

Regulatory T Cells and Viral Infections

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TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Natural vs inducible CD4⁺CD25⁺ Treg cells
 - 3.1. Natural CD4⁺CD25⁺ Treg cells.
 - 3.2. Inducible CD4⁺CD25⁺ Treg cells.
4. Induction of CD4⁺CD25⁺ Treg cells in viral infections
5. Role of virus-induced CD4⁺CD25⁺ Treg cells
 - 5.1. Role of CD4⁺CD25⁺ Treg cells in viral pathogenesis
 - 5.2. Role of CD4⁺CD25⁺ Treg cells in host defense
 - 5.2.1. Control of inflammatory responses
 - 5.2.2. Control of hyperactivation of immune responses
 - 5.2.3. Control of virus-associated autoimmune disorders
6. Mechanisms of CD4⁺CD25⁺ Treg cell induction in viral infection
 - 6.1. Viral antigens
 - 6.2. Virus-induced cytokines
 - 6.3. Virus-induced DC
7. Mechanisms of suppression by virus-induced CD4⁺CD25⁺ Treg cells
 - 7.1. FOXP3
 - 7.2. CTLA-4
 - 7.3. TGF β
 - 7.4. IL-10
 - 7.5. IL-2
 - 7.6. GITR
8. Conclusions and perspective
9. Acknowledgement
10. References

1. ABSTRACT

Viral infections often lead to generalized immunosuppression characterized by the downregulation of virus-specific CD4⁺ and CD8⁺ T cell responses and/or non-specific inflammation. One of the mechanisms for the virus-induced immunosuppression is the induction of CD4⁺CD25⁺ regulatory T (Treg) cells that act to suppress effector T cell functions during infection. Depending on the situation, the CD4⁺CD25⁺ Treg cell-mediated suppression can be either beneficial or detrimental to the host. On one hand, they play a critical role in maintaining host homeostasis by controlling exaggerated and destructive inflammations paralleling strong antiviral immune responses and thus contribute to host protection. On the other hand, suppression of virus-specific T cell responses by the Treg cells depresses host antiviral immune responses and thus facilitates viral persistence and disease progression. Despite numerous reports on induction of CD4⁺CD25⁺ Treg cells in viral infections, it is still not fully clear how they are induced and how they act to suppress effector T cell functions in viral infections. This review discusses recent progress in our understanding of the role of virus-induced CD4⁺CD25⁺ Treg cells and the mechanisms by which they are induced and exert their function during infection.

2. INTRODUCTION

Regulatory T cells, particularly CD4⁺CD25⁺ Treg cells, have been shown to play an important role in regulating immune responses and maintaining homeostasis under various disease conditions including autoimmune disease, inflammation, cancer, and microbial infections. Indeed, a growing body of studies has indicated that CD4⁺CD25⁺ Treg cells actively regulate host antiviral responses by suppressing effector T cell activation and functions during viral infections. They are initially recognized by their roles in mediating immunologic self-tolerance to prevent the activation of self-reactive T cells and thus prevent harmful autoimmune responses to self and foreign antigens (1, 2). The finding by Belkaid *et al* in 2002 on the ability of CD4⁺CD25⁺ Treg cells to control *Leishmania major* persistence and immunity uncovered a new role of these Treg cells in regulation of T cell responses to foreign pathogens (3), and thus extended their role in maintaining host homeostasis beyond autoimmunity. Accordingly, their target cell repertoire for suppression also expanded to include pathogen-specific CD4⁺ and CD8⁺ T cells that are not self-reactive in nature. Because of their involvement in a wide variety of disease types and their unique role in helping both pathogens and the host, CD4⁺CD25⁺ Treg cells have been under intensive

investigation in recent years, and are the best characterized member in the regulatory T cell family which includes several subpopulations of T cells with similar suppressive activities but distinct phenotypes, origins, and mechanisms of action, such as T regulatory type 1 (Tr1) (4), Th3 (5), and CD8⁺ (6, 7) Treg cells. Although all these groups of regulatory T cells have been shown to be involved in suppression of T cell immune responses during viral infections, this review will mainly focus on the CD4⁺CD25⁺ Treg cells, since they are likely the major player in the Treg cell family and most of the Treg cell-associated suppressive effects observed in viral infections are mediated by this group of Treg cells.

Effective control of viral infection relies on the function of effector T cells to mount strong virus-specific immune responses characterized by the activation of cytotoxic T cells (CTL). However, this function is often suppressed or impaired in viral infections, especially in chronic or persistent viral infections such as those caused by human immunodeficiency virus (HIV), hepatitis C virus (HCV), and Epstein-Barr virus (EBV). This suppression enables viruses to evade host immune surveillance, prevents them from being eradicated by host immune cells, and facilitates the establishment of their persistence or latency. A number of mechanisms have been proposed for the virus-induced immunosuppression, and different mechanisms have been defined in different viral infections. Some examples include direct infection of immune cells leading to the impairment in their capability to mount vigorous antiviral immune responses, induction of immunosuppressive soluble factors elaborated by host immune cells or infected cells of other origins, induction of immunoregulatory cells, suppressive effect of viral proteins, or dysregulation of immune responses such as cytokine shift from Th1 to Th2 responses (8-10). During the recent years, CD4⁺CD25⁺ Treg cells that are expanded and activated upon virus infection have been found to possess strong suppressive effect on virus-specific effector CD4⁺ helper (Th) and CD8⁺ cytotoxic T cells (11-15). With their potent immunosuppressive properties, CD4⁺CD25⁺ Treg cells represent another mechanism by which viruses induce immunosuppression to aid their efforts to overcome or evade host antiviral responses. By suppressing host antiviral responses, CD4⁺CD25⁺ Treg cells contribute to the development of chronic or persistent infections and thus are beneficial to the virus.

However, the combat between viruses and the host is a dynamic process. While viruses endeavor every effort to overcome or evade host antiviral defensive responses, the host immune system strives to mount vigorous antiviral responses to eliminate invading viruses. The activation of immune cells to generate responses to viral antigens involves a series of interactions and events that culminates in the formation of both non-specific inflammatory and specific humoral and/or cell-mediated immune responses. These responses need to be strong so that viruses can be eliminated and the infection can be confined quickly. However, strong responses, both non-specific inflammatory and specific immune responses, can be harmful to the host if the activated immune cells

overreact. Obviously, these antiviral responses require a high degree of regulation to be effective. CD4⁺CD25⁺ Treg cells are likely to play a key role in this process to control excessive inflammatory and immune responses. This has been demonstrated by an increasing number of studies showing that CD4⁺CD25⁺ Treg cells function to suppress immune-mediated pathogenesis resulting from viral infections (14, 16-19). Apparently, they function to benefit the host.

Thus, CD4⁺CD25⁺ Treg cells induced during viral infections can be a double-sided sword that, depending on the situation, can be either beneficial or detrimental to the host. Their suppressive function is essential for the maintenance of host homeostasis during the course of antiviral immune responses and, therefore, constitutes an indispensable component of host defense to viral infections. Despite the large number of studies showing the activation and involvement of CD4⁺CD25⁺ Treg cells in viral infections, the mechanisms for their induction and action are still not fully understood. This review intends to provide a brief summary of recent studies on how these cells are induced and how they act to suppress effector T cell functions in viral infections. A better understand of the dynamics and mechanisms of CD4⁺CD25⁺ Treg cell induction and action during infection will help us develop appropriate strategies to harness these cells towards desired clinical outcomes that benefit the host.

3. NATURAL VS INDUCIBLE CD4⁺CD25⁺ TREG CELLS

Based on their origins, CD4⁺CD25⁺ Treg cells can be further divided into two subpopulations, the natural Treg cells that emerge from the thymus (1, 20-22) and the inducible Treg cells that are induced and converted in the periphery from CD4⁺CD25⁻ naive T cells (22-24). The CD4⁺CD25⁺ Treg cells isolated from clinical samples of viral infections may contain either or both of these two subsets of Treg cells. Phenotypically, they are indistinguishable in peripheral blood or tissues.

3.1. Natural CD4⁺CD25⁺ Treg cells

This type of Treg cells are generated and matured in the thymus and then exported to the periphery. They are known as natural Treg cells. They represent 5 - 10% of peripheral CD4⁺ T cells in mice and less than 5% in humans and constitutively express CD25, the α subunit of the IL-2 receptor, on their surface (1, 25, 26). The thymus origin of these cells was demonstrated by Asano *et al* who showed that neonatal mice thymectomized on day 3 after birth lacked CD4⁺CD25⁺ Treg cells and developed autoimmune gastritis and high titers of circulating antiparietal cell autoantibodies (25). Further studies have demonstrated that natural CD4⁺CD25⁺ Treg cells are potent suppressor T cells that mediate strong immunosuppression in a contact-dependent but cytokine-independent manner (26-30). *In vitro* studies have shown that they are anergic to polyclonal, allogenic, or antigen-specific stimulation (27, 31, 32). However, they are able to proliferate *in vivo* in response to T cell receptor (TCR)-mediated stimulation

(33). Their suppressive function requires their activation via TCR stimulation, and this activation is independent of co-stimulation mediated by CD28/CTLA-4 interactions with CD80/CD86 (26-29, 34-36). In addition to CD25, they also express, either constitutively or specifically, several receptors and molecules that may participate in their development and function. These include cytotoxic T lymphocyte-associated antigen-4 (CTLA-4) (37), glucocorticoid-induced tumor necrosis factor receptor (GITR) (38), and Forkhead box P3 transcription factor (FOXP3) (39).

3.2. Inducible CD4⁺CD25⁺ Treg cells

This type of Treg cells are generated in the peripheral and are converted from CD4⁺CD25⁻ T cells under certain conditions. Generation of CD4⁺CD25⁺ Treg cells from CD4⁺CD25⁻ precursor has been demonstrated both *in vitro* (40, 41) and *in vivo* (42). The conversion can be induced by cytokines such as IL-10 (43) and TGFβ (24, 41, 44), dendritic cells (DC) (23, 45), alloantigens (42), superantigens (46), or costimulatory signals (40). Unlike thymus-derived natural Treg cells that show exclusively cytokine-independent suppression, peripheral-converted CD4⁺CD25⁺ Treg cells may act through a cytokine-dependent mechanism (41). Notably, not all conditions that induce CD25 expression to convert the phenotype from CD4⁺CD25⁻ to CD4⁺CD25⁺ will result in the generation of regulatory T cells with suppressive function. This is illustrated by Thornton *et al* who showed that CD4⁺CD25⁺ T cells generated by Con A stimulation of CD4⁺CD25⁻ T cells purified from normal mice failed to suppress the proliferative responses of the CD4⁺CD25⁻ cells (27). This suggests that the conversion of naïve CD4⁺CD25⁻ T cells to CD4⁺CD25⁺ Treg cells is likely a complex process which may require appropriate signals and cytokine milieu, and that mere expression of CD25 is insufficient to render the cells capable to mediate suppression.

4. INDUCTION OF CD4⁺CD25⁺ TREG CELLS IN VIRAL INFECTIONS

The existence of Treg cells and their involvement in immunosuppression during viral infection was observed decades ago in several animal models. In 1975, Toy *et al* observed that cellular immunity was suppressed *in vitro* by Friend virus-infected mouse spleen cells (47). Similarly, Carpenter *et al* observed that suppressor cells from the spleens of reticuloendotheliosis virus (REV)-infected chicken severely inhibited the ability of spleen cells from uninfected chickens to respond to PHA, and the inhibition required cell-cell contact between suppressor and target cells (48). In fact, such virus-induced suppressor cells were reported in a large number of viral infections in various animal models such as in mice infected by Moloney sarcoma virus (49, 50), Friend leukemia virus (47) and AK virus (51), in cats infected by feline leukemia virus (52) and feline immunodeficiency virus (FIV) (53), and in chickens infected by REV (54) and Marek's disease virus (55). In humans, suppressor cells were found in patients infected with measles virus (56), herpes simplex virus (HSV) (57), and reovirus (58). Despite the clear evidence for the presence of such suppressor cells in various types of

viral infections, the nature of these suppressor cells were not clear at that time.

It was not until recent years that the role of some T cells, particularly CD4⁺ T cells, in mediating immunosuppressive effects during virus infection was demonstrated. In 2001, Iwashiro *et al* showed that CD4⁺ T cells from mice chronically infected with Friend virus were able to suppress antitumor immune responses *in vivo* after being adoptively transferred into uninfected mice and also inhibit CTL generation in mixed lymphocyte cultures *in vitro* (11). This was the first evidence showing the induction of CD4⁺ Treg cells in viral infection. Since then, the induction of CD4⁺ Treg cells, particularly CD4⁺CD25⁺ Treg cells, has been demonstrated in many human and animal viral infections including HIV (13, 46), hepatitis B virus (HBV) (59, 60), HCV (15, 61, 62), HSV (12, 16), EBV (63) cytomegalovirus (CMV) (46), human T cell lymphotropic virus type 1 (HTLV-1) (64), FIV (53), murine acquired immunodeficiency syndrome (MAIDS) virus (65), Coxsackievirus B3 (CVB3) (66), dengue virus (19), and simian immunodeficiency virus (SIV) (67). The induction is often manifested by the increased number of CD4⁺CD25⁺ Treg cells in the peripheral blood or local tissues of virus-infected patients, and this increase is always associated with certain changes in host antiviral immune responses or disease status. For example, in HIV infection, the increased frequency of CD4⁺CD25⁺ Treg cells in patient peripheral blood has been related to low peripheral blood CD4⁺ T cell counts and polarization toward a Th2 immune response (68). In chronic severe HBV infection, the frequencies of CD4⁺CD25⁺ Treg cells in both peripheral blood and liver-infiltrating lymphocytes were significantly increased, and this increase correlated with serum viral load (69). Since it is impossible to differentiate natural from inducible CD4⁺CD25⁺ Treg cells in the peripheral, it is not known whether the increase in CD4⁺CD25⁺ Treg cell frequency after virus infection results from the expansion of natural Treg cells or the conversion of naïve CD4⁺CD25⁻ T cells, or both. Another indication of CD4⁺CD25⁺ Treg cell induction in viral infections is their activation. Normal CD4⁺CD25⁺ Treg cells require activation through the engagement of their TCR to become suppressive (27). However, CD4⁺CD25⁺ Treg cells isolated from virus-infected animals or patients are fully functional and do not need additional TCR-mediated activation. Evidently, these cells have been activated *in vivo* after virus infection, most likely through the engagement of their TCR with viral antigens or other undefined mechanisms.

5. ROLE OF VIRUS-INDUCED CD4⁺CD25⁺ TREG CELLS

As a component of the immune system, CD4⁺CD25⁺ Treg cells are expected to participate in host antiviral immune responses with the goal to protect the host from undue damage by viruses and restore homeostasis. However, their role in viral infections appears complicated and it is still difficult to predict the exact role that they will play in any given virus infection. Whether they act to benefit the host or virus may depend on various factors

such as the nature and the stage of the infection and immune activation.

5.1. Role of CD4⁺CD25⁺ Treg cells in viral pathogenesis

Numerous studies have shown that CD4⁺CD25⁺ Treg cells suppress host antiviral immune responses. Although it is not known whether the suppression represents an active strategy of viruses to weaken host defense against them or just a “side-effect” of host antiviral immune responses, it provides viruses with the opportunity to evade host immune attack and facilitate their spread and pathogenic effects. The targets of suppression by CD4⁺CD25⁺ Treg cells include both CD4⁺ and CD8⁺ effector T cells as well as other cells that may be involved in antiviral immune responses such as dendritic cells (70), monocytes/macrophages (71), and NK cells (72, 73). Since CD8⁺ cytotoxic T cells play a major role in antiviral immune responses, it is not surprising that they are the prime target of CD4⁺CD25⁺ Treg cell suppression. Indeed, a large body of studies have shown that virus-specific CD8⁺ T cell responses including their proliferation, IFN γ production, perforin expression, and cytotoxic activities are suppressed by CD4⁺CD25⁺ Treg cells in many human and animal viral infections (14, 15, 46, 60, 69, 74-77). As a result, host antiviral immune responses are impaired and infections progress. Interestingly, not all functions of CD8⁺ T cells are suppressed in every virus infection. For example, suppression of CD8⁺ T cell effector functions without affecting their proliferation and activation was reported in mouse chronically infected with Friend retrovirus (74, 77), although suppression of CD8⁺ T cell proliferation was observed in many other viral infections. The question is what influences the CD4⁺CD25⁺ Treg cells to selectively suppress one function of CD8⁺ T cells but not another. In addition to CD8⁺ T cells, the proliferation and effector functions of CD4⁺ T cells are also suppressed by virus-induced CD4⁺CD25⁺ Treg cells in many viral infections (13-15, 18, 19).

For many chronic or persistent infections, how viruses escape host immune surveillance and what leads to the development of persistent infection have been the hot topics of investigation. One possible mechanism appears to be the induction of CD4⁺CD25⁺ Treg cells at the early stage of acute infection that dampens the T cell antiviral responses resulting in their inability to completely clear the virus. For example, in HCV infection, virus-specific T cells have been shown to arise early after infection and strong T cell responses are induced during acute phase of infection (78, 79). This rapid response of immune system is necessary for quick clearance of the virus and control of the infection. However, complete clearance of the virus and full recovery only occur in a small percentage of HCV-infected patients and most patients develop chronic disease with persistent infection. The reason for the failure to clear the virus in most of the patients during the initial antiviral T cell immune responses still remains unknown. However, weak or unsustained acute CTL responses have been linked to the development of chronic disease (79, 80). One possible explanation for the weak CD8⁺ T cell responses during acute viral infection is the early induction of CD4⁺CD25⁺ Treg cells upon infection. Recently, Perrella *et*

al observed that the frequency and suppressive activity of CD4⁺CD25⁺ Treg cells were increased in the majority of the patients with acute HCV infection (81). Similarly, in SIV-infected rhesus macaque and African green monkey models, Estes *et al* (82) and Kornfeld *et al* (67) also observed a rapid induction of CD4⁺CD25⁺ Treg cells during acute SIV infections. More significantly, the early induction of CD4⁺CD25⁺ Treg cells during acute infection was associated with the failure of viral clearance, as patients with elevated frequency and function of CD4⁺CD25⁺ Treg cells progressed to chronic infection, whereas those who showed no significant increase in Treg frequency or function during acute phase of infection cleared the virus and fully recovered from the disease (81). It is highly likely that these Treg cells induced at the early stages of infection suppressed CD8⁺ T cell functions during acute infection, resulting in the failure of viral clearance. In support of this hypothesis, Suvas *et al* demonstrated in a mouse model of acute HSV infection that depletion of CD4⁺CD25⁺ Treg cells before infection noticeably accelerated the viral clearance (12). These observations provided support for the proposition of early induction of CD4⁺CD25⁺ Treg cells as one of the mechanisms for developing chronic or persistent viral infections and raised the possibility to use this induction as a clinical predictor for the prognosis of acute viral infection.

Early induction of CD4⁺CD25⁺ Treg cells is likely programmed to control the magnitude of CD8⁺ T cell activity to ensure that no host damage will result from the strong antiviral immune responses which are often accompanied by local or systemic inflammation. However, the induction may have been exaggerated or occurred too early, as in the case demonstrated by Estes *et al* in acute SIV infections (82), where it dampens the CD8⁺ T cell response before the viruses have been completely cleared. What triggers the premature induction of these Treg cells remains to be determined. Rapid replication and spread of viruses, together with acute inflammation or massive activation of antiviral immune responses during acute infections, may create a microenvironment that facilitates premature expansion and activation of CD4⁺CD25⁺ Treg cells. Thus, it appears that the magnitude and timing of the early induction of CD4⁺CD25⁺ Treg cell activity may play a decisive role in the fate of the acute infection.

Amazingly, viruses can not only induce CD4⁺CD25⁺ Treg cells to suppress host antiviral immune responses to aid their infection, but also inhibit the function of these cells leading to the dysregulation of host immune response that favors their pathogenic effects and development of latency. This was demonstrated in a mouse model with γ -herpesvirus 68 infection, in which direct viral infection of CD4⁺CD25⁺ Treg cells decreased the frequency and function of these cells and resulted in an uncontrolled expansion of leukocytes and the establishment of viral latency (83). Such inhibition of CD4⁺CD25⁺ Treg cell function was also observed in a human retroviral infection in which CD4⁺CD25⁺ Treg cells defective in their suppressive function were found in patients infected with HTLV-1 virus (17). An HTLV-1 viral protein, Tax, was identified as responsible for the direct inhibition of Treg

cell function. These patients showed high levels of immune activation and developed a neurological inflammatory disease termed HTLV-1-associated myelopathy/tropical spastic paraparesis (HAM/TSP), which may result from a break in immunological self tolerance. In these studies, both γ -herpesvirus 68 and HTLV-1 viruses directly infected CD4⁺CD25⁺ Treg cells. Conceivably, the expression of viral proteins, such as HTLV-1 Tax protein, or replication of these viruses within the Treg cells inhibited the expression of certain cellular proteins essential for the suppressive activities of CD4⁺CD25⁺ Treg cells, such as FOXP3, GITR and HTLA-4 (17), and thus impaired the function of these cells. Interestingly, another virus, FIV, that also directly infects CD4⁺CD25⁺ Treg cells, did not show suppressive effects on the cells (53). Instead, infection by FIV phenotypically and functionally activated the Treg cells.

5.2. Role of CD4⁺CD25⁺ Treg cells in host defense

5.2.1. Control of inflammatory responses

Virtually all viral infections induce host inflammatory responses, which may represent an essential component of host defense mechanisms against viral infections. However, excessive inflammation can cause tissue damage and thus be harmful to the host if left uncontrolled. CD4⁺CD25⁺ Treg cells have been suggested to play a critical role in controlling exaggerated inflammatory responses accompanied with many acute and chronic viral infections. A recent study on acute dengue virus infection showed that CD4⁺CD25⁺ Treg cells, which were induced during the acute dengue virus infection, are able to suppress the production of inflammatory cytokines including IFN γ , TNF α , and IL-6 from T cells and monocytes *in vitro* in response to dengue virus antigens (19). The disproportionate production of these inflammatory cytokines has been thought to contribute to the severity of dengue disease since they cause plasma leakage in the patients (84). More importantly, a significant increase in CD4⁺CD25⁺ Treg cells was found in patients with mild disease symptoms but not in those with severe disease. This is a strong indication that the activity of CD4⁺CD25⁺ Treg cells contributes to the control of inflammatory phase of the disease and is beneficial for disease outcome. The role of CD4⁺CD25⁺ Treg cells described here in acute dengue virus infection seems not in agreement with that described above in acute HCV infection. This may be due to the difference between the two viruses and the infections they cause. Unlike HCV, Dengue virus rarely causes chronic infection and the virus-induced inflammation is much more severe during the acute infection. This is perhaps a good example for the conception that the role of CD4⁺CD25⁺ Treg cells varies with the nature of the virus and infection.

If CD4⁺CD25⁺ Treg cells are critical in controlling virus-induced inflammation, one would expect that the absence of these cells would negatively impact the disease outcome. Indeed, in a mouse model of human HSV stromal keratitis, which usually results from a T cell-mediated immunoinflammatory response to HSV infection in the corneal stroma, the pathological lesion is significantly more severe if mice were depleted of

CD4⁺CD25⁺ Treg cells before infection (16). Similarly, in an influenza virus hemagglutinin (HA)-induced mouse uveoretinitis model, depletion of CD4⁺CD25⁺ Treg cells exacerbated intraocular inflammation, whereas injection of mice with HA-specific CD4⁺CD25⁺ T cells controlled the disease (85).

One negative aspect for the function of CD4⁺CD25⁺ Treg cells in controlling virus-induced inflammatory responses is that the suppression is not selective or specific. Thus, while CD4⁺CD25⁺ Treg cells exert their suppression on inflammatory responses, simultaneous suppression may occur on virus-specific T cell responses necessary for virus clearance. This is best illustrated in chronic HCV infection. While a significant inverse correlation was found between the frequency and HCV-specific TGF β response of CD4⁺CD25⁺ Treg cells and liver inflammation (15, 18), the same group of chronic HCV patients also showed a positive correlation between these Treg cells and viral load (15). Thus, CD4⁺CD25⁺ Treg cells acted to protect the host through controlling viral infection-associated liver inflammation. However, this protection is at the expense of reduced antiviral T cell responses and increased risk for virus spread.

5.2.2. Control of hyperactivation of immune responses

Chronic or persistent immune activation is a characteristic of certain human and animal retroviral infections such as HIV (86), and has been suggested to be responsible for the disease deterioration. In HIV and SIV infections, elevated immune activation induced by virus infection has been thought to result in CD4⁺ T cell depletion (87-89), a hallmark of AIDS progression. An increasing line of evidence indicates that the hyperactivation of T cells may be controlled by CD4⁺CD25⁺ Treg cells, which are induced in most retroviral infections (13, 46, 53, 64, 65, 67, 82). The induced CD4⁺CD25⁺ Treg cells are able to suppress virus-specific CD4⁺ and CD8⁺ T cell responses *in vitro* in a cell contact-dependent and cytokine-independent manner. Decrease in the frequency of these Treg cells as HIV infection progresses has been found to be associated with immune activation of CD4⁺ and CD8⁺ T cells in HIV patients (90). Importantly, patients with strong HIV-specific CD4⁺CD25⁺ Treg cell functions *in vitro* showed favorable clinical markers of disease status such as significantly lower levels of plasma viremia and higher CD4⁺ to CD8⁺ T cell ratios than those without Treg cell activity (14). This suggests that CD4⁺CD25⁺ Treg cells may play an important role in protecting HIV patients from progression to AIDS. In support of this, a recent study found a direct correlation between the preservation of CD4⁺ T cell counts and high percentage of CD4⁺CD25⁺ Treg cells in SIV-infected sooty mangabeys (91) which, in marked contrast to HIV-infected humans, usually show normal CD4⁺ T cell counts and lack of generalized immune activation, and rarely progress to AIDS despite chronic high levels of virus replication (92).

Another piece of evidence supporting the beneficial role of CD4⁺CD25⁺ Treg cell suppression of HIV-specific T cell responses is the study of HIV-1

infection in young children. It has been observed that vertical transmission of HIV-1 *in utero* only occurs in a very low percentage (only 3% - 15%) of HIV-1-exposed infants (93). A recent study has found that the virus-specific T cell responses are weak in HIV-1-exposed children. The weak T cell responses are likely due to the suppression by CD4⁺CD25⁺ Treg cells, since the responses become strong after the Treg cells are removed (94). This suggests that CD4⁺CD25⁺ Treg cells may contribute to suppress the activation of virus-specific T cells and thus protect against infection in young children.

The studies described above demonstrate the benefit of CD4⁺CD25⁺ Treg cell induction in viral infections that induce immune hyperactivation. Conceivably, their absence or a deficiency in their suppressive function may lead to progression of the infection and deterioration of the disease. Indeed, in patients with HTLV-1-associated HAM/TSP, the frequency of CD4⁺CD25⁺ Treg cells actually increased when compared to the HTLV-1-infected asymptomatic carriers and uninfected healthy donors (17). However, these cells are defective in their suppressive functions and unable to control the hyperactivation of virus-specific T cell immune responses in HTLV-1 infection (17). This leads to the progression of HTLV-1 infection and the manifestation of HTLV-1-associated HAM/TSP. As discussed above, the loss of suppressive function may result from their direct infection by HTLV-1 virus. This could also represent one of the potential mechanisms for the failure of these Treg cells to successfully control chronic activation of T cells in AIDS patients.

5.2.3. Control of virus-associated autoimmune disorders

Viral infection has long been associated with the development of autoimmune diseases (95). For example, patients with chronic HCV infection often develop mixed cryoglobulinemia (MC), an autoimmune disorder characterized by polyclonal B cell activation and autoantibody production (96). Other autoimmune disorders have also been associated with chronic HCV infection, including membranoproliferative glomerulonephritis, autoimmune thyroiditis and lymphoproliferative disorders, autoimmune thrombocytopenia, pruritus, and type II diabetes mellitus (97-99). In addition, many viruses are associated with the development of type I diabetes (100), multiple sclerosis (101, 102), myocarditis (66, 103, 104), and systemic lupus erythematosus (105) in human and animals. One mechanism for the onset of the autoimmune diseases after virus infection may be the reduction or impairment of the CD4⁺CD25⁺ Treg cell suppressive function following virus infection, which leads to the breakdown of self tolerance and the enhancement of autoimmune T cell responses. This is implicated by the finding that the frequency of CD4⁺CD25⁺ Treg cells was significantly reduced in chronic HCV-infected patients who developed MC symptoms as compared to those with asymptomatic MC, no MC, or healthy controls (106). In this particular case, the CD4⁺CD25⁺ Treg cell deficiency likely enhanced the CD4⁺ T cell help to autoantibody-secreting B cells leading to the development of MC symptoms. Consistently, in an animal model, depletion of

CD4⁺CD25⁺ Treg cells in reovirus-infected mice resulted in the development of severe insulinitis leading to an overt early diabetes (107). Thus, failure to maintain Treg cell function may be a critical determinant for disease expression in virus-induced autoimmune diseases.

6. MECHANISMS OF CD4⁺CD25⁺ TREG CELL INDUCTION IN VIRAL INFECTION

To date, many questions regarding the mechanisms for CD4⁺CD25⁺ Treg cell induction during viral infections still remain to be answered. Direct evidence on the induction of these cells by viruses or viral proteins is very limited. How these cells are generated and activated following virus infection is largely unknown, especially for the natural Treg cells since they are generated in thymus and are assumed to respond only to self-antigens (108). The induction of CD4⁺CD25⁺ Treg cells is not likely a selective event, since other CD4⁺ and/or CD8⁺ T cells are also induced at the onset of infection. In addition, both CD4⁺CD25⁺ Treg cells and CD4⁺ helper T cells have been induced against the same epitopes on the viral protein in HCV-infected patients (61), suggesting that the same signal that induces regular CD4⁺ T cells can also induce Treg cells simultaneously. One possibility is that, instead of selective induction of the Treg cell population, all CD4⁺ T cells are induced upon infection, and then the activated CD4⁺CD25⁺ Treg cells suppress effector CD4⁺ T cell functions. If this is the case, a kinetic change in activation of the CD4⁺ T cell populations should be detectable. Data from many studies have suggested that CD4⁺CD25⁺ Treg cells can be induced through a number of mechanisms during virus infection.

6.1. Viral antigens

To date, information on direct induction of CD4⁺CD25⁺ Treg cells *in vivo* by viral antigens is still limited, although *in vitro* responses of these cells to viral proteins and peptides have been reported in several studies (13, 15, 109). Whether viral antigens directly activate and expand CD4⁺CD25⁺ Treg cells *in vivo* still remains to be demonstrated. However, this possibility is strongly suggested by observations from a large number of studies. First, virus-specific CD4⁺CD25⁺ Treg cells can be rapidly induced *in vitro* by culturing PBMC from virus-infected patients with viral peptides, as shown in HCV infection (110), suggesting that these cells have previously been primed by cognate HCV viral antigens *in vivo*. Furthermore, it has been found that only a small number of peptides from a peptide pool can induce Treg cells and the actual peptides that can stimulate Treg cells vary between patients, indicating the existence of dominant epitopes for CD4⁺CD25⁺ Treg cells. Second, CD4⁺CD25⁺ Treg cells specific to viral antigens can be found in virus-infected patients. Studies from Weiss *et al* showed that HIV viral protein p24 readily induced IL-10 production and TGFβ mRNA expression by CD4⁺CD25⁺ Treg cells from HIV-infected patients (13). This induction was p24-specific since CMV and PPD showed no or low stimulation of IL-10 and TGFβ expression in these cells. Similar specific responses to viral antigens has also been observed for CD4⁺CD25⁺ Treg cells from patients with HCV infection (15). These studies demonstrated the existence of virus-

specific CD4⁺CD25⁺ Treg cells *in vivo* and suggested that expansion and activation of CD4⁺CD25⁺ Treg cells *in vivo* may be triggered by viral antigens. Third, as discussed above, CD4⁺CD25⁺ Treg cells isolated from virus-infected patients are fully functional and do not need additional activation for their suppressive function. Apparently, they have been activated *in vivo* following virus infection. Since the acquisition of their suppressive function requires their activation via TCR (27), it is highly likely that this TCR-dependent activation *in vivo* is achieved through the engagement of their TCR with viral antigens. Finally, a more direct implication comes from a study by Huber *et al*, who showed that a single nonconserved amino acid difference in the nuclear capsid proteins of Coxsackievirus B3 H3 and H310A1 strains completely changed their capabilities to induce CD4⁺CD25⁺ Treg cells (66). Thus, a direct interaction between viral proteins or peptides and Treg cells is likely involved in Treg cell induction.

Several possibilities exist for direct induction of CD4⁺CD25⁺ Treg cells by viral antigens *in vivo*. As discussed above, viral infections often trigger autoimmune responses, and one of the ways this could occur is through the mechanism of molecular mimicry. It has been shown that viral peptides with sufficient structural similarity to self-peptides can activate T cell clones that are specific only for autoantigens such as myelin basic protein (111, 112). Such molecular mimicry between viral and host proteins has been observed in a number of viruses such as measles virus (113), HBV (111), HSV (112, 113), EBV (63, 112), and Adenovirus type 12 (112). Autoantibodies have been found frequently in the sera of patients infected with hepatitis, chickenpox, herpes, mumps, or measles viruses (114). In fact, it was concluded a long time ago that molecular mimicry is a common phenomenon following the finding that many human and animal viruses share antigenic determinants with host cell proteins (114, 115). Although the mechanism of molecular mimicry has not been confirmed for CD4⁺CD25⁺ Treg cell induction in viral infections, it still remains a plausible hypothesis for viral antigen-mediated direct induction of CD4⁺CD25⁺ Treg cell activation, especially for the induction of thymus-derived natural Treg cells since they are considered to only respond to autoantigens. In addition, many human and animal retroviruses contain highly conserved immunosuppressive peptides in their envelope proteins (116-119). These peptides are able to suppress immune responses of lymphocytes, monocytes, NK cells and macrophages. It remains to be determined whether these immunosuppressive viral peptides directly induce CD4⁺CD25⁺ Treg cell generation and/or activation. However, this proposition is encouraged by the study of Walker *et al*, who demonstrated that antigen-specific CD4⁺CD25⁺ Treg cells can be generated *in vitro* from either naïve or memory CD4⁺ T cells using influenza virus hemagglutinin epitopes (120). Moreover, viral antigen may induce generation of CD4⁺CD25⁺ Treg cells from regular CD4⁺ T cells in chronic viral infections by long-term, low-dose stimulation of these cells. This resembles the so-called "low-zone tolerance" that describes the antigen-specific tolerance resulting from long-term stimulation with sub-immunogenic doses of antigens (121). Finally, the

induction may also be achieved by superantigens expressed by some viruses such as mouse mammary tumor virus (122).

6.2. Virus-induced cytokines

The cytokines that have been studied extensively in the context of Treg cell generation and activation are TGFβ and IL-10, due to their pronounced immunosuppressive nature. Accumulating data indicate that TGFβ plays an important role in induction of CD4⁺CD25⁺ Treg cells. However, this induction seems limited only in peripheral through either expansion of a small number of thymus-derived natural CD4⁺CD25⁺ Treg cells maintained in peripheral (123, 124) or conversion of peripheral naïve CD4⁺CD25⁺ T cells into FOXP3-expressing CD4⁺CD25⁺ Treg cells (24, 125). The latter may be due, at least partly, to the capability of TGFβ to upregulate the expression of CD25, FOXP3, and CTLA-4 on activated naïve CD4⁺ T cells (24, 41, 44, 125-127). It seems clear that TGFβ is not required for thymic generation of CD4⁺CD25⁺ Treg cells, since several studies have shown that CD4⁺CD25⁺ Treg cells develop normally in thymus in both TGFβ1^{-/-} mice (128, 129) and transgenic mice defective in TGFβ1 signaling specifically in T cells and thymocytes (127, 130, 131). Thus, the role of TGFβ is most likely to maintain the peripheral CD4⁺CD25⁺ Treg cell pool and expand the cells when necessary.

The effect of virus-induced cytokines on the induction of CD4⁺CD25⁺ Treg cells during viral infections still remains to be fully illustrated. Elevated levels of TGFβ expression and production have been detected in many viral infections including HIV (132, 133), SIV (82), HCV (134, 135), EBV (136), CMV (137), dengue virus (138), influenza virus (139), HTLV-1 (140), and vaccinia virus (141) infections. The elevated TGFβ levels may represent either a strategy utilized by virus to induce immunosuppression for their invasion, or host protective anti-inflammatory response against virus-induced inflammation, or both. The induction of TGFβ may occur very rapidly after infection, as shown in a SIV-infected Africa green monkey model, in which the upregulated TGFβ expression was detected as early as 24 hours after infection (67). Importantly, the strong induction of TGFβ1 was correlated with both upregulation of FOXP3 expression and increased levels of CD25⁺ T cells in the infected animals, indicating an active role of virus-induced TGFβ in expansion and activation of CD4⁺CD25⁺ Treg cells in viral infection.

The capability of IL-10 to induce CD4⁺CD25⁺ Treg cells *in vivo* was demonstrated in a mouse model of type I diabetes, in which overexpression of IL-10 using an IL-10-expressing adeno-associated virus vector induced CD4⁺CD25⁺ Treg cells *in vivo* and ameliorated the disease (142). The induction of the Treg cells by IL-10 seems to be an indirect event mediated through dendritic cells. IL-10 has been shown to inhibit the upregulation of the costimulatory molecules including CD40, CD80, CD86 on DC (143, 144). Consequently, it arrests the development of fully matured DC but leads to the generation of the so-

called “regulatory DC” with reduced expression of the costimulatory molecules similar to immature DC. These regulatory DC have been shown to induce CD4⁺CD25⁺ Treg cells (23, 144). Similar to TGFβ, IL-10 production is also induced in many human and animal viral infections (145-150). Notably, some viruses, such as EBV (151) and parapoxvirus orf virus (152) and equine herpesvirus 2 (153), encode a viral homolog of cellular IL-10. Like cellular IL-10, viral IL-10 is also able to inhibit DC maturation and function and promote generation of immature DC that promote T cell anergy (154, 155).

6.3. Virus-induced DC

DC are potent antigen-presenting cells (APC) essential for activation of both virus-specific CD4⁺ and CD8⁺ T cells. Like TGFβ, DC have no effect on thymic generation of CD4⁺CD25⁺ Treg cells. Increasing evidence from recent studies have shown that DC, under appropriate conditions, are able to either directly induce antigen-specific expansion of functional CD4⁺CD25⁺ Treg cells both *in vitro* and *in vivo* by stimulating extensive proliferation of these Treg cells (156, 157), or induce naïve CD4⁺ T cells to differentiate into CD4⁺CD25⁺ Treg cells with strong suppressive function (45). Thus it is perceivable to expect the involvement of DC, whether active or passive, in expansion and activation of CD4⁺CD25⁺ Treg cells in viral infections. In fact, many viral infections have shown dysregulation of DC during infection. This includes reduced expression of MHC class II and co-stimulatory molecules such as CD40, CD80, and CD86, and impaired function in effector T cell stimulation (158-161). The dysregulation of DC may result from direct infection by viruses (162) or from the suppression by virus-induced immunosuppressive cytokines such as IL-10, as described above. These DC are able to induce CD4⁺CD25⁺ Treg cells (23, 144). Interestingly, the study by Moseman *et al* showed that activated allogeneic plasmacytoid dendritic cells (PDC) can efficiently induce naïve CD4⁺CD25⁺ T cells to differentiate into suppressive CD4⁺CD25⁺ Treg cells (45). This raised the possibility for the existence of a specialized DC subset dedicated to the CD4⁺CD25⁺ Treg cell induction. Similar function of virus- or CpG-stimulated PDC to induce other types of Treg cells such as Tr1 and CD4⁺ cytotoxic Treg cells has also been reported (163, 164). PDC have been shown to play a central role in innate immunity against viral infections (165) and many studies have shown that PDC are activated by viral infections (164, 166-168). A striking feature of the activated PDC is their production of high levels of IFNα upon viral infections (169), which is an essential component of innate immunity against viral infections (170). Interestingly, IFNα has been shown to synergize with IL-10 in priming CD4⁺ T cells to differentiate into Tr1 regulatory T cells (164, 171).

7. MECHANISMS OF SUPPRESSION BY VIRUS-INDUCED CD4⁺CD25⁺ TREG CELLS

The molecular mechanisms by which Treg cells suppress effector T cells are still not fully understood. However, enormous progress has been made towards our understanding on how virus-induced Treg cells exert their suppressive functions on effector T cells. It has been

generally accepted that CD4⁺CD25⁺ Treg cells exert their suppressive function through direct cell-to-cell contact with effector T cells or APC. However, controversial results have been obtained regarding cytokine involvement in mediating CD4⁺CD25⁺ Treg cell suppression. While most *in vitro* studies have shown that suppression by CD4⁺CD25⁺ Treg cells is cytokine-independent, a large body of *in vivo* studies has demonstrated the requirement of TGFβ and IL-10 for suppression by CD4⁺CD25⁺ Treg cells (3, 130, 172-176). Nevertheless, accumulating evidence has suggested that suppression by CD4⁺CD25⁺ Treg cells may be mediated by a variety of mechanisms and that the molecules and pathways involved in mediating the suppression may vary with conditions or stages of disease and/or immune responses.

7.1. FOXP3

FOXP3 is specifically expressed in CD4⁺CD25⁺ Treg cells and has been suggested to be a master regulator for the development and function of CD4⁺CD25⁺ Treg cells (39, 177-179). This is based on the observations that induction of FOXP3 expression is usually accompanied by an acquisition of suppressive function of CD4⁺CD25⁺ Treg cells, and loss of FOXP3 expression abrogates the ability of the cells to suppress effector T cell responses. In viral infections, this is best illustrated in patients infected with HTLV-1 virus. Infection of CD4⁺CD25⁺ Treg cells by HTLV-1 virus leads to the reduction of FOXP3 mRNA expression and protein production. This abrogates the ability of HTLV-1-infected Treg cells to suppress the proliferation of CD4⁺CD25⁺ T cells, resulting in the development of HAM/TSP in HTLV-1-infected patients (17, 180). Since FOXP3 is essential for the development of peripheral CD4⁺CD25⁺ Treg cells (39, 177-179, 181, 182), it is not clear whether the loss of CD4⁺CD25⁺ Treg cell activity is due to a decrease in the frequency of the CD4⁺CD25⁺ T cells with regulatory capability in the total CD4⁺CD25⁺ T cell population, or due to the loss of function of CD4⁺CD25⁺ Treg cells. However, increased production of IL-2 was detected in these cells, suggesting that reduction in FOXP3 expression may lead to a defect in their regulatory function rather than a decrease in their frequency, since normal functional CD4⁺CD25⁺ Treg cells do not produce IL-2 (27, 28, 31). Interestingly, Sereti *et al* found that high levels of FOXP3 expression can be induced in a long-lived population of cytokine-expanded naïve (CEN) CD4⁺CD25⁺ T cells in HIV-infected patients receiving IL-2 immunotherapy (183). These IL-2-expanded FOXP3⁺CD4⁺CD25⁺ T cells are phenotypically and functionally distinct from naturally occurring CD4⁺CD25⁺ Treg cells and do not display strong suppressive activity. This raised the question about the previous statement that only CD4⁺CD25⁺ Treg cells express FOXP3 and suggested that expression of FOXP3 may not necessarily confer immunosuppressive activity on the cells.

7.2. CTLA-4

Because CD4⁺CD25⁺ Treg cell suppression is dependent on direct cell contact, the surface molecules on Treg cells, target cells, and the cells with potential involvement in CD4⁺CD25⁺ Treg cell function have gained much attention in recent years. Among them, CTLA-4 has

drawn widespread interest because of its high constitutive expression on CD4⁺CD25⁺ and other Treg cells (37). A number of studies have implicated that CTLA-4 expression plays a key role in mediating the suppressive function of CD4⁺CD25⁺ Treg cells, as *in vivo* blockade of CTLA-4 abolished the suppressive activities of these Treg cells to control inflammatory (174, 175) or autoimmune (37) diseases. A similar role of CTLA-4 has also been observed in viral infections. In mice infected with Friend virus, virus infection induced CD4⁺CD25⁺ Treg cells and these cells suppressed allogeneic CTL generation *in vitro* (11). Blocking CTLA-4 activity with a monoclonal anti-CTLA-4 antibody reversed this Treg cell-mediated suppression and restored the CTL generation. In another study with SIV-infected macaques, increased effector function of both SIV-specific CD4⁺ and CD8⁺ T cells and decreased viral load in lymph nodes were detected after treating the animals with the anti-CTLA-4 antibody that blunted CD4⁺CD25⁺ Treg cell suppression (184). These studies provide evidence for the essential role of CTLA-4 in mediating the suppressive effect of CD4⁺CD25⁺ Treg cells on host antiviral immune response. In many viral infections, such as in HIV (185, 186) (187), SIV (82), HCV (188, 189), influenza (190), and EBV (191) infections, upregulated expression of CTLA-4 has been observed in parallel to the induction of CD4⁺CD25⁺ Treg cells. There is also a recent report on the direct effect of HIV-1 viral protein Vpr on the induction of CTLA-4 in HIV-infected T cells (192).

How CTLA-4 mediates the suppressive effect of CD25⁺CD4⁺ Treg cells is still under investigation. One commonly accepted mechanism is that it functions through its interaction with costimulatory molecules CD80 and CD86 on APC and/or effector T cells (193). Ligation of CTLA-4 to CD80/CD86 on APC either induces the production of the immunosuppressive enzyme indoleamine 2,3-dioxygenase (IDO) (194, 195) and the generation of inhibitory tryptophan metabolites (196-198), or competes with CD28 on effector T cells to bind to CD80/CD86 on APC and thus blocks the costimulatory signaling pathway for effector T cell activation. This seems true for CTLA-4 in viral infections, as elevated IDO expression was detected in viral infections in both animal model (188, 189) and patients infected with HCV (188). More convincingly, decreased IDO expression was detected in SIV-infected macaques following CTLA-4 blockade, which apparently diminished the suppressive effect of CD4⁺CD25⁺ Treg cells, as increased SIV-specific effector T cell function and decreased viral load were detected in these animals (184).

7.3. TGFβ

Because of its marked immunosuppressive property and elevated production following CD4⁺CD25⁺ Treg cell activation in many situations, TGFβ has been examined extensively for its role in mediating CD4⁺CD25⁺ Treg cell suppression. The results obtained are rather controversial or confusing. While a number of *in vitro* studies have shown that suppression by CD4⁺CD25⁺ Treg cells does not involve the activity of TGFβ (27, 28, 31, 199, 200), several *in vivo* studies have demonstrated the importance of TGFβ in CD4⁺CD25⁺ Treg cell suppression (130, 172, 174-176). However, the latter is debatable as

well, as some studies have also shown the independence of CD4⁺CD25⁺ Treg cell function on TGFβ *in vivo* (66, 128, 200). In an effort to explain these above discrepancies, Nakamura *et al* made an interesting finding that mouse CD4⁺CD25⁺ Treg cells express the latent form of TGFβ1 on their surface, and suggested that the cell contact-dependent suppression by CD4⁺CD25⁺ Treg cells was mediated by this cell membrane-bound TGFβ1, which is activated upon cell contact between the Treg and target T cells (34). However, this mechanism was not confirmed by others in either human (35) and mouse systems (200), as CD4⁺CD25⁺ Treg cells strongly suppressed CD4⁺ T cells from mice deficient in Smad3, a signaling protein essential for TGFβ suppression of T cell proliferation and activation, and the CD4⁺CD25⁺ T cells from Smad3^{-/-} mice were as efficient in mediating suppression as those from wild-type mice, indicating that no TGFβ signaling is involved in the suppression (200).

To date, data about the role of TGFβ in virus-induced CD4⁺CD25⁺ T cell suppressive function are derived exclusively from *in vitro* studies, and inconsistent results similar to those mentioned above have also been observed for CD4⁺CD25⁺ T cells from virus-infected subjects. Most *in vitro* studies using these Treg cells indicate that TGFβ is not essential for CD4⁺CD25⁺ Treg cell-mediated suppression because anti-TGFβ antibody cannot block the suppressive effect of these Treg cells on virus-specific CD4⁺ or CD8⁺ T cell responses (13, 14, 75, 76, 201). However, unlike the *in vitro* results described above in other systems, there is also a report that *in vitro* suppression by CD4⁺CD25⁺ Treg cells is TGFβ-dependent. In this study, addition of anti-TGFβ antibody to the cell culture abrogated CD4⁺CD25⁺ Treg cell-mediated suppression and increased HCV-specific IFNγ production in HCV-stimulated PBMC (15). Thus, further studies, particularly *in vivo* studies, are needed to clarify the role of TGFβ in mediating CD4⁺CD25⁺ Treg cell function in viral infections.

Although there is still a lack of *in vivo* demonstration for the role of TGFβ in mediating CD4⁺CD25⁺ Treg cell function in viral infections, it can be anticipated that TGFβ may contribute to the suppression by virus-induced CD4⁺CD25⁺ Treg cells *in vivo*. This is based on the fact that almost all viral infections lead to immunosuppression and virus-induced TGFβ has been suggested as the key mediator of immunosuppression in viral infections (137, 202, 203). Many studies have shown that CD4⁺CD25⁺ Treg cells from virus-infected patients produce high levels of TGFβ (13, 15, 67, 82, 134). In addition, a wide variety of cell types are capable to produce TGFβ upon virus infection, including other types of regulatory T cells such as Th3 (204) and CD8⁺ Treg cells (205). Thus, the suppressive effect observed *in vivo* represents a total effect of TGFβ from all sources. It is difficult to conceive that the TGFβ produced by virus-induced CD4⁺CD25⁺ Treg cells does not join the total TGFβ pool in mediating immunosuppression during viral infections.

7.4. IL-10

Another immunosuppressive cytokine potentially involved in mediating CD4⁺CD25⁺ Treg cell function is IL-10. Similar to TGF β , studies on the role of IL-10 in mediating CD4⁺CD25⁺ Treg cell function in viral infections have shown controversial results. Most *in vitro* studies using CD4⁺CD25⁺ Treg cells from virus-infected patients showed that blocking IL-10 activity *in vitro* by using anti-IL-10 or anti-IL-10 receptor antibodies could not abrogate the suppression by CD4⁺CD25⁺ Treg cells, and thus concluded that the suppression was not mediated by IL-10 (13, 14, 201). However, a study by Huber *et al* showed that coxsackievirus B3 variant H310A1-induced CD4⁺CD25⁺ Treg cells were dependent on IL-10 for their suppressive function *in vivo*, and anti-IL-10 treatment abrogated the protection by CD4⁺CD25⁺ Treg cells against viral myocarditis (66). This seems consistent with the results from other *in vivo* studies that showed the dependence of CD4⁺CD25⁺ Treg cells on IL-10 for their suppressive functions in controlling expansion of peripheral CD4⁺ T cells (206), over-production of cytokines (207), Th1-mediated inflammation (208), and autoimmune pneumonitis (209).

7.5. IL-2

Inhibition of IL-2 production was the first mechanism suggested for CD4⁺CD25⁺ Treg cell-mediated suppression when Thornton *et al* observed that CD4⁺CD25⁺ Treg cells inhibited the proliferation and IL-2 production of CD4⁺CD25⁻ effector T cells, and that addition of exogenous IL-2 not only markedly enhanced the effector CD4⁺ T cell responses but almost completely abrogated the suppressive effects of CD4⁺CD25⁺ Treg cells (27). However, this mechanism was questioned since IL-2 is not required for the development and function of T cells *in vivo* (210, 211), except for Treg cells themselves (212). In viral infections, little is known about the role of IL-2 in mediating the suppressive function of virus-induced CD4⁺CD25⁺ Treg cells. The only information is a study on HIV infection showing that suppression of HIV p24-stimulated CD4⁺ T cell proliferation by CD4⁺CD25⁺ Treg cells is associated with the inhibition of IL-2 production (14). Whether IL-2 plays a role in virus-induced CD4⁺CD25⁺ Treg cell function still remains to be determined.

7.6. GITR

GTIR has been shown to be constitutively and predominantly expressed at high levels on CD4⁺CD25⁺ Treg cells (38, 213). Initially, data from both *in vitro* and *in vivo* studies suggested an essential role of GITR in mediating CD4⁺CD25⁺ Treg cell function. This was based on the observations that CD4⁺CD25⁺ Treg cell-mediated suppression of anti-CD3-stimulated or peptide-specific CD4⁺ T cell proliferation was abrogated after blocking GITR signaling with anti-GITR antibodies *in vitro* (38, 213), and that injection of normal mice with the antibodies led to the development of autoimmune gastritis and autoantibodies against parietal cells (38), a typical consequence from the loss of CD4⁺CD25⁺ Treg cell function in maintaining self-tolerance. However, it was later found that addition of anti-GITR antibody could not

reverse the CD4⁺CD25⁺ Treg cell-mediated suppression of CD4⁺ T cells from GITR^{-/-} mice (214), although these Treg cells from normal mice expressed GITR. Furthermore, CD4⁺CD25⁺ Treg cells from GITR^{-/-} mice showed comparably potent suppressive activity to those from normal mice, and the suppression by CD4⁺CD25⁺GITR^{-/-} Treg cells could be reversed by the addition of anti-GITR antibody. Collectively, these results clearly indicate that suppression by CD4⁺CD25⁺ Treg cells is mediated through the GITR on the target cells and not on the CD4⁺CD25⁺ Treg cells. Thus, engagement of the GITR on CD4⁺CD25⁺ Treg cells plays no role in abrogation of the CD4⁺CD25⁺ T cell-mediated suppression. Instead, engagement of GITR on target T cells, which turned out to render the target T cell resistant to the suppression by CD4⁺CD25⁺ Treg cells, is required to overcome CD4⁺CD25⁺ Treg cell-mediated suppression. Nonetheless, engagement of GITR with its ligand or antibody will likely change the status of effector T cell responses to pathologic stimuli, and thus may possess therapeutic potential. In support of this, studies on a mouse Friend virus model of retrovirus infection showed that administration of anti-GITR antibody to the infected mice increased Th1 cytokine production by both CD4⁺ and CD8⁺ T cells, ameliorated disease manifestation, and reduced virus load (74, 215). Consistently, La *et al* demonstrated that a single injection of anti-GITR antibody to HSV-infected mice immediately after viral infection significantly increased both the number of activated, antigen-specific CD4⁺ Th1 and CD8⁺ T cells and their IFN γ production (216). Thus, signaling through GITR enhances the antiviral immune responses. This concept has been tested in animal models for its potential clinical application in antiviral therapy. One approach is to use GITR ligand (GITRL) as an adjuvant to enhance viral vaccine efficacy. In a recent study on the development of DNA vaccine for HIV infection, Stone *et al* generated plasmids encoding a multimeric soluble GITRL fusion protein to use as an adjuvant (217). Immunization of mice with HIV DNA vaccine plus GITRL adjuvant plasmids significantly augmented virus-specific CD4⁺ and CD8⁺ T cell responses including their proliferation, IFN γ production, cytolytic activity, and antibody production. Since ligation of GITR provides costimulatory signals to both CD4⁺CD25⁺ Treg cells and CD4⁺CD25⁻ effector T cells (218), the enhanced effector T cell responses may represent combined effects of direct activation upon ligation of their GITR with GITRL and indirect activation through the reduction of Treg cell suppressive effect following the ligation of GITR on Treg cells.

In summary, more studies are still needed to clarify the mechanisms by which CD4⁺CD25⁺ Treg cells execute their suppressive functions. Whether there are more molecules potentially involved in the function of these Treg cells remains to be determined. Giving the fact that the suppression of CD4⁺CD25⁺ Treg cells can be abrogated by blocking one molecule but not another and that all molecules described above are potentially capable of mediating CD4⁺CD25⁺ Treg cell function, it is likely that the suppression of CD4⁺CD25⁺ Treg cells is mediated by different molecules under different conditions such as the nature of virus and infection, or the stage, magnitude, and

nature of inflammation and immune activation. It is important to find out whether the suppression observed is mediated by a single molecule or results from the collective effects of multiple molecules. In fact, the role of these molecules in mediating Treg cell function may change during the course of infection as disease condition changes. For example, as shown in the study on *Leishmania major* infection, IL-10 is not involved in suppression by CD4⁺CD25⁺ Treg cells at the early stage of infection, but is required at a later stage when chronic infections are well-established (3).

8. CONCLUSIONS AND PERSPECTIVE

In a productive and successful immune response to a viral infection, the processes of mounting a vigorous antiviral immune response and restricting excessive inflammatory responses to minimize collateral damage to host tissues are appropriately regulated and homeostasis is maintained. Because of their unique suppressive function, CD4⁺CD25⁺ Treg cells appear to play a key role in controlling undue inflammatory and hyperimmune responses in viral infections. However, this benefit is achieved at the cost of reduced antiviral immune responses and increased risk of developing chronic or persistent viral infection. The active role of CD4⁺CD25⁺ Treg cells in regulating antiviral immune responses makes them promising targets for clinical antiviral therapy. The biggest challenge is how to direct the function of these Treg cells precisely towards a desired therapeutic outcome to benefit the host, and, at the same time, keep their detrimental effect to the minimum. This requires a clear understanding of the biological and functional properties of CD4⁺CD25⁺ Treg cells in viral infections. To date, information on many aspects of CD4⁺CD25⁺ Treg cells in viral infections is still very limited. Many questions still remain to be answered and controversial results to be clarified. Further studies are needed to depict precise mechanisms on how CD4⁺CD25⁺ Treg cells are induced and regulated during viral infections and how they execute their function. Special attention should be paid to the discrepancy between *in vitro* and *in vivo* observations. To date, there is still no explanation for this discrepancy. Since *in vitro* assay conditions usually do not reflect the actual physiological environment *in vivo*, data from *in vitro* studies should be interpreted with caution. Accordingly, the functional property of CD4⁺CD25⁺ Treg cells *in vivo* will very likely differ from that predicted from *in vitro* studies. This is particularly important in the design of therapeutic strategies targeting CD4⁺CD25⁺ Treg cells for clinical application since strategies based on *in vitro* observations may not generate the anticipated effect *in vivo*. Finally, it should also be kept in mind that other types of regulatory T cells, such as Tr1, Th3, CD8⁺, and CD45RB^{low} Treg cells, may also regulate immune responses in viral infections. The relationship between these Treg cells and CD4⁺CD25⁺ Treg cells and how they affect each other in viral infections are unclear. Detailed *in vivo* analysis should provide information for our understanding of the biological effect of these Treg cells in viral infections and help with the development of strategies for the prevention and treatment of viral diseases.

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10. REFERENCES

1. Sakaguchi, S., N. Sakaguchi, M. Asano, M. Itoh & M. Toda: Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol*, 155, 1151-1164 (1995)
2. Suri-Payer, E., A. Z. Amar, A. M. Thornton & E. M. Shevach: CD4⁺CD25⁺ T cells inhibit both the induction and effector function of autoreactive T cells and represent a unique lineage of immunoregulatory cells. *J Immunol*, 160, 1212-1218 (1998)
3. Belkaid, Y., C. A. Piccirillo, S. Mendez, E. M. Shevach & D. L. Sacks: CD4⁺CD25⁺ regulatory T cells control *Leishmania major* persistence and immunity. *Nature*, 420, 502-507 (2002)
4. Groux, H., A. O'Garra, M. Bigler, M. Rouleau, S. Antonenko, J. E. de Vries & M. G. Roncarolo: A CD4⁺ T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. *Nature*, 389, 737-742 (1997)
5. Chen, Y., V. K. Kuchroo, J. Inobe, D. A. Hafler & H. L. Weiner: Regulatory T cell clones induced by oral tolerance: suppression of autoimmune encephalomyelitis. *Science*, 265, 1237-1240 (1994)
6. Jiang, S., S. Tugulea, G. Pennesi, Z. Liu, A. Mulder, S. Lederman, P. Harris, R. Cortesini & N. Suci-Foca: Induction of MHC-class I restricted human suppressor T cells by peptide priming *in vitro*. *Hum Immunol*, 59, 690-699 (1998)
7. Rifa'i, M., Y. Kawamoto, I. Nakashima & H. Suzuki: Essential roles of CD8⁺CD122⁺ regulatory T cells in the maintenance of T cell homeostasis. *J Exp Med*, 200, 1123-1134 (2004)
8. Rouse, B. T. & D. W. Horohov: Immunosuppression in viral infections. *Rev Infect Dis*, 8, 850-873 (1986)
9. Horohov, D. W. & B. T. Rouse: Virus-induced immunosuppression. *Vet Clin North Am Small Anim Pract*, 16, 1097-1127 (1986)
10. Borrow, P., C. F. Evans & M. B. Oldstone: Virus-induced immunosuppression: immune system-mediated destruction of virus-infected dendritic cells results in generalized immune suppression. *J Virol*, 69, 1059-1070 (1995)
11. Iwashiro, M., R. J. Messer, K. E. Peterson, I. M. Stromnes, T. Sugie & K. J. Hasenkrug: Immunosuppression by CD4⁺ regulatory T cells induced by chronic retroviral infection. *Proc Natl Acad Sci U S A*, 98, 9226-9230 (2001)
12. Suvas, S., U. Kumaraguru, C. D. Pack, S. Lee & B. T. Rouse: CD4⁺CD25⁺ T cells regulate virus-specific primary and memory CD8⁺ T cell responses. *J Exp Med*, 198, 889-901 (2003)
13. Weiss, L., V. Donkova-Petrini, L. Caccavelli, M. Balbo, C. Carbonneil & Y. Levy: Human

- immunodeficiency virus-driven expansion of CD4⁺CD25⁺ regulatory T cells, which suppress HIV-specific CD4 T-cell responses in HIV-infected patients. *Blood*, 104, 3249-3256 (2004)
14. Kinter, A. L., M. Hennessey, A. Bell, S. Kern, Y. Lin, M. Daucher, M. Planta, M. McGlaughlin, R. Jackson, S. F. Ziegler & A. S. Fauci: CD25(+)CD4(+) regulatory T cells from the peripheral blood of asymptomatic HIV-infected individuals regulate CD4(+) and CD8(+) HIV-specific T cell immune responses *in vitro* and are associated with favorable clinical markers of disease status. *J Exp Med*, 200, 331-343 (2004)
15. Cabrera, R., Z. Tu, Y. Xu, R. J. Firpi, H. R. Rosen, C. Liu & D. R. Nelson: An immunomodulatory role for CD4(+)CD25(+) regulatory T lymphocytes in hepatitis C virus infection. *Hepatology*, 40, 1062-1071 (2004)
16. Suvas, S., A. K. Azkur, B. S. Kim, U. Kumaraguru & B. T. Rouse: CD4⁺CD25⁺ regulatory T cells control the severity of viral immunoinflammatory lesions. *J Immunol*, 172, 4123-4132 (2004)
17. Yamano, Y., N. Takenouchi, H. C. Li, U. Tomaru, K. Yao, C. W. Grant, D. A. Maric & S. Jacobson: Virus-induced dysfunction of CD4⁺CD25⁺ T cells in patients with HTLV-1-associated neuroimmunological disease. *J Clin Invest*, 115, 1361-1368 (2005)
18. Bolacchi, F., A. Sinistro, C. Ciaprini, F. Demin, M. Capozzi, F. C. Carducci, C. M. Drapeau, G. Rocchi & A. Bergamini: Increased hepatitis C virus (HCV)-specific CD4⁺CD25⁺ regulatory T lymphocytes and reduced HCV-specific CD4⁺ T cell response in HCV-infected patients with normal versus abnormal alanine aminotransferase levels. *Clin Exp Immunol*, 144, 188-196 (2006)
19. Luhn, K., C. P. Simmons, E. Moran, N. T. Dung, T. N. Chau, N. T. Quyen, L. T. Thao, T. Van Ngoc, N. M. Dung, B. Wills, J. Farrar, A. J. McMichael, T. Dong & S. Rowland-Jones: Increased frequencies of CD4⁺CD25^{high} regulatory T cells in acute dengue infection. *J Exp Med* (2007)
20. Papiernik, M., M. L. de Moraes, C. Pontoux, F. Vasseur & C. Penit: Regulatory CD4 T cells: expression of IL-2R alpha chain, resistance to clonal deletion and IL-2 dependency. *Int Immunol*, 10, 371-378 (1998)
21. Itoh, M., T. Takahashi, N. Sakaguchi, Y. Kuniyasu, J. Shimizu, F. Otsuka & S. Sakaguchi: Thymus and autoimmunity: production of CD25⁺CD4⁺ naturally anergic and suppressive T cells as a key function of the thymus in maintaining immunologic self-tolerance. *J Immunol*, 162, 5317-5326 (1999)
22. Bluestone, J. A. & A. K. Abbas: Natural versus adaptive regulatory T cells. *Nat Rev Immunol*, 3, 253-257 (2003)
23. Jonuleit, H., E. Schmitt, G. Schuler, J. Knop & A. H. Enk: Induction of interleukin 10-producing, nonproliferating CD4(+) T cells with regulatory properties by repetitive stimulation with allogeneic immature human dendritic cells. *J Exp Med*, 192, 1213-1222 (2000)
24. Chen, W., W. Jin, N. Hardegen, K. J. Lei, L. Li, N. Marinos, G. McGrady & S. M. Wahl: Conversion of peripheral CD4⁺CD25⁻ naive T cells to CD4⁺CD25⁺ regulatory T cells by TGF-beta induction of transcription factor Foxp3. *J Exp Med*, 198, 1875-1886 (2003)
25. Asano, M., M. Toda, N. Sakaguchi & S. Sakaguchi: Autoimmune disease as a consequence of developmental abnormality of a T cell subpopulation. *J Exp Med*, 184, 387-396 (1996)
26. Baecher-Allan, C., J. A. Brown, G. J. Freeman & D. A. Hafler: CD4⁺CD25^{high} regulatory cells in human peripheral blood. *J Immunol*, 167, 1245-1253 (2001)
27. Thornton, A. M. & E. M. Shevach: CD4⁺CD25⁺ immunoregulatory T cells suppress polyclonal T cell activation *in vitro* by inhibiting interleukin 2 production. *J Exp Med*, 188, 287-296 (1998)
28. Jonuleit, H., E. Schmitt, M. Stassen, A. Tuettenberg, J. Knop & A. H. Enk: Identification and functional characterization of human CD4(+)CD25(+) T cells with regulatory properties isolated from peripheral blood. *J Exp Med*, 193, 1285-1294 (2001)
29. Dieckmann, D., H. Plottner, S. Berchtold, T. Berger & G. Schuler: *Ex vivo* isolation and characterization of CD4(+)CD25(+) T cells with regulatory properties from human blood. *J Exp Med*, 193, 1303-1310 (2001)
30. Shevach, E. M.: CD4⁺ CD25⁺ suppressor T cells: more questions than answers. *Nat Rev Immunol*, 2, 389-400 (2002)
31. Takahashi, T., Y. Kuniyasu, M. Toda, N. Sakaguchi, M. Itoh, M. Iwata, J. Shimizu & S. Sakaguchi: Immunologic self-tolerance maintained by CD25⁺CD4⁺ naturally anergic and suppressive T cells: induction of autoimmune disease by breaking their anergic/suppressive state. *Int Immunol*, 10, 1969-1980 (1998)
32. Kuniyasu, Y., T. Takahashi, M. Itoh, J. Shimizu, G. Toda & S. Sakaguchi: Naturally anergic and suppressive CD25(+)CD4(+) T cells as a functionally and phenotypically distinct immunoregulatory T cell subpopulation. *Int Immunol*, 12, 1145-1155 (2000)
33. Walker, L. S., A. Chodos, M. Eggena, H. Doms & A. K. Abbas: Antigen-dependent proliferation of CD4⁺CD25⁺ regulatory T cells *in vivo*. *J Exp Med*, 198, 249-258 (2003)
34. Nakamura, K., A. Kitani & W. Strober: Cell contact-dependent immunosuppression by CD4(+)CD25(+) regulatory T cells is mediated by cell surface-bound transforming growth factor beta. *J Exp Med*, 194, 629-644 (2001)
35. Jonuleit, H., E. Schmitt, H. Kakirman, M. Stassen, J. Knop & A. H. Enk: Infectious tolerance: human CD25(+) regulatory T cells convey suppressor activity to conventional CD4(+) T helper cells. *J Exp Med*, 196, 255-260 (2002)
36. Shevach, E. M., R. S. McHugh, C. A. Piccirillo & A. M. Thornton: Control of T-cell activation by CD4⁺CD25⁺ suppressor T cells. *Immunol Rev*, 182, 58-67 (2001)
37. Takahashi, T., T. Tagami, S. Yamazaki, T. Uede, J. Shimizu, N. Sakaguchi, T. W. Mak & S. Sakaguchi: Immunologic self-tolerance maintained by CD25(+)CD4(+) regulatory T cells constitutively expressing cytotoxic T lymphocyte-associated antigen 4. *J Exp Med*, 192, 303-310 (2000)
38. Shimizu, J., S. Yamazaki, T. Takahashi, Y. Ishida & S. Sakaguchi: Stimulation of CD25(+)CD4(+) regulatory T cells by TGF-beta induction of

- regulatory T cells through GITR breaks immunological self-tolerance. *Nat Immunol*, 3, 135-142 (2002)
39. Hori, S., T. Nomura & S. Sakaguchi: Control of regulatory T cell development by the transcription factor Foxp3. *Science*, 299, 1057-1061 (2003)
40. Masuyama, J., S. Kaga, S. Kano & S. Minota: A novel costimulation pathway via the 4C8 antigen for the induction of CD4⁺ regulatory T cells. *J Immunol*, 169, 3710-3716 (2002)
41. Pyzik, M. & C. A. Piccirillo: TGF- β 1 modulates Foxp3 expression and regulatory activity in distinct CD4⁺ T cell subsets. *J Leukoc Biol* (2007)
42. Karim, M., C. I. Kingsley, A. R. Bushell, B. S. Sawitzki & K. J. Wood: Alloantigen-induced CD25⁺CD4⁺ regulatory T cells can develop *in vivo* from CD25⁺CD4⁺ precursors in a thymus-independent process. *J Immunol*, 172, 923-928 (2004)
43. Chen, Z. M., M. J. O'Shaughnessy, I. Gramaglia, A. Panoskaltis-Mortari, W. J. Murphy, S. Narula, M. G. Roncarolo & B. R. Blazar: IL-10 and TGF- β induce alloreactive CD4⁺CD25⁺ T cells to acquire regulatory cell function. *Blood*, 101, 5076-5083 (2003)
44. Park, H. B., D. J. Paik, E. Jang, S. Hong & J. Youn: Acquisition of anergic and suppressive activities in transforming growth factor- β -costimulated CD4⁺CD25⁺ T cells. *Int Immunol*, 16, 1203-1213 (2004)
45. Moseman, E. A., X. Liang, A. J. Dawson, A. Panoskaltis-Mortari, A. M. Krieg, Y. J. Liu, B. R. Blazar & W. Chen: Human plasmacytoid dendritic cells activated by CpG oligodeoxynucleotides induce the generation of CD4⁺CD25⁺ regulatory T cells. *J Immunol*, 173, 4433-4442 (2004)
46. Aandahl, E. M., J. Michaelsson, W. J. Moretto, F. M. Hecht & D. F. Nixon: Human CD4⁺ CD25⁺ regulatory T cells control T-cell responses to human immunodeficiency virus and cytomegalovirus antigens. *J Virol*, 78, 2454-2459 (2004)
47. Toy, S. T. & E. F. Whellock: *In vitro* depression of cellular immunity by Friend virus leukemic spleen cells. *Cell Immunol*, 17, 57-73 (1975)
48. Carpenter, C. R., A. S. Rubin & H. R. Bose, Jr.: Suppression of the mitogen-stimulated blastogenic response during reticuloendotheliosis virus-induced tumorigenesis: investigations into the mechanism of action of the suppressor. *J Immunol*, 120, 1313-1320 (1978)
49. Kirchner, H., T. M. Chused, R. B. Herberman, H. T. Holden & D. H. Lavrin: Evidence of suppressor cell activity in spleens of mice bearing primary tumors induced by Moloney sarcoma virus. *J Exp Med*, 139, 1473-1487 (1974)
50. Cerny, J., K. D. Grinwich & R. A. Stiller: Immunosuppression by spleen cells from Moloney leukemia. III. Evidence for a suppressor cell that is not the leukemic, virus-producing cell. *J Immunol*, 119, 1097-1101 (1977)
51. Roman, J. M. & E. S. Golub: Leukemia in AKR mice. I. Effects of leukemic cells on antibody-forming potential of syngeneic and allogeneic normal cells. *J Exp Med*, 143, 482-496 (1976)
52. Langweiler, M., G. L. Cockerell & F. deNoronha: Role of suppressor cells in feline leukemia virus-associated immunosuppression. *Cancer Res*, 43, 1957-1960 (1983)
53. Vahlenkamp, T. W., M. B. Tompkins & W. A. Tompkins: Feline immunodeficiency virus infection phenotypically and functionally activates immunosuppressive CD4⁺CD25⁺ T regulatory cells. *J Immunol*, 172, 4752-4761 (2004)
54. Scofield, V. L. & H. R. Bose, Jr.: Depression of mitogen response in spleen cells from reticuloendotheliosis virus-infected chickens and their suppressive effect on normal lymphocyte response. *J Immunol*, 120, 1321-1325 (1978)
55. Theis, G. A.: Effects of lymphocytes from Marek's disease-infected chickens on mitogen responses of syngeneic normal chicken spleen cells. *J Immunol*, 118, 887-894 (1977)
56. Ilonen, J., M. Lanning, E. Herva & A. Salmi: Lymphocyte blast transformation responses in measles infection. *Scand J Immunol*, 12, 383-391 (1980)
57. Kirchner, H., M. Schwentek, H. Northoff & E. Schopf: Defective *in vitro* lymphoproliferative responses to herpes simplex virus in patients with frequently recurring herpes infections during the disease-free interval. *Clin Immunol Immunopathol*, 11, 267-274 (1978)
58. Fontana, A. & H. L. Weiner: Interaction of reovirus with cell surface receptors. II. Generation of suppressor T cells by the hemagglutinin of reovirus type 3. *J Immunol*, 125, 2660-2664 (1980)
59. Franzese, O., P. T. Kennedy, A. J. Gehring, J. Gotto, R. Williams, M. K. Maini & A. Bertolotti: Modulation of the CD8⁺-T-cell response by CD4⁺ CD25⁺ regulatory T cells in patients with hepatitis B virus infection. *J Virol*, 79, 3322-3328 (2005)
60. Stoop, J. N., R. G. van der Molen, C. C. Baan, L. J. van der Laan, E. J. Kuipers, J. G. Kusters & H. L. Janssen: Regulatory T cells contribute to the impaired immune response in patients with chronic hepatitis B virus infection. *Hepatology*, 41, 771-778 (2005)
61. MacDonald, A. J., M. Duffy, M. T. Brady, S. McKiernan, W. Hall, J. Hegarty, M. Curry & K. H. Mills: CD4 T helper type 1 and regulatory T cells induced against the same epitopes on the core protein in hepatitis C virus-infected persons. *J Infect Dis*, 185, 720-727 (2002)
62. Sugimoto, K., F. Ikeda, J. Stadanlick, F. A. Nunes, H. J. Alter & K. M. Chang: Suppression of HCV-specific T cells without differential hierarchy demonstrated *ex vivo* in persistent HCV infection. *Hepatology*, 38, 1437-1448 (2003)
63. Marshall, N. A., M. A. Vickers & R. N. Barker: Regulatory T cells secreting IL-10 dominate the immune response to EBV latent membrane protein 1. *J Immunol*, 170, 6183-6189 (2003)
64. Kohno, T., Y. Yamada, N. Akamatsu, S. Kamihira, Y. Imaizumi, M. Tomonaga & T. Matsuyama: Possible origin of adult T-cell leukemia/lymphoma cells from human T lymphotropic virus type-1-infected regulatory T cells. *Cancer Sci*, 96, 527-533 (2005)
65. Beilharz, M. W., L. M. Samuels, A. Paun, K. Shaw, P. van Eeden, M. W. Watson & M. L. Ashdown: Timed ablation of regulatory CD4⁺ T cells can prevent murine AIDS progression. *J Immunol*, 172, 4917-4925 (2004)
66. Huber, S. A., A. M. Feldman & D. Sartini: Cocksackievirus B3 induces T regulatory cells, which

- inhibit cardiomyopathy in tumor necrosis factor- α transgenic mice. *Circ Res*, 99, 1109-1116 (2006)
67. Kornfeld, C., M. J. Ploquin, I. Pandrea, A. Faye, R. Onanga, C. Apetrei, V. Poaty-Mavoungou, P. Rouquet, J. Estaquier, L. Mortara, J. F. Desoutter, C. Butor, R. Le Grand, P. Roques, F. Simon, F. Barre-Sinoussi, O. M. Diop & M. C. Muller-Trutwin: Antiinflammatory profiles during primary SIV infection in African green monkeys are associated with protection against AIDS. *J Clin Invest*, 115, 1082-1091 (2005)
68. Tsunemi, S., T. Iwasaki, T. Imado, S. Higasa, E. Kakishita, T. Shirasaka & H. Sano: Relationship of CD4⁺CD25⁺ regulatory T cells to immune status in HIV-infected patients. *Aids*, 19, 879-886 (2005)
69. Xu, D., J. Fu, L. Jin, H. Zhang, C. Zhou, Z. Zou, J. M. Zhao, B. Zhang, M. Shi, X. Ding, Z. Tang, Y. X. Fu & F. S. Wang: Circulating and liver resident CD4⁺CD25⁺ regulatory T cells actively influence the antiviral immune response and disease progression in patients with hepatitis B. *J Immunol*, 177, 739-747 (2006)
70. Misra, N., J. Bayry, S. Lacroix-Desmazes, M. D. Kazatchkine & S. V. Kaveri: Cutting edge: human CD4⁺CD25⁺ T cells restrain the maturation and antigen-presenting function of dendritic cells. *J Immunol*, 172, 4676-4680 (2004)
71. Taams, L. S., J. M. van Amelsfort, M. M. Tiemessen, K. M. Jacobs, E. C. de Jong, A. N. Akbar, J. W. Bijlsma & F. P. Lafeber: Modulation of monocyte/macrophage function by human CD4⁺CD25⁺ regulatory T cells. *Hum Immunol*, 66, 222-230 (2005)
72. Trzonkowski, P., E. Szmit, J. Mysliwska, A. Dobyszyk & A. Mysliwski: CD4⁺CD25⁺ T regulatory cells inhibit cytotoxic activity of T CD8⁺ and NK lymphocytes in the direct cell-to-cell interaction. *Clin Immunol*, 112, 258-267 (2004)
73. Ralainirina, N., A. Poli, T. Michel, L. Poos, E. Andres, F. Hentges & J. Zimmer: Control of NK cell functions by CD4⁺CD25⁺ regulatory T cells. *J Leukoc Biol*, 81, 144-153 (2007)
74. Dittmer, U., H. He, R. J. Messer, S. Schimmer, A. R. Olbrich, C. Ohlen, P. D. Greenberg, I. M. Stromnes, M. Iwashiro, S. Sakaguchi, L. H. Evans, K. E. Peterson, G. Yang & K. J. Hasenkrug: Functional impairment of CD8(+) T cells by regulatory T cells during persistent retroviral infection. *Immunity*, 20, 293-303 (2004)
75. Boettler, T., H. C. Spangenberg, C. Neumann-Haefelin, E. Panther, S. Urbani, C. Ferrari, H. E. Blum, F. von Weizsacker & R. Thimme: T cells with a CD4⁺CD25⁺ regulatory phenotype suppress *in vitro* proliferation of virus-specific CD8⁺ T cells during chronic hepatitis C virus infection. *J Virol*, 79, 7860-7867 (2005)
76. Rushbrook, S. M., S. M. Ward, E. Unitt, S. L. Vowler, M. Lucas, P. Klenerman & G. J. Alexander: Regulatory T cells suppress *in vitro* proliferation of virus-specific CD8⁺ T cells during persistent hepatitis C virus infection. *J Virol*, 79, 7852-7859 (2005)
77. Robertson, S. J., R. J. Messer, A. B. Carmody & K. J. Hasenkrug: *In vitro* suppression of CD8⁺ T cell function by Friend virus-induced regulatory T cells. *J Immunol*, 176, 3342-3349 (2006)
78. Cucchiari, M., A. R. Kammer, B. Grabscheid, H. M. Diepolder, T. J. Gerlach, N. Gruner, T. Santantonio, J. Reichen, G. R. Pape & A. Cerny: Vigorous peripheral blood cytotoxic T cell response during the acute phase of hepatitis C virus infection. *Cell Immunol*, 203, 111-123 (2000)
79. Lechner, F., N. H. Gruener, S. Urbani, J. Uggeri, T. Santantonio, A. R. Kammer, A. Cerny, R. Phillips, C. Ferrari, G. R. Pape & P. Klenerman: CD8⁺ T lymphocyte responses are induced during acute hepatitis C virus infection but are not sustained. *Eur J Immunol*, 30, 2479-2487 (2000)
80. Cooper, S., A. L. Erickson, E. J. Adams, J. Kansopon, A. J. Weiner, D. Y. Chien, M. Houghton, P. Parham & C. M. Walker: Analysis of a successful immune response against hepatitis C virus. *Immunity*, 10, 439-449 (1999)
81. Perrella, A., L. Vitiello, L. Atripaldi, P. Conti, C. Sbriglia, S. Altamura, T. Patarino, R. Vela, G. Morelli, P. Bellopede, C. Alone, L. Racioppi & O. Perrella: Elevated CD4⁺/CD25⁺ T cell frequency and function during acute hepatitis C presage chronic evolution. *Gut*, 55, 1370-1371 (2006)
82. Estes, J. D., Q. Li, M. R. Reynolds, S. Wietgreffe, L. Duan, T. Schacker, L. J. Picker, D. I. Watkins, J. D. Lifson, C. Reilly, J. Carlis & A. T. Haase: Premature induction of an immunosuppressive regulatory T cell response during acute simian immunodeficiency virus infection. *J Infect Dis*, 193, 703-712 (2006)
83. Gasper-Smith, N., I. Marriott & K. L. Bost: Murine gamma-herpesvirus 68 limits naturally occurring CD4⁺CD25⁺ T regulatory cell activity following infection. *J Immunol*, 177, 4670-4678 (2006)
84. Kurane, I., A. L. Rothman, P. G. Livingston, S. Green, S. J. Gagnon, J. Janus, B. L. Innis, S. Nimmannitya, A. Nisalak & F. A. Ennis: Immunopathologic mechanisms of dengue hemorrhagic fever and dengue shock syndrome. *Arch Virol Suppl*, 9, 59-64 (1994)
85. Terrada, C., S. Fisson, Y. De Kozak, M. Kaddouri, P. Lehoang, D. Klatzmann, B. L. Salomon & B. Bodaghi: Regulatory T cells control uveoretinitis induced by pathogenic Th1 cells reacting to a specific retinal neoantigen. *J Immunol*, 176, 7171-7179 (2006)
86. Hazenberg, M. D., S. A. Otto, B. H. van Benthem, M. T. Roos, R. A. Coutinho, J. M. Lange, D. Hamann, M. Prins & F. Miedema: Persistent immune activation in HIV-1 infection is associated with progression to AIDS. *Aids*, 17, 1881-1888 (2003)
87. Sousa, A. E., J. Carneiro, M. Meier-Schellersheim, Z. Grossman & R. M. Victorino: CD4 T cell depletion is linked directly to immune activation in the pathogenesis of HIV-1 and HIV-2 but only indirectly to the viral load. *J Immunol*, 169, 3400-3406 (2002)
88. Grossman, Z., M. Meier-Schellersheim, A. E. Sousa, R. M. Victorino & W. E. Paul: CD4⁺ T-cell depletion in HIV infection: are we closer to understanding the cause? *Nat Med*, 8, 319-323 (2002)
89. Deeks, S. G., C. M. Kitchen, L. Liu, H. Guo, R. Gascon, A. B. Narvaez, P. Hunt, J. N. Martin, J. O. Kahn, J. Levy, M. S. McGrath & F. M. Hecht: Immune activation set point during early HIV infection predicts subsequent CD4⁺ T-cell changes independent of viral load. *Blood*, 104, 942-947 (2004)
90. Eggena, M. P., B. Barugahare, N. Jones, M. Okello, S. Mutalya, C. Kityo, P. Mugenyi & H. Cao: Depletion of

- regulatory T cells in HIV infection is associated with immune activation. *J Immunol*, 174, 4407-4414 (2005)
91. Sumpter, B., R. Dunham, S. Gordon, J. Engram, M. Hennessy, A. Kinter, M. Paiardini, B. Cervasi, N. Klatt, H. McClure, J. M. Milush, S. Staprans, D. L. Sodora & G. Silvestri: Correlates of preserved CD4(+) T cell homeostasis during natural, nonpathogenic simian immunodeficiency virus infection of sooty mangabeys: implications for AIDS pathogenesis. *J Immunol*, 178, 1680-1691 (2007)
 92. Silvestri, G.: Naturally SIV-infected sooty mangabeys: are we closer to understanding why they do not develop AIDS? *J Med Primatol*, 34, 243-252 (2005)
 93. Magder, L. S., L. Mofenson, M. E. Paul, C. D. Zorrilla, W. A. Blattner, R. E. Tuomala, P. LaRussa, S. Landesman & K. C. Rich: Risk factors for *in utero* and intrapartum transmission of HIV. *J Acquir Immune Defic Syndr*, 38, 87-95 (2005)
 94. Legrand, F. A., D. F. Nixon, C. P. Loo, E. Ono, J. M. Chapman, M. Miyamoto, R. S. Diaz, A. M. Santos, R. C. Succi, J. Abadi, M. G. Rosenberg, M. I. de Moraes-Pinto & E. G. Kallas: Strong HIV-1-Specific T Cell Responses in HIV-1-Exposed Uninfected Infants and Neonates Revealed after Regulatory T Cell Removal. *PLoS ONE*, 1, e102 (2006)
 95. Olson, J. K., J. L. Croxford & S. D. Miller: Virus-induced autoimmunity: potential role of viruses in initiation, perpetuation, and progression of T-cell-mediated autoimmune disease. *Viral Immunol*, 14, 227-250 (2001)
 96. Lunel, F., L. Musset, P. Cacoub, L. Frangeul, P. Cresta, M. Perrin, P. Grippon, C. Hoang, D. Valla, J. C. Piette & *et al.*: Cryoglobulinemia in chronic liver diseases: role of hepatitis C virus and liver damage. *Gastroenterology*, 106, 1291-1300 (1994)
 97. Gordon, S. C.: Extrahepatic manifestations of hepatitis C. *Dig Dis*, 14, 157-168 (1996)
 98. Sene, D., N. Limal & P. Cacoub: Hepatitis C virus-associated extrahepatic manifestations: a review. *Metab Brain Dis*, 19, 357-381 (2004)
 99. Vigani, A. G., A. Macedo-de-Oliveira, M. H. Pavan, M. N. Pedro & F. L. Goncalves Jr: Hepatitis C virus infection, cryoglobulinemia, and peripheral neuropathy: a case report. *Braz J Med Biol Res*, 38, 1729-1734 (2005)
 100. Jun, H. S. & J. W. Yoon: The role of viruses in type I diabetes: two distinct cellular and molecular pathogenic mechanisms of virus-induced diabetes in animals. *Diabetologia*, 44, 271-285 (2001)
 101. Haase, A. T., P. Ventura, C. J. Gibbs, Jr. & W. W. Tourtellotte: Measles virus nucleotide sequences: detection by hybridization *in situ*. *Science*, 212, 672-675 (1981)
 102. Bergstrom, T., O. Andersen & A. Vahlne: Isolation of herpes simplex virus type 1 during first attack of multiple sclerosis. *Ann Neurol*, 26, 283-285 (1989)
 103. Bowles, N. E., J. Ni, F. Marcus & J. A. Towbin: The detection of cardiotropic viruses in the myocardium of patients with arrhythmogenic right ventricular dysplasia/cardiomyopathy. *J Am Coll Cardiol*, 39, 892-895 (2002)
 104. Fairweather, D., S. Frisancho-Kiss & N. R. Rose: Viruses as adjuvants for autoimmunity: evidence from Coxsackievirus-induced myocarditis. *Rev Med Virol*, 15, 17-27 (2005)
 105. Harley, J. B. & J. A. James: Epstein-Barr virus infection induces lupus autoimmunity. *Bull NYU Hosp Jt Dis*, 64, 45-50 (2006)
 106. Boyer, O., D. Saadoun, J. Abriol, M. Dodille, J. C. Piette, P. Cacoub & D. Klatzmann: CD4⁺CD25⁺ regulatory T-cell deficiency in patients with hepatitis C-mixed cryoglobulinemia vasculitis. *Blood*, 103, 3428-3430 (2004)
 107. Hayashi, T., K. Hasegawa, Y. Sasaki & T. Onodera: Elimination of CD4⁺CD25⁺ regulatory T cells breaks down reovirus type 2-triggered and CpG ODN-induced prolonged mild autoimmune insulinitis in DBA/1 mice. *Scand J Immunol*, 63, 116-124 (2006)
 108. Maloy, K. J. & F. Powrie: Regulatory T cells in the control of immune pathology. *Nat Immunol*, 2, 816-822 (2001)
 109. Tajimi, M., T. Ugajin, M. Ota, K. Hiroishi, I. Nakamura & M. Imawari: Immune responses of liver-infiltrating lymphocytes and peripheral blood mononuclear cells to hepatitis C virus core and NS3 antigens. *Hepatol Res*, 35, 250-255 (2006)
 110. Li, S., K. L. Jones, D. J. Woollard, J. Dromey, G. Paukovics, M. Plebanski & E. J. Gowans: Defining target antigens for CD25(+)FOXP3(+)IFN-gamma(-) regulatory T cells in chronic hepatitis C virus infection. *Immunol Cell Biol*, 85, 197-204 (2007)
 111. Fujinami, R. S. & M. B. Oldstone: Amino acid homology between the encephalitogenic site of myelin basic protein and virus: mechanism for autoimmunity. *Science*, 230, 1043-1045 (1985)
 112. Wucherpfennig, K. W. & J. L. Strominger: Molecular mimicry in T cell-mediated autoimmunity: viral peptides activate human T cell clones specific for myelin basic protein. *Cell*, 80, 695-705 (1995)
 113. Fujinami, R. S., M. B. Oldstone, Z. Wroblewska, M. E. Frankel & H. Koprowski: Molecular mimicry in virus infection: crossreaction of measles virus phosphoprotein or of herpes simplex virus protein with human intermediate filaments. *Proc Natl Acad Sci U S A*, 80, 2346-2350 (1983)
 114. Toh, B. H., A. Yildiz, J. Sotelo, O. Osung, E. J. Holborow, F. Kanakoudi & J. V. Small: Viral infections and IgM autoantibodies to cytoplasmic intermediate filaments. *Clin Exp Immunol*, 37, 76-82 (1979)
 115. Srinivasappa, J., J. Saegusa, B. S. Prabhakar, M. K. Gentry, M. J. Buchmeier, T. J. Wiktor, H. Koprowski, M. B. Oldstone & A. L. Notkins: Molecular mimicry: frequency of reactivity of monoclonal antiviral antibodies with normal tissues. *J Virol*, 57, 397-401 (1986)
 116. Cianciolo, G. J., T. D. Copeland, S. Oroszlan & R. Snyderman: Inhibition of lymphocyte proliferation by a synthetic peptide homologous to retroviral envelope proteins. *Science*, 230, 453-455 (1985)
 117. Cauda, R., M. Tumbarello, L. Ortona, P. Kanda, R. C. Kennedy & T. C. Chanh: Inhibition of normal human natural killer cell activity by human immunodeficiency virus synthetic transmembrane peptides. *Cell Immunol*, 115, 57-65 (1988)
 118. Chanh, T. C., R. C. Kennedy & P. Kanda: Synthetic peptides homologous to HIV transmembrane glycoprotein suppress normal human lymphocyte blastogenic response. *Cell Immunol*, 111, 77-86 (1988)
 119. Ruegg, C. L., C. R. Monell & M. Strand: Identification, using synthetic peptides, of the minimum

- amino acid sequence from the retroviral transmembrane protein p15E required for inhibition of lymphoproliferation and its similarity to gp21 of human T-lymphotropic virus types I and II. *J Virol*, 63, 3250-3256 (1989)
120. Walker, M. R., B. D. Carson, G. T. Nepom, S. F. Ziegler & J. H. Buckner: De novo generation of antigen-specific CD4⁺CD25⁺ regulatory T cells from human CD4⁺CD25⁻ cells. *Proc Natl Acad Sci U S A*, 102, 4103-4108 (2005)
121. Apostolou, I. & H. von Boehmer: *In vivo* instruction of suppressor commitment in naive T cells. *J Exp Med*, 199, 1401-1408 (2004)
122. Papiernik, M.: Natural CD4⁺ CD25⁺ regulatory T cells. Their role in the control of superantigen responses. *Immunol Rev*, 182, 180-189 (2001)
123. Yamagiwa, S., J. D. Gray, S. Hashimoto & D. A. Horwitz: A role for TGF-beta in the generation and expansion of CD4⁺CD25⁺ regulatory T cells from human peripheral blood. *J Immunol*, 166, 7282-7289 (2001)
124. Peng, Y., Y. Laouar, M. O. Li, E. A. Green & R. A. Flavell: TGF-beta regulates *in vivo* expansion of Foxp3-expressing CD4⁺CD25⁺ regulatory T cells responsible for protection against diabetes. *Proc Natl Acad Sci U S A*, 101, 4572-4577 (2004)
125. Fantini, M. C., C. Becker, G. Monteleone, F. Pallone, P. R. Galle & M. F. Neurath: Cutting edge: TGF-beta induces a regulatory phenotype in CD4⁺CD25⁻ T cells through Foxp3 induction and down-regulation of Smad7. *J Immunol*, 172, 5149-5153 (2004)
126. de Jong, R., R. A. van Lier, F. W. Ruscetti, C. Schmitt, P. Debre & M. D. Mossalayi: Differential effect of transforming growth factor-beta 1 on the activation of human naive and memory CD4⁺ T lymphocytes. *Int Immunol*, 6, 631-638 (1994)
127. Schramm, C., S. Huber, M. Protschka, P. Czochra, J. Burg, E. Schmitt, A. W. Lohse, P. R. Galle & M. Blessing: TGFbeta regulates the CD4⁺CD25⁺ T-cell pool and the expression of Foxp3 *in vivo*. *Int Immunol*, 16, 1241-1249 (2004)
128. Mamura, M., W. Lee, T. J. Sullivan, A. Felici, A. L. Sowers, J. P. Allison & J. J. Letterio: CD28 disruption exacerbates inflammation in Tgf-beta1^{-/-} mice: *in vivo* suppression by CD4⁺CD25⁺ regulatory T cells independent of autocrine TGF-beta1. *Blood*, 103, 4594-4601 (2004)
129. Marie, J. C., J. J. Letterio, M. Gavin & A. Y. Rudensky: TGF-beta1 maintains suppressor function and Foxp3 expression in CD4⁺CD25⁺ regulatory T cells. *J Exp Med*, 201, 1061-1067 (2005)
130. Huber, S., C. Schramm, H. A. Lehr, A. Mann, S. Schmitt, C. Becker, M. Protschka, P. R. Galle, M. F. Neurath & M. Blessing: Cutting edge: TGF-beta signaling is required for the *in vivo* expansion and immunosuppressive capacity of regulatory CD4⁺CD25⁺ T cells. *J Immunol*, 173, 6526-6531 (2004)
131. Fahlen, L., S. Read, L. Gorelik, S. D. Hurst, R. L. Coffman, R. A. Flavell & F. Powrie: T cells that cannot respond to TGF-beta escape control by CD4(+)CD25(+) regulatory T cells. *J Exp Med*, 201, 737-746 (2005)
132. Kekow, J., W. Wachsman, J. A. McCutchan, M. Cronin, D. A. Carson & M. Lotz: Transforming growth factor beta and noncytopathic mechanisms of immunodeficiency in human immunodeficiency virus infection. *Proc Natl Acad Sci U S A*, 87, 8321-8325 (1990)
133. Navikas, V., J. Link, B. Wahren, C. Persson & H. Link: Increased levels of interferon-gamma (IFN-gamma), IL-4 and transforming growth factor-beta (TGF-beta) mRNA expressing blood mononuclear cells in human HIV infection. *Clin Exp Immunol*, 96, 59-63 (1994)
134. Nelson, D. R., R. P. Gonzalez-Peralta, K. Qian, Y. Xu, C. G. Marousis, G. L. Davis & J. Y. Lau: Transforming growth factor-beta 1 in chronic hepatitis C. *J Viral Hepat*, 4, 29-35 (1997)
135. Roulot, D., H. Durand, T. Coste, J. Rautureau, A. D. Strosberg, R. Benarous & S. Marullo: Quantitative analysis of transforming growth factor beta 1 messenger RNA in the liver of patients with chronic hepatitis C: absence of correlation between high levels and severity of disease. *Hepatology*, 21, 298-304 (1995)
136. Xu, J., J. Menezes, U. Prasad & A. Ahmad: Elevated serum levels of transforming growth factor beta1 in Epstein-Barr virus-associated nasopharyngeal carcinoma patients. *Int J Cancer*, 84, 396-399 (1999)
137. Michelson, S., J. Alcamí, S. J. Kim, D. Danielpour, F. Bachelier, L. Picard, C. Bessia, C. Paya & J. L. Virelizier: Human cytomegalovirus infection induces transcription and secretion of transforming growth factor beta 1. *J Virol*, 68, 5730-5737 (1994)
138. Agarwal, R., E. A. Elbishbishi, U. C. Chaturvedi, R. Nagar & A. S. Mustafa: Profile of transforming growth factor-beta 1 in patients with dengue haemorrhagic fever. *Int J Exp Pathol*, 80, 143-149 (1999)
139. Schultz-Cherry, S. & V. S. Hinshaw: Influenza virus neuraminidase activates latent transforming growth factor beta. *J Virol*, 70, 8624-8629 (1996)
140. Kim, S. J., J. H. Kehrl, J. Burton, C. L. Tandler, K. T. Jeang, D. Danielpour, C. Thevenin, K. Y. Kim, M. B. Sporn & A. B. Roberts: Transactivation of the transforming growth factor beta 1 (TGF-beta 1) gene by human T lymphotropic virus type 1 tax: a potential mechanism for the increased production of TGF-beta 1 in adult T cell leukemia. *J Exp Med*, 172, 121-129 (1990)
141. Liu, L., Z. Xu, R. C. Fuhlbrigge, V. Pena-Cruz, J. Lieberman & T. S. Kupper: Vaccinia virus induces strong immunoregulatory cytokine production in healthy human epidermal keratinocytes: a novel strategy for immune evasion. *J Virol*, 79, 7363-7370 (2005)
142. Goudy, K. S., B. R. Burkhardt, C. Wasserfall, S. Song, M. L. Campbell-Thompson, T. Brusko, M. A. Powers, M. J. Clare-Salzler, E. S. Sobel, T. M. Ellis, T. R. Flotte & M. A. Atkinson: Systemic overexpression of IL-10 induces CD4⁺CD25⁺ cell populations *in vivo* and ameliorates type 1 diabetes in nonobese diabetic mice in a dose-dependent fashion. *J Immunol*, 171, 2270-2278 (2003)
143. Steinbrink, K., M. Wolf, H. Jonuleit, J. Knop & A. H. Enk: Induction of tolerance by IL-10-treated dendritic cells. *J Immunol*, 159, 4772-4780 (1997)
144. Sato, K., N. Yamashita, M. Baba & T. Matsuyama: Modified myeloid dendritic cells act as regulatory dendritic cells to induce anergic and regulatory T cells. *Blood*, 101, 3581-3589 (2003)
145. Reiser, M., C. G. Marousis, D. R. Nelson, G. Lauer, R. P. Gonzalez-Peralta, G. L. Davis & J. Y. Lau: Serum interleukin 4 and interleukin 10 levels in patients with

- chronic hepatitis C virus infection. *J Hepatol*, 26, 471-478 (1997)
146. Villinger, F., P. E. Rollin, S. S. Brar, N. F. Chikkala, J. Winter, J. B. Sundstrom, S. R. Zaki, R. Swanepoel, A. A. Ansari & C. J. Peters: Markedly elevated levels of interferon (IFN)-gamma, IFN-alpha, interleukin (IL)-2, IL-10, and tumor necrosis factor-alpha associated with fatal Ebola virus infection. *J Infect Dis*, 179 Suppl 1, S188-191 (1999)
147. Stylianou, E., P. Aukrust, D. Kvale, F. Muller & S. S. Froland: IL-10 in HIV infection: increasing serum IL-10 levels with disease progression--down-regulatory effect of potent anti-retroviral therapy. *Clin Exp Immunol*, 116, 115-120 (1999)
148. Bozkaya, H., M. Bozdayi, R. Turkyilmaz, M. Sarioglu, H. Cetinkaya, K. Cinar, K. Kose, C. Yurdaydin & O. Uzunlimoglu: Circulating IL-2, IL-10 and TNF-alpha in chronic hepatitis B: their relations to HBeAg status and the activity of liver disease. *Hepatogastroenterology*, 47, 1675-1679 (2000)
149. Suradhat, S., R. Thanawongnuwech & Y. Poovorawan: Upregulation of IL-10 gene expression in porcine peripheral blood mononuclear cells by porcine reproductive and respiratory syndrome virus. *J Gen Virol*, 84, 453-459 (2003)
150. Murai, H., A. Terada, M. Mizuno, M. Asai, Y. Hirabayashi, S. Shimizu, T. Morishita, H. Kakita, M. H. Hussein, T. Ito, I. Kato, K. Asai & H. Togari: IL-10 and RANTES are Elevated in Nasopharyngeal Secretions of Children with Respiratory Syncytial Virus Infection. *Allergol Int*, 56, 157-163 (2007)
151. Moore, K. W., P. Vieira, D. F. Fiorentino, M. L. Trounstein, T. A. Khan & T. R. Mosmann: Homology of cytokine synthesis inhibitory factor (IL-10) to the Epstein-Barr virus gene BCRF1. *Science*, 248, 1230-1234 (1990)
152. Fleming, S. B., C. A. McCaughan, A. E. Andrews, A. D. Nash & A. A. Mercer: A homolog of interleukin-10 is encoded by the poxvirus orf virus. *J Virol*, 71, 4857-4861 (1997)
153. Rode, H. J., W. Janssen, A. Rosen-Wolff, J. J. Bugert, P. Thein, Y. Becker & G. Darai: The genome of equine herpesvirus type 2 harbors an interleukin 10 (IL10)-like gene. *Virus Genes*, 7, 111-116 (1993)
154. de Waal Malefyt, R., J. Haanen, H. Spits, M. G. Roncarolo, A. te Velde, C. Figdor, K. Johnson, R. Kastelein, H. Yssel & J. E. de Vries: Interleukin 10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T cell proliferation by diminishing the antigen-presenting capacity of monocytes via downregulation of class II major histocompatibility complex expression. *J Exp Med*, 174, 915-924 (1991)
155. Takayama, T., Y. Nishioka, L. Lu, M. T. Lotze, H. Tahara & A. W. Thomson: Retroviral delivery of viral interleukin-10 into myeloid dendritic cells markedly inhibits their allostimulatory activity and promotes the induction of T-cell hyporesponsiveness. *Transplantation*, 66, 1567-1574 (1998)
156. Yamazaki, S., T. Iyoda, K. Tarbell, K. Olson, K. Velinzon, K. Inaba & R. M. Steinman: Direct expansion of functional CD25⁺ CD4⁺ regulatory T cells by antigen-processing dendritic cells. *J Exp Med*, 198, 235-247 (2003)
157. Fehervari, Z. & S. Sakaguchi: Control of Foxp3⁺ CD25⁺CD4⁺ regulatory cell activation and function by dendritic cells. *Int Immunol*, 16, 1769-1780 (2004)
158. Engelmayer, J., M. Larsson, M. Subklewe, A. Chahroudi, W. I. Cox, R. M. Steinman & N. Bhardwaj: Vaccinia virus inhibits the maturation of human dendritic cells: a novel mechanism of immune evasion. *J Immunol*, 163, 6762-6768 (1999)
159. Salio, M., M. Cella, M. Suter & A. Lanzavecchia: Inhibition of dendritic cell maturation by herpes simplex virus. *Eur J Immunol*, 29, 3245-3253 (1999)
160. Fugier-Vivier, I., C. Servet-Delprat, P. Rivailler, M. C. Rissouan, Y. J. Liu & C. Rabourdin-Combe: Measles virus suppresses cell-mediated immunity by interfering with the survival and functions of dendritic and T cells. *J Exp Med*, 186, 813-823 (1997)
161. Moutafsi, M., A. M. Mehl, L. K. Borysiewicz & Z. Tabi: Human cytomegalovirus inhibits maturation and impairs function of monocyte-derived dendritic cells. *Blood*, 99, 2913-2921 (2002)
162. Senechal, B., A. M. Boruchov, J. L. Reagan, D. N. Hart & J. W. Young: Infection of mature monocyte-derived dendritic cells with human cytomegalovirus inhibits stimulation of T-cell proliferation via the release of soluble CD83. *Blood*, 103, 4207-4215 (2004)
163. Bilsborough, J., T. C. George, A. Norment & J. L. Viney: Mucosal CD8alpha⁺ DC, with a plasmacytoid phenotype, induce differentiation and support function of T cells with regulatory properties. *Immunology*, 108, 481-492 (2003)
164. Kawamura, K., N. Kadowaki, T. Kitawaki & T. Uchiyama: Virus-stimulated plasmacytoid dendritic cells induce CD4⁺ cytotoxic regulatory T cells. *Blood*, 107, 1031-1038 (2006)
165. Liu, Y. J.: Dendritic cell subsets and lineages, and their functions in innate and adaptive immunity. *Cell*, 106, 259-262 (2001)
166. Cella, M., F. Facchetti, A. Lanzavecchia & M. Colonna: Plasmacytoid dendritic cells activated by influenza virus and CD40L drive a potent TH1 polarization. *Nat Immunol*, 1, 305-310 (2000)
167. Fonteneau, J. F., M. Gilliet, M. Larsson, I. Dasilva, C. Munz, Y. J. Liu & N. Bhardwaj: Activation of influenza virus-specific CD4⁺ and CD8⁺ T cells: a new role for plasmacytoid dendritic cells in adaptive immunity. *Blood*, 101, 3520-3526 (2003)
168. McKenna, K., A. S. Beignon & N. Bhardwaj: Plasmacytoid dendritic cells: linking innate and adaptive immunity. *J Virol*, 79, 17-27 (2005)
169. Siegal, F. P., N. Kadowaki, M. Shodell, P. A. Fitzgerald-Bocarsly, K. Shah, S. Ho, S. Antonenko & Y. J. Liu: The nature of the principal type 1 interferon-producing cells in human blood. *Science*, 284, 1835-1837 (1999)
170. Muller, U., U. Steinhoff, L. F. Reis, S. Hemmi, J. Pavlovic, R. M. Zinkernagel & M. Aguet: Functional role of type I and type II interferons in antiviral defense. *Science*, 264, 1918-1921 (1994)
171. Levings, M. K., R. Sangregorio, F. Galbiati, S. Squadrone, R. de Waal Malefyt & M. G. Roncarolo: IFN-alpha and IL-10 induce the differentiation of human type 1 T regulatory cells. *J Immunol*, 166, 5530-5539 (2001)

172. Green, E. A., L. Gorelik, C. M. McGregor, E. H. Tran & R. A. Flavell: CD4⁺CD25⁺ T regulatory cells control anti-islet CD8⁺ T cells through TGF-beta-TGF-beta receptor interactions in type 1 diabetes. *Proc Natl Acad Sci U S A*, 100, 10878-10883 (2003)
173. Klein, L., K. Khazaie & H. von Boehmer: *In vivo* dynamics of antigen-specific regulatory T cells not predicted from behavior *in vitro*. *Proc Natl Acad Sci U S A*, 100, 8886-8891 (2003)
174. Read, S., V. Malmstrom & F. Powrie: Cytotoxic T lymphocyte-associated antigen 4 plays an essential role in the function of CD25(+)CD4(+) regulatory cells that control intestinal inflammation. *J Exp Med*, 192, 295-302 (2000)
175. Liu, H., B. Hu, D. Xu & F. Y. Liew: CD4⁺CD25⁺ regulatory T cells cure murine colitis: the role of IL-10, TGF-beta, and CTLA4. *J Immunol*, 171, 5012-5017 (2003)
176. Nakamura, K., A. Kitani, I. Fuss, A. Pedersen, N. Harada, H. Nawata & W. Strober: TGF-beta 1 plays an important role in the mechanism of CD4⁺CD25⁺ regulatory T cell activity in both humans and mice. *J Immunol*, 172, 834-842 (2004)
177. Fontenot, J. D., M. A. Gavin & A. Y. Rudensky: Foxp3 programs the development and function of CD4⁺CD25⁺ regulatory T cells. *Nat Immunol*, 4, 330-336 (2003)
178. Khattri, R., T. Cox, S. A. Yasayko & F. Ramsdell: An essential role for Scurfin in CD4⁺CD25⁺ T regulatory cells. *Nat Immunol*, 4, 337-342 (2003)
179. Yagi, H., T. Nomura, K. Nakamura, S. Yamazaki, T. Kitawaki, S. Hori, M. Maeda, M. Onodera, T. Uchiyama, S. Fujii & S. Sakaguchi: Crucial role of FOXP3 in the development and function of human CD25⁺CD4⁺ regulatory T cells. *Int Immunol*, 16, 1643-1656 (2004)
180. Oh, U., C. Grant, C. Griffith, K. Fugo, N. Takenouchi & S. Jacobson: Reduced Foxp3 protein expression is associated with inflammatory disease during human t lymphotropic virus type 1 infection. *J Infect Dis*, 193, 1557-1566 (2006)
181. Walker, M. R., D. J. Kasproicz, V. H. Gersuk, A. Benard, M. Van Landeghen, J. H. Buckner & S. F. Ziegler: Induction of FoxP3 and acquisition of T regulatory activity by stimulated human CD4⁺CD25⁺ T cells. *J Clin Invest*, 112, 1437-1443 (2003)
182. Williams, L. M. & A. Y. Rudensky: Maintenance of the Foxp3-dependent developmental program in mature regulatory T cells requires continued expression of Foxp3. *Nat Immunol*, 8, 277-284 (2007)
183. Sereti, I., H. Imamichi, V. Natarajan, T. Imamichi, M. S. Ramchandani, Y. Badralmaa, S. C. Berg, J. A. Metcalf, B. K. Hahn, J. M. Shen, A. Powers, R. T. Davey, J. A. Kovacs, E. M. Shevach & H. C. Lane: *In vivo* expansion of CD4CD45RO-CD25 T cells expressing foxP3 in IL-2-treated HIV-infected patients. *J Clin Invest*, 115, 1839-1847 (2005)
184. Hryniewicz, A., A. Boasso, Y. Edghill-Smith, M. Vaccari, D. Fuchs, D. Venzon, J. Nacsa, M. R. Betts, W. P. Tsai, J. M. Heraud, B. Beer, D. Blanset, C. Chougnet, I. Lowy, G. M. Shearer & G. Franchini: CTLA-4 blockade decreases TGF-beta, IDO, and viral RNA expression in tissues of SIVmac251-infected macaques. *Blood*, 108, 3834-3842 (2006)
185. Steiner, K., I. Waase, T. Rau, M. Dietrich, B. Fleischer & B. M. Broker: Enhanced expression of CTLA-4 (CD152) on CD4⁺ T cells in HIV infection. *Clin Exp Immunol*, 115, 451-457 (1999)
186. Leng, Q., Z. Bentwich, E. Magen, A. Kalinkovich & G. Borkow: CTLA-4 upregulation during HIV infection: association with anergy and possible target for therapeutic intervention. *Aids*, 16, 519-529 (2002)
187. Andersson, J., A. Boasso, J. Nilsson, R. Zhang, N. J. Shire, S. Lindback, G. M. Shearer & C. A. Chougnet: The prevalence of regulatory T cells in lymphoid tissue is correlated with viral load in HIV-infected patients. *J Immunol*, 174, 3143-3147 (2005)
188. Larrea, E., J. I. Riezu-Boj, L. Gil-Guerrero, N. Casares, R. Aldabe, P. Sarobe, M. P. Civeira, J. L. Heeney, C. Rollier, B. Verstrepen, T. Wakita, F. Borrás-Cuesta, J. J. Lasarte & J. Prieto: Upregulation of indoleamine 2,3-dioxygenase in hepatitis C virus infection. *J Virol*, 81, 3662-3666 (2007)
189. Rollier, C. S., G. Paranhos-Baccala, E. J. Verschoor, B. E. Verstrepen, J. A. Drexhage, Z. Fagrouch, J. L. Berland, F. Komurian-Pradel, B. Duverger, N. Himoudi, C. Staib, M. Meyr, M. Whelan, J. A. Whelan, V. A. Adams, E. Larrea, J. I. Riezu, J. J. Lasarte, B. Bartosch, F. L. Cosset, W. J. Spaan, H. M. Diepolder, G. R. Pape, G. Sutter, G. Inchauspe & J. L. Heeney: Vaccine-induced early control of hepatitis C virus infection in chimpanzees fails to impact on hepatic PD-1 and chronicity. *Hepatology*, 45, 602-613 (2007)
190. Ayukawa, H., T. Matsubara, M. Kaneko, M. Hasegawa, T. Ichiyama & S. Furukawa: Expression of CTLA-4 (CD152) in peripheral blood T cells of children with influenza virus infection including encephalopathy in comparison with respiratory syncytial virus infection. *Clin Exp Immunol*, 137, 151-155 (2004)
191. Matsubara, T., R. Anwar, M. Fujiwara, T. Ichiyama & S. Furukawa: CTLA-4 (CD152) expression in peripheral blood T cells in Kawasaki disease. *Clin Exp Immunol*, 132, 169-173 (2003)
192. Venkatachari, N. J., B. Majumder & V. Ayyavoo: Human immunodeficiency virus (HIV) type 1 Vpr induces differential regulation of T cell costimulatory molecules: direct effect of Vpr on T cell activation and immune function. *Virology*, 358, 347-356 (2007)
193. Paust, S., L. Lu, N. McCarty & H. Cantor: Engagement of B7 on effector T cells by regulatory T cells prevents autoimmune disease. *Proc Natl Acad Sci U S A*, 101, 10398-10403 (2004)
194. Fallarino, F., U. Grohmann, K. W. Hwang, C. Orabona, C. Vacca, R. Bianchi, M. L. Belladonna, M. C. Fioretti, M. L. Alegre & P. Puccetti: Modulation of tryptophan catabolism by regulatory T cells. *Nat Immunol*, 4, 1206-1212 (2003)
195. Grohmann, U., C. Orabona, F. Fallarino, C. Vacca, F. Calcinaro, A. Falorni, P. Candeloro, M. L. Belladonna, R. Bianchi, M. C. Fioretti & P. Puccetti: CTLA-4-Ig regulates tryptophan catabolism *in vivo*. *Nat Immunol*, 3, 1097-1101 (2002)
196. Munn, D. H., E. Shafizadeh, J. T. Attwood, I. Bondarev, A. Pashine & A. L. Mellor: Inhibition of T cell proliferation by macrophage tryptophan catabolism. *J Exp Med*, 189, 1363-1372 (1999)

197. Terness, P., T. M. Bauer, L. Rose, C. Dufter, A. Watzlik, H. Simon & G. Opelz: Inhibition of allogeneic T cell proliferation by indoleamine 2,3-dioxygenase-expressing dendritic cells: mediation of suppression by tryptophan metabolites. *J Exp Med*, 196, 447-457 (2002)
198. Frumento, G., R. Rotondo, M. Tonetti, G. Damonte, U. Benatti & G. B. Ferrara: Tryptophan-derived catabolites are responsible for inhibition of T and natural killer cell proliferation induced by indoleamine 2,3-dioxygenase. *J Exp Med*, 196, 459-468 (2002)
199. Taams, L. S., J. Smith, M. H. Rustin, M. Salmon, L. W. Poulter & A. N. Akbar: Human anergic/suppressive CD4(+)CD25(+) T cells: a highly differentiated and apoptosis-prone population. *Eur J Immunol*, 31, 1122-1131 (2001)
200. Piccirillo, C. A., J. J. Letterio, A. M. Thornton, R. S. McHugh, M. Mamura, H. Mizuhara & E. M. Shevach: CD4(+)CD25(+) regulatory T cells can mediate suppressor function in the absence of transforming growth factor beta1 production and responsiveness. *J Exp Med*, 196, 237-246 (2002)
201. Nesburn, A. B., I. Bettahi, G. Dasgupta, A. A. Chentoufi, X. Zhang, S. You, N. Morishige, A. J. Wahlert, D. J. Brown, J. V. Jester, S. L. Wechsler & L. Benmohamed: Functional Foxp3⁺ CD4⁺CD25(Bright+) "Natural" Regulatory T Cells are Abundant in Rabbit Conjunctiva and Suppress Virus-Specific CD4⁺ and CD8⁺ Effector T Cells During Ocular Herpes Infection. *J Virol* (2007)
202. Bauer, G., M. Gotschl & P. Hofler: Tumor-promoting activity of Epstein-Barr-virus-inducing factor transforming growth factor type beta (EIF/TGF-beta) is due to the induction of irreversible transformation. *Int J Cancer*, 47, 881-888 (1991)
203. Charan, S., K. Palmer, P. Chester, A. R. Mire-Sluis, A. Meager & N. Edington: Transforming growth factor-beta induced by live or ultraviolet-inactivated equid herpes virus type-1 mediates immunosuppression in the horse. *Immunology*, 90, 586-591 (1997)
204. Weiner, H. L.: Induction and mechanism of action of transforming growth factor-beta-secreting Th3 regulatory cells. *Immunol Rev*, 182, 207-214 (2001)
205. Garba, M. L., C. D. Pilcher, A. L. Bingham, J. Eron & J. A. Frelinger: HIV antigens can induce TGF-beta(1)-producing immunoregulatory CD8⁺ T cells. *J Immunol*, 168, 2247-2254 (2002)
206. Annacker, O., R. Pimenta-Araujo, O. Burlen-Defranoux, T. C. Barbosa, A. Cumano & A. Bandeira: CD25⁺ CD4⁺ T cells regulate the expansion of peripheral CD4 T cells through the production of IL-10. *J Immunol*, 166, 3008-3018 (2001)
207. Pontoux, C., A. Banz & M. Papiernik: Natural CD4 CD25(+) regulatory T cells control the burst of superantigen-induced cytokine production: the role of IL-10. *Int Immunol*, 14, 233-239 (2002)
208. Asseman, C., S. Mauze, M. W. Leach, R. L. Coffman & F. Powrie: An essential role for interleukin 10 in the function of regulatory T cells that inhibit intestinal inflammation. *J Exp Med*, 190, 995-1004 (1999)
209. Huang, X., J. Zhu & Y. Yang: Protection against autoimmunity in nonlymphopenic hosts by CD4⁺ CD25⁺ regulatory T cells is antigen-specific and requires IL-10 and TGF-beta. *J Immunol*, 175, 4283-4291 (2005)
210. Schorle, H., T. Holtschke, T. Hunig, A. Schimpl & I. Horak: Development and function of T cells in mice rendered interleukin-2 deficient by gene targeting. *Nature*, 352, 621-624 (1991)
211. Sadlack, B., R. Kuhn, H. Schorle, K. Rajewsky, W. Muller & I. Horak: Development and proliferation of lymphocytes in mice deficient for both interleukins-2 and -4. *Eur J Immunol*, 24, 281-284 (1994)
212. Nelson, B. H.: IL-2, regulatory T cells, and tolerance. *J Immunol*, 172, 3983-3988 (2004)
213. McHugh, R. S., M. J. Whitters, C. A. Piccirillo, D. A. Young, E. M. Shevach, M. Collins & M. C. Byrne: CD4(+)CD25(+) immunoregulatory T cells: gene expression analysis reveals a functional role for the glucocorticoid-induced TNF receptor. *Immunity*, 16, 311-323 (2002)
214. Stephens, G. L., R. S. McHugh, M. J. Whitters, D. A. Young, D. Luxenberg, B. M. Carreno, M. Collins & E. M. Shevach: Engagement of glucocorticoid-induced TNFR family-related receptor on effector T cells by its ligand mediates resistance to suppression by CD4⁺CD25⁺ T cells. *J Immunol*, 173, 5008-5020 (2004)
215. He, H., R. J. Messer, S. Sakaguchi, G. Yang, S. J. Robertson & K. J. Hasenkrug: Reduction of retrovirus-induced immunosuppression by *in vivo* modulation of T cells during acute infection. *J Virol*, 78, 11641-11647 (2004)
216. La, S., E. Kim & B. Kwon: *In vivo* ligation of glucocorticoid-induced TNF receptor enhances the T-cell immunity to herpes simplex virus type 1. *Exp Mol Med*, 37, 193-198 (2005)
217. Stone, G. W., S. Barzee, V. Snarsky, K. Kee, C. A. Spina, X. F. Yu & R. S. Kornbluth: Multimeric soluble CD40 ligand and GITR ligand as adjuvants for human immunodeficiency virus DNA vaccines. *J Virol*, 80, 1762-1772 (2006)
218. Ronchetti, S., O. Zollo, S. Bruscoli, M. Agostini, R. Bianchini, G. Nocentini, E. Ayroldi & C. Riccardi: GITR, a member of the TNF receptor superfamily, is costimulatory to mouse T lymphocyte subpopulations. *Eur J Immunol*, 34, 613-622 (2004)

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