

Obesity and Breast Cancer: mechanisms and therapeutic implications

Kristy A. Brown^{1,2}, Evan R. Simpson^{1,3}

¹*Metabolism and Cancer Laboratory, Prince Henry's Institute, Clayton, Australia, 3168*, ²*Department of Physiology, Monash University, Clayton, Australia, 3168*, ³*Department of Biochemistry and Molecular Biology, Monash University, Clayton, Australia, 3168*

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Pre- versus post-menopausal breast cancer
4. Common features and risk factors
 - 4.1. Dysregulated metabolism
 - 4.2. Inflammation
 - 4.3. Insulin and IGF-I
 - 4.4. Estrogens
5. Perspectives and emerging therapies
6. Acknowledgements
7. References

1. ABSTRACT

Obesity is a known risk factor for postmenopausal breast cancer, whereby factors produced by the adipose tissue are known to directly and indirectly affect tumour growth. It is now becoming increasingly clear that both obesity and cancer arise as a consequence of dysregulated metabolism, both in response to altered energy status and endocrine factors, and to intrinsic changes within cells. Moreover, both obesity and cancer lead to localised inflammation, whereby inflammatory cytokines and prostaglandins are produced by adipose tissue and tumour cells. Obesity is also a risk factor for type 2 diabetes, with a consequential increase in circulating insulin and insulin-like growth factor-I (IGF-I) known to promote tumour cell growth. Interestingly, these factors converge to increase aromatase expression within the breast and hence, estrogen production, thereby increasing the risk of breast cancer and the growth of breast tumour cells. Therapies aimed at treating obesity/diabetes are therefore attractive options for the treatment of postmenopausal breast cancer.

2. INTRODUCTION

Obesity is a risk factor for breast cancer. Factors produced by the adipose tissue are known to directly and indirectly affect tumour cell growth. Moreover, both obesity and breast cancer are accompanied by a number of common features, including being a consequence of dysregulated metabolism, resulting in inflammation, causing and responding to changes in adipokines, insulin, IGF-1 and estrogen signalling. This review will examine how these common features and risk factors develop in and affect function of cells within the adipose and breast tumour epithelium. We will not go in depth into the role of the adipokines leptin and adiponectin in breast cancer as it has recently been reviewed in this journal (1). Suffice it to say that recent work has shown that higher leptin levels such as those found in obesity are significantly associated with an increase in breast cancer risk. On the other hand, with increased obesity, adiponectin serum levels decrease and these levels are inversely correlated with breast cancer risk. We will, however, discuss how these adipokines have been shown to affect estrogen production within the breast,

a topic mentioned only briefly in the previous review. The present review also aims to provide the reader with an insight into how these common features and risk factors may impact future therapy development.

3. PRE- VERSUS POST-MENOPAUSAL BREAST CANCER

In developed countries, over two thirds of newly diagnosed cancers are in individuals aged over 55 (2), and obesity is a recognised risk factor for breast cancer in postmenopausal women (reviewed in 3, 4). In particular, a body mass index (BMI) of 30, categorised as obese, carries a two-fold increased risk of developing breast cancer, and the risk increases exponentially with increasing BMI (4-7). Changes in the hormonal milieu at the menopause are associated with an increase in total adiposity (8, 9) and increased breast cancer risk (10). Moreover, the majority of obesity-related postmenopausal breast cancers are hormone receptor positive (reviewed in 11, 12, 13). As estrogens are no longer produced in postmenopausal ovaries, this suggests that locally produced estrogens and/or other endocrine factors are affecting tumour cell proliferation within the breast (14, 15). Conversely, data for premenopausal women has been conflicting. Although it was once believed that obesity was associated with a decreased risk of breast cancer in younger women, a number of studies now indicate that a BMI > 30 increases the risk of hormone receptor negative tumours in women prior to menopause (16-18), indicating that these tumour cells are not dependent on estrogens for proliferation but rather, may rely more heavily on insulin, IGF-I and adipose-derived factors for growth. Interestingly, gaining weight prior to menopause, particularly from 30 years of age to menopause, has been associated with an increased risk of developing breast cancer postmenopausally (19).

4. COMMON FEATURES AND RISK FACTORS

4.1. Dysregulated metabolism

Adipocytes within white adipose depots are responsible for storing energy in the form of triglycerides in times of high energy supply and mobilising these lipids in times of high energy demand. The storage of lipids in adipocytes helps to prevent excess circulating free fatty acids (FFA) being stored ectopically as triglycerides in tissues such as the pancreas, liver, muscle and kidneys which would lead to various pathologies, including hypertension, dyslipidemia, glucose intolerance and insulin resistance (20). Catecholamines, which increase intracellular cAMP, cause an increase in lipolysis, whereas insulin inhibits it (21). Obesity results when there is an imbalance between energy intake and energy expenditure and is accompanied by an increase in the number and size of adipocytes, leading to hyperplasia and hypertrophy, respectively (22). In obesity, leptin no longer elicits satiety signals from the central nervous system, thereby leading to increased energy uptake despite already high stores of energy in the adipocytes and further favouring triglyceride accumulation (23). Furthermore, the protein expression of lipolytic enzymes ATGL (adipocyte triglyceride lipase) and HSL (hormone sensitive lipase) are decreased in obese

patients compared to lean individuals. AMP activated protein kinase (AMPK) is a master regulator of energy homeostasis in a number of tissues including the muscle, liver, central nervous system and adipose tissue and is suppressed in many diseases, including obesity, metabolic syndrome and cancer (24). AMPK functions as an energy sensor, directly binding AMP and ATP, and responding to energy deficiencies by conformational changes allowing the upstream kinase Liver Kinase B1 (LKB1) to phosphorylate the alpha subunit at Thr172. This leads to an increase in activity of pathways of energy production (oxidation of fatty acids, for example) and inhibition of pathways of energy utilisation (e.g. fatty acid and cholesterol synthesis and gluconeogenesis). In adipocytes of lean individuals, adiponectin activates AMPK and stimulates glucose uptake and oxidation of fatty acids and leads to an increase in available energy (25). Sex hormones have also been shown to influence AMPK activity in adipocytes where estrogens activate and androgens inactivate, and the relationship between the two appears to be a crucial determinant of fatty acid accumulation (26, 27). These findings provide one mechanism whereby hormonal changes during the menopausal transition may lead to dysregulated metabolism and weight gain.

Dysregulated metabolism is also a common feature of tumour cells. Much work has been done to characterise ideas first proposed by Warburg 80 years ago whereby it was suggested that the origin of cancer depended not on genetic changes but rather on a shift in how energy was produced (28). Although it is clear that genetic changes to oncogenes and tumour suppressors, for example, play an important role in tumour development and growth, the idea that tumour cells also acquire a metabolic advantage is now largely supported. The complexities involved in a cell's ability to shift from mitochondrial respiration to aerobic glycolysis (Warburg effect) are beyond the scope of this review, but they have been beautifully reviewed in (29). Rather, we will focus on common features of obese adipocytes and breast tumour cells, centralising around fatty acid metabolism. Noteworthy is the observation that fatty acid synthase expression, important for *de novo* lipogenesis, is increased in many types of cancer, including breast (30, 31) and consistent with this observation, LKB1 and pAMPK, known to inhibit lipogenesis, are suppressed in breast cancer cells (32-34). The inhibited activity of AMPK in obesity may be involved in carcinogenesis of breast cancer. In addition to its role in energy homeostasis, AMPK can suppress cell proliferation by inhibiting protein biosynthesis via inhibition of the mTOR (mammalian target of rapamycin) pathway, and activation of p53 and cell cycle arrest. Much work has been done to characterise the role of p53 in mediating cell senescence and apoptosis, however, it also has an established role in maintaining energy homeostasis (35). Namely, in states of limited nutrient availability, p53 expression is induced, and p53 protein is stabilised and has an increased binding affinity to DNA, and as a consequence its downstream targets promote oxidative phosphorylation to increase energy production and inhibit glycolysis. Mutations in p53 are frequent in several types of cancer, including breast.

Inactivation of p53 thereby increases cell proliferation not only due to impairments in its pro-apoptotic capacity, but also as a consequence of shifting from mitochondrial respiration to aerobic glycolysis. Little is known of the role of p53 in adipocyte biology although it has been suggested to be anti-adipogenic (36).

4.2. Inflammation

It is now clear that obesity is associated with low grade chronic inflammation as a direct result of energy accumulation within the body. Excessive fat storage leads to stress reactions within adipocytes which in turn lead to the release of pro-inflammatory cytokines, including TNF-alpha and IL-6, from adipocytes themselves and immune cells within the adipose tissue (37). One mechanism whereby this occurs is via the increased secretion of leptin, a pro-inflammatory adipokine, which induces the production of chemokines, TNF-alpha and IL-6 from macrophages and monocytes (38, 39). Indeed, adipocytes are not the only cell type within the adipose tissue to initiate and respond to inflammatory signals. The stromal vascular fraction, which includes preadipocytes or adipose stromal cells (ASCs), endothelial cells, smooth muscle cells, multipotent stem cells, and immune cells, surrounds the adipocytes and actively participates in inflammatory responses within the adipose tissue. Macrophages in particular have been shown to infiltrate adipose tissue of obese individuals and directly associate with overly stressed or necrotic adipocytes, leading to histologically visible crown-like structures (40, 41). The number of these crown-like structures is directly related to the degree of inflammation within the adipose tissue, and is associated with the release of free fatty acids, pro-inflammatory cytokines, activation of NF-kB signalling and the increased expression of aromatase (42). Furthermore, the increased local production of IL-6 leads to an increase in circulating levels of the pro-inflammatory marker C-reactive protein, a strong predictor of the development of type II diabetes (T2D) (43). Moreover, TNF-alpha has been shown to decrease fatty acid oxidation via the downregulation of AMPK (44). Interestingly, the release of fatty acids from adipocytes has also been shown to stimulate the release of TNF-alpha and increase the expression of the inducible cyclooxygenase, COX-2, the rate limiting enzyme in prostaglandin synthesis (45). Prostaglandins act via their cognate G-protein coupled receptors to mediate autocrine and paracrine actions and have been shown to act on many physiological systems, including the adipose tissue (46). Namely, *in vitro* studies have demonstrated that prostaglandins stimulate adipocyte differentiation by increasing the clonal expansion phase (47); (48, 49) and *in vivo* data using knockout mice suggests that inhibiting prostaglandin E2 (PGE2) results in the disinhibition of lipolysis, and protects animals from high fat diet-induced obesity (50).

Inflammation also occurs in many tumour types and in some instances, precedes tumour formation (reviewed in 51). Many lines of evidence have been offered to support a direct and causative link between inflammation and cancer, and the increased prevalence of cancer at sites of chronic inflammation suggest that the inflammatory

nature of obesity may contribute to the predisposition of obese women to develop breast cancer. TNF-alpha and IL-6 are well characterised for their tumour-promoting effects via the activation of oncogenic transcription factors NF-kB, AP-1 and STAT3 in epithelial cells (reviewed in 52). The intrinsic pathway of inflammation, however, is activated as a consequence of the activation of oncogenes, which then leads to the secretion of chemokines, cytokines and prostaglandins from the tumour cells themselves. This process is also accompanied by the recruitment of inflammatory cells, the increased vascularisation and the remodelling of the affected tissue (reviewed in 51, 53). Interestingly, tumour cells from invasive and *in situ* breast cancer also have increased expression of COX-2 (54). Moreover, inflammatory mediators act to stimulate cell proliferation, migration and invasion, and have been shown to affect hormone production in the adjacent stroma (reviewed below).

4.3. Insulin and IGF-I

It is well accepted that obesity is associated with an increased risk of insulin resistance and diabetes. In 2010, it was estimated that 285 million people between the ages of 20 and 79 had diabetes worldwide (reviewed in 2). Type II diabetes (T2D) which accounts for 95% of cases is characterised by hyperinsulinemia, hyperglycemia and insulin resistance, and is a risk factor for several types of cancer, including breast, colon and pancreas. Insulin and insulin-like growth factor-I (IGF-I) are peptide hormones which mediate metabolic and mitogenic effects via binding to their cognate receptors, the insulin receptor (IR) and the IGF-I receptor (IGF-IR), although actions via hybrid receptors (insulin/IGF-I receptor) have also been described.

Changes in receptor abundance and the increased formation of hybrid receptors on tumour cells have been associated with decreased patient survival. Insulin has been shown to increase the proliferation of breast cancer cells, perhaps via activation of the hybrid receptors. Interestingly, the increased growth of tumour cells in T2D appears to be independent of obesity. A-ZIP/F-1 mice, which are a lipotrophic model of T2D demonstrate increased tumour incidence despite having no white fat and the MKR model of T2D, which is hyperinsulinemic, insulin-resistant and glucose-intolerant, displays increased growth of orthotopically inoculated mouse mammary carcinoma cells compared to control mice, despite not being obese (reviewed in 55). Furthermore, elevated fasting insulin levels are positively associated with breast cancer risk, independent of obesity in women. Interestingly, IGF-I is expressed at a higher level in ER-positive tumours than in ER-negative breast tumours (56).

4.4. Estrogens

The regulation of estrogen production within the breast is emerging as a central linking factor between obesity and breast cancer. Total and free plasma estrogen concentrations are elevated in postmenopausal obese women compared to their healthy weight counterparts (5, 57-59). Moreover, the majority of postmenopausal breast cancers are estrogen receptor (ER)-positive and their dependence of estrogens is emphasised by the efficacy of

current endocrine therapy. Noteworthy is the fact that the ovaries no longer produce measurable levels of estrogens after menopause, and the major source then becomes the adipose tissue. Aging is also associated with an increase in the ability of adipose tissue to intrinsically synthesise estrogens via the increased expression of aromatase, the enzyme responsible for catalysing the rate-limiting and one of the last steps in estrogen biosynthesis. This does not occur within adipocytes, rather, in the undifferentiated preadipocytes or adipose stromal cells (ASCs). Not only is the expression of aromatase under the control of adipose-derived factors, its expression is largely increased in response to tumour-derived factors, thereby creating a positive feedback loop between the tumour and the adipose stroma. Noteworthy is the observation that obese breast cancer patients have 130% higher circulating estradiol than their normal weight counterparts (58). Interestingly, adipokines, insulin and IGF-I, inflammatory factors and dysregulated metabolic pathways have all been shown to affect aromatase expression.

The structure of the gene which encodes aromatase, *CYP19A1*, is complex (reviewed in 60). It is composed of 9 coding exons, as well as a number of untranslated first exons driven by tissue-specific promoters. In normal breast adipose tissue, activity of promoter I.4 accounts for approximately half of total aromatase transcripts, with the remaining half being under the control of promoters I.3 and PII (61). The relative contribution of these promoters changes in the presence of a tumour, where promoters I.3 and PII are the main promoters utilised to drive aromatase expression (62).

Inflammatory factors, including prostaglandins and class I cytokines, produced by both the adipose and tumour cells are known to potently induce aromatase expression in human breast ASCs. TNF- α and a number of class I cytokines, including IL-6, IL-11, leukemia inhibitory factor, oncostatin-M, induces the promoter I.4-driven expression of aromatase in the presence of glucocorticoids, whereas PGE2 causes the upregulation of promoter I.3 and PII-specific transcripts (63-65).

This review has already emphasised the role of LKB1/AMPK signalling in energy homeostasis. Interestingly, LKB1/AMPK is also a potent inhibitor of aromatase expression within breast ASCs (66). The mechanism whereby this occurs relies on the fact that AMPK inhibits the expression of CREB-target genes by directly phosphorylating the CREB co-activator CRTC2, thereby preventing its nuclear translocation and ability to bind CREB (67). Tumour-derived factors such as PGE2 are dependent on CREB activity for increased activation of promoters I.3 and PII and hence, expression of aromatase (68, 69). Interestingly, both leptin and PGE2 decrease the expression of LKB1 and the phosphorylation of AMPK, and lead to the nuclear translocation of CRTC2 and its increased binding and activation of aromatase PII in human breast ASCs (66), providing one mechanism behind the observation that obese cancer patients have higher estrogens than normal weight women. Interestingly,

research performed in p53-inactivated primary cultures of mouse mammary epithelial cells demonstrated that aromatase expression and CREB phosphorylation and nuclear accumulation were increased compared to control cells (70).

The effect of insulin and IGF-I on estrogen production has also been characterised. In addition to increasing the bioavailability of estrogens via the downregulation of sex-hormone binding globulin (SHBG), insulin also acts directly on estrogen production and significantly increases aromatase activity in human ASCs in culture (71, 72). Moreover, IGF-I increases the activity of aromatase in stably transfected breast cancer cell lines (73). Interestingly, insulin and IGF-1 potentiate the actions of dexamethasone but inhibit the stimulatory effect of cAMP on aromatase activity (74). *In vivo*, SHBG also correlates negatively with insulin and IGF-I (75).

5. PERSPECTIVES AND EMERGING THERAPIES

In a world where 1.5 billion adults are overweight and of these, over 300 million women are obese, it is difficult to imagine the economic and societal impacts of aging and increased susceptibility to disease. The need to take action to manage obesity and prevent the emergence of a breast cancer epidemic is upon us. Furthermore and due to the commonalities between obesity and breast cancer, it is predictable that therapies which target obesity may also play an important role in treating and preventing breast cancer.

Weight loss has been shown to lower the level of pro-inflammatory proteins in obese women (76-79), decrease the incidence of diabetes and restore euglycemia in patients with T2D (reviewed in 2), decrease circulating leptin (80) and increase SHBG levels (81). Curiously, one study reported no change in sex hormone levels after weight loss (81) whereas an earlier study examining the effect of weight loss on sex hormones levels in women with PCOS, demonstrated a significant decrease (82). However, the association between weight loss and breast cancer risk is less clear and this is largely due to difficulties in defining the amount of weight loss and the duration necessary when designing clinical studies, and results in difficulties in interpreting conflicting studies and more so in designing more comprehensive ones. Moreover, many individuals struggle to lose weight via traditional methods focussed on healthy eating and exercise, and resorting to alternative methods of weight loss or disease control becomes a necessity. Weight loss as a result of bariatric surgery is also associated with improvement of insulin resistance, decreased inflammation, as well as beneficial modulation of adipokines and sex hormones (reviewed in 83). As a result women who have undergone this procedure have a reduced risk of developing many obesity-related cancers, including that of the breast (84).

Laboratory-based evidence suggests that adiponectin protects against obesity-related dysregulated metabolism, by increasing insulin sensitivity (85) and reducing blood glucose, triglycerides and free fatty acids

(86). Furthermore, adiponectin prevents the PGE2-mediated induction in aromatase expression in human breast ASCs (66).

Anti-inflammatory and anti-diabetic drugs are now attractive therapies to treat and prevent breast cancer. Not only do they act to reduce symptoms of metabolic syndrome, but also tend to have direct and indirect effects on tumour growth. Direct effects occur via alterations in energy production and protein synthesis pathways, and indirect effects by decreasing insulin, IGF-I, leptin, estrogens, and increasing adiponectin. Case-control and cohort studies suggest that aspirin and ibuprofen, widely used non-selective COX-1/COX-2 inhibitors, and other non-steroidal anti-inflammatory drugs (NSAIDs) decrease the incidence of breast cancer by 10-40% (reviewed in 87).

AMPK-activating drugs are also attractive therapies for obesity and breast cancer. It has been suggested that use of AMPK agonists may mimic the effect of exercise to enhance running endurance (88). The AMP analogue 5-Aminoimidazole-4-carboxamide 1- β -D-ribofuranoside (AICAR) has also been shown to decrease adiposity (89) and have anti-inflammatory properties (90) in rodents. Moreover, treatment of the breast cancer cell line MDA-MB-231 with AICAR results in the inhibition of lipogenesis, protein translation, and DNA synthesis, and as a consequence, inhibits cell proliferation (91). AICAR also inhibits the PGE2-mediated expression of aromatase in human breast ASCs (66). The biguanide metformin is the most commonly prescribed anti-diabetic drug and its use causes a decrease in circulating glucose by inhibiting gluconeogenesis in the liver via the activation of AMPK. Metformin also decreases plasma lipids and improves insulin sensitivity in patients with T2D (92). Moreover, metformin has also been shown to decrease TNF-alpha and C-reactive protein (93), which has a number of implications with regards to inflammation, insulin resistance and estrogen biosynthesis. Interestingly, recent epidemiological evidence also suggests that metformin may lower cancer risk and reduce the incidence of cancer-associated death in diabetic patients treated with the drug (94). A number of clinical trials are currently underway (95). Moreover, recent studies from our laboratory suggest that metformin would inhibit aromatase expression in a promoter-specific manner, thereby targeting adipose and tumour estrogen production and protecting other sites of estrogen action, including the brain and bone where other promoters are employed (1f and I.4, respectively) (96, 97). Consequent to these findings, we have initiated a prevention study aimed at determining the effects of metformin on estrogen production within the breast of mid-life women and are in the process of designing a neo-adjuvant study to explore the effect of metformin on aromatase expression in breast cancer.

6. ACKNOWLEDGEMENTS

This work was supported by the Victorian Government, through the Victorian Cancer Agency funding of the Victorian Breast Cancer Research Consortium, NHMRC Project Grants 494819 and 1005735, and Program

Grant 494802, and by the Victorian Government's Operational Infrastructure Support Program. K.A.B. is supported by an NHMRC Career Development Award.

7. REFERENCES

1. G. Paz-Filho, E.L. Lim, M.L. Wong and J. Licinio: Associations between adipokines and obesity-related cancer. *Front Biosci*, 16, 1634-1650 (2011)
2. E. Giovannucci, D.M. Harlan, M.C. Archer, R.M. Bergenstal, S.M. Gapstur, L.A. Habel, M. Pollak, J.G. Regensteiner and D. Yee: Diabetes and cancer: a consensus report. *CA Cancer J Clin*, 60, 207-221 (2010)
3. G. Ursin, M.P. Longnecker, R.W. Haile and S. Greenland: A meta-analysis of body mass index and risk of premenopausal breast cancer. *Epidemiology*, 6, 137-141 (1995)
4. P.A. van den Brandt, D. Spiegelman, S.S. Yaun, H.O. Adami, L. Beeson, A.R. Folsom, G. Fraser, R.A. Goldbohm, S. Graham, L. Kushi, J.R. Marshall, A.B. Miller, T. Rohan, S.A. Smith-Warner, F.E. Speizer, W.C. Willett, A. Wolk and D.J. Hunter: Pooled analysis of prospective cohort studies on height, weight, and breast cancer risk. *Am J Epidemiol*, 152, 514-527 (2000)
5. S. Rinaldi, T.J. Key, P.H. Peeters, P.H. Lahmann, A. Lukanova, L. Dossus, C. Biessy, P. Vineis, C. Sacerdote, F. Berrino, S. Panico, R. Tumino, D. Palli, G. Nagel, J. Linseisen, H. Boeing, A. Roddam, S. Bingham, K.T. Khaw, J. Chloutos, A. Trichopoulou, D. Trichopoulos, B. Tehard, F. Clavel-Chapelon, C.A. Gonzalez, N. Larranaga, A. Barricarte, J.R. Quiros, M.D. Chirlaque, C. Martinez, E. Monninkhof, D.E. Grobbee, H.B. Bueno-de-Mesquita, P. Ferrari, N. Slimani, E. Riboli and R. Kaaks: Anthropometric measures, endogenous sex steroids and breast cancer risk in postmenopausal women: a study within the EPIC cohort. *Int J Cancer*, 118, 2832-2839 (2006)
6. A.R. Carmichael: Obesity as a risk factor for development and poor prognosis of breast cancer. *Bjog*, 113, 1160-1166 (2006)
7. C. La Vecchia, E. Negri, S. Franceschi, R. Talamini, P. Bruzzi, D. Palli and A. Decarli: Body mass index and postmenopausal breast cancer: an age-specific analysis. *Br J Cancer*, 75, 441-444 (1997)
8. J.C. Lovejoy and A. Sainsbury: Sex differences in obesity and the regulation of energy homeostasis. *Obes Rev*, 10, 154-167 (2009)
9. E.T. Poehlman, M.J. Toth and A.W. Gardner: Changes in energy balance and body composition at menopause: a controlled longitudinal study. *Ann Intern Med*, 123, 673-675 (1995)
10. E.R. Simpson: Sources of estrogen and their importance. *J Steroid Biochem Mol Biol*, 86, 225-230 (2003)

11. M.D. Althuis, J.H. Fergenbaum, M. Garcia-Closas, L.A. Brinton, M.P. Madigan and M.E. Sherman: Etiology of hormone receptor-defined breast cancer: a systematic review of the literature. *Cancer Epidemiol Biomarkers Prev*, 13, 1558-1568 (2004)
12. R. Suzuki, T. Rylander-Rudqvist, W. Ye, S. Saji and A. Wolk: Body weight and postmenopausal breast cancer risk defined by estrogen and progesterone receptor status among Swedish women: A prospective cohort study. *Int J Cancer*, 119, 1683-1689 (2006)
13. X.R. Yang, J. Chang-Claude, E.L. Goode, F.J. Couch, H. Nevanlinna, R.L. Milne, M. Gaudet, M.K. Schmidt, A. Broeks, A. Cox, P.A. Fasching, R. Hein, A.B. Spurdle, F. Blows, K. Driver, D. Flesch-Janys, J. Heinz, P. Sinn, A. Vrieling, T. Heikkinen, K. Aittomaki, P. Heikkila, C. Blomqvist, J. Lissowska, B. Peplonska, S. Chanock, J. Figueroa, L. Brinton, P. Hall, K. Czene, K. Humphreys, H. Darabi, J. Liu, L.J. Van 't Veer, F.E. van Leeuwen, I.L. Andrulis, G. Glendon, J.A. Knight, A.M. Mulligan, F.P. O'Malley, N. Weerasooriya, E.M. John, M.W. Beckmann, A. Hartmann, S.B. Weibrecht, D.L. Wachter, S.M. Jud, C.R. Loehberg, L. Baglietto, D.R. English, G.G. Giles, C.A. McLean, G. Severi, D. Lambrechts, T. Vondorp, C. Weltens, R. Paridaens, A. Smeets, P. Neven, H. Wildiers, X. Wang, J.E. Olson, V. Caouere, Z. Fredericksen, M. Kosel, C. Vachon, H.E. Cramp, D. Connley, S.S. Cross, S.P. Balasubramanian, M.W. Reed, T. Dork, M. Bremer, A. Meyer, J.H. Karstens, A. Ay, T.W. Park-Simon, P. Hilleman, J.I. Arias Perez, P. Menendez Rodriguez, P. Zamora, J. Benitez, Y.D. Ko, H.P. Fischer, U. Hamann, B. Pesch, T. Bruning, C. Justenhoven, H. Brauch, D.M. Eccles, W.J. Tapper, S.M. Gerty, E.J. Sawyer, I.P. Tomlinson, A. Jones, M. Kerin, N. Miller, N. McInerney, H. Anton-Culver, A. Ziogas, C.Y. Shen, C.N. Hsiung, P.E. Wu, S.L. Yang, J.C. Yu, S.T. Chen, G.C. Hsu, C.A. Haiman, B.E. Henderson, L. Le Marchand, L.N. Kolonel, A. Lindblom, S. Margolin, A. Jakubowska, J. Lubinski, T. Huzarski, T. Byrski, B. Gorski, J. Gronwald, M.J. Hooning, A. Hollestelle, A.M. van den Ouweland, A. Jager, M. Kriege, M.M. Tilanus-Linthorst, M. Collee, S. Wang-Gohrke, K. Pytkas, A. Jukkola-Vuorinen, K. Mononen, M. Grip, P. Hirvikoski, R. Winqvist, A. Mannermaa, V.M. Kosma, J. Kauppinen, V. Kataja, P. Auvinen, Y. Soini, R. Sironen, S.E. Bojesen, D.D. Orsted, D. Kaur-Knudsen, H. Flyger, B.G. Nordestgaard, H. Holland, G. Chenevix-Trench, S. Manoukian, M. Barile, P. Radice, S.E. Hankinson, D.J. Hunter, R. Tamimi, S. Sangrajrang, P. Brennan, J. McKay, F. Odefrey, V. Gaborieau, P. Devilee, P.E. Huijts, R.A. Tollenaar, C. Seynaeve, G.S. Dite, C. Apicella, J.L. Hopper, F. Hammet, H. Tsimiklis, L.D. Smith, M.C. Southey, M.K. Humphreys, D. Easton, P. Pharoah, M.E. Sherman and M. Garcia-Closas: Associations of breast cancer risk factors with tumor subtypes: a pooled analysis from the Breast Cancer Association Consortium studies. *J Natl Cancer Inst*, 103, 250-263 (2011)
14. M. Harvie, L. Hooper and A.H. Howell: Central obesity and breast cancer risk: a systematic review. *Obes Rev*, 4, 157-173 (2003)
15. D.P. Rose, D. Komninou and G.D. Stephenson: Obesity, adipocytokines, and insulin resistance in breast cancer. *Obes Rev*, 5, 153-165 (2004)
16. J.R. Daling, K.E. Malone, D.R. Doody, L.G. Johnson, J.R. Galloway and P.L. Porter: Relation of body mass index to tumor markers and survival among young women with invasive ductal breast carcinoma. *Cancer*, 92, 720-729 (2001)
17. M. Cotterchio, N. Kreiger, B. Theis, M. Sloan and S. Bahl: Hormonal factors and the risk of breast cancer according to estrogen- and progesterone-receptor subgroup. *Cancer Epidemiol Biomarkers Prev*, 12, 1053-1060 (2003)
18. G. Fagherazzi, N. Chabbert-Buffet, A. Fabre, G. Guillas, M.C. Boutron-Ruault, S. Mesrine and F. Clavel-Chapelon: Hip circumference is associated with the risk of premenopausal ER-/PR- breast cancer. *Int J Obes (Lond)* (2011)
19. A. Howell, M. Chapman and M. Harvie: Energy restriction for breast cancer prevention. *Recent Results Cancer Res*, 181, 97-111 (2009)
20. K.G. Hofbauer: Molecular pathways to obesity. *Int J Obes Relat Metab Disord*, 26 Suppl 2, S18-27 (2002)
21. R.E. Duncan, M. Ahmadian, K. Jaworski, E. Sarkadi-Nagy and H.S. Sul: Regulation of lipolysis in adipocytes. *Annu Rev Nutr*, 27, 79-101 (2007)
22. R. Drolet, C. Richard, A.D. Sniderman, J. Mailloux, M. Fortier, C. Huot, C. Rheume and A. Tchernof: Hypertrophy and hyperplasia of abdominal adipose tissues in women. *Int J Obes (Lond)*, 32, 283-291 (2008)
23. A. Kleinridders, D. Schenten, A.C. Konner, B.F. Belgardt, J. Mauer, T. Okamura, F.T. Wunderlich, R. Medzhitov and J.C. Bruning: MyD88 signaling in the CNS is required for development of fatty acid-induced leptin resistance and diet-induced obesity. *Cell Metab*, 10, 249-259 (2009)
24. G.R. Steinberg and B.E. Kemp: AMPK in Health and Disease. *Physiol Rev*, 89, 1025-1078 (2009)
25. X. Wu, H. Motoshima, K. Mahadev, T.J. Stalker, R. Scalia and B.J. Goldstein: Involvement of AMP-activated protein kinase in glucose uptake stimulated by the globular domain of adiponectin in primary rat adipocytes. *Diabetes*, 52, 1355-1363 (2003)
26. K.J. McInnes, A. Corbould, E.R. Simpson and M.E. Jones: Regulation of adenosine 5',monophosphate-activated protein kinase and lipogenesis by androgens contributes to visceral obesity in an estrogen-deficient state. *Endocrinology*, 147, 5907-5913 (2006)
27. K.J. McInnes, K.A. Brown, N.I. Hunger and E.R. Simpson: Regulation of LKB1 expression by sex hormones in adipocytes. *Int J Obes (Lond)* (2011)

28. O. Warburg: On the origin of cancer cells. *Science*, 123, 309-314 (1956)
29. B.J. Cairns, T.Y. Yang and V. Beral: That rising obesity levels will greatly add to the burden of cancer: misconceptions I. *Br J Cancer*, 104, 4-5 (2011)
30. J.A. Menendez and R. Lupu: Fatty acid synthase and the lipogenic phenotype in cancer pathogenesis. *Nat Rev Cancer*, 7, 763-777 (2007)
31. A. Vazquez-Martin, S. Ropero, J. Brunet, R. Colomer and J.A. Menendez: Inhibition of Fatty Acid Synthase (FASN) synergistically enhances the efficacy of 5-fluorouracil in breast carcinoma cells. *Oncol Rep*, 18, 973-980 (2007)
32. K.A. Brown, K.J. McInnes, K. Takagi, K. Ono, N.I. Hunger, L. Wang, H. Sasano and E.R. Simpson: LKB1 expression is inhibited by estradiol-17beta in MCF-7 cells. *J Steroid Biochem Mol Biol* (2011)
33. H. Fenton, B. Carlile, E.A. Montgomery, H. Carraway, J. Herman, F. Sahin, G.H. Su and P. Argani: LKB1 protein expression in human breast cancer. *Appl Immunohistochem Mol Morphol*, 14, 146-153 (2006)
34. S.M. Hadad, L. Baker, P.R. Quinlan, K.E. Robertson, S.E. Bray, G. Thomson, D. Kellock, L.B. Jordan, C.A. Purdie, D.G. Hardie, S. Fleming and A.M. Thompson: Histological evaluation of AMPK signalling in primary breast cancer. *BMC Cancer*, 9, 307 (2009)
35. K.H. Vousden and K.M. Ryan: p53 and metabolism. *Nat Rev Cancer*, 9, 691-700 (2009)
36. P. Hallenborg, S. Feddersen, L. Madsen and K. Kristiansen: The tumor suppressors pRB and p53 as regulators of adipocyte differentiation and function. *Expert Opin Ther Targets*, 13, 235-246 (2009)
37. A.I. Monteiro R: Chronic inflammation in obesity and the metabolic syndrome. *Mediators Inflamm* (2010)
38. J. Santos-Alvarez, R. Goberna and V. Sanchez-Margalet: Human leptin stimulates proliferation and activation of human circulating monocytes. *Cell Immunol*, 194, 6-11 (1999)
39. N. Kiguchi, T. Maeda, Y. Kobayashi, Y. Fukazawa and S. Kishioka: Leptin enhances CC-chemokine ligand expression in cultured murine macrophage. *Biochem Biophys Res Commun*, 384, 311-315 (2009)
40. J.E. Davis, N.K. Gabler, J. Walker-Daniels and M.E. Spurlock: Tlr-4 deficiency selectively protects against obesity induced by diets high in saturated fat. *Obesity (Silver Spring)*, 16, 1248-1255 (2008)
41. I. Murano, G. Barbatelli, V. Parisani, C. Latini, G. Muzzonigro, M. Castellucci and S. Cinti: Dead adipocytes, detected as crown-like structures, are prevalent in visceral fat depots of genetically obese mice. *J Lipid Res*, 49, 1562-1568 (2008)
42. P.G. Morris, C.A. Hudis, D. Giri, M. Morrow, D.J. Falcone, X.K. Zhou, B. Du, E. Brogi, C.B. Crawford, L. Kopelovich, K. Subbaramaiah and A.J. Dannenberg: Inflammation and increased aromatase expression occur in the breast tissue of obese women with breast cancer. *Cancer Prev Res (Phila)*, 4, 1021-1029 (2011)
43. A.D. Pradhan, J.E. Manson, N. Rifai, J.E. Buring and P.M. Ridker: C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA*, 286, 327-334 (2001)
44. G.R. Steinberg, B.J. Michell, B.J. van Denderen, M.J. Watt, A.L. Carey, B.C. Fam, S. Andrikopoulos, J. Proietto, C.Z. Gorgun, D. Carling, G.S. Hotamisligil, M.A. Febbraio, T.W. Kay and B.E. Kemp: Tumor necrosis factor alpha-induced skeletal muscle insulin resistance involves suppression of AMP-kinase signaling. *Cell Metab*, 4, 465-474 (2006)
45. K. Subbaramaiah, L.R. Howe, P. Bhardwaj, B. Du, C. Gravaghi, R.K. Yantiss, X.K. Zhou, V.A. Blaho, T. Hla, P. Yang, L. Kopelovich, C.A. Hudis and A.J. Dannenberg: Obesity is associated with inflammation and elevated aromatase expression in the mouse mammary gland. *Cancer Prev Res (Phila)*, 4, 329-346 (2011)
46. B. Richelsen: Release and effects of prostaglandins in adipose tissue. *Prostaglandins Leukot Essent Fatty Acids*, 47, 171-182 (1992)
47. J. Aubert, P. Saint-Marc, N. Belmonte, C. Dani, R. Negrel and G. Ailhaud: Prostacyclin IP receptor up-regulates the early expression of C/EBPbeta and C/EBPdelta in preadipose cells. *Mol Cell Endocrinol*, 160, 149-156 (2000)
48. B.M. Forman, P. Tontonoz, J. Chen, R.P. Brun, B.M. Spiegelman and R.M. Evans: 15-Deoxy-delta 12, 14-prostaglandin J2 is a ligand for the adipocyte determination factor PPAR gamma. *Cell*, 83, 803-812 (1995)
49. L. Fajas, S. Miard, M.R. Briggs and J. Auwerx: Selective cyclo-oxygenase-2 inhibitors impair adipocyte differentiation through inhibition of the clonal expansion phase. *J Lipid Res*, 44, 1652-1659 (2003)
50. K. Jaworski, M. Ahmadian, R.E. Duncan, E. Sarkadi-Nagy, K.A. Varady, M.K. Hellerstein, H.Y. Lee, V.T. Samuel, G.I. Shulman, K.H. Kim, S. de Val, C. Kang and H.S. Sul: AdPLA ablation increases lipolysis and prevents obesity induced by high-fat feeding or leptin deficiency. *Nat Med*, 15, 159-168 (2009)
51. A. Mantovani, P. Allavena, A. Sica and F. Balkwill: Cancer-related inflammation. *Nature*, 454, 436-444 (2008)
52. F. Balkwill: Tumour necrosis factor and cancer. *Nat Rev Cancer*, 9, 361-371 (2009)

53. F. Balkwill, K.A. Charles and A. Mantovani: Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell*, 7, 211-217 (2005)
54. E. Half, X.M. Tang, K. Gwyn, A. Sahin, K. Wathen and F.A. Sinicrope: Cyclooxygenase-2 expression in human breast cancers and adjacent ductal carcinoma *in situ*. *Cancer Res*, 62, 1676-1681 (2002)
55. D. Cannata, Y. Fierz, A. Vijayakumar and D. LeRoith: Type 2 diabetes and cancer: what is the connection? *Mt Sinai J Med*, 77, 197-213 (2010)
56. Y.M. Chong, K. Colston, W.G. Jiang, A.K. Sharma and K. Mokbel: The relationship between the insulin-like growth factor-I system and the oestrogen metabolising enzymes in breast cancer tissue and its adjacent non-cancerous tissue. *Breast Cancer Res Treat*, 99, 275-288 (2006)
57. R. Kaaks, S. Rinaldi, T.J. Key, F. Berrino, P.H. Peeters, C. Biessy, L. Dossus, A. Lukanova, S. Bingham, K.T. Khaw, N.E. Allen, H.B. Bueno-de-Mesquita, C.H. van Gils, D. Grobbee, H. Boeing, P.H. Lahmann, G. Nagel, J. Chang-Claude, F. Clavel-Chapelon, A. Fournier, A. Thiebaut, C.A. Gonzalez, J.R. Quiros, M.J. Tormo, E. Ardanaz, P. Amiano, V. Krogh, D. Palli, S. Panico, R. Tumino, P. Vineis, A. Trichopoulou, V. Kalapothaki, D. Trichopoulos, P. Ferrari, T. Norat, R. Saracci and E. Riboli: Postmenopausal serum androgens, oestrogens and breast cancer risk: the European prospective investigation into cancer and nutrition. *Endocr Relat Cancer*, 12, 1071-1082 (2005)
58. A. McTiernan, K.B. Rajan, S.S. Tworoger, M. Irwin, L. Bernstein, R. Baumgartner, F. Gilliland, F.Z. Stanczyk, Y. Yasui and R. Ballard-Barbash: Adiposity and sex hormones in postmenopausal breast cancer survivors. *J Clin Oncol*, 21, 1961-1966 (2003)
59. L. Baglietto, D.R. English, J.L. Hopper, R.J. MacInnis, H.A. Morris, W.D. Tilley, K. Krishnan and G.G. Giles: Circulating steroid hormone concentrations in postmenopausal women in relation to body size and composition. *Breast Cancer Res Treat*, 115, 171-179 (2009)
60. S.E. Bulun, Z. Lin, G. Imir, S. Amin, M. Demura, B. Yilmaz, R. Martin, H. Utsunomiya, S. Thung, B. Gurates, M. Tamura, D. Langoi and S. Deb: Regulation of aromatase expression in estrogen-responsive breast and uterine disease: from bench to treatment. *Pharmacol Rev*, 57, 359-383 (2005)
61. V.R. Agarwal, C.I. Ashanullah, E.R. Simpson and S.E. Bulun: Alternatively spliced transcripts of the aromatase cytochrome P450 (CYP19) gene in adipose tissue of women. *J Clin Endocrinol Metab*, 82, 70-74 (1997)
62. C. Zhou, D. Zhou, J. Esteban, J. Murai, P.K. Siiteri, S. Wilczynski and S. Chen: Aromatase gene expression and its exon I usage in human breast tumors. Detection of aromatase messenger RNA by reverse transcription-polymerase chain reaction. *J Steroid Biochem Mol Biol*, 59, 163-171 (1996)
63. Y. Zhao, C.R. Mendelson and E.R. Simpson: Characterization of the sequences of the human CYP19 (aromatase) gene that mediate regulation by glucocorticoids in adipose stromal cells and fetal hepatocytes. *Mol Endocrinol*, 9, 340-349 (1995)
64. Y. Zhao, J.E. Nichols, S.E. Bulun, C.R. Mendelson and E.R. Simpson: Aromatase P450 gene expression in human adipose tissue. Role of a Jak/STAT pathway in regulation of the adipose-specific promoter. *J Biol Chem*, 270, 16449-16457 (1995)
65. Y. Zhao, V.R. Agarwal, C.R. Mendelson and E.R. Simpson: Estrogen biosynthesis proximal to a breast tumor is stimulated by PGE2 via cyclic AMP, leading to activation of promoter II of the CYP19 (aromatase) gene. *Endocrinology*, 137, 5739-5742 (1996)
66. K.A. Brown, K.J. McInnes, N.I. Hunger, J.S. Oakhill, G.R. Steinberg and E.R. Simpson: Subcellular localization of cyclic AMP-responsive element binding protein-regulated transcription coactivator 2 provides a link between obesity and breast cancer in postmenopausal women. *Cancer Res*, 69, 5392-5399 (2009)
67. S.H. Koo, L. Flechner, L. Qi, X. Zhang, R.A. Screation, S. Jeffries, S. Hedrick, W. Xu, F. Boussouar, P. Brindle, H. Takemori and M. Montminy: The CREB coactivator TORC2 is a key regulator of fasting glucose metabolism. *Nature*, 437, 1109-1111 (2005)
68. M. Sofi, M.J. Young, T. Papamakarios, E.R. Simpson and C.D. Clyne: Role of CRE-binding protein (CREB) in aromatase expression in breast adipose. *Breast Cancer Res Treat*, 79, 399-407 (2003)
69. S. Chen, D. Zhou, C. Yang, T. Okubo, Y. Kinoshita, B. Yu, Y.C. Kao and T. Itoh: Modulation of aromatase expression in human breast tissue. *J Steroid Biochem Mol Biol*, 79, 35-40 (2001)
70. H.K. Choi, S.H. Roh, H.G. Kim, E.H. Han, H.G. Jeong and K.W. Kang: Enhanced expression of aromatase in p53-inactivated mammary epithelial cells. *Endocr Relat Cancer*, 15, 139-147 (2008)
71. P.G. McTernan, A. Anwar, M.C. Eggo, A.H. Barnett, P.M. Stewart and S. Kumar: Gender differences in the regulation of P450 aromatase expression and activity in human adipose tissue. *Int J Obes Relat Metab Disord*, 24, 875-881 (2000)
72. M. Schmidt and G. Loffler: Induction of aromatase in stromal vascular cells from human

- breast adipose tissue depends on cortisol and growth factors. *FEBS Lett*, 341, 177-181 (1994)
73. B. Su, C. Wong, Y. Hong and S. Chen: Growth factor signaling enhances aromatase activity of breast cancer cells via post-transcriptional mechanisms. *J Steroid Biochem Mol Biol*, 123, 101-108 (2011)
74. P. Lueprasitsakul, D. Latour and C. Longcope: Aromatase activity in human adipose tissue stromal cells: effect of growth factors. *Steroids*, 55, 540-544 (1990)
75. P.E. Lonning, S.I. Helle, D.C. Johannessen, H. Adlercreutz, E.A. Lien, M. Tally, D. Ekse, T. Fotsis, G.B. Anker and K. Hall: Relations between sex hormones, sex hormone binding globulin, insulin-like growth factor-I and insulin-like growth factor binding protein-1 in post-menopausal breast cancer patients. *Clin Endocrinol (Oxf)*, 42, 23-30 (1995)
76. K. Esposito, A. Pontillo, C. Di Palo, G. Giugliano, M. Masella, R. Marfella and D. Giugliano: Effect of weight loss and lifestyle changes on vascular inflammatory markers in obese women: a randomized trial. *JAMA*, 289, 1799-1804 (2003)
77. P. Dandona, R. Weinstock, K. Thusu, E. Abdel-Rahman, A. Aljada and T. Wadden: Tumor necrosis factor-alpha in sera of obese patients: fall with weight loss. *J Clin Endocrinol Metab*, 83, 2907-2910 (1998)
78. J.P. Bastard, C. Jardel, E. Bruckert, P. Blondy, J. Capeau, M. Laville, H. Vidal and B. Hainque: Elevated levels of interleukin 6 are reduced in serum and subcutaneous adipose tissue of obese women after weight loss. *J Clin Endocrinol Metab*, 85, 3338-3342 (2000)
79. R. Cencello, C. Henegar, N. Viguerie, S. Taleb, C. Poitou, C. Rouault, M. Coupaye, V. Pelloux, D. Hugol, J.L. Bouillot, A. Bouloumie, G. Barbatelli, S. Cinti, P.A. Svensson, G.S. Barsh, J.D. Zucker, A. Basdevant, D. Langin and K. Clement: Reduction of macrophage infiltration and chemoattractant gene expression changes in white adipose tissue of morbidly obese subjects after surgery-induced weight loss. *Diabetes*, 54, 2277-2286 (2005)
80. E.F. van Rossum, B.J. Nicklas, K.E. Dennis, D.M. Berman and A.P. Goldberg: Leptin responses to weight loss in postmenopausal women: relationship to sex-hormone binding globulin and visceral obesity. *Obes Res*, 8, 29-35 (2000)
81. D. Schapira, P. Wolff, N. Kumar, J. Anderson, N. Aziz, G. Lyman and M. Swanson: The effect of weight-loss on estimated breast-cancer risk and sex-hormone levels. *Oncol Rep*, 1, 613-617 (1994)
82. R. Pasquali, R. Fabbri, S. Venturoli, R. Paradisi, D. Antenucci and N. Melchionda: Effect of weight loss and antiandrogenic therapy on sex hormone blood levels and insulin resistance in obese patients with polycystic ovaries. *Am J Obstet Gynecol*, 154, 139-144 (1986)
83. H. Ashrafian, K. Ahmed, S.P. Rowland, V.M. Patel, N.J. Gooderham, E. Holmes, A. Darzi and T. Athanasiou: Metabolic surgery and cancer: protective effects of bariatric procedures. *Cancer*, 117, 788-799 (2011)
84. N.V. Christou, M. Lieberman, F. Sampalis and J.S. Sampalis: Bariatric surgery reduces cancer risk in morbidly obese patients. *Surg Obes Relat Dis*, 4, 691-695 (2008)
85. A.H. Berg and P.E. Scherer: Adipose tissue, inflammation, and cardiovascular disease. *Circ Res*, 96, 939-949 (2005)
86. J. Fruebis, T.S. Tsao, S. Javorschi, D. Ebbets-Reed, M.R. Erickson, F.T. Yen, B.E. Bihain and H.F. Lodish: Proteolytic cleavage product of 30-kDa adipocyte complement-related protein increases fatty acid oxidation in muscle and causes weight loss in mice. *Proc Natl Acad Sci USA*, 98, 2005-2010 (2001)
87. Y.S. Zhao, S. Zhu, X.W. Li, F. Wang, F.L. Hu, D.D. Li, W.C. Zhang and X. Li: Association between NSAIDs use and breast cancer risk: a systematic review and meta-analysis. *Breast Cancer Res Treat*, 117, 141-150 (2009)
88. V.A. Narkar, M. Downes, R.T. Yu, E. Embler, Y.X. Wang, E. Banayo, M.M. Mihaylova, M.C. Nelson, Y. Zou, H. Juguilon, H. Kang, R.J. Shaw and R.M. Evans: AMPK and PPARdelta agonists are exercise mimetics. *Cell*, 134, 405-415 (2008)
89. M.P. Gaidhu, A. Frontini, S. Hung, K. Pistor, S. Cinti and R.B. Ceddia: Chronic AMP-kinase activation with AICAR reduces adiposity by remodeling adipocyte metabolism and increasing leptin sensitivity. *J Lipid Res*, 52, 1702-1711 (2011)
90. A. Bai, M. Yong, A.G. Ma, Y. Ma, C.R. Weiss, Q. Guan, C.N. Bernstein and Z. Peng: Novel anti-inflammatory action of 5-aminoimidazole-4-carboxamide ribonucleoside with protective effect in dextran sulfate sodium-induced acute and chronic colitis. *J Pharmacol Exp Ther*, 333, 717-725 (2010)
91. J.V. Swinnen, A. Beckers, K. Brusselmans, S. Organe, J. Segers, L. Timmermans, F. Vanderhoydonc, L. Deboel, R. Derua, E. Waelkens, E. De Schrijver, T. Van de Sande, A. Noel, F. Fougelle and G. Verhoeven: Mimicry of a cellular low energy status blocks tumor cell anabolism and suppresses the malignant phenotype. *Cancer Res*, 65, 2441-2448 (2005)
92. W.C. Knowler, E. Barrett-Connor, S.E. Fowler, R.F. Hamman, J.M. Lachin, E.A. Walker and D.M. Nathan: Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med*, 346, 393-403 (2002)

93. S. Haffner, M. Temprosa, J. Crandall, S. Fowler, R. Goldberg, E. Horton, S. Marcovina, K. Mather, T. Orchard, R. Ratner and E. Barrett-Connor: Intensive lifestyle intervention or metformin on inflammation and coagulation in participants with impaired glucose tolerance. *Diabetes*, 54, 1566-1572 (2005)
94. M. Cazzaniga, B. Bonanni, A. Guerrieri-Gonzaga and A. Decensi: Is it time to test metformin in breast cancer clinical trials? *Cancer Epidemiol Biomarkers Prev*, 18, 701-705 (2009)
95. B. Martin-Castillo, A. Vazquez-Martin, C. Oliveras-Ferraros and J.A. Menendez: Metformin and cancer: doses, mechanisms and the dandelion and hormetic phenomena. *Cell Cycle*, 9, 1057-1064 (2010)
96. K.A. Brown, N.I. Hunger, M. Docanto and E.R. Simpson: Metformin inhibits aromatase expression in human breast adipose stromal cells via stimulation of AMP-activated protein kinase. *Breast Cancer Res Treat*, 123, 591-596 (2010)
97. N.U. Samarajeewa, S. Ham, F. Yang, E.R. Simpson and K.A. Brown: Promoter-specific effects of metformin on aromatase transcript expression. *Steroids*, 76, 768-771 (2011)

Key Words: Obesity, Breast cancer, Adipokines, Inflammation, Insulin, IGF-I, Estrogens, Metabolism, Review

Send correspondence to: Kristy A. Brown, Metabolism and Cancer Laboratory, Prince Henry's Institute, 246 Clayton Road, Clayton, Victoria, 3168, Tel: 61 3 9594 4333, Fax: 61 3 9594 6125, E-mail: kristy.brown@princehenrys.org