Transforming growth factor-beta signaling in breast cancer

Ching-Fang Chang 1, Reyhan Westbrook 2, Jun Ma 1, Deliang Cao 1

¹ Department of Medical Microbiology, Immunology, and Cell Biology, SimmonsCooper Cancer Institute, Southern Illinois University School of Medicine. 913 N. Rutledge Street, Springfield, IL 62702, ² Department of Medicine, Geriatric Research, Southern Illinois University School of Medicine. 801 N. Rutledge Street, Springfield, IL 62794

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. TGF-β and its receptors
- 4. TGF-β signaling in mammary gland development
- 5. TGF-\(\beta\) signaling in mammary gland tumorigenesis
- 6. TGF-β pathway as a target for cancer therapy
- 7. Conclusion
- 8. References

1. ABSTRACT

Transforming growth factor- β (TGF- β) is a multifunctional polypeptide that regulates cell growth, differentiation, and extracellular matrix formation. Studies on genetically engineered animal models have demonstrated that TGF- β -mediated signaling pathway plays a critical role in both normal development and tumorigenesis of the breast. In pathogenesis of breast cancer, the role of TGF- β appears featured with growth-inhibitory effects at early stages of carcinogenesis, but aggressive oncogenesis with transition to more advanced malignant states. The TGF- β signaling pathway is also tissue-context and ligand content-dependent. Therein, therapeutic modulation of TGF- β signaling may be a multifactorial event.

2. INTRODUCTION

The transforming growth factor β (TGF- β) ligand family is composed of several multifunctional growth factors, including TGF- β (TGF- β 1/2/3 isoforms), activins, and bone morphogenetic proteins (BMPs) (1). The TGF- β was first discovered in relation to its capability of inducing a transformed morphology and clonogenic growth in soft agar of non-transformed, anchorage-dependent normal rat kidney cells and fibroblasts (2). However, subsequent studies demonstrated that TGF- β inhibits the growth of normal epithelial cells, demonstrating the complexity of the function (3). TGF- β ligands and receptors are expressed in nearly all types of cells, including epithelial, stromal, immune, lymphoid, and endothelial cells. In these cells, the TGF- β signaling regulates development, differentiation,

Table 1. TGF- β signaling molecules

Signaling Molecules	Function
Type I receptor (TβRI)	Bind to TGF-β/TβRII complex
	and phosphorylates Smad2/3
Type II receptor (TβRII)	Bind to TGF-β and activate TβRI
Type III receptor (TβRIII)	Unclear
Receptor-activated Smad (Smad2,	Signal transmission/DNA binding
Smad3)	
Common mediator Smad (Smad4)	Smad2/3 mediator
Inhibitory Smad (Smad6, Smad7)	TGF-β signaling inhibitors

References: 4, 5, 26, 55

extracellular matrix formation, cell cycle, angiogenesis, hematopoiesis, chemotaxis, and immune functions. Recent studies indicate that the TGF- β signaling demonstrates dual functions in mammary tumor development. TGF- β appears inhibitory at early stage of tumorigenesis, whereas tumor cells at advanced stages can evade antiproliferative control and undergo tumorigenic progression in response to TGF- β . In this review, discussion will focus on recent progress on the role of the TGF- β in mammary gland development and tumorigenesis, and its potential avenues toward cancer therapies.

3. TGF-B LIGANDS AND THEIR RECEPTORS

The TGF- β ligands consist of three isoforms, TGF- β 1/2/3 (1, 4). In the TGF- β signaling pathway, biological signals are transmitted via binding of the TGF- β ligands to two types of receptors, TGF- β type I (T β RI) and TGF- β type II (T β RII). The three TGF- β isoforms often elicit similar responses (5). Table 1 summarizes the key signaling molecules and their functions in this pathway.

Active TGF-β is produced through the maturation of a propeptide (6). For instance, TGF-β1 gene encodes a 390 amino acid polypeptide. At the dibasic cleavage site (residue 278), this polypeptide is cleaved into a latencyassociated peptide (LAP) and TGF-\(\beta\)1 during posttranslational modification (6, 7). LAP and TGF-\$1 form an inactive, noncovalently associated small latent TGF-B complex (SLC), which is secreted. Alternatively, this complex can be linked via a disulfide bond to a latent TGFβ binding protein (LTBP) for storage. This trimolecular aggregate is called large latent complex (LLC). LTBP secures the ligands in extracellular matrix (ECM) via a cross-link at the N-terminus catalyzed by transglutaminase (8). This latent complex primarily mediates the biological activity of TGF-β, and therefore, TGF-β message RNA levels do not usually reflect the protein production or activity (9-11). Latent TGF-β can be activated by heat, chaotropic agents, pH, proteases, urokinase plasminogen activator, integrins, fibronectin fibrils, and thrombospondin-1 (12-16).Thrombospondin-1 stimulates TGF-β release by the interaction of a motif (K412RFK415) in thrombospondin-1 with the LSKL motif in LAP at the amino terminus (15).

There are three TGF- β receptors identified thus far. T β RI and T β RII are transmembrane serine-threonine kinase receptors (5). TGF- β binds to T β RII and then recruits T β RI, forming the heterodimeric T β RII/T β RI complex with bound TGF- β (Figure 1). Formation of the heterodimer allows the transphosphorylation of specific serine and threonine residues in juxtamembrane segment of

TβRI by TβRII kinase, thereby activating TβRI kinase and initiating the signaling process (17). TGF- β receptor III (ΤβRIII), also known as betaglycan (18), is not signaling. The biological function of the TβRIII is unclear, but its short cytoplasmic domain and subsequent loss of intrinsic kinase activity may indicate its role as a co-receptor, enhancing ligand binding to TβRII (19).

Activated $T\beta RI$ kinase phosphorylates C-terminal serine residues of Smad2 and Smad3, two distinct proteins in a subclass of R-Smads (receptor-activated Smads) (20). Phosphorylated R-Smads bind to a common mediator Smad4, also referred to as Co-Smad, forming a functional trimeric protein complex. This complex is translocated into the nucleus, binds to the Smad-binding element in promoter of the target genes, recruits transcription factors, and controls the transcription of these genes (20, 21).

The activity of the TGF- β signaling pathway is regulated by a negative regulatory feedback loop mediated by inhibitory Smads (I-Smad), Smad6 and Smad7. I-Smads competitively bind to TGF- β /receptor complex and inhibit the phosphorylation of R-Smad (22). In addition, Smad7 can recruit phosphatases to dephosphorylate and thus inactivate the receptor complex.

Recent studies revealed that TGF- β mediates cell cycle through the RhoA/p160^{ROCK} signaling pathway (23). In mammary gland epithelial cells, by activating RhoA, TGF- β stimulates p160^{ROCK} translocation to the nucleus, alters the phosphorylation of the linker region of Smad2/3 at Ser²⁰³ and Ser²⁰⁷ residues, and triggers downstream gene expression, such as p15^{INK4B} and p21^{cip1/waf1}. This results in inhibition of pRb phosphorylation and cell cycle arrest or triggers apoptosis through the regulation of various proapoptotic and anti-apoptotic molecules: p53, Bad, Bax, Bik, Bcl-2, and Bcl-XL (24, 25).

4. $TGF-\beta$ SIGNALING IN MAMMARY GLAND DEVELOPMENT

The role of the TGF- β signaling pathway in cell proliferation and tissue formation is complicated. In mammalian breast development, the TGF- β signaling pathway is involved in establishing proper gland structures, maintaining epithelium in functionally undifferentiated status, and inducing apoptosis in the involuting gland (26).

In mammary glands, the expression of TGF- β isoforms is strictly regulated with the development of mammary glands. In mice, all three TGF- β isoforms are expressed in epithelium in all developmental stages of the breast, where TGF- β acts as an inhibitor of ductal elongation and branch formation. During puberty, the TGF- β activity is decreased in mammary ducts in response to proliferation signals, preparing for proliferation (27). In transgenic mice expressing TGF- β 1 s223/225 (a constitutively active TGF- β) specifically in mammary epithelial cells, ductal tree hypoplasia occurred (28). TGF- β also regulates development of the alveolar epithelium. Transgenic mice harboring active TGF- β were unable to lactate because of the inhibition of secretory epithelium

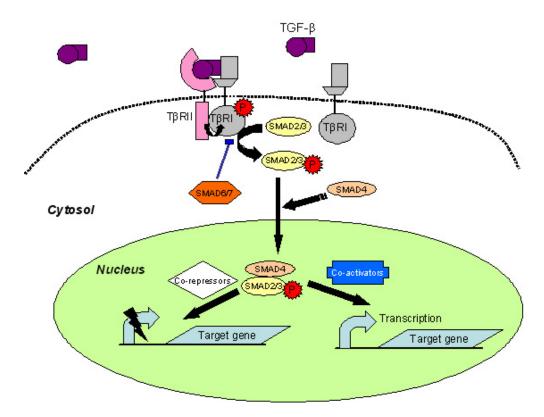


Figure 1. TGF- β /Smad signaling transduction pathway. TGF- β binds to TGF- β receptor II (T β RII) on cell surface, which recruits and activates TGF- β receptor I (T β RI), a serine-threonine kinase. Activated TGF- β /T β RII/T β RI complex phosphorylates Smad2/3 that in turn associates with Smad4 and moves to nucleus. In the nucleus, Smad complex binds to DNA and regulates targeting gene expression through recruitment of transcriptional co-repressors or co-activators. Smad6 or Smad7 are the inhibitory Smads which can competitively bind to TGF- β /T β RII/T β RI complex, inhibiting Smad2/3 phosphorylation.

stemming from early apoptosis in differentiating alveolar cells (29). On the contrary, disrupting TGF-β signaling also causes inappropriate alveolar development. Mice carrying TβRII antisense RNA controlled by the MMTV promoter displayed precocious lobuloalveolar development, indicating a critical role of TBRII in maintaining non-differentiated status of virgin mammary gland epithelium (30). Furthermore, treating mice with slow-release plastic pellets containing TGF-β resulted in reversible regression of the end-buds in developing mammary gland during puberty, but not in the alveolar buds in pregnancy (31). This selective regression action indicates that TGF- β functions in cell type and/or tissue context-dependent manners and that TGF-β activity is differentially regulated during distinct stages of mammary gland development (32).

During pregnancy, TGF- β 1 expression is decreased while TGF- β 2 and TGF- β 3 are elevated until the onset of lactation (33). Studies using specific antibodies that recognize latent and active TGF- β 1 indicated that TGF- β 1 activation was primarily localized in luminal epithelial cells, not in cap and myoepithelial cells (27). TGF- β 1 promotes apoptosis during involution. In the involuting gland, TGF- β 1 arises from days 1 to 10 after weaning, with an expression peak at day 6. This expression profile is consistent with bulk mammary epithelial cell death during

post-lactational mammary gland involution (34). Finally, the response of the cultured TAC-2.1 epithelial cells varies with the concentrations of TGF- β . At picomolar levels, the TGF- β inhibited branching morphogenesis, whereas it stimulated at fentomolar levels (35). Therefore, the TGF- β signaling not only holds a pleiotropic role in mammary gland development, but is also tissue context- and ligand dose-dependent.

5. $TGF-\beta$ SIGNALING IN MAMMARY GLAND TUMORIGENESIS

TGF- β promotes cell growth inhibition, apoptosis, and differentiation, and therefore, is considered as a potent tumor suppressor (36). However, recent studies indicate that TGF- β plays a dual role in mammary tumorigenesis. During advanced stages, TGF- β factually stimulates cancer cell invasion and metastasis (37, 38).

The involvement of TGF- β signaling pathway in tumorigenesis was documented first by functional changes of the signaling molecules in this pathway, such as TGF- β receptor and Smad mutations (37, 39-41). In human breast cancer, however, the alterations of TGF- β signaling molecules are relatively rare, except for T β RII downregulation (42, 43). Pathological studies of archived breast samples, including benign lesions, ductal carcinoma

in situ (DCIS), and invasive mammary carcinomas (IMC), indicated that TβRII downregulation correlated with progression and aggression of both in-situ and invasive breast carcinomas (44, 45). In mice expressing dominant-negative TβRII in mammary epithelium, spontaneous epithelial tumor occurrence was significantly increased (46). TβRI may also prevent mammary gland tumor formation. In mice carrying the Neu oncogene, active TβRI expressed specifically in the mammary epithelium diminished epithelial tumor appearance (47). In TGF- β 1 and TGF- α double-transgenic mice, the frequency of tumors was significantly reduced compared to that in TGF- α transgene alone. In addition, mammary gland tumorigenesis induced by 7,12-dimethylbenz[a]anthracene was prevented by TGF- β expression (48).

Although its biological function remains unclear, TGF- β receptor III (T β RIII) may act as a suppressor of breast cancer. T β RIII has a short cytoplasmic domain, and therefore, its intrinsic kinase activity and role in TGF- β signaling need to be defined. However, the decrease or loss of T β RIII expression occurred in approximately 90% of breast cancer at mRNA levels and 70% at protein levels. In addition, T β RIII loss occurred at substantially high levels in advanced, invasive breast carcinomas. Therefore, T β RIII loss may be a negative prognostic factor for patients with invasive breast cancer (49).

Premature stem cells are involved in pathogenesis of mammary gland cancer (50). TGF- β induces premature senescence of mammary stem cells and thus may suppress tumorigenesis (50-52). This hypothesis was confirmed by a telomerase study. Telomerase underpins stem cell renewal and proliferation and thus enhances the occurrence of breast cancer. Li's report indicated that TGF- β can repress

median latency of tumors induced by polyomavirus middle-T (47, 59). Constitutive expression of active TGF- β 1 did not affect tumor latency in transgenic mice but enhanced the tumor invasiveness and metastasis to lungs (62, 63). All these findings indicate a tumor stage-related dual function of TGF- β signaling in mammary tumorigenesis.

The dual role of TGF- β in tumorigenesis also appears in human breast cancer. T β RII inactivation enhances the invasiveness of premalignant or low-grade breast tumor cells, but reduces the metastasis of high-grade tumors (64). As for TGF- β 1, this ligand induces mammary epithelial-to-mesenchymal transition (EMT), resulting in the loss of epithelial polarity, disruption of cellular adhesion, and tumor cell invasion (65-67). TGF- β 1 also upregulates integrin-linked kinase, increasing cellular motility (68). In addition, high levels of TGF- β 1 mRNA correlated with enhanced angiogenesis and poor prognosis of breast cancer (69). Therefore, TGF- β 8 may be an important regulating factor of tumor invasion and metastasis at later stages (63).

Tumor metastasis is a multifactorial event, including tumor cell invasion into stroma and formation of blood and lymphatic vessels (43). Investigations in the past decades have demonstrated the importance of peritumoral

telomerase reverse transcriptase (TERT) expression by stimulating rapid entrance of Smad3 into nucleus (53). In the nucleus, Smad3 associates with c-myc, binds to the promoter region of TERT gene, and suppresses its expression. This negative regulation of telomerase activity can be interrupted by the Smad3 antagonist, Smad7 (53). Currently, TERT is proposed as a diagnostic and prognostic biomarker of breast cancer, as well as a potential therapeutic target of this disease (54).

On the contrary, considerable evidence indicates that TGF- β functions as a tumor promoter through the autocrine and paracrine actions, favoring tumor cell growth, invasion, and metastasis (55). In Neu transgenic mice, T β RI kinase activated c-myc and ligand-independent phosphatidylinositol-3 kinase (PI3K)/Akt in mammary cells, rendering cellular resistance to TGF- β -mediated growth arrest (56). In this model, TGF- β may synergize with the oncogene Neu to enhance survival and transformation of mammary epithelial cells.

A reduced response of tumor cells to TGF- β signaling often accompanies an increase in secretion of this ligand (57, 58). In breast cancer patients with poor prognosis, TGF- β 1 levels were often elevated in plasma, tumor cells, and associated stroma (59-61). Factually, in transgenic mice expressing either activated or dominant-negative TGF- β ligands or receptors, a biphasic role of TGF- β signaling appeared in mammary tumor progression. In Neu transgenic mice, constitutively active T β RI increased the latency of mammary tumor formation, but enhanced the frequency of extravascular lung metastasis (47). On the contrary, the dominant-negative form of T β RII reduced tumor metastasis in Neu mice, but shortened the

stroma in the development and/or evolution of tumors (70). Stromal cells support and facilitate tumor growth by secreting growth factors and/or proteases, such as vascular endothelial growth factor (VEGF), metalloproteinases (MMPs), and TGF-β. These factors constitute a microenvironment that benefits tumor cell growth and progression (71, 72). Cheng's work (73) demonstrated that co-transplanting TBRII knock-out mammary fibroblasts with carcinoma cells promoted growth and invasion of tumor cells, associated with an increase in activity of several tyrosine kinase receptors: erbB1, erbB2, RON, and c-Met. In an in vitro assay, cancer cells cultured under fibroblast-conditioned medium showed increased proliferation and motility (73), indicating the role of stromal TGF-β signaling in neoplastic progression.

6. TGF- β PATHWAY AS A TARGET FOR CANCER THERAPY

Therapeutic potential of the TGF- β signaling pathway is derived from its supportive function in late-stage tumors, enhancing tumor invasion, neoangiogenesis, metastasis, and the escape of immunosurveillance (74). In addition, TGF- β signaling is also involved in anti-tumor drug resistance in *in vivo* and *in vitro* studies (75, 76). Current cancer therapeutic approaches that target the TGF-

 β pathway include antagonism of TGF- β ligand binding to heteromeric receptor complex, intracellular inhibition of T β RI kinase, and sequence-specific degradation of TGF- β mRNA. Among them, the most extensively investigated intervention agents are sorted into small-molecule and large-molecule inhibitors (77, 78).

Ki26894, a TβRI kinase inhibitor, is a representative of small-molecule inhibitors. Systemic administration of Ki26894 via intraperitoneal injection effectively reduces the number and size of lung metastasis in both orthotropic xenografts and experimental metastasis models of human breast carcinomas (79, 80). Other small compounds such as SB-203580, an inhibitor of TBRI kinase (81), and SD-093 and LY580276, inhibitors of epithelialto-mesenchymal transition (82, 83), all showed promising potency in suppression of tumor cell invasion and metastasis. In addition, TGF-β signaling often promotes metastasis by activating survival signals, such as epidermal growth factor receptors (84, 85); therefore, Gleevec, a specific tyrosine kinase receptor inhibitor, effectively TGF-β-induced proliferation of blocks human osteosarcoma cells (86).

Large-molecule inhibitors of the TGF-β signaling include peptides, monoclonal antibodies, and antisense oligonucleotides/antisense RNA (78). Short phospho-Smad peptide [pSmad3(-3), KVLTQMGSPSIRCSS(PO4)VS] is a specific substrate of active TβRI, inhibiting TGF-β-induced Smad2 phosphorylation in mouse mammary epithelial cells (82). Bioengineered protein composed of extracellular domain of TBRII and Fc domain of murine IgG1 heavy chain (Fc:TβRII) demonstrated capability of enhancing the apoptosis of primary tumors and inhibiting tumor cell motility, intravasation, and lung metastases (87). In Fc:TβRII transgenic mice, tumor metastasis to distant organs was significantly less than in wild type animals (87). In addition, a monoclonal anti-TGF-β antibody (1D11) significantly suppressed metastasis of highly metastatic 4T1 murine breast cancer cells to the lungs in animal studies (88).

DNA vaccine is another immunological approach in cancer therapy. TGF-β is known as an important factor regulating tumor cell migration toward blood vessels, the first step of metastasis (89). Tumor-associated macrophages (TAMs) are key players in this process through the production of a variety of factors, including TGF-β (89). Legumain is a protein specifically expressed in TAMs (90). An ongoing approach immunologically targets legumain to induce the destruction of TAMs, remodeling tumor microenvironments and inhibiting tumor growth and metastasis (91). In the 4T1 breast carcinoma metastatic model, a legumain-based DNA vaccine administered after surgical resection of primary tumors significantly increased the lifespan in 75% (6/8) of the experimental mice up to 3 months, and 62% of mice were completely free from metastases, demonstrating the effectiveness of TAMtargeted tumor growth and metastasis inhibition (92).

TGF- β antisense RNA is a novel strategy targeting TGF- β signaling pathway by triggering mRNA

degradation (93). AP12009 is a complementary antisense RNA of TGF- β 2 mRNA and showed promising therapeutic efficacy in animal tumor models with TGF- β 2 overexpression, such as malignant glioma and pancreatic cancer (94). A recent study further demonstrated that combining the TGF- β 2 antibody and antisense RNA completely regressed 4T1 tumors in 40% of the mice tested (95).

Compared to small-molecule inhibitors, these large-molecules are characterized with specificity and prolonged duration. However, their limited tissue penetration is a major concern for clinical applications. Small-molecule inhibitors penetrate tissue better, but their tumor-selectivity is usually low. Circumventing the shortcomings of large- and small-molecules and choosing appropriate treatment modalities for patients with different TGF- β signaling responses is a major challenge that needs to be more extensively investigated in the future.

7. CONCLUSION

In this article, we update the current understanding of TGF-β signaling, with focus on mammary gland development and tumorigenesis, and discuss the therapeutic perspectives of this pathway. TGF-\beta signaling pathway is complicated and functions in mammary gland in ligand content- and tissue context-dependent manners. TGF-β signaling also functions biphasically in tumor suppression and progression. TGF-β normally prevents uncontrolled cell proliferation; however, once aberrant genetic or epigenetic events abolish the cytostatic function of TGF-β, tumor cells evade TGF-β control and acquire the ability to proliferate, invade, and metastasize. Therefore, understanding the bifunctional features of TGF-β signaling in tumorigenesis is important to the development and the clinical practice of antitumor agents targeting this pathway. In addition, development of more specific agents with better tissue penetration would be the effort of the investigators in cancer therapeutic studies.

8. REFERENCES

- 1. Roberts, A. B., N. L. Thompson, U. Heine, C. Flanders & M. B. Sporn: Transforming growth factor-beta: possible roles in carcinogenesis. *Br J Cancer*, 57, 594-600 (1988)
- 2. Moses, H. L., E. L. Branum, J. A. Proper & R. A. Robinson: Transforming growth factor production by chemically transformed cells. *Cancer Res*, 41, 2842-8 (1981)
- 3. Koff, A., M. Ohtsuki, K. Polyak, J. M. Roberts & J. Massague: Negative regulation of G1 in mammalian cells: inhibition of cyclin E-dependent kinase by TGF-beta. *Science*, 260, 536-9 (1993)
- 4. Wakefield, L. M., E. Piek & E. P. Bottinger: TGF-beta signaling in mammary gland development and tumorigenesis. *J Mammary Gland Biol Neoplasia*, 6, 67-82 (2001)
- 5. Massague, J.: TGF-beta signal transduction. *Annu Rev Biochem*, 67, 753-91 (1998)
- 6. Gentry, L. E., M. N. Lioubin, A. F. Purchio & H. Marquardt: Molecular events in the processing of

- recombinant type 1 pre-pro-transforming growth factor beta to the mature polypeptide. *Mol Cell Biol*, 8, 4162-8 (1988)
- 7. Gentry, L. E., N. R. Webb, G. J. Lim, A. M. Brunner, J. E. Ranchalis, D. R. Twardzik, M. N. Lioubin, H. Marquardt & A. F. Purchio: Type 1 transforming growth factor beta: amplified expression and secretion of mature and precursor polypeptides in Chinese hamster ovary cells. *Mol Cell Biol*, 7, 3418-27 (1987)
- 8. Saharinen, J., J. Taipale & J. Keski-Oja: Association of the small latent transforming growth factor-beta with an eight cysteine repeat of its binding protein LTBP-1. *Embo J*, 15, 245-53 (1996)
- 9. Ge, G. & D. S. Greenspan: BMP1 controls TGFbeta1 activation via cleavage of latent TGFbeta-binding protein. *J Cell Biol*, 175, 111-20 (2006)
- 10. Arteaga, C. L., R. J. Coffey, Jr., T. C. Dugger, C. M. McCutchen, H. L. Moses & R. M. Lyons: Growth stimulation of human breast cancer cells with anti-transforming growth factor beta antibodies: evidence for negative autocrine regulation by transforming growth factor beta. *Cell Growth Differ*, 1, 367-74 (1990)
- 11. Knabbe, C., M. E. Lippman, L. M. Wakefield, K. C. Flanders, A. Kasid, R. Derynck & R. B. Dickson: Evidence that transforming growth factor-beta is a hormonally regulated negative growth factor in human breast cancer cells. *Cell*, 48, 417-28 (1987)
- 12. Munger, J. S., J. G. Harpel, F. G. Giancotti & D. B. Rifkin: Interactions between growth factors and integrins: latent forms of transforming growth factor-beta are ligands for the integrin alphavbeta1. *Mol Biol Cell*, 9, 2627-38 (1998)
- 13. Fontana, L., Y. Chen, P. Prijatelj, T. Sakai, R. Fassler, L. Y. Sakai & D. B. Rifkin: Fibronectin is required for integrin alphavbeta6-mediated activation of latent TGF-beta complexes containing LTBP-1. *Faseb J*, 19, 1798-808 (2005)
- 14. Hyytiainen, M. & J. Keski-Oja: Latent TGF-beta binding protein LTBP-2 decreases fibroblast adhesion to fibronectin. *J Cell Biol*, 163, 1363-74 (2003)
- 15. Annes, J. P., J. S. Munger & D. B. Rifkin: Making sense of latent TGFbeta activation. *J Cell Sci*, 116, 217-24 (2003)
- 16. Rifkin, D. B.: Latent transforming growth factor-beta (TGF-beta) binding proteins: orchestrators of TGF-beta availability. *J Biol Chem*, 280, 7409-12 (2005)
- 17. Zhu, H. J. & A. M. Sizeland: A pivotal role for the transmembrane domain in transforming growth factor-beta receptor activation. *J Biol Chem*, 274, 11773-81 (1999)
- 18. Andres, J. L., K. Stanley, S. Cheifetz & J. Massague: Membrane-anchored and soluble forms of betaglycan, a polymorphic proteoglycan that binds transforming growth factor-beta. *J Cell Biol*, 109, 3137-45 (1989)
- 19. De Crescenzo, G., S. Grothe, J. Zwaagstra, M. Tsang & M. D. O'Connor-McCourt: Real-time monitoring of the interactions of transforming growth factor-beta (TGF-beta) isoforms with latency-associated protein and the ectodomains of the TGF-beta type II and III receptors reveals different kinetic models and stoichiometries of binding. *J Biol Chem*, 276, 29632-43 (2001)
- 20. Massague, J., J. Seoane & D. Wotton: Smad transcription factors. *Genes Dev*, 19, 2783-810 (2005)

- 21. Schmierer, B. & C. S. Hill: Kinetic analysis of Smad nucleocytoplasmic shuttling reveals a mechanism for transforming growth factor beta-dependent nuclear accumulation of Smads. *Mol Cell Biol*, 25, 9845-58 (2005) 22. Park, S. H.: Fine tuning and cross-talking of TGF-beta signal by inhibitory Smads. *J Biochem Mol Biol*, 38, 9-16 (2005)
- 23. Brown, K. & N. A. Bhowmick: Linking TGF-betamediated Cdc25A inhibition and cytoskeletal regulation through RhoA/p160 (ROCK) signaling. *Cell Cycle*, 3, 408-10 (2004)
- 24. Schuster, N. & K. Krieglstein: Mechanisms of TGF-beta-mediated apoptosis. *Cell Tissue Res*, 307, 1-14 (2002) 25. Ferrari, G., G. Pintucci, G. Seghezzi, K. Hyman, A. C. Galloway & P. Mignatti: VEGF, a prosurvival factor, acts in concert with TGF-beta1 to induce endothelial cell apoptosis. *Proc Natl Acad Sci U S A*, 103, 17260-17265 (2006)
- 26. Serra, R. & M. R. Crowley: Mouse models of transforming growth factor beta impact in breast development and cancer. *Endocr Relat Cancer*, 12, 749-60 (2005)
- 27. Ewan, K. B., G. Shyamala, S. A. Ravani, Y. Tang, R. Akhurst, L. Wakefield & M. H. Barcellos-Hoff: Latent transforming growth factor-beta activation in mammary gland: regulation by ovarian hormones affects ductal and alveolar proliferation. *Am J Pathol*, 160, 2081-93 (2002)
- 28. Pierce, D. F., Jr., M. D. Johnson, Y. Matsui, S. D. Robinson, L. I. Gold, A. F. Purchio, C. W. Daniel, B. L. Hogan & H. L. Moses: Inhibition of mammary duct development but not alveolar outgrowth during pregnancy in transgenic mice expressing active TGF-beta 1. *Genes Dev*, 7, 2308-17 (1993)
- 29. Jhappan, C., A. G. Geiser, E. C. Kordon, D. Bagheri, L. Hennighausen, A. B. Roberts, G. H. Smith & G. Merlino: Targeting expression of a transforming growth factor beta 1 transgene to the pregnant mammary gland inhibits alveolar development and lactation. *Embo J*, 12, 1835-45 (1993)
- 30. Lenferink, A. E., J. Magoon, M. C. Pepin, A. Guimond & M. D. O'Connor-McCourt: Expression of TGF-beta type II receptor antisense RNA impairs TGF-beta signaling in vitro and promotes mammary gland differentiation in vivo. *Int J Cancer*, 107, 919-28 (2003)
- 31. Silberstein, G. B. & C. W. Daniel: Reversible inhibition of mammary gland growth by transforming growth factor-beta. *Science*, 237, 291-3 (1987)
- 32. Daniel, C. W., G. B. Silberstein, K. Van Horn, P. Strickland & S. Robinson: TGF-beta 1-induced inhibition of mouse mammary ductal growth: developmental specificity and characterization. *Dev Biol*, 135, 20-30 (1989)
- 33. Robinson, S. D., G. B. Silberstein, A. B. Roberts, K. C. Flanders & C. W. Daniel: Regulated expression and growth inhibitory effects of transforming growth factorbeta isoforms in mouse mammary gland development. *Development*, 113, 867-78 (1991)
- 34. Strange, R., F. Li, S. Saurer, A. Burkhardt & R. R. Friis: Apoptotic cell death and tissue remodelling during mouse mammary gland involution. *Development*, 115, 49-58 (1992)

- 35. Soriano, J. V., L. Orci & R. Montesano: TGF-beta1 induces morphogenesis of branching cords by cloned mammary epithelial cells at subpicomolar concentrations. *Biochem Biophys Res Commun*, 220, 879-85 (1996)
- 36. Bachman, K. E. & B. H. Park: Duel nature of TGF-beta signaling: tumor suppressor vs. tumor promoter. *Curr Opin Oncol*, 17, 49-54 (2005)
- 37. Huo, Y. Y., K. T. Zhang, B. Y. Li, R. F. Duan, B. X. Fan, X. Q. Xiang, Y. C. Hu, L. Xie & D. C. Wu: [Regulation of Smad7 gene by TGF-beta 1 in process of malignant transformation]. *Ai Zheng*, 21, 117-21 (2002)
- 38. Derynck, R., R. J. Akhurst & A. Balmain: TGF-beta signaling in tumor suppression and cancer progression. *Nat Genet*, 29, 117-29 (2001)
- 39. Chung, Y. J., J. M. Song, J. Y. Lee, Y. T. Jung, E. J. Seo, S. W. Choi & M. G. Rhyu: Microsatellite instability-associated mutations associate preferentially with the intestinal type of primary gastric carcinomas in a high-risk population. *Cancer Res*, 56, 4662-5 (1996)
- 40. Han, S. U., H. T. Kim, D. H. Seong, Y. S. Kim, Y. S. Park, Y. J. Bang, H. K. Yang & S. J. Kim: Loss of the Smad3 expression increases susceptibility to tumorigenicity in human gastric cancer. *Oncogene*, 23, 1333-41 (2004)
- 41. Riggins, G. J., S. Thiagalingam, E. Rozenblum, C. L. Weinstein, S. E. Kern, S. R. Hamilton, J. K. Willson, S. D. Markowitz, K. W. Kinzler & B. Vogelstein: Mad-related genes in the human. *Nat Genet*, 13, 347-9 (1996)
- 42. Schmitt, F. C., R. Soares, H. Gobbi, F. Milanezzi, F. Santos-Silva, L. Cirnes, C. Costa & R. Seruca: Microsatellite instability in medullary breast carcinomas. *Int J Cancer*, 82, 644-7 (1999)
- 43. Jakowlew, S. B.: Transforming growth factor-beta in cancer and metastasis. *Cancer Metastasis Rev* (2006)
- 44. Gobbi, H., W. D. Dupont, J. F. Simpson, W. D. Plummer, Jr., P. A. Schuyler, S. J. Olson, C. L. Arteaga & D. L. Page: Transforming growth factor-beta and breast cancer risk in women with mammary epithelial hyperplasia. *J Natl Cancer Inst*, 91, 2096-101 (1999)
- 45. Gobbi, H., C. L. Arteaga, R. A. Jensen, J. F. Simpson, W. D. Dupont, S. J. Olson, P. A. Schuyler, W. D. Plummer, Jr. & D. L. Page: Loss of expression of transforming growth factor beta type II receptor correlates with high tumour grade in human breast in-situ and invasive carcinomas. *Histopathology*, 36, 168-77 (2000)
- 46. Gorska, A. E., R. A. Jensen, Y. Shyr, M. E. Aakre, N. A. Bhowmick & H. L. Moses: Transgenic mice expressing a dominant-negative mutant type II transforming growth factor-beta receptor exhibit impaired mammary development and enhanced mammary tumor formation. *Am J Pathol*, 163, 1539-49 (2003)
- 47. Siegel, P. M., W. Shu, R. D. Cardiff, W. J. Muller & J. Massague: Transforming growth factor beta signaling impairs Neu-induced mammary tumorigenesis while promoting pulmonary metastasis. *Proc Natl Acad Sci U S A*, 100, 8430-5 (2003)
- 48. Pierce, D. F., Jr., A. E. Gorska, A. Chytil, K. S. Meise, D. L. Page, R. J. Coffey, Jr. & H. L. Moses: Mammary tumor suppression by transforming growth factor beta 1 transgene expression. *Proc Natl Acad Sci U S A*, 92, 4254-8 (1995)
- 49. Dong, M., T. How, K. C. Kirkbride, K. J. Gordon, J. D. Lee, N. Hempel, P. Kelly, B. J. Moeller, J. R. Marks &

- G. C. Blobe: The type III TGF-beta receptor suppresses breast cancer progression. *J Clin Invest* (2006)
- 50. Boulanger, C. A. & G. H. Smith: Reducing mammary cancer risk through premature stem cell senescence. *Oncogene*, 20, 2264-72 (2001)
- 51. Kordon, E. C., R. A. McKnight, C. Jhappan, L. Hennighausen, G. Merlino & G. H. Smith: Ectopic TGF beta 1 expression in the secretory mammary epithelium induces early senescence of the epithelial stem cell population. *Dev Biol*, 168, 47-61 (1995)
- 52. Boulanger, C. A., K. U. Wagner & G. H. Smith: Parity-induced mouse mammary epithelial cells are pluripotent, self-renewing and sensitive to TGF-beta1 expression. *Oncogene*, 24, 552-60 (2005)
- 53. Li, H., D. Xu, J. Li, M. C. Berndt & J. P. Liu: Transforming growth factor beta suppresses human telomerase reverse transcriptase (hTERT) by Smad3 interactions with c-Myc and the hTERT gene. *J Biol Chem*, 281, 25588-600 (2006)
- 54. Hines, W. C., A. M. Fajardo, N. E. Joste, M. Bisoffi & J. K. Griffith: Quantitative and spatial measurements of telomerase reverse transcriptase expression within normal and malignant human breast tissues. *Mol Cancer Res*, 3, 503-9 (2005)
- 55. Dumont, N. & C. L. Arteaga: Transforming growth factor-beta and breast cancer: Tumor promoting effects of transforming growth factor-beta. *Breast Cancer Res*, 2, 125-32 (2000)
- 56. Muraoka-Cook, R. S., I. Shin, J. Y. Yi, E. Easterly, M. H. Barcellos-Hoff, J. M. Yingling, R. Zent & C. L. Arteaga: Activated type I TGFbeta receptor kinase enhances the survival of mammary epithelial cells and accelerates tumor progression. *Oncogene*, 25, 3408-23 (2006)
- 57. Reiss, M.: TGF-beta and cancer. *Microbes Infect*, 1, 1327-47 (1999)
- 58. Akhurst, R. J. & A. Balmain: Genetic events and the role of TGF beta in epithelial tumour progression. *J Pathol*, 187, 82-90 (1999)
- 59. Forrester, E., A. Chytil, B. Bierie, M. Aakre, A. E. Gorska, A. R. Sharif-Afshar, W. J. Muller & H. L. Moses: Effect of conditional knockout of the type II TGF-beta receptor gene in mammary epithelia on mammary gland development and polyomavirus middle T antigen induced tumor formation and metastasis. *Cancer Res*, 65, 2296-302 (2005)
- 60. Sheen-Chen, S. M., H. S. Chen, C. W. Sheen, H. L. Eng & W. J. Chen: Serum levels of transforming growth factor beta1 in patients with breast cancer. *Arch Surg*, 136, 937-40 (2001)
- 61. Ivanovic, V., N. Todorovic-Rakovic, M. Demajo, Z. Neskovic-Konstantinovic, V. Subota, O. Ivanisevic-Milovanovic & D. Nikolic-Vukosavljevic: Elevated plasma levels of transforming growth factor-beta 1 (TGF-beta 1) in patients with advanced breast cancer: association with disease progression. *Eur J Cancer*, 39, 454-61 (2003)
- 62. Muraoka, R. S., Y. Koh, L. R. Roebuck, M. E. Sanders, D. Brantley-Sieders, A. E. Gorska, H. L. Moses & C. L. Arteaga: Increased malignancy of Neu-induced mammary tumors overexpressing active transforming growth factor beta1. *Mol Cell Biol*, 23, 8691-703 (2003)
- 63. Muraoka-Cook, R. S., H. Kurokawa, Y. Koh, J. T. Forbes, L. R. Roebuck, M. H. Barcellos-Hoff, S. E. Moody, L. A. Chodosh & C. L. Arteaga: Conditional

- overexpression of active transforming growth factor beta1 in vivo accelerates metastases of transgenic mammary tumors. *Cancer Res*, 64, 9002-11 (2004)
- 64. Tang, B., M. Vu, T. Booker, S. J. Santner, F. R. Miller, M. R. Anver & L. M. Wakefield: TGF-beta switches from tumor suppressor to prometastatic factor in a model of breast cancer progression. *J Clin Invest*, 112, 1116-24 (2003)
- 65. Knudsen, K. A., C. Frankowski, K. R. Johnson & M. J. Wheelock: A role for cadherins in cellular signaling and differentiation. *J Cell Biochem Suppl*, 30-31, 168-76 (1998) 66. Oft, M., J. Peli, C. Rudaz, H. Schwarz, H. Beug & E. Reichmann: TGF-beta1 and Ha-Ras collaborate in modulating the phenotypic plasticity and invasiveness of epithelial tumor cells. *Genes Dev*, 10, 2462-77 (1996)
- 67. Miettinen, P. J., R. Ebner, A. R. Lopez & R. Derynck: TGF-beta induced transdifferentiation of mammary epithelial cells to mesenchymal cells: involvement of type I receptors. *J Cell Biol*, 127, 2021-36 (1994)
- 68. Janji, B., C. Melchior, V. Gouon, L. Vallar & N. Kieffer: Autocrine TGF-beta-regulated expression of adhesion receptors and integrin-linked kinase in HT-144 melanoma cells correlates with their metastatic phenotype. *Int J Cancer*, 83, 255-62 (1999)
- 69. Relf, M., S. LeJeune, P. A. Scott, S. Fox, K. Smith, R. Leek, A. Moghaddam, R. Whitehouse, R. Bicknell & A. L. Harris: Expression of the angiogenic factors vascular endothelial cell growth factor, acidic and basic fibroblast growth factor, tumor growth factor beta-1, platelet-derived endothelial cell growth factor, placenta growth factor, and pleiotrophin in human primary breast cancer and its relation to angiogenesis. *Cancer Res*, 57, 963-9 (1997)
- 70. Hagedorn, H. G., B. E. Bachmeier & A. G. Nerlich: Synthesis and degradation of basement membranes and extracellular matrix and their regulation by TGF-beta in invasive carcinomas (Review). *Int J Oncol*, 18, 669-81 (2001)
- 71. Noel, A. & J. M. Foidart: The role of stroma in breast carcinoma growth in vivo. *J Mammary Gland Biol Neoplasia*, 3, 215-25 (1998)
- 72. Tuxhorn, J. A., S. J. McAlhany, T. D. Dang, G. E. Ayala & D. R. Rowley: Stromal cells promote angiogenesis and growth of human prostate tumors in a differential reactive stroma (DRS) xenograft model. *Cancer Res*, 62, 3298-307 (2002)
- 73. Cheng, N., N. A. Bhowmick, A. Chytil, A. E. Gorksa, K. A. Brown, R. Muraoka, C. L. Arteaga, E. G. Neilson, S. W. Hayward & H. L. Moses: Loss of TGF-beta type II receptor in fibroblasts promotes mammary carcinoma growth and invasion through upregulation of TGF-alpha-, MSP- and HGF-mediated signaling networks. *Oncogene*, 24, 5053-68 (2005)
- 74. Pinkas, J. & B. A. Teicher: TGF-beta in cancer and as a therapeutic target. *Biochem Pharmacol*, 72, 523-9 (2006) 75. hmori, T., J. L. Yang, J. O. Price & C. L. Arteaga: Blockade of tumor cell transforming growth factor-betas enhances cell cycle progression and sensitizes human breast carcinoma cells to cytotoxic chemotherapy. *Exp Cell Res*, 245, 350-9 (1998)
- 76. Teicher, B. A., M. Ikebe, G. Ara, S. R. Keyes & R. S. Herbst: Transforming growth factor-beta 1 overexpression

- produces drug resistance in vivo: reversal by decorin. *In Vivo*, 11, 463-72 (1997)
- 77. Pardali, K. & A. Moustakas: Actions of TGF-beta as tumor suppressor and pro-metastatic factor in human cancer. *Biochim Biophys Acta* (2006)
- 78. Yingling, J. M., K. L. Blanchard & J. S. Sawyer: Development of TGF-beta signalling inhibitors for cancer therapy. *Nat Rev Drug Discov*, 3, 1011-22 (2004)
- 79. Ehata, S., A. Hanyu, M. Fujime, Y. Katsuno, E. Fukunaga, K. Goto, Y. Ishikawa, K. Nomura, H. Yokoo, T. Shimizu, E. Ogata, K. Miyazono, K. Shimizu & T. Imamura: Ki26894, a novel transforming growth factorbeta type I receptor kinase inhibitor, inhibits in vitro invasion and in vivo bone metastasis of a human breast cancer cell line. *Cancer Sci* (2006)
- 80. Bandyopadhyay, A., J. K. Agyin, L. Wang, Y. Tang, X. Lei, B. M. Story, J. E. Cornell, B. H. Pollock, G. R. Mundy & L. Z. Sun: Inhibition of pulmonary and skeletal metastasis by a transforming growth factor-beta type I receptor kinase inhibitor. *Cancer Res*, 66, 6714-21 (2006)
- 81. Yakymovych, I., U. Engstrom, S. Grimsby, C. H. Heldin & S. Souchelnytskyi: Inhibition of transforming growth factor-beta signaling by low molecular weight compounds interfering with ATP- or substrate-binding sites of the TGF beta type I receptor kinase. *Biochemistry*, 41, 11000-7 (2002)
- 82. Peng, S. B., L. Yan, X. Xia, S. A. Watkins, H. B. Brooks, D. Beight, D. K. Herron, M. L. Jones, J. W. Lampe, W. T. McMillen, N. Mort, J. S. Sawyer & J. M. Yingling: Kinetic characterization of novel pyrazole TGF-beta receptor I kinase inhibitors and their blockade of the epithelial-mesenchymal transition. *Biochemistry*, 44, 2293-304 (2005)
- 83. Subramanian, G., R. E. Schwarz, L. Higgins, G. McEnroe, S. Chakravarty, S. Dugar & M. Reiss: Targeting endogenous transforming growth factor beta receptor signaling in SMAD4-deficient human pancreatic carcinoma cells inhibits their invasive phenotype1. *Cancer Res*, 64, 5200-11 (2004)
- 84. Caja, L., C. Ortiz, E. Bertran, M. M. Murillo, M. J. Miro-Obradors, E. Palacios & I. Fabregat: Differential intracellular signalling induced by TGF-beta in rat adult hepatocytes and hepatoma cells: Implications in liver carcinogenesis. *Cell Signal* (2006)
- 85. Jechlinger, M., A. Sommer, R. Moriggl, P. Seither, N. Kraut, P. Capodiecci, M. Donovan, C. Cordon-Cardo, H. Beug & S. Grunert: Autocrine PDGFR signaling promotes mammary cancer metastasis. *J Clin Invest*, 116, 1561-70 (2006)
- 86. Matsuyama, S., M. Iwadate, M. Kondo, M. Saitoh, A. Hanyu, K. Shimizu, H. Aburatani, H. K. Mishima, T. Imamura, K. Miyazono & K. Miyazawa: SB-431542 and Gleevec inhibit transforming growth factor-beta-induced proliferation of human osteosarcoma cells. *Cancer Res*, 63, 7791-8 (2003)
- 87. Muraoka, R. S., N. Dumont, C. A. Ritter, T. C. Dugger, D. M. Brantley, J. Chen, E. Easterly, L. R. Roebuck, S. Ryan, P. J. Gotwals, V. Koteliansky & C. L. Arteaga: Blockade of TGF-beta inhibits mammary tumor cell viability, migration, and metastases. *J Clin Invest*, 109, 1551-9 (2002)

- 88. Nam, J. S., A. M. Suchar, M. J. Kang, C. H. Stuelten, B. Tang, A. M. Michalowska, L. W. Fisher, N. S. Fedarko, A. Jain, J. Pinkas, S. Lonning & L. M. Wakefield: Bone sialoprotein mediates the tumor cell-targeted prometastatic activity of transforming growth factor beta in a mouse model of breast cancer. *Cancer Res*, 66, 6327-35 (2006)
- 89. De Wever, O. & M. Mareel: Role of tissue stroma in cancer cell invasion. *J Pathol*, 200, 429-47 (2003)
- 90. Condeelis, J. & J. W. Pollard: Macrophages: obligate partners for tumor cell migration, invasion, and metastasis. *Cell*, 124, 263-6 (2006)
- 91. Oosterling, S. J., G. J. van der Bij, G. A. Meijer, C. W. Tuk, E. van Garderen, N. van Rooijen, S. Meijer, J. R. van der Sijp, R. H. Beelen & M. van Egmond: Macrophages direct tumour histology and clinical outcome in a colon cancer model. *J Pathol*, 207, 147-55 (2005)
- 92. Luo, Y., H. Zhou, J. Krueger, C. Kaplan, S. H. Lee, C. Dolman, D. Markowitz, W. Wu, C. Liu, R. A. Reisfeld & R. Xiang: Targeting tumor-associated macrophages as a novel strategy against breast cancer. *J Clin Invest*, 116, 2132-2141 (2006)
- 93. Iyer, S., Z. G. Wang, M. Akhtari, W. Zhao & P. Seth: Targeting TGFbeta signaling for cancer therapy. *Cancer Biol Ther*, 4, 261-6 (2005)
- 94. Schlingensiepen, R., M. Goldbrunner, M. N. Szyrach, G. Stauder, P. Jachimczak, U. Bogdahn, F. Schulmeyer, P. Hau & K. H. Schlingensiepen: Intracerebral and intrathecal infusion of the TGF-beta 2-specific antisense phosphorothioate oligonucleotide AP 12009 in rabbits and primates: toxicology and safety. *Oligonucleotides*, 15, 94-104 (2005)
- 95. Kobie, J. J., R. S. Wu, R. A. Kurt, S. Lou, M. K. Adelman, L. J. Whitesell, L. V. Ramanathapuram, C. L. Arteaga & E. T. Akporiaye: Transforming growth factor beta inhibits the antigen-presenting functions and antitumor activity of dendritic cell vaccines. *Cancer Res*, 63, 1860-4 (2003)

Abbreviations: TGF: Transforming growth factor; T β RII: TGF- β type II receptor; T β RII: TGF- β type II receptor; MMTV: mouse mammary tumor virus; TERT: telomerase reverse transcriptase; and TAMs, Tumor-associated macrophages

Key Words: TGF-β ligand, TGF-β receptor, cancer therapy, mammary gland, tumor-associated macrophages, breast cancer, animal model, Smad protein, tumor suppressor, tumor promotion, and review

Send correspondence to: Deliang Cao, Ph.D. Department of Medical Microbiology, Immunology, and Cell Biology, Simmons Cooper Cancer Institute, Southern Illinois University School of Medicine, 913 N. Rutledge Street, Springfield, IL 62702, Tel: 217-545-9703, Fax: 217-545-9718, E-mail: dcao@siumed.edu

http://www.bioscience.org/current/vol12.htm