talamini, E. Vaccher, M. Spina, C. Simonelli, A. Steffan, M. Berretta, E. Chimienti, U. Tirelli: MUC-1 (CA 15-3 antigen) as a highly reliable predictor of response to EGFR inhibitors in patients with bronchioloalveolar carcinoma: an experience on 26 patients. *Int J Biol Markers* 22, 307-311 (2007)
30. T. Muley, T.H. Fetz, H. Dienemann, H. Hoffmann, F.J. Herth, M. Meister and W. Ebert: Tumor volume and tumor marker index based on CYFRA 21-1 and CEA are strong prognostic factors in operated early stage NSCLC. *Lung Cancer* 60, 408-415 (2008)
31. F. Blankenburg, R. Hatz, D. Nagel, D. Ankerst, J. Reinmiedl, C. Gruber, D. Seidel and P. Stieber: Preoperative CYFRA 21-1 and CEA as prognostic factors in patients with stage I non-small cell lung cancer: external validation of a prognostic score. *Tumour Biol* 29, 272-277 (2008)
32. J. Niklinski, M. Furman, J. Laudanski and M. Kozlowski: Prognostic value of pretreatment CEA, SCC-Ag and CA 19-9 levels in sera of patients with non-small cell lung cancer. *Eur J Cancer Prev* 1, 1401-1406 (1992)
33. K. Kagohashi, H. Satoh, H. Ishikawa, M. Ohtsuka and K. Sekizawa: A re-evaluation of squamous cell carcinoma antigen (SCC) as a serum marker for non-small cell lung cancer. *Med Oncol* 25, 187-189 (2008)
34. M. Volker, M.J. Mone, P. Karmakar, A. van Hoffen, W. Schul, W. Vermeulen, J.H. Hoeijmakers, R. van Driel, A.A. van Zeeland and L.H. Mullenders: Sequential assembly of the nucleotide excision repair factors *in vivo*. *Molec Cell* 8, 213-224 (2001)
35. G.R. Simon, S. Sharma, A. Cantor, P. Smith and G. Bepler: ERCC1 expression is a predictor of survival in resected patients with non-small cell lung cancer. *Chest* 127, 978-983 (2005)
36. R.V. Lord, J. Brabender, D. Gandara, V. Alberola, C. Camps, M. Domine, F. Cardenal, J.M. Sánchez, P.H. Gumerlock, M. Tarón, J.J. Sánchez, K.D. Danenberg, P.V. Danenberg and R. Rosell: Low ERCC1 expression correlates with prolonged survival after cisplatin plus gemcitabine chemotherapy in non-small cell lung cancer. *Clin Cancer Res* 8, 2286-2291 (2002)
37. K. Olaussen, A. Dunant, P. Fouret, E. Brambilla, F. André, V. Haddad, E. Taranchon, M. Filipits, R. Pirker, H.H. Popper, R. Stahel, L. Sabatier, J.P. Pignon, T. Tursz, T. Le Chevalier and J.C. Soria; IALT Bio Investigators: DNA repair by ERCC1 in non-small-cell lung cancer and cisplatin-based adjuvant chemotherapy. *Engl J Med* 355, 983-991 (2006)
38. J. Stubbe: Ribonucleotide reductase in the twenty-first century: *Proc Natl Acad Sci USA* 95, 2723-2724 (1998)
39. C. Reynolds, C. Obasaju, M.J. Schell, X. Li, Z. Zheng, D. Boulware, J.R. Caton, L.C. Demarco, M.A. O'Rourke, G. Shaw Wright, K.A. Boehm, L. Asmar, J. Bromund, G. Peng, M.J. Monberg and G. Bepler: Randomized phase III trial of gemcitabine-based chemotherapy with *in situ* RRM1 and ERCC1 protein levels for response prediction in non-small-cell lung cancer. *J Clin Oncol* 27, 5808-5815 (2009)
40. S. Lafarge, V. Sylvain, M. Ferrara and Y.J. Bignon: Inhibition of BRCA1 leads to increased chemoresistance to microtubule-interfering agents, an effect that involves the JNK pathway. *Oncogene* 20, 6597-6606 (2001)
41. A. Husain, G. He, E.S. Venkatraman and D.R. Spriggs: BRCA1 up-regulation is associated with repair-mediated resistance to cis-diamminedichloroplatinum (II). *Cancer Res* 58, 1120-1123 (1998)
42. A. Bhattacharyya, U.S. Ear, B.H. Koller, R.R. Weichselbaum and D.K. Bishop: The breast cancer susceptibility gene BRCA1 is required for subnuclear assembly of Rad51 and survival following treatment with the DNA cross-linking agent cisplatin. *J Biol Chem* 275, 23899-23903 (2000)
43. A. Potti, S. Mukherjee, R. Peterson, H.K. Dressman and A. Bild: A genomic strategy to refine prognosis in early stage non-small cell lung cancer. *N Engl J Med* 355, 570-580 (2006)
44. R. Rosell, M. Skrzypski, E. Jassem, M. Taron, R. Bartolucci, J.J. Sanchez, P. Mendez, I. Chaib, L. Perez-Roca, A. Szymanowska, W. Rzyman, F. Puma, G. Kobińska-Gulida, F. Farabi and J. Jassem: BRCA1: a

Markers in lung cancer

novel prognostic factor in resected non-small cell lung cancer. *PLoS One* 2, e1129 (2007)

45. J. Schlessinger: Ligand-induced, receptor-mediated dimerization and activation of EGF receptor. *Cell* 110, 669-672 (2002)

46. C.L. Arteaga: Overview of epidermal growth factor receptor biology and its role as a therapeutic target in human neoplasia. *Semin Oncol* 29 (Suppl 14), 3-9 (2002)

47. R.S. Herbst and P.A. Bunn Jr: Targeting the epidermal growth factor receptor in non-small cell lung cancer. *Clin Cancer Res* 9, 5813-5824 (2003)

48. T.J. Lynch, D.W. Bell, R. Sordella, S. Gurubhagavatula, R.A. Okimoto, B.W. Brannigan, P.L. Harris, S.M. Hasserlat, J.G. Supko, F.G. Haluska, D.N. Louis, D.C. Christiani, J. Settleman and D.A. Haber: Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* 350, 2129-2139 (2004)

49. J.G. Paez, P.A. Jänne, J.C. Lee, S. Tracy, H. Greulich, S. Gabriel, P. Herman, F.J. Kaye, N. Lindeman, T.J. Boggon, K. Naoki, H. Sasaki, Y. Fujii, M.J. Eck, W.R. Sellers, B.E. Johnson and M. Meyerson: EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* 304, 1497-1500 (2004)

50. T.S. Mok, Y.L. Wu, C.J. Yu, C. Zhou, Y.M. Chen, L. Zhang, J. Ignacio, M. Liao, V. Srimuninnimit, M.J. Boyer, M. Chua-Tan, V. Sriuranpong, A.W. Sudoyo, K. Jin, M. Johnston, W. Chui and J.S. Lee: Randomized, placebo-controlled, phase II study of sequential erlotinib and chemotherapy as first-line treatment for advanced non-small-cell lung cancer. *J Clin Oncol* 27, 5080-5087 (2009)

51. M.S. Tsao, A. Sakurada, J.C. Cutz, C.Q. Zhu, S. Kamel-Reid, J. Squire, I. Lorimer, T. Zhang, N. Liu, M. Daneshmand, P. Marrano, G. da Cunha Santos, A. Lagarde, F. Richardson, L. Seymour, M. Whitehead, K. Ding, J. Pater and F.A. Shepherd: Erlotinib in lung cancer - molecular and clinical predictors of outcome. *N Engl J Med* 353, 133-144 (2005)

52. H. Linardou, I.J. Dahabreh, D. Kanakoupiti, F. Siannis, D. Bafaloukos, P. Kosmidis, C.A. Papadimitriou and S. Murray: Assessment of somatic k-RAS mutations as a mechanism associated with resistance to EGFR-targeted agents: a systematic review and meta-analysis of studies in advanced non-small-cell lung cancer and metastatic colorectal cancer. *Lancet Oncol* 9, 962-972 (2008)

53. P. Stephens, C. Hunter, G. Bignell, S. Edkins, H. Davies, J. Teague, C. Stevens, S. O'Meara, R. Smith, A. Parker, A. Barthorpe, M. Blow, L. Brackenbury, A. Butler, O. Clarke, J. Cole, E. Dicks, A. Dike, A. Drozd, K. Edwards, S. Forbes, R. Foster, K. Gray, C. Greenman, K. Halliday, K. Hills, V. Kosmidou, R. Lugg, A. Menzies, J. Perry, R. Petty, K. Raine, L. Ratford, R. Shepherd, A. Small, Y. Stephens, C. Tofts, J. Varian, S. West, S. Widaa,

A. Yates, F. Brasseur, C.S. Cooper, A.M. Flanagan, M. Knowles, S.Y. Leung, D.N. Louis, L.H. Looijenga, B. Malkowicz, M.A. Pierotti, B. Teh, G. Chenevix-Trench, B.L. Weber, S.T. Yuen, G. Harris, P. Goldstraw, A.G. Nicholson, P.A. Futreal, R. Wooster and M.R. Stratton: Lung cancer: intragenic ERBB2 kinase mutations in tumours. *Nature* 431, 525-526 (2004)

54. E.E. Cohen, M.W. Lingen, L.E. Martin, P.L. Harris, B.W. Brannigan, S.M. Hasserlat, R.A. Okimoto, D.C. Sgroi, S. Dahiya, B. Muir, J.R. Clark, J.W. Rocco, E.E. Vokes, D.A. Haber and D.W. Bell: Response of some head and neck cancers to epidermal growth factor receptor tyrosine kinase inhibitors may be linked to mutation of ERBB2 rather than EGFR. *Clin Cancer Res* 11, 8105-8108 (2005)

55. F. Hirsch, M. Varella-Garcia and F. Cappuzzo: Predictive value of EGFR and HER2 overexpression in advanced non-small-cell lung cancer. *Oncogene* 28 (Suppl 1), S32-37 (2009)

Abbreviations: SCLC: small cell lung cancer, NSCLC: non small cell lung cancer, NSE: neuron specific enolase, CEA: carcinoembryonic antigen, TPA: tissue polypeptide antigen, NE: neuroendocrine, CgA: Chromogranin A, TPS: tissue polypeptide specific, ECM: extra cellular matrix, EGFR: epidermal growth factor receptor, TKI: tyrosine kinase inhibitor, SCC: squamous cell carcinoma, ERCC1: excision repair cross-complementing-1, BRAC1: breast cancer susceptibility gene 1, TK: tyrosine kinase, m-CRC: metastatic colorectal cancer.

Key Words: Markers, Lung, Cancer, Tumor, Review

Send correspondence to: Alessandra Bearz, Department of Medical Oncology, National Cancer Institute, via Franco Gallini 2, 33081 Aviano (PN), Italy, Tel: 39-0434-659294, Fax: 39-0434-659531, E-mail: abearz@cro.it

<http://www.bioscience.org/current/vol3E.htm>