

## ALK inhibitors for clinical use in cancer therapy

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### TABLE OF CONTENTS

1. Abstract
2. Introduction
3. ALK inhibitors
  - 3.1. The development
  - 3.2. Crizotinib
4. Mechanisms of resistance to ALK inhibitors
5. "Second generation" ALK inhibitors
6. Other approaches: HSP90 inhibitors
7. "Third generation" inhibitors
8. Conclusions
9. Acknowledgements
10. References

### 1. ABSTRACT

Anaplastic lymphoma kinase (ALK) is a receptor tyrosine kinase protein implicated in a variety of tumors, both solid and hematological. Few years ago crizotinib, an inhibitor of the receptor tyrosine kinases c-Met and ALK, demonstrated its activity in ALK positive non-small-cell lung cancer and other tumors with excellent toxicity profile. Subsequently several ALK inhibitors have been developed, offering new personalized treatment options. This review addresses some clinical considerations on the use of ALK inhibitors in ALK positive tumors and on the development of resistance to them.

### 2. INTRODUCTION

In 1994 Steve Morris identified ALK as a novel oncogene involved in a specific 2;5 chromosomal translocation and fused to the *nucleophosmin* (*NPM*) gene in Anaplastic Large Cell Lymphoma (ALCL) (1). Clinical development of ALK inhibitors took off however only in 2007, when aberrant activation of *anaplastic lymphoma kinase* (ALK) gene was found in non-small-cell lung cancer (NSCLC) and was identified as an *echinoderm microtubule-associated protein-like 4* (*EML4*)-ALK rearrangement (2). From this point the interest in ALK has increased and a number of inhibitor entered in preclinical and clinical studies, offering new treatment options in tumors driven by abnormal ALK signalling.

ALK is a tyrosine kinase (TK) receptor (1,3), normally expressed only in the nervous system (4,5) with a role in neural development and

differentiation (6-8). ALK is expressed in various type of cancers such as inflammatory myofibroblastic tumor (IMT) (9), glioblastoma (6,10), inflammatory breast cancer (11), neuroblastoma (12), Ewing sarcoma (13), retinoblastoma (14), diffuse-large-B-cell lymphoma (DLBCL) (15), urothelial carcinoma (16), fetal lung interstitial tumor (17) and melanoma (18).

ALK pathological expression is due to chromosomal translocations that lead to the formation of ALK-containing oncogenic fusion proteins (19) that interrupt the chromosome at the level of ALK gene at 2p23. Currently 16 different ALK fusion proteins have been identified (17,20) with different predominance in each type of tumors. ALK fusions cause constitutive activation of its kinase domain (21) so, ALK fusion proteins are deregulated, ectopically expressed and constitutively activated in neoplastic cells altering the phosphorylation of intracellular substrates (4,22) and activating several transduction pathways (23-31).

For this reason, the relationship between the development ALK-containing fusions and malignant transformation makes ALK a potential therapeutic target. This evidence is less solid when a fusion or an activating point mutation is missing. During last years several ALK inhibitors have been developed (32-35): these molecules represent an excellent proof-of-principle for targeted therapy (36). As it has been observed with other TK inhibitors (TKIs), resistance has recently emerged in patients treated with ALK inhibitors too (37-41).

### 3. ALK INHIBITORS

#### 3.1. The development

ALK is a good candidate for the development of targeted treatment not only because the lack of widely expression in normal adult tissues that should not give potentially important toxic effects (31,38,42) but also because it is causally linked to the transformation process. Potential strategies for targeting ALK include immunotherapy, gene silencing, inhibition of downstream signalling pathways and direct inhibition of its catalytic activity through small-molecules inhibitors in order to inhibit ALK dependent cancer cell growth (20). The aim of this target therapy is to obtain a maximum tumor specific effect with low toxicities, opposite to a conventional cytotoxic chemotherapy (43). Kinase inhibitors are directed against the ATP-binding site of the catalytic domain, which is highly conserved in ALK TK, with the goal of obtain a successful ALK inhibition.

However, the presence of different ALK fusions confers different sensitivity to ALK inhibitors (44) and also point mutations may be the cause of secondary drug resistance as found first in neuroblastoma (45,46).

Initial testing of ALK inhibitors were performed using inhibitors derived from natural products such as staurosporine derivatives which are not specific inhibitors of ALK (47). Synergies with heat shock protein 90 (HSP-90) inhibitors were also observed (48). Subsequently, almost 20 different classes of small molecule inhibitors of ALK have been developed (Table 1).

#### 3.2. Crizotinib

The largest volume of available knowledge on ALK inhibitors relates to crizotinib. Crizotinib (PF-02341066, Pfizer) is the first in human ALK inhibitor developed. It is a derivate of aminopyridine and was originally developed as a potent, orally bioavailable, ATP-competitive small molecule inhibitor of mesenchymal epithelial transition growth factor/hepatocyte growth factor receptor (c-MET) (49). In 2011 it was approved by the Food and Drugs Administration in ALK positive NSCLC.

The phase I trial established 250 mg twice daily as the maximum tolerated dose (MTD) in patients with advanced cancer (excluding leukemias) (50). Preclinical and single-arm phase I studies have shown that patients with advanced stage ALK positive NSCLC can be successfully treated with crizotinib with overall response rate (ORR) of 61%. Progression free survival (PFS) was 10-11 months and there were very few grade 3 and 4 adverse events (AEs) (51,52). The most common AEs were: ocular flashes, nausea, emesis, fatigue, and diarrhoea; all manageable and reversible. Fatigue was the dose-limiting toxicity (DLT), occurring at grade 3 in 2/6 patients treated with crizotinib at

300 mg twice a day. Recently published post-marketing AEs described nephrotoxicity in a patient with previous idiopathic chronic kidney disease (53), esophagitis (54,55), alveolar hemorrhage (56) and interstitial lung disease (57). However, the combined administration of crizotinib with steroids has been shown to overcome this last AE (58). A recent phase III trial (NCT00932893) confirm the phase I-results with a PFS of 8 months with crizotinib versus 3 months with platinum-based chemotherapy and a ORR 65% versus 20% respectively (59). Others randomized phase III trials are testing crizotinib versus standard chemotherapy in first-line treatment in ALK positive non squamous lung cancer (NCT01154140) and adjuvant crizotinib versus placebo in NSCLC removed by surgery (NCT02201992).

Crizotinib has shown its efficacy also in rarer tumor like ALCL, DLBCL, IMT and other type of ALK-rearranged cancers. The first treatment of an ALK positive ALCL patient occurred in 2010. A report from *New England Journal of Medicine* described 2 adult patients with recurrent ALK positive ALCLs that achieved complete response (CR) shortly after receiving crizotinib as single agent, administered under a compassionate drug program (60). The final results of the study were recently published demonstrating clearly the activity of crizotinib in ALK positive ALCLs. ORR was 91% with 82% CR and 1/11 partial response (PR); 2-year PFS of 64% (61). All relapses developed within the initial 3 months of treatment with no parameter predicting for the development of a durable response, other than having failed an ABMT. These results were recently confirmed in a sponsored trial in which reported an ORR of 64% among 14 ALK positive lymphoma patients (62). A recent phase I study also reported this good safety profile with high response rates in children with relapsed ALK positive ALCL (63).

Crizotinib can be efficacious also in the treatment of ALK positive DLBCL: the compassionate study previously cited (61) reported 2 patients with ALK positive DLBCL treated with crizotinib with one rapid but transient response. Some experiences shows that crizotinib can be used as maintenance after allogenic bone marrow transplant, confirming its good safety profile (64).

Recently two cases of IMT and its epithelioid variant with systemic manifestations and ALK translocation were successfully treated with crizotinib (65,66). There were also reported some case reports on other ALK positive tumour successfully treated with crizotinib: an advanced pretreated sarcomatoid carcinoma of the upper aerodigestive tract has achieved with crizotinib clinical improvement and stable disease with minimal tumor shrinkage lasted for 4 months (67). A phase I/II trial of crizotinib in tumors other than NSCLC (NCT01121588) and in young patients (NCT00939770) is presently ongoing.

**Clinical use of ALK inhibitors**

**Table 1.** ALK inhibitors in development

Name	Mechanism of action	Clinical trials in tumors	Responses	Adverse events	References
Crizotinib (PF-02341066 - Pfizer)	Selective ATP competitive oral inhibitor of ALK and MET tyrosine kinases and their variants	Phase I-II Phase III in second line Ongoing phase III in first line in NSCLC Studies with crizotinib as single agent or in combination	In NSCLC: 61-65% ORR, PFS 7.7. months In ALCL: ORR 64-91%, 2-years-PFS 64% Experiences in other solid tumors	Gastrointestinal adverse events (diarrhoea, emesis and nausea) Fatigue ALT elevation Ocular flashes	(51,52,59,61,62)
Alectinib (CH5424802 - Chugai Pharmaceutical, Roche)	Potent and selective orally available ALK and ROS1 inhibitor Activity against L1196M	Phase I/II trial in NSCLC patients in Japan Ongoing phase III in Japan versus crizotinib resistant NSCLC Ongoing phase I/II in US	In NSCLC: ORR 93.5. % with 60% ORR in crizotinib resistant NSCLC	Dysgeusia, ALT and creatinine elevation, rush, neutropenia	(94-96,117,118)
Ceritinib (LDK 378 - Novartis)	Selective ALK, IFG-1R and c-MET inhibitor Activity against L1196M	Phase I . in crizotinib naïve and resistant Phase II Phase III ongoing both as a first- and second-line therapy for ALK positive NSCLC	In NSCLC ORR 58% with 56% in crizotinib refractory, PFS 8.6. months	Nausea, diarrhea, fatigue and ALT elevation	(84,90,91,119)
AP-26113 (Ariad)	Dual ALK/EGFR inhibitor and ROS1 inhibitor AP-26113 is 10 fold more potent compared to PF-02341066	Phase I/II in crizotinib naïve and resistant: ongoing	In NSCLC ORR 63% with ORR 73% in crizotinib resistance	Well tolerated, fatigue, nausea, diarrhea, dyspnea, ALT elevation	(98,99,120)
ASP 3026 (Astellas Pharma)	ALK, ROS1 and EGFR inhibitor Activity against L1196M	Phase I: promising safety and tolerability	In crizotinib resistant NSCLC: 44% PR and 50% SD, PFS 5.9. months	Nausea, vomiting, fatigue, constipation, rush, AST/ALT elevation	(113,121)
PHA-E429 (nerviano medical science)	ATP competitive, crystal structure of the ALK kinase domain in a complex with PHA-E429	Phase I			(122)
CEP-14083 and CEP-14513 (Cephalon)	Selective ALK inhibitor	Phase I			
PF06463922 (Pfizer)	Selective inhibitor of anaplastic lymphoma kinase (ALK), EGFR and c-Ros oncogene 1 (ROS1)	Phase I/II			(109)
TSR-011 (Tesaro)	Inhibits ALK and tropomyosin-related kinase (TRK) A, B, and C receptor and oncogenic echinoderm microtubule associated protein like 4 (EML4)-ALK and tropomyosin (TPM)-TRKA dependent tumor growth in mice Activity against L1196M	Phase I/II	In NSCLC, 3/4 (all crizotinib-resistant) achieved PR Stable disease was observed in ALK- papillary thyroid, pancreatic and colorectal patients		(110)

(Contd...)

**Table 1.** (Continued...)

Name	Mechanism of action	Clinical trials in tumors	Responses	Adverse events	References
RXDX101 (Ignyta Inc.)	Inhibitor of TrkA, TrkB and TrkC, as well as ROS1 and ALK	Phase I in different type of advanced tumors	1/1 PR in ALK positive neuroblastoma	Well tolerated. Grade 1-2 paresthesias, nausea, dysgeusia, and diarrhoea	(114)
X396 (Xcovery)	ALK inhibitor and synergic activity with mTOR inhibitor rapamycin Activity against L1196M	Phase I in NSCLC ongoing	Preliminary data: SD is 28% and PR 28%	Preliminary data: rash fatigue, nausea, vomiting and edema	(106, 107)
CEP-37440 (Cephalon)		Phase I in NSCLC			(123)
NMS-E628 (Ariad)	ALK inhibitor	Phase I			
X376 (Xcovery)	ALK inhibitor Preclinical data available only				(107)
NVP-TAE684 (Novartis)	Selective ALK inhibitor	Phase I discontinued			(84)
WZ-5-126 (Ambit Biosciences)	ALK small molecules inhibitor Preclinical data available only				(124)
CRL151104A (chebridge st. jude)	ATP competitor Preclinical data available only				(125)
CEP-28122 (Cephalon)	Selective ALK inhibitor Preclinical data available only				(126)
F91873 and F91874 (Institute de rechercher Pierre Fabre)	ATP noncompetitive inhibitors, pyridoisoquinoline derivatives Preclinical data available only				
GSK1838705A (GlaxoSmithKlein)	Small molecule kinase inhibitor of insulin-like growth factor receptor. Preclinical data available only				(127)

#### 4. MECHANISMS OF RESISTANCE TO ALK INHIBITORS

The development of drug resistance remains one of the major limitations of successful treatment of advanced cancers: the study mentioned above(61) have shown that resistance can emerge in some patients who have demonstrated initial response to ALK inhibition. Mechanism of acquired resistance can be divided in 3 groups, very similar to what we know on chronic myeloid leukaemia (CML) (68). First is ALK mutation, followed by resistance due to mechanisms that upregulates ALK (such as gene amplification or copy number gain), and by the activation of alternative signal transduction pathways (69-71). Mutations in ALK kinase domain confer a structural change in the ATP-binding site (72) and reduced drugs sensitivity both *in vitro* and *in vivo* with variable level of drug resistance like for BCR-ABL in EML4-ALK. Mutations can involve the "gatekeeper" residue L1196, (for example L1196M (72)), and other "non-gatekeeper" residues distributed throughout the kinase domain: G1202R, S1206Y, G1269A, I1151Tins, F1174L, L1152R and C1156Y (69,73). The prevalence and the significance of these point mutations need yet

to be clarified but, it is known that C1156Y, G1269A and L1196M, conferred clinical resistance to crizotinib in NSCLC patients and another one, F1174L, causes resistance to the same drug in an IMT patient (72,74). The acquired resistance to ALK-targeted therapy has been distinguished in "ALK-dominant" and "ALK-non-dominant" mechanisms.

In "ALK-dominant" mechanisms, the acquired resistance is due to the development of novel ALK kinase domain mutation alone or in combination with the increase of rearranged ALK gene copies in cancer cells, so the signalling remains dominant. Cells are still dependent on that pathway for survival and "second generation" inhibitors may be sufficient to bypass the resistance. Multiple ALK inhibitors have been tested against crizotinib-resistant mutants (75). These authors found that L1196Q substitution conferred resistance to crizotinib but not to AP26113 and NVP-TAE 694, while cells carrying I1171N mutation were resistant to all tested inhibitors (37). In the clinical report described above (61) deep sequencing of 2 blood samples of relapsed ALCL revealed the presence of different mutations: Q1064R at high prevalence (95%) in 1 patient, and I1171N

(33%) plus M1328I (14%) in the other patient. All these mutations were not present in samples obtained before crizotinib treatment. I1171N was already discovered in an *in vitro* screening (5): it commands an intermediate level of resistance to crizotinib which however is cross resistant with other ALK inhibitors such as AP26113 and NVP-TAE684. Mechanisms of acquired resistance are heterogeneous and may evolve dynamically in response to different ALK TKIs. Recently, consistent with preclinical data, clinical evidence was found that ALK mutations in two residues, G1202R and F1174V, can mediate resistance also to ceritinib (76) and alectinib (77).

In contrast, “ALK-non-dominant” mechanisms are cases in which different second oncogenic drivers were activated (for example, KRAS, KIT, NTRK1 or EGFR), coexisting in the same cell with the ALK rearrangement (78,79). These events should, in theory, bypass the dominance of ALK signalling and replace its oncogenic potential (37); in such a situation, “second generation” inhibitors would be ineffective. In order to overcome resistance, it will be important to differentiate patients that preserve ALK dominance versus those that have diminished ALK dominance.

It is reassuring however that “second generation” inhibitors work better than what could be expected on the basis of “non-dominant” models, with ORR from 55 to 60% (80). This complexity of mechanisms of acquired resistance suggests that other therapeutic options, including the use of radiation to treat isolated areas of progression and adding or switching to cytotoxic chemotherapy could be required (81), or the combination of some of these TKIs with other therapies such as for example, HSP-90 inhibitors or immunotherapy (82,83). None of these approaches have been subjected to controlled studies.

## 5. “SECOND GENERATION” ALK INHIBITORS

Ceritinib (LDK378, Novartis Inc.), alectinib (CH5434802/RO5424802, Chugai Pharmaceutical/Roche), AP26113 (ARIAD Pharmaceuticals) are some examples of “new generation” inhibitors (Table 1). Ceritinib specifically inhibits the ALK fusion protein and is not active against C-Met. It is 20-times more potent than crizotinib (84,85): (39). In particular, ceritinib effectively inhibits ALK harboring L1196M, G1269A (the two most common crizotinib-resistant mutations), I1171T, and S1206Y mutations. However, we observed that ceritinib did not overcome 2 crizotinib-resistant ALK mutations, G1202R and F1174C (86). The first-phase I trial in human, limited to relapsed/refractory NSCLC patients demonstrated objective responses (87,88), confirming preclinical data (89). ORR was 58% at the dose of 400 mg once daily, in particular ORR of 56% was reported in crizotinib resistant patients (90,91): this response rate is

similar for patients not previously treated with ALK inhibitor or treated with crizotinib. Also PFS can support a front-line treatment scenario: PFS was 8.6. months, and was 3 months longer in patients who had not received crizotinib previously. Major adverse events were gastrointestinal AEs (primarily grade 1 or 2 nausea and diarrhea), fatigue and elevation of ALT very similar to crizotinib; however only half the patients required dose modification when compared with crizotinib(90). However ceritinib showed a higher drug-related grade 3 or 4 diarrhea and nausea (respectively 7% and 5%) compared with crizotinib (0% and 1%) (52,59,90). Ceritinib has been approved in the US under ‘Breakthrough Therapy’ designation for the second-line treatment of ALK-positive NSCLC; EMEA response is still pending (92). Currently phase II and III trials (NCT01964157, NCT01947608, NCT01828112 and NCT01828099) are ongoing both in crizotinib naïve or not patients. There is also a phase I study in paediatric patient investigated ceritinib (NCT01742286).

Alectinib is a potent and selective ALK inhibitor (93). Preclinical data have demonstrated that alectinib specifically inhibits EML4-ALK with higher potency than crizotinib, and is also active in cell lines with the most common crizotinib resistance mutations, including L1196M, C1156Y, and F1174L (94). A phase I/II trial in crizotinib-naïve patients with ALK positive NSCLC demonstrated objective responses in 93.5.% patients enrolled at the MTD (300 mg twice day without DLT), with 2 CR and 41 PR and not yet determined PFS (95). The activity was associated with mild adverse events (no grade 4 were reported): mostly dysgeusia, liver dysfunction, neutropenia, rash and creatinine elevation. Visual effects and gastrointestinal disorders (diarrhea, vomiting, and nausea) were rare. During a phase I study there were an ORR of 60% and good safety profile in patients who are refractory to crizotinib (96): for this reason, a single-arm phase I/II study in crizotinib resistant patients is ongoing (NCT01801111). There is also an ongoing phase III trial comparing alectinib versus crizotinib in advanced NSCLC (NCT02075840).

AP26113 (Ariad) is a dual ALK and EGFR inhibitor: in preclinical setting it has shown efficacy also in crizotinib resistant L1196M (97). It has clinical activity in crizotinib-naïve and crizotinib-resistant NSCLC, with an ORR of 63% or 73% in crizotinib resistant patients (98). Increased ALT and dyspnea have been noted as dose-limiting toxicities, and the most common grade 3 and 4 adverse events were pulmonary symptoms like hypoxia and dyspnea (at 180 mg/day), with also grade 1 or 2 fatigue and gastrointestinal AEs (99). The phase II study (NCT01449461 and NCT02094573) is ongoing at the dose of 180 mg daily both in crizotinib-resistant and crizotinib-naïve patients and in patients with ALK wild-type NSCLC who have EGFR mutations resistant to available TKIs.

## 6. OTHER APPROACHES: HSP90 INHIBITORS

HSP-90 inhibitors and other natural-derived compounds inhibit ALK by increasing the proteosome-mediated degradation of ALK protein through binding to HSP-90 (100,101). This strategy could be useful to manage acquired resistance to second-generation ALK inhibitors (102). Various inhibitors such as ganetespib (STA-9090), AUY922, IPI504 and retaspamycin are under investigation. They have shown ORR between 66% and 40% for IPI504 (103) and ganetespib (104) respectively, with PFS of 7-8 months. AEs are constituted mainly by fatigue, diarrhea and nausea. AUY922 (105) and ganetespib (101) has demonstrated activity also in crizotinib-resistant patients. There are several phase I/II trials examining all HSP-90 inhibitors alone or in combination with crizotinib or ceritinib in patients with advanced ALK positive tumors: IPI-504 (Infinity pharmaceuticals, NCT01228435), ganetespib (Syntax pharmaceuticals, NCT01579994 and NCT01562015), AT13387 (Astex pharmaceutical, NCT01712217), AUY922 (Novartis, NCT01772797, NCT01124864 and NCT01752400) and DS 2248 (Daiichi Sankyo, NCT01288430).

## 7. "THIRD GENERATION" INHIBITORS

Other new ALK TKIs, such as ASP-3026 (Astellas Pharma), NMS-E628 (Nerviano Medical Sciences), X-396 (Xcovery), CEP-37440 (Teva Pharmaceutical Industries), TSR-011 (Tesaro), PF-06463922 (Pfizer) and RXDX101 (Ignya Inc.) are currently starting clinical trials.

X-396 is a novel, potent ALK small molecule TKI with significant anti-tumor activity. Preliminary data on phase I (NCT01625234) study shows that it is generally well tolerated at doses up to 250 mg daily and induces responses in both crizotinib naïve and crizotinib resistant ALK positive NSCLC patients (106). Moreover, it can inhibit ALK kinase also when it is associated with its resistance point mutation L1196M and C1156Y (107).

PF06463922 is a new ALK inhibitor and a potent ROS1 kinase inhibitor and preclinical data show its great potential to overcome the resistance associated with ROS1 mutation (108,109). A phase I/II trial is ongoing (NCT01970865).

TSR-011 is currently used in a phase I/IIa trial (NCT02048488): its preclinical data show a high affinity for the ALK domain and preliminary data show promising results (110).

Recently ASP 3026 has shown activity in ALCL mice models (111,112). Preliminary data on "fast following" design trial (NCT01401504) demonstrated good activity in crizotinib resistant NSCLC with low toxicity (113). The phase II trial is recruiting patients with advanced malignancies (NCT01284192).

RXDX-101 is an oral small molecule inhibitor of TrkA, TrkB and TrkC, as well as ROS1 and ALK, with high potency and selectivity. It has demonstrated potent pharmacological activity in preclinical studies and it is well tolerated in patients with advanced solid tumors (114): The phase I/IIa is ongoing (NCT02097810).

## 8. CONCLUSIONS

Personalised treatment of cancer patients has become a reality in the last few years, with many drugs having been developed that target specific altered pathways. The initial identification of the genetic lesion at the basis of malignant transformation in ALK positive ALCLs, originally obtained in 1994 (1), was successfully exploited and brought to patient bedside in 2010 (60). This compares favorably with the time which elapsed between the discovery of the Philadelphia (Ph) chromosome in CML and the clinical development of imatinib (115). The success in identifying the ALK translocations and rapidly developing targeted drugs to exploit it paves the way for a better understanding of NSCLC and other tumors biology. The entry of crizotinib in the treatment of ALK positive tumors marked a new era in the therapy of these malignancies. The demonstration of high response rates, even in the setting of advanced and resistant disease, should prompt the development of clinical studies in less advanced conditions and in combinations with already active drugs. A particular emphasis should be placed on the need to decrease as much as possible the use of cytotoxic drugs. However, the best way in which ALK inhibitors should be administered in the setting of ALK-rearranged tumors remain to be fully elucidated (116). Indeed the future availability of several ALK TKIs will require studies to investigate the best use (sequential vs. combination) of these powerful tools. The possibility of obtaining responses in a substantial fraction of advanced, heavily pretreated ALK positive tumors illustrates our present inability to forecast the level of heterogeneity present inside the disease, such as a metastatic NSCLC, relapsed blast crisis CML or relapsed Ph positive acute lymphoblastic leukemia, in which monotherapy with TKIs seldom obtain durable responses. Further studies, such as exome sequencing of pre and post treatment samples could hopefully shed light and provide an useful indicator of the level of heterogeneity present inside a tumor at any given time. The next couple of years will hopefully see the fading of regimens based only on unspecific cytotoxic drugs in favor of more specific and hopefully less toxic approaches. The use of crizotinib and other ALK TKIs represent an useful example of how personalized medicine has improved patient care through the use of molecular-targeted therapy.

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## 10. REFERENCES

1. S. W. Morris, M. N. Kirstein, M. B. Valentine, K. G. Dittmer, D. N. Shapiro, D. L. Saltman and A. T. Look: Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. *Science*, 263(5151), 1281-4 (1994) DOI: 10.1126/science.8122112
2. M. Soda, Y. L. Choi, M. Enomoto, S. Takada, Y. Yamashita, S. Ishikawa, S. Fujiwara, H. Watanabe, K. Kurashina, H. Hatanaka, M. Bando, S. Ohno, Y. Ishikawa, H. Aburatani, T. Niki, Y. Sohara, Y. Sugiyama and H. Mano: Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature*, 448(7153), 561-6 (2007) DOI: 10.1038/nature05945
3. M. Shiota, J. Fujimoto, T. Semba, H. Satoh, T. Yamamoto and S. Mori: Hyperphosphorylation of a novel 80 kDa protein-tyrosine kinase similar to Ltk in a human Ki-1 lymphoma cell line, AMS3. *Oncogene*, 9(6), 1567-74 (1994)
4. L. Passoni and C. Gambacorti-Passerini: ALK a novel lymphoma-associated tumor antigen for vaccination strategies. *Leuk Lymphoma*, 44(10), 1675-81 (2003) DOI: 10.1080/1042819031000099625
5. F. Y. Hsu, Y. Zhao, W. F. Anderson and P. B. Johnston: Downregulation of NPM-ALK by siRNA causes anaplastic large cell lymphoma cell growth inhibition and augments the anti cancer effects of chemotherapy *in vitro*. *Cancer Invest*, 25(4), 240-8 (2007) DOI: 10.1080/07357900701206372
6. W. G. Dirks, S. Fahrrich, Y. Lis, E. Becker, R. A. MacLeod and H. G. Drexler: Expression and functional analysis of the anaplastic lymphoma kinase (ALK) gene in tumor cell lines. *Int J Cancer*, 100(1), 49-56 (2002) DOI: 10.1002/ijc.10435
7. C. Powers, A. Aigner, G. E. Stoica, K. McDonnell and A. Wellstein: Pleiotrophin signaling through anaplastic lymphoma kinase is rate-limiting for glioblastoma growth. *J Biol Chem*, 277(16), 14153-8 (2002) DOI: 10.1074/jbc.M112354200
8. R. Li and S. W. Morris: Development of anaplastic lymphoma kinase (ALK) small-molecule inhibitors for cancer therapy. *Med Res Rev*, 28(3), 372-412 (2008) DOI: 10.1002/med.20109
9. A. S. Patel, K. M. Murphy, A. L. Hawkins, J. S. Cohen, P. P. Long, E. J. Perlman and C. A. Griffin: RANBP2 and CLTC are involved in ALK rearrangements in inflammatory myofibroblastic tumors. *Cancer Genet Cytogenet*, 176(2), 107-14 (2007) DOI: 10.1016/j.cancergenryo.2007.04.004
10. K. V. Lu, K. A. Jong, G. Y. Kim, J. Singh, E. Q. Dia, K. Yoshimoto, M. Y. Wang, T. F. Cloughesy, S. F. Nelson and P. S. Mischel: Differential induction of glioblastoma migration and growth by two forms of pleiotrophin. *J Biol Chem*, 280(29), 26953-64 (2005) DOI: 10.1074/jbc.M502614200
11. R. S. Tuma: ALK gene amplified in most inflammatory breast cancers. *J Natl Cancer Inst*, 104(2), 87-8 (2012) DOI: 10.1093/jnci/djr553
12. L. Lamant, K. Pulford, D. Bischof, S. W. Morris, D. Y. Mason, G. Delsol and B. Mariame: Expression of the ALK tyrosine kinase gene in neuroblastoma. *Am J Pathol*, 156(5), 1711-21 (2000) DOI: 10.1016/S0002-9440(10)65042-0
13. A. Zoubeka, I. Simonitschb, E. R. Panzer-Grümayera, D. Ghalia, C. Pfleiderera, O.A. Haasa, G. Manna, S. Langb, T. Radaszkiewiczb, H. Gadnera, H. Kovar: Ewing tumor after treatment of Ki-1+ anaplastic large cell lymphoma. Therapy-associated secondary neoplasm or unrelated coincidence? *Cancer Genet Cytogenet*, 83(1), 5-11 (1995) DOI: 10.1016/S0165-4608(95)00014-3
14. G. Z. Rassidakis, R. Lai, M. Herling, C. Cromwell, A. Schmitt-Graeff and L. J. Medeiros: Retinoblastoma protein is frequently absent or phosphorylated in anaplastic large-cell lymphoma. *Am J Pathol*, 164(6), 2259-67 (2004) DOI: 10.1016/S0002-9440(10)63782-0
15. G. Delsol, L. Lamant, B. Mariame, K. Pulford, N. Dastugue, P. Brousset, F. Rigal-Huguet, T. al Saati, D. P. Cerretti, S. W. Morris and D. Y. Mason: A new subtype of large B-cell lymphoma expressing the ALK kinase and lacking the 2; 5 translocation. *Blood*, 89(5), 1483-90 (1997)
16. J. Bellmunt, S. Selvarajah, S. Rodig, M. Salido, S. de Muga, I. Costa, B. Bellosillo, L. Werner, S. Mullane, A. P. Fay, R. O'Brien, J. Barretina,

- A. E. Minoche, S. Signoretti, C. Montagut, H. Himmelbauer, D. M. Berman, P. Kantoff, T. K. Choueiri and J. E. Rosenberg: Identification of ALK Gene Alterations in Urothelial Carcinoma. *PLoS One*, 9(8), e103325 (2014)  
DOI: 10.1371/journal.pone.0103325
17. T. Onoda, M. Kanno, H. Sato, N. Takahashi, H. Izumino, H. Ohta, T. Emura, H. Katoh, H. Ohizumi, H. Otake, H. Asao, L. P. Dehner, A. D. Hill, K. Hayasaka and T. Mitsui: Identification of novel ALK rearrangement A2M-ALK in a neonate with fetal lung interstitial tumor. *Genes Chromosomes Cancer*, 53(10), 865-74 (2014)  
DOI: 10.1002/gcc.22199
18. F. Czubayko, A. M. Schulte, G. J. Berchem and A. Wellstein: Melanoma angiogenesis and metastasis modulated by ribozyme targeting of the secreted growth factor pleiotrophin. *Proc Natl Acad Sci U S A*, 93(25), 14753-8 (1996)  
DOI: 10.1073/pnas.93.25.14753
19. M. Ladanyi: The NPM/ALK gene fusion in the pathogenesis of anaplastic large cell lymphoma. *Cancer Surv*, 30, 59-75 (1997)
20. E. Ardini, P. Magnaghi, P. Orsini, A. Galvani and M. Menichincheri: Anaplastic Lymphoma Kinase: role in specific tumours, and development of small molecule inhibitors for cancer therapy. *Cancer Lett*, 299(2), 81-94 (2010)  
DOI: 10.1016/j.canlet.2010.09.001
21. K. Pulford, L. Lamant, E. Espinos, Q. Jiang, L. Xue, F. Turturro, G. Delsol and S. W. Morris: The emerging normal and disease-related roles of anaplastic lymphoma kinase. *Cell Mol Life Sci*, 61(23), 2939-53 (2004)  
DOI: 10.1007/s00018-004-4275-9
22. H. M. Amin and R. Lai: Pathobiology of ALK+ anaplastic large-cell lymphoma. *Blood*, 110(7), 2259-67 (2007)  
DOI: 10.1182/blood-2007-04-060715
23. R. Y. Bai, P. Dieter, C. Peschel, S. W. Morris and J. Duyster: Nucleophosmin-anaplastic lymphoma kinase of large-cell anaplastic lymphoma is a constitutively active tyrosine kinase that utilizes phospholipase C-gamma to mediate its mitogenicity. *Mol Cell Biol*, 18(12), 6951-61 (1998)
24. M. Marzec, M. Kasprzycka, A. Ptaszniak, P. Włodarski, Q. Zhang, N. Odum and M. A. Wasik: Inhibition of ALK enzymatic activity in T-cell lymphoma cells induces apoptosis and suppresses proliferation and STAT3 phosphorylation independently of Jak3. *Lab Invest*, 85(12), 1544-54 (2005)  
DOI: 10.1038/labinvest.3700348.
25. D. Polgar, C. Leisser, S. Maier, S. Strasser, B. Ruger, M. Dettke, M. Khorchide, I. Simonitsch, C. Cerni and G. Krupitza: Truncated ALK derived from chromosomal translocation t(2;5)(p23;q35) binds to the SH3 domain of p85-PI3K. *Mutat Res*, 570(1), 9-15 (2005)  
DOI: 10.1016/j.mrfmmm.2004.09.011
26. A. Slupianek, M. Nieborowska-Skorska, G. Hoser, A. Morrione, M. Majewski, L. Xue, S. W. Morris, M. A. Wasik and T. Skorski: Role of phosphatidylinositol 3-kinase-Akt pathway in nucleophosmin/anaplastic lymphoma kinase-mediated lymphomagenesis. *Cancer Res*, 61(5), 2194-9 (2001)
27. R. Y. Bai, T. Ouyang, C. Miething, S. W. Morris, C. Peschel and J. Duyster: Nucleophosmin-anaplastic lymphoma kinase associated with anaplastic large-cell lymphoma activates the phosphatidylinositol 3-kinase/Akt antiapoptotic signaling pathway. *Blood*, 96(13), 4319-27 (2000)
28. M. Nieborowska-Skorska, A. Slupianek, L. Xue, Q. Zhang, P. N. Raghunath, G. Hoser, M. A. Wasik, S. W. Morris and T. Skorski: Role of signal transducer and activator of transcription 5 in nucleophosmin/anaplastic lymphoma kinase-mediated malignant transformation of lymphoid cells. *Cancer Res*, 61(17), 6517-23 (2001)
29. A. Zamo, R. Chiarle, R. Piva, J. Howes, Y. Fan, M. Chilosi, D. E. Levy and G. Inghirami: Anaplastic lymphoma kinase (ALK) activates Stat3 and protects hematopoietic cells from cell death. *Oncogene*, 21(7), 1038-47 (2002)  
DOI: 10.1038/sj.onc.1205152
30. Q. Zhang, P. N. Raghunath, L. Xue, M. Majewski, D. F. Carpenteri, N. Odum, S. Morris, T. Skorski and M. A. Wasik: Multilevel dysregulation of STAT3 activation in anaplastic lymphoma kinase-positive T/null-cell lymphoma. *J Immunol*, 168(1), 466-74 (2002)  
DOI: 10.4049/jimmunol.168.1.466
31. R. Chiarle, C. Voena, C. Ambrogio, R. Piva and G. Inghirami: The anaplastic lymphoma kinase in the pathogenesis of cancer. *Nat Rev Cancer*, 8(1), 11-23 (2008)  
DOI: 10.1038/nrc2291

32. P. Minoo and H. Y. Wang: ALK-immunoreactive neoplasms. *Int J Clin Exp Pathol*, 5(5), 397-410 (2012)
33. K. V. Foyil and N. L. Bartlett: Brentuximab vedotin and crizotinib in anaplastic large-cell lymphoma. *Cancer J*, 18(5), 450-6 (2012) DOI: 10.1097/PPO.0b013e31826aef4a
34. C. J. Tartari, L. Scapozza and C. Gambacorti-Passerini: The ALK gene, an attractive target for inhibitor development. *Curr Top Med Chem*, 11(11), 1406-19 (2011) DOI: 10.2174/156802611795589593
35. J. A. Lee, L. Bubendorf, R. Stahel and S. Peters: Testing for anaplastic lymphoma kinase rearrangement to target crizotinib therapy: oncology, pathology and health economic perspectives. *Expert Rev Anticancer Ther*, 13(5), 625-36 (2013) DOI: 10.1586/era.13.42
36. L. Mologni: Inhibitors of the anaplastic lymphoma kinase. *Expert Opin Investig Drugs*, 21(7), 985-94 (2012) DOI: 10.1517/13543784.2012.690031
37. M. Ceccon, L. Mologni, W. Bisson, L. Scapozza and C. Gambacorti-Passerini: Crizotinib-resistant NPM-ALK mutants confer differential sensitivity to unrelated Alk inhibitors. *Mol Cancer Res*, 11(2), 122-32 (2013) DOI: 10.1158/1541-7786.MCR-12-0569
38. B. Hallberg and R. H. Palmer: Mechanistic insight into ALK receptor tyrosine kinase in human cancer biology. *Nat Rev Cancer*, 13(10), 685-700 (2013) DOI: 10.1038/nrc3580
39. J. M. Heuckmann, M. Holzel, M. L. Sos, S. Heynck, H. Balke-Want, M. Koker, M. Peifer, J. Weiss, C. M. Lovly, C. Grutter, D. Rauh, W. Pao and R. K. Thomas: ALK mutations conferring differential resistance to structurally diverse ALK inhibitors. *Clin Cancer Res*, 17(23), 7394-401 (2011) DOI: 10.1158/1078-0432.CCR-11-1648
40. D. Huang, D. W. Kim, A. Kotsakis, S. Deng, P. Lira, S. N. Ho, N. V. Lee, P. Vizcarra, J. Q. Cao, J. G. Christensen, T. M. Kim, J. M. Sun, J. S. Ahn, M. J. Ahn, K. Park and M. Mao: Multiplexed deep sequencing analysis of ALK kinase domain identifies resistance mutations in relapsed patients following crizotinib treatment. *Genomics*, 102(3), 157-62 (2013) DOI: 10.1016/j.ygeno.2013.02.006
41. C. Voena and R. Chiarle: The battle against ALK resistance: successes and setbacks. *Expert Opin Investig Drugs*, 21(12), 1751-4 (2012) DOI: 10.1517/13543784.2012.717930
42. R. Piva, R. Chiarle, A. D. Manazza, R. Taulli, W. Simmons, C. Ambrogio, V. D'Escamard, E. Pellegrino, C. Ponzetto, G. Palestro and G. Inghirami: Ablation of oncogenic ALK is a viable therapeutic approach for anaplastic large-cell lymphomas. *Blood*, 107(2), 689-97 (2006) DOI: 10.1182/blood-2005-05-2125
43. A. M. Coluccia, R. H. Gunby, C. J. Tartari, L. Scapozza, C. Gambacorti-Passerini and L. Passoni: Anaplastic lymphoma kinase and its signalling molecules as novel targets in lymphoma therapy. *Expert Opin Ther Targets*, 9(3), 515-32 (2005) DOI: 10.1517/14728222.9.3.515
44. K. Van Roosbroeck, J. Cools, D. Dierickx, J. Thomas, P. Vandenberghe, M. Stul, J. Delabie, C. De Wolf-Peeters, P. Marynen and I. Wlodarska: ALK-positive large B-cell lymphomas with cryptic SEC31A-ALK and NPM1-ALK fusions. *Haematologica*, 95(3), 509-13 (2010) DOI: 10.3324/haematol.2009.014761
45. A. K. Murugan and M. Xing: Anaplastic thyroid cancers harbor novel oncogenic mutations of the ALK gene. *Cancer Res*, 71(13), 4403-11 (2011) DOI: 10.1158/0008-5472.CAN-10-4041
46. L. Passoni, L. Longo, P. Collini, A. M. Coluccia, F. Bozzi, M. Podda, A. Gregorio, C. Gambini, A. Garaventa, V. Pistoia, F. Del Grosso, G. P. Tonini, M. Cheng, C. Gambacorti-Passerini, A. Anichini, F. Fossati-Bellani, M. Di Nicola and R. Luksch: Mutation-independent anaplastic lymphoma kinase overexpression in poor prognosis neuroblastoma patients. *Cancer Res*, 69(18), 7338-46 (2009) DOI: 10.1158/0008-5472.CAN-08-4419
47. R. H. Gunby, C. J. Tartari, F. Porchia, A. Donella-Deana, L. Scapozza and C. Gambacorti-Passerini: An enzyme-linked immunosorbent assay to screen for inhibitors of the oncogenic anaplastic lymphoma kinase. *Haematologica*, 90(7), 988-90 (2005)
48. F. Turturro, M. D. Arnold, A. Y. Frist and K. Fulford: Model of inhibition of the NPM-ALK

- kinase activity by herbimycin A. *Clin Cancer Res*, 8(1), 240-5 (2002)
49. H. Y. Zou, Q. Li, J. H. Lee, M. E. Arango, S. R. McDonnell, S. Yamazaki, T. B. Koudriakova, G. Alton, J. J. Cui, P. P. Kung, M. D. Nambu, G. Los, S. L. Bender, B. Mroczkowski and J. G. Christensen: An orally available small-molecule inhibitor of c-Met, PF-2341066, exhibits cytoreductive antitumor efficacy through antiproliferative and antiangiogenic mechanisms. *Cancer Res*, 67(9), 4408-17 (2007)  
DOI: 10.1158/0008-5472.CAN-06-4443
50. E. L. Kwak, D. R. Camidge, J. Clark, G. I. Shapiro, R. G. Maki, M. J. Ratain, B. Solomon, Y. Bang, S. Ou, R. Salgia: Clinical activity observed in a phase I dose escalation trial of an oral c-MET and ALK inhibitor, PF-02341066. *J Clin Oncol* 27(Suppl)(148s) (2009)
51. E. L. Kwak, Y. J. Bang, D. R. Camidge, A. T. Shaw, B. Solomon, R. G. Maki, S. H. Ou, B. J. Dezube, P. A. Janne, D. B. Costa, M. Varella-Garcia, W. H. Kim, T. J. Lynch, P. Fidias, H. Stubbs, J. A. Engelmann, L. V. Sequist, W. Tan, L. Gandhi, M. Mino-Kenudson, G. C. Wei, S. M. Shreeve, M. J. Ratain, J. Settleman, J. G. Christensen, D. A. Haber, K. Wilner, R. Salgia, G. I. Shapiro, J. W. Clark and A. J. Iafrate: Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med*, 363(18), 1693-703 (2010)  
DOI: 10.1056/NEJMoa1006448
52. D. R. Camidge, Y. J. Bang, E. L. Kwak, A. J. Iafrate, M. Varella-Garcia, S. B. Fox, G. J. Riely, B. Solomon, S. H. Ou, D. W. Kim, R. Salgia, P. Fidias, J. A. Engelmann, L. Gandhi, P. A. Janne, D. B. Costa, G. I. Shapiro, P. Lorusso, K. Ruffner, P. Stephenson, Y. Tang, K. Wilner, J. W. Clark and A. T. Shaw: Activity and safety of crizotinib in patients with ALK-positive non-small-cell lung cancer: updated results from a phase 1 study. *Lancet Oncol*, 13(10), 1011-9 (2012)  
DOI: 10.1016/S1470-2045(12)70344-3
53. P. Martin Martorell, M. Huerta Alvaro, M. A. Solis Salguero and A. Insa Molla: Crizotinib and renal insufficiency: a case report and review of the literature. *Lung Cancer*, 84(3), 310-3 (2014)  
DOI: 10.1016/j.lungcan.2014.03.001
54. J. Park, K. Yoshida, C. Kondo, J. Shimizu, Y. Horio, S. Hijioka and T. Hida: Crizotinib-induced esophageal ulceration: a novel adverse event of crizotinib. *Lung Cancer*, 81(3), 495-6 (2013)  
DOI: 10.1016/j.lungcan.2013.06.017
55. N. Srivastava, P. A. VanderLaan, C. P. Kelly and D. B. Costa: Esophagitis: a novel adverse event of crizotinib in a patient with ALK-positive non-small-cell lung cancer. *J Thorac Oncol*, 8(3), e23-4 (2013)  
DOI: 10.1097/JTO.0b013e31827e2451
56. A. Ono, T. Takahashi, T. Oishi, T. Sugino, H. Akamatsu, T. Shukuya, T. Taira, H. Kenmotsu, T. Naito, H. Murakami, T. Nakajima, M. Endo and N. Yamamoto: Acute lung injury with alveolar hemorrhage as adverse drug reaction related to crizotinib. *J Clin Oncol*, 31(26), e417-9 (2013)  
DOI: 10.1200/JCO.2012.47.1110
57. A. Tamiya, I. Okamoto, M. Miyazaki, S. Shimizu, M. Kitaichi and K. Nakagawa: Severe acute interstitial lung disease after crizotinib therapy in a patient with EML4-ALK-positive non-small-cell lung cancer. *J Clin Oncol*, 31(1), e15-7 (2013)  
DOI: 10.1200/JCO.2012.43.3730
58. M. Tachihara, K. Kobayashi, Y. Ishikawa, S. Hori, D. Tamura, H. Otera, Y. Funada and Y. Nishimura: Successful Crizotinib Rechallenge After Crizotinib-induced Interstitial Lung Disease. *Jpn J Clin Oncol*, 44(8), 762-4 (2014)  
DOI: 10.1093/jjco/hyu074
59. A. T. Shaw, D. W. Kim, K. Nakagawa, T. Seto, L. Crino, M. J. Ahn, T. De Pas, B. Besse, B. J. Solomon, F. Blackhall, Y. L. Wu, M. Thomas, K. J. O'Byrne, D. Moro-Sibilot, D. R. Camidge, T. Mok, V. Hirsh, G. J. Riely, S. Iyer, V. Tassell, A. Polli, K. D. Wilner and P. A. Janne: Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med*, 368(25), 2385-94 (2013)  
DOI: 10.1056/NEJMoa1214886
60. C. Gambacorti-Passerini, C. Messa and E. M. Pogliani: Crizotinib in anaplastic large-cell lymphoma. *N Engl J Med*, 364(8), 775-6
61. C. Gambacorti Passerini, F. Farina, A. Stasia, S. Redaelli, M. Ceccon, L. Mologni, C. Messa, L. Guerra, G. Giudici, E. Sala, L. Mussolin, D. Deeren, M. H. King, M. Steurer, R. Ordemann, A. M. Cohen, M. Grube, L. Bernard, G.

- Chiriano, L. Antolini and R. Piazza: Crizotinib in advanced, chemoresistant anaplastic lymphoma kinase-positive lymphoma patients. *J Natl Cancer Inst*, 106(2), djt378 (2014)
62. C. Gambacorti-Passerini, Horibe, Braiteh, Huang, Shi, Taylor, Brega, Paolini, Selaru and T. M. Kim: Safety and Clinical Activity Of Crizotinib In Patients With ALK-Rearranged Hematologic Malignancies. In: *ASH annual meeting, New Orleans 2013 (abs 4342)*. (2013)
63. Y. P. Mosse, M. S. Lim, S. D. Voss, K. Wilner, K. Ruffner, J. Laliberte, D. Rolland, F. M. Balis, J. M. Maris, B. J. Weigel, A. M. Ingle, C. Ahern, P. C. Adamson and S. M. Blaney: Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: a Children's Oncology Group phase 1 consortium study. *Lancet Oncol*, 14(6), 472-80 (2013)  
DOI: 10.1016/S1470-2045(13)70095-0
64. J. M. Cleary, S. Rodig, P. M. Barr, A. B. Shinagare, J. W. Clark, G. I. Shapiro and P. Armand: Crizotinib as salvage and maintenance with allogeneic stem cell transplantation for refractory anaplastic large cell lymphoma. *J Natl Compr Canc Netw*, 12(3), 323-6; quiz 326 (2014)
65. S. V. Jacob, J. D. Reith, A. Y. Kojima, W. D. Williams, C. Liu and L. Vila Duckworth: An Unusual Case of Systemic Inflammatory Myofibroblastic Tumor with Successful Treatment with ALK-Inhibitor. *Case Rep Pathol*, 2014, 470340 (2014)
66. S. Kimbara, K. Takeda, H. Fukushima, T. Inoue, H. Okada, Y. Shibata, U. Katsushima, A. Tsuya, S. Tokunaga, H. Daga and T. Okuno: A Case Report of Epithelioid Inflammatory Myofibroblastic Sarcoma with RANBP2-ALK Fusion Gene Treated with the ALK Inhibitor, Crizotinib. *Jpn J Clin Oncol* (2014)
67. S. Kim, J. Sun, Y. Go, K. Park, C. Baek, Y. Choi, Y. Son and M. J. Ahn: The presence of ALK translocation in sarcomatoid carcinoma of head and neck and treatment effect of crizotinib: A case series. *J Clin Oncol* 32, 2014 (suppl; abstr e17048), Abstract: e17048 (2014)
68. F. Farina, A. Stasia and C. Gambacorti Passerini: Developments in anaplastic large-cell lymphoma: targeting the anaplastic lymphoma kinase. *Blood and Lymphatic Cancer: Targets and Therapy*, 4, 69-79 (2014)
69. R. C. Doebele, A. B. Pilling, D. L. Aisner, T. G. Kutateladze, A. T. Le, A. J. Weickhardt, K. L. Kondo, D. J. Linderman, L. E. Heasley, W. A. Franklin, M. Varella-Garcia and D. R. Camidge: Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin Cancer Res*, 18(5), 1472-82 (2012)  
DOI: 10.1158/1078-0432.CCR-11-2906
70. R. Katayama, A. T. Shaw, T. M. Khan, M. Mino-Kenudson, B. J. Solomon, B. Halmos, N. A. Jessop, J. C. Wain, A. T. Yeo, C. Benes, L. Drew, J. C. Saeh, K. Crosby, L. V. Sequist, A. J. Iafrate and J. A. Engelman: Mechanisms of acquired crizotinib resistance in ALK-rearranged lung Cancers. *Sci Transl Med*, 4(120), 120ra17 (2012)
71. Y. P. Mosse, A. Wood and J. M. Maris: Inhibition of ALK signaling for cancer therapy. *Clin Cancer Res*, 15(18), 5609-14 (2009)  
DOI: 10.1158/1078-0432.CCR-08-2762
72. Y. L. Choi, M. Soda, Y. Yamashita, T. Ueno, J. Takashima, T. Nakajima, Y. Yatabe, K. Takeuchi, T. Hamada, H. Haruta, Y. Ishikawa, H. Kimura, T. Mitsudomi, Y. Tanio and H. Mano: EML4-ALK mutations in lung cancer that confer resistance to ALK inhibitors. *N Engl J Med*, 363(18), 1734-9 (2010)  
DOI: 10.1056/NEJMoa1007478
73. K. Esfahani, J. S. Agulnik and V. Cohen: A Systemic Review of Resistance Mechanisms and Ongoing Clinical Trials in ALK-Rearranged Non-Small Cell Lung Cancer. *Front Oncol*, 4, 174 (2014)
74. B. Hallberg and R. H. Palmer: Crizotinib-latest champion in the cancer wars? *N Engl J Med*, 363(18), 1760-2 (2010)
75. D. R. Camidge and R. C. Doebele: Treating ALK-positive lung cancer--early successes and future challenges. *Nat Rev Clin Oncol*, 9(5), 268-77 (2012)  
DOI: 10.1038/nrclinonc.2012.43
76. J. Gainor, L. Friboulet, R. Katayama, M. Awad, E. Lockerman, K. Schultz, S. Mahmood, M. Nishio, N. Yanagitani, L. Sequist, M. Mino-Kenudson, J. Engelman and A. T. Shaw: Evolution of resistance in ALK-positive patients treated with ALK tyrosine kinase

- inhibitors (TKIs). *J Clin Oncol* 32:5s, 2014 (suppl; abstr 8031) (2014)
77. S. H. Ignatius Ou, M. Azada, D. J. Hsiang, J. M. Herman, T. S. Kain, C. Siwak-Tapp, C. Casey, J. He, S. M. Ali, S. J. Klempner and V. A. Miller: Next-generation sequencing reveals a Novel NSCLC ALK F1174V mutation and confirms ALK G1202R mutation confers high-level resistance to alectinib (CH5424802/ RO5424802) in ALK-rearranged NSCLC patients who progressed on crizotinib. *J Thorac Oncol*, 9(4), 549-53 (2014)  
DOI: 10.1097/JTO.0000000000000094
  78. R. Schwab, I. Petak, M. Kollar, F. Pinter, E. Varkondi, A. Kohanka, H. Barti-Juhasz, J. Schonleber, D. Brauswetter, L. Kopper and L. Urban: Major partial response to crizotinib, a dual MET/ALK inhibitor, in a squamous cell lung (SCC) carcinoma patient with de novo c-MET amplification in the absence of ALK rearrangement. *Lung Cancer* (2013)
  79. N. Yamaguchi, A. R. Lucena-Araujo, S. Nakayama, L. L. de Figueiredo-Pontes, D. A. Gonzalez, H. Yasuda, S. Kobayashi and D. B. Costa: Dual ALK and EGFR inhibition targets a mechanism of acquired resistance to the tyrosine kinase inhibitor crizotinib in ALK rearranged lung cancer. *Lung Cancer* (2014)
  80. R. C. Doebele: A nice problem to have: when ALK inhibitor therapy works better than expected. *J Thorac Oncol*, 9(4), 433-5 (2014)  
DOI: 10.1097/JTO.0000000000000124
  81. A. Kruczynski, G. Delsol, C. Laurent, P. Brousset and L. Lamant: Anaplastic lymphoma kinase as a therapeutic target. *Expert Opin Ther Targets*, 16(11), 1127-38 (2012)  
DOI: 10.1517/14728222.2012.719498
  82. D. R. Camidge, W. Pao and L. V. Sequist: Acquired resistance to TKIs in solid tumours: learning from lung cancer. *Nat Rev Clin Oncol*, 11(8), 473-81 (2014)  
DOI: 10.1038/nrclinonc.2014.104
  83. C. M. Lovly and A. T. Shaw: Molecular pathways: resistance to kinase inhibitors and implications for therapeutic strategies. *Clin Cancer Res*, 20(9), 2249-56 (2014)  
DOI: 10.1158/1078-0432.CCR-13-1610
  84. T. H. Marsilje, W. Pei, B. Chen, W. Lu, T. Uno, Y. Jin, T. Jiang, S. Kim, N. Li, M. Warmuth, Y. Sarkisova, F. Sun, A. Steffy, A. C. Pferdekamper, A. G. Li, S. B. Joseph, Y. Kim, B. Liu, T. Tuntland, X. Cui, N. S. Gray, R. Steensma, Y. Wan, J. Jiang, G. Chopiuk, J. Li, W. P. Gordon, W. Richmond, K. Johnson, J. Chang, T. Groessl, Y. Q. He, A. Phimister, A. Aycinena, C. C. Lee, B. Bursulaya, D. S. Karanewsky, H. M. Seidel, J. L. Harris and P. Y. Michelllys: Synthesis, Structure-Activity Relationships, and *in vivo* Efficacy of the Novel Potent and Selective Anaplastic Lymphoma Kinase (ALK) Inhibitor 5-Chloro-N2-(2-isopropoxy-5-methyl-4-(piperidin-4-yl)phenyl)-N4-(2-(isopropylsulfonyl)phenyl)pyrimidine-2,4-diamine (LDK378) Currently in Phase 1 and Phase 2 Clinical Trials. *J Med Chem*, 56(14), 5675-90 (2013)  
DOI: 10.1021/jm400402q
  85. J. F. Vansteenkiste: Ceritinib for treatment of ALK-rearranged advanced non-small-cell lung cancer. *Future Oncol*, 1-15 (2014)
  86. L. Fribolet, N. Li, R. Katayama, C. C. Lee, J. F. Gainor, A. S. Crystal, P. Y. Michelllys, M. M. Awad, N. Yanagitani, S. Kim, A. C. Pferdekamper, J. Li, S. Kasibhatla, F. Sun, X. Sun, S. Hua, P. McNamara, S. Mahmood, E. L. Lockerman, N. Fujita, M. Nishio, J. L. Harris, A. T. Shaw and J. A. Engelman: The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discov*, 4(6), 662-73 (2014)  
DOI: 10.1158/2159-8290.CD-13-0846
  87. M. P.-Y. Li N., Sungjon K., Culazzo Pferdekamper A., Li J., Kasibhatla S., Tompkins C. S., Steffy A., Li A., Sun F., Sun X., Hua S., Tiedt R., Sarkisova Y., Marsilje T. H., McNamara P., Harris J.: Activity of a potent and selective phase I ALK inhibitor LDK378 in naïve and crizotinib-resistant preclinical tumor models. In: AACR-NCI-EORTC International Conference: Molecular Targets and Cancer Therapeutics., San Francisco (2011)
  88. R. Mehra, D. R. Camidge, S. Sharma, E. Felip, D. Shao-Weng Tan, J. F. Vansteenkiste, T.M. De Pas, D.W. Kim, A. Santoro, G. Liu, M. Goldwasser, D. Dai, M. Radona, A. Boral, A. Tsang Shaw: First-in-human phase I study of the ALK inhibitor LDK378 in advanced solid tumors. *J Clin Oncol* 30 (suppl; abstr 3007). (2012)
  89. J. Chen, C. Jiang and S. Wang: LDK378: a promising anaplastic lymphoma kinase (ALK) inhibitor. *J Med Chem*, 56(14), 5673-4 (2013)  
DOI: 10.1021/jm401005u

90. A. T. Shaw, D. W. Kim, R. Mehra, D. S. Tan, E. Felip, L. Q. Chow, D. R. Camidge, J. Vansteenkiste, S. Sharma, T. De Pas, G. J. Riely, B. J. Solomon, J. Wolf, M. Thomas, M. Schuler, G. Liu, A. Santoro, Y. Y. Lau, M. Goldwasser, A. L. Boral and J. A. Engelman: Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med*, 370(13), 1189-97 (2014)  
DOI: 10.1056/NEJMoa1311107
91. D. Kim, R. Mehra, D. Tan, W. Felip, L. Chow, D. Camidge, J. Vansteenkiste, S. Sharma, T. De Pas, G. Riely, B. Solomon, J. Wolf, M. Thomas, M. Schuler, G. Liu, A. Santoro, M. Geraldes, A. Boral, A. Yovine and A. T. Shaw: Ceritinib in advanced anaplastic lymphoma kinase (ALK)-rearranged (ALK+) non-small cell lung cancer (NSCLC): Results of the ASCEND-1 trial. *J Clin Oncol* 32:5s, 2014 (suppl; abstr 8003<sup>A</sup>) (2014)
92. S. Dhillon and M. Clark: Ceritinib: first global approval. *Drugs*, 74(11), 1285-91 (2014)  
DOI: 10.1007/s40265-014-0251-3
93. T. Kodama, T. Tsukaguchi, M. Yoshida, O. Kondoh and H. Sakamoto: Selective ALK inhibitor alectinib with potent antitumor activity in models of crizotinib resistance. *Cancer Lett*, 351(2), 215-21 (2014)  
DOI: 10.1016/j.canlet.2014.05.020
94. H. Sakamoto, T. Tsukaguchi, S. Hiroshima, T. Kodama, T. Kobayashi, T. A. Fukami, N. Oikawa, T. Tsukuda, N. Ishii and Y. Aoki: CH5424802, a selective ALK inhibitor capable of blocking the resistant gatekeeper mutant. *Cancer Cell*, 19(5), 679-90 (2011)  
DOI: 10.1016/j.ccr.2011.04.004
95. T. Seto, K. Kiura, M. Nishio, K. Nakagawa, M. Maemondo, A. Inoue, T. Hida, N. Yamamoto, H. Yoshioka, M. Harada, Y. Ohe, N. Nogami, K. Takeuchi, T. Shimada, T. Tanaka and T. Tamura: CH5424802 (RO5424802) for patients with ALK-rearranged advanced non-small-cell lung cancer (AF-001JP study): a single-arm, open-label, phase 1-2 study. *Lancet Oncol*, 14(7), 590-8 (2013)  
DOI: 10.1016/S1470-2045(13)70142-6
96. K. Nakagawa, T. Hida, T. Seto, M. Satouchi, M. Nishio, K. Hotta, H. Murakami, Y. Ohe, K. Takeda, M. Tatsuno, N. Yoshikawa, T. Tanaka and T. Tamura: Antitumor activity of alectinib (CH5424802/RO5424802) for ALK-rearranged NSCLC with or without prior crizotinib treatment in bioequivalence study. *J Clin Oncol* 32:5s, 2014 (suppl; abstr 8103) (2014)
97. R. Katayama, T. M. Khan, C. Benes, E. Lifshits, H. Ebi, V. M. Rivera, W. C. Shakespeare, A. J. Iafrate, J. A. Engelman and A. T. Shaw: Therapeutic strategies to overcome crizotinib resistance in non-small cell lung cancers harboring the fusion oncogene EML4-ALK. *Proc Natl Acad Sci U S A*, 108(18), 7535-40 (2011)  
DOI: 10.1073/pnas.1019559108
98. S. Gettinger, G.J. Weiss, R. Salgia, L. Bazhenova, N.I. Narasimhan, D.J. Dorer, V. Rivera, J. Zhang, T. Clackson, F.G. Haluska, A. Shaw, R. Camidge: A first-in-human dose-finding study of the ALK/EGFR inhibitor AP26113 in patients with advanced malignancies. In: *37th Annual ESMO Conference*. Vienna (2012)
99. S. Gettinger, L. Bazhenova, R. Salgia, C. Langer, K. Gold, R. Rosell, A. T. Shaw, G. Weiss, N. Narasimhan, D. Dorer, V. Rivera, T. Clackson, F. Haluska and D. Camidge: Updated efficacy and safety of the ALK inhibitor AP26113 in patients (pts) with advanced malignancies, including ALK+ non-small cell lung cancer (NSCLC). *J Clin Oncol* 32:5s, 2014 (suppl; abstr 8047) (2014)
100. G. V. Georgakis, Y. Li, G. Z. Rassidakis, L. J. Medeiros and A. Younes: The HSP90 inhibitor 17-AAG synergizes with doxorubicin and U0126 in anaplastic large cell lymphoma irrespective of ALK expression. *Exp Hematol*, 34(12), 1670-9 (2006)  
DOI: 10.1016/j.exphem.2006.07.002
101. J. Sang, J. Acquaviva, J. C. Friedland, D. L. Smith, M. Sequeira, C. Zhang, Q. Jiang, L. Xue, C. M. Lovly, J. P. Jimenez, A. T. Shaw, R. C. Doebele, S. He, R. C. Bates, D. R. Camidge, S. W. Morris, I. El-Hariry and D. A. Proia: Targeted inhibition of the molecular chaperone Hsp90 overcomes ALK inhibitor resistance in non-small cell lung cancer. *Cancer Discov*, 3(4), 430-43 (2013)  
DOI: 10.1158/2159-8290.CD-12-0440
102. E. Normant, G. Paez, K. A. West, A. R. Lim, K. L. Slocum, C. Tunkey, J. McDougall, A. A. Wylie, K. Robison, K. Caliri, V. J. Palombella and C. C. Fritz: The Hsp90 inhibitor IPI-504

- rapidly lowers EML4-ALK levels and induces tumor regression in ALK-driven NSCLC models. *Oncogene*, 30(22), 2581-6 (2011)  
DOI: 10.1038/onc.2010.625
103. L. V. Sequist, S. Gettinger, N. N. Senzer, R. G. Martins, P. A. Janne, R. Lilenbaum, J. E. Gray, A. J. Iafrate, R. Katayama, N. Hafeez, J. Sweeney, J. R. Walker, C. Fritz, R. W. Ross, D. Grayzel, J. A. Engelman, D. R. Borger, G. Paez and R. Natale: Activity of IPI-504, a novel heat-shock protein 90 inhibitor, in patients with molecularly defined non-small-cell lung cancer. *J Clin Oncol*, 28(33), 4953-60 (2010)  
DOI: 10.1200/JCO.2010.30.8338
104. M. A. Socinski, J. Goldman, I. El-Hariry, M. Koczywas, V. Vukovic, L. Horn, E. Paschold, R. Salgia, H. West, L. V. Sequist, P. Bonomi, J. Brahmer, L. C. Chen, A. Sandler, C. P. Belani, T. Webb, H. Harper, M. Huberman, S. Ramalingam, K. K. Wong, F. Teofilovici, W. Guo and G. I. Shapiro: A multicenter phase II study of ganetespib monotherapy in patients with genotypically defined advanced non-small cell lung cancer. *Clin Cancer Res*, 19(11), 3068-77 (2013)  
DOI: 10.1158/1078-0432.CCR-12-3381
105. E. Felip, E. Carcereny, F. Barlesi, L. Gandhi, L. Sequist and S. Kim: Phase II activity of the HSP90 inhibitor AUY922 in patients with ALK-rearranged or EGFR-mutated advanced non-small-cell lung cancer. *Ann Oncol*, 23 (ix152):Abstr.4380 (2012)
106. L. Horn, J. Infante, G. Blumenschein, H. Wakelee, H. Arkenau, G. Dukart, C. Liang, K. Harrow, J. Gibbons, C. Lovly and W. Pao: A phase I trial of X-396, a novel ALK inhibitor, in patients with advanced solid tumors. *J Clin Oncol* 32:5s, 2014 (suppl; abstr 8030^) (2014)
107. C. M. Lovly, J. M. Heuckmann, E. de Stanchina, H. Chen, R. K. Thomas, C. Liang and W. Pao: Insights into ALK-driven cancers revealed through development of novel ALK tyrosine kinase inhibitors. *Cancer Res*, 71(14), 4920-31 (2011)  
DOI: 10.1158/0008-5472.CAN-10-3879
108. X. Lu and K. Ding: Novel anaplastic lymphoma kinase inhibitors targeting clinically acquired resistance. *J Med Chem*, 57(4), 1167-9 (2014)  
DOI: 10.1021/jm500178r
109. S. Yamazaki, J. Lam, H. Y. Zou, H. Wang, T. Smeal and P. Vicini: Translational Pharmacokinetic-Pharmacodynamic Modeling for An Orally Available Novel Inhibitor of Anaplastic Lymphoma Kinase and c-Ros Oncogene 1. *J Pharmacol Exp Ther* (2014)
110. G. Weiss, J. Sachdev, J. Infante, M. Mita, R. Natale, H. Arkenau, K. Wilcoxen, V. Kansra, H. Laken, L. Hughes, D. Brooks, R. Martell and S. Anthony: Phase (Ph) 1/2 study of TSR-011, a potent inhibitor of ALK and TRK, including crizotinib-resistant ALK mutations. *J Clin Oncol* 32, 2014 (suppl; abstr e19005) (2014)
111. S. K. George, D. Vishwamitra, R. Mansouri, P. Shi and H. M. Amin: The ALK inhibitor ASP3026 eradicates NPM-ALK(+) T-cell anaplastic large-cell lymphoma *in vitro* and in a systemic xenograft lymphoma model. *Oncotarget*, 5(14), 5750-63 (2014)
112. M. Mori, Y. Ueno, S. Konagai, H. Fushiki, I. Shimada, Y. Kondoh, R. Saito, K. Mori, N. Shindou, T. Soga, H. Sakagami, T. Furutani, H. Doihara, M. Kudoh and S. Kuromitsu: The selective anaplastic lymphoma receptor tyrosine kinase inhibitor ASP3026 induces tumor regression and prolongs survival in non-small cell lung cancer model mice. *Mol Cancer Ther*, 13(2), 329-40 (2014)  
DOI: 10.1158/1535-7163.MCT-13-0395
113. M. Maitland, S. Ou, A. Tolcher, P. LoRusso, E. Bahceci, H. Ball, J. Park, G. Yuen, L. Koplowitz and T. Li: Safety, activity, and pharmacokinetics of an oral anaplastic lymphoma kinase (ALK) inhibitor, ASP3026, observed in a "fast follower" phase 1 trial design. *J Clin Oncol* 32:5s, 2014 (suppl; abstr 2624^) (2014)
114. F. De Braud, L. Pilla, M. Niger, S. Damian, B. Bardazza, A. Martinetti, G. Pelosi, G. Marrapese, L. Palmeri, G. Cerea, E. Valtorta, S. Veronese, A. Sartore-Bianchi, E. Ardini, M. Martignoni, A. Galvani, P. Pearson, D. Luo, J. Freddo and S. Siena: Phase 1 open label, dose escalation study of RXDX101, an oral pan-trk, ROS1, and ALK inhibitor, in patients with advanced solid tumors with relevant molecular alterations. *J Clin Oncol* 32:5s, 2014 (suppl; abstr 2502) (2014)
115. C. Gambacorti-Passerini: Part I: Milestones in personalised medicine--imatinib. *Lancet Oncol*, 9(6), 600 (2008)  
DOI: 10.1016/S1470-2045(08)70152-9

116. G. Toyokawa and T. Seto: ALK Inhibitors: What Is the Best Way to Treat Patients With ALK Non-Small-Cell Lung Cancer? *Clin Lung Cancer* (2014)
117. M. Latif, A. Saeed and S. H. Kim: Journey of the ALK-inhibitor CH5424802 to phase II clinical trial. *Arch Pharm Res*, 36(9), 1051-4 (2013)  
DOI: 10.1007/s12272-013-0157-8
118. K. Nakagawa: A phase I/II study with a highly selective ALK inhibitor CH5424802 in ALK-positive non-small cell lung cancer (NSCLC) patients: Updated safety and efficacy results from AF-001JP. *J Clin Oncol* 31, 2013 (suppl; abstr 8033) (2013)
119. A. T. Shaw: Clinical activity of the ALK inhibitor LDK378 in advanced, ALK-positive NSCLC. *J Clin Oncol* 31, 2013 (suppl; abstr TPS8119 and abstract 8010) (2013)
120. R. Camidge: First-in-human dose-finding study of the ALK/EGFR inhibitor AP26113 in patients with advanced malignancies: Updated results. *J Clin Oncol* 31, 2013 (suppl; abstr 8031). (2013)
121. A. Patnaik: Pharmacokinetics and safety of an oral ALK inhibitor, ASP3026, observed in a phase I dose escalation trial. *J Clin Oncol* 31, 2013 (suppl; abstr 2602) (2013)
122. R. T. Bossi, M. B. Saccardo, E. Ardini, M. Menichincheri, L. Rusconi, P. Magnaghi, P. Orsini, N. Avanzi, A. L. Borgia, M. Nesi, T. Bandiera, G. Fogliatto and J. A. Bertrand: Crystal structures of anaplastic lymphoma kinase in complex with ATP competitive inhibitors. *Biochemistry*, 49(32), 6813-25 (2010)  
DOI: 10.1021/bi1005514
123. E. Grande, M. V. Bolos and E. Arriola: Targeting oncogenic ALK: a promising strategy for cancer treatment. *Mol Cancer Ther*, 10(4), 569-79 (2011)  
DOI: 10.1158/1535-7163.MCT-10-0615
124. U. McDermott, A. J. Iafrate, N. S. Gray, T. Shioda, M. Classon, S. Maheswaran, W. Zhou, H. G. Choi, S. L. Smith, L. Dowell, L. E. Ulkus, G. Kuhlmann, P. Greninger, J. G. Christensen, D. A. Haber and J. Settleman: Genomic alterations of anaplastic lymphoma kinase may sensitize tumors to anaplastic lymphoma kinase inhibitors. *Cancer Res*, 68(9), 3389-95 (2008)  
DOI: 10.1158/0008-5472.CAN-07-6186
125. T. R. Webb, J. Slavish, R. E. George, A. T. Look, L. Xue, Q. Jiang, X. Cui, W. B. Rentrop and S. W. Morris: Anaplastic lymphoma kinase: role in cancer pathogenesis and small-molecule inhibitor development for therapy. *Expert Rev Anticancer Ther*, 9(3), 331-56 (2009)  
DOI: 10.1586/14737140.9.3.331
126. M. Cheng, M. R. Quail, D. E. Gingrich, G. R. Ott, L. Lu, W. Wan, M. S. Albom, T. S. Angeles, L. D. Aimone, F. Cristofani, R. Machiorlatti, C. Abele, M. A. Ator, B. D. Dorsey, G. Inghirami and B. A. Ruggeri: CEP-28122, a highly potent and selective orally active inhibitor of anaplastic lymphoma kinase with antitumor activity in experimental models of human cancers. *Mol Cancer Ther*, 11(3), 670-9 (2012)  
DOI: 10.1158/1535-7163.MCT-11-0776
127. P. Sabbatini, S. Korenchuk, J. L. Rowand, A. Groy, Q. Liu, D. Leperi, C. Atkins, M. Dumble, J. Yang, K. Anderson, R. G. Kruger, R. R. Gontarek, K. R. Maksimchuk, S. Suravajjala, R. R. Lapierre, J. B. Shotwell, J. W. Wilson, S. D. Chamberlain, S. K. Rabindran and R. Kumar: GSK1838705A inhibits the insulin-like growth factor-1 receptor and anaplastic lymphoma kinase and shows antitumor activity in experimental models of human cancers. *Mol Cancer Ther*, 8(10), 2811-20 (2009)  
DOI: 10.1158/1535-7163.MCT-09-0423

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