# NF-kappaB: a potential target for cancer chemoprevention and therapy

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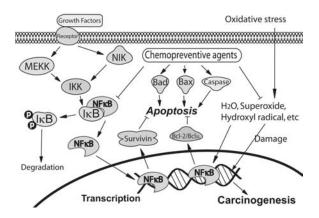
# 1. ABSTRACT

Nuclear factor-kappaB pathway plays important roles in the control of cell proliferation, differentiation. apoptosis, inflammation, stress response, cell signaling transduction, and other physiological processes. Because the disorder of these physiological processes has been linked with the onset of cancers, NF-kappaB has been described as a major culprit in cancer. Experimental in vitro and in vivo studies have shown that down-regulation of NFkappaB activity by natural and synthetic NF-kappaB inhibitors suppresses the development of carcinogeninduced tumors, inhibits the growth of cancer cells, and induces apoptosis with alternation of gene expression which is critical for the control of carcinogenesis and cancer cell survival. Moreover, recent studies indicate that the effects of conventional cancer therapeutics could be enhanced by natural and synthetic NF-kappaB inhibitors, suggesting that down-regulation of NF-kappaB could sensitize cancer cells to conventional therapeutics. Therefore, targeting NF-kappaB is a novel preventive and therapeutic strategy against human cancers. In this brief review article, we will summarize the state of our knowledge for the role of NF-kappaB in relation to cancer prevention and therapy.

### 2. INTRODUCTION

The nuclear factor-kappaB (NF-kappaB) pathway is one of the most important cellular signaling transduction pathways involved in both physiological processes and disease conditions. It plays important roles in the control of immune function, differentiation, inflammation, stress response, apoptosis, and cell survival (1, 2). Moreover, NF-kappaB is critically involved in the processes of development and progression of cancers, raising its importance in cancer research (3, 4).

The NF-kappaB family is composed of five proteins: RelA (p65), RelB, c-Rel, NF-kappaB1(p50), and NF-kappaB2(p52), each of which may form homo- or heterodimers. In human cells without specific extracellular signal, NF-kappaB is sequestered in the cytoplasm through tight association with its inhibitors: IkappaBalpha which acts as a NF-kappaB inhibitor and p100 proteins which serves as both an inhibitor and precursor of NF-kappaB DNA-binding subunits. NF-kappaB can be activated by many types of stimuli including TNF-alpha, UV radiation, free radicals, etc. The activation of NF-kappaB occurs through site-specific phosphorylation of IkappaBalpha by IKKbeta and/or phosphorylation of p100 by IKKalpha,



**Figure 1.** NF-kappaB as a target of chemopreventive agents in cancer prevention.

leading to degradation of IkappaBalpha by the 26S proteasome and/or the processing of p100 into smaller forms (p52). This process allows two forms of NF-kappaB (p50-p65 and p52-RelB) to become free, resulting in the translocation of active NF-kappaB into the nucleus for binding to NF-kappaB-specific DNA-binding sites and, in turn, regulating the transcription of target genes (5, 6). In this way, NF-kappaB controls the expression of its target genes that are involved in cellular physiological processes including cell proliferation, differentiation, apoptosis, inflammation, stress response, etc. The disorder of these physiological processes has been linked with the onset of cancers; therefore, NF-kappaB has been described as a major culprit in cancer.

# 3. NF-kappaB AND CANCERS

# 3.1. Oxidative stress and NF-kappaB activation

NF-kappaB is critically involved in the processes of oxidative stress response. Oxidative stress is defined as an increase in intracellular reactive oxygen species (ROS) such as  $H_2O_2$ , superoxide, hydroxyl radical, etc. Direct addition of  $H_2O_2$  to cell culture medium could activate NF-kappaB in several types of cell lines (7). ROS in cells are increased in response to agents that also activate NF-kappaB. These findings suggest that oxidative stress activates NF-kappaB activity in the cells (8).

When ROS are formed intracellularly, they have the capacity to induce DNA damage and alter cellular signal transduction pathways, among which NF-kappaB signaling is the most important (Figure 1). Since DNA damage plays a central role in carcinogenesis, it is conceivable that oxidative stress could be carcinogenic (9). Hence, DNA repair is an important process in preventing mutagenesis. However, under oxidative stress, the repair of DNA damage can be inhibited by several redox-dependent metals, resulting in carcinogenesis. Moreover, the activation of NF-kappaB by ROS under oxidative stress has been known as a key event in carcinogenesis (3, 4). Therefore, it has been believed that the inhibition of NFkappaB could suppress carcinogenesis. In addition, inhibition of ROS-stimulated NF-kappaB activation is widely recognized as a strategy to combat inflammatory disease. Evidence from many studies suggests that inhibition of NF-kappaB activity is not only beneficial for the treatment of inflammation but also for cancer therapy (6, 10). Examination of the inflammatory microenvironment in neoplastic tissues has supported the hypothesis that inflammation is a cofactor in oncogenesis for a variety of cancers. Many anti-inflammation drugs and antioxidants are known to inhibit NF-kappaB activity and induce apoptosis; therefore, they could also be used in the prevention and treatment of cancers.

## 3.2. NF-kappaB activation in cancers

NF-kappaB have shown its proliferative and antiapoptotic effects in normal and malignant cells. Studies have shown that NF-kappaB is over-expressed and activated in various cancers, especially in the poorly differentiated cancers (3). It has been reported that NFkappaB is constitutively activated in Hodgkin's tumor cells whereas inhibition of NF-kappaB blocks the tumor cell growth (11). Studies have also demonstrated that NFkappaB regulates growth and survival of multiple myeloma, and that NF-kappaB is a novel therapeutics target in multiple myeloma (12). NF-kappaB is constitutively activated in most human pancreatic cancer tissues and cell lines but not in normal pancreatic tissues and immortalized pancreatic ductal epithelial cells, suggesting that the activation of NF-kappaB is involved in the carcinogenesis of pancreatic cancers (13, 14). The activation of NF-kappaB is also frequently observed in breast and prostate cancer cells (15, 16). Recent reports have shown that NF-kappaB is activated in gastric carcinomas and squamous cell carcinoma of the head and neck (17, 18). Therefore, it is becoming obvious that inhibition of NF-kappaB activity is highly desirable in the prevention and treatment of cancer and, as such, the chemopreventive agents that are known to down-regulate the activity of NF-kappaB could potentially inhibit the development and progression of cancers (Figure 1).

# 4. NF-kappaB AS A TARGET OF CHEMOPREVENTION

Experimental studies have shown that natural antioxidant compounds including isoflavones, I3C, DIM, curcumin, EGCG, etc, inhibit the activity of NF-kappaB and the growth of cancer cells, and also induce apoptosis in cancer cells (19-23), suggesting that NF-kappaB could be a target for cancer prevention. Moreover, synthetic antioxidants or NF-kappaB inhibitors also suppress tumor growth through inhibition of NF-kappaB activity. Thus, natural or synthetic antioxidants that inactivate NF-kappaB activity could serve as molecularly targeting agents against cancers (Figure 1).

### 4.1. Isoflavones

Isoflavones are a subclass of flavonoids and much more readily found in soybeans. Genistein, daidzein, and glycitein are three main isoflavones found in soybeans. Evidences from epidemiologic and *in vivo* studies have shown a decreased risk of cancer with soy consumption (24-26). Soy isoflavone genistein has been believed to be responsible for the decreased risk of cancer; however, some

controversies exist in the field. Experimental studies have revealed that isoflavones, particularly genistein, exert antioxidant effects on various human cells. It has been found that genistein protects cells against ROS by scavenging free radicals and reducing the expression of stress-response related genes (27, 28). Isoflavones also stimulate antioxidant protein gene expression in Caco-2 cells (29). Isoflavone supplementation has been found to reduce hydrogen peroxide-induced DNA damage in sperm, suggesting the antioxidant effects of isoflavone (30). Moreover, it has been demonstrated that genistein inhibits tumor promoter, 12-O-tetradecanoylphorbol-13-acetate induced hydrogen peroxide production polymorphonuclear leukocytes and HL-60 cells (31), suggesting the inhibitory effect of genistein on carcinogenesis through antioxidant mechanism.

We have investigated whether genistein treatment could modulate NF-kappaB DNA binding activity in prostate, breast, and pancreatic cancer cells. We found that 50 µM genistein treatment for 24-72 hours significantly inhibited NF-kappaB DNA-binding activity in all cell lines we tested (22, 32-34). We also found that genistein significantly inhibited cancer cell growth and induced apoptotic cell death through the down-regulation of NFkappaB (22, 32-34). Other investigators also demonstrated a similar effect of genistein on NF-kappaB in different types of cells. Baxa et al. showed that genistein reduced NF-kappaB in T lymphoma cells via a caspase-mediated cleavage of IkappaBalpha (35). Tabary et al. also found that genistein inhibited constitutive and inducible NFkappaB activation and decreased IL-8 production in human cystic fibrosis bronchial gland cells (36). These results suggested that genistein functions as an antioxidant, which could be a potent agent for the inhibition of oxidative stress and the prevention of cancers.

Using *in vivo* studies, we found that isoflavone supplementation inhibited NF-kappaB DNA binding activity by 56% and abrogated TNF-alpha induced NF-kappaB activity by 50% in lymphocytes harvested from human subjects receiving soy isoflavone supplements (37). We also measured the levels of oxidative DNA damage in the blood of the subjects before and after supplementation with isoflavone. We found that isoflavone is very effective in reducing the level of 5-OhmdU, a modified DNA base that represents the endogenous status of cellular oxidative stress (37). These results demonstrated that isoflavone inhibits NF-kappaB activation and decreases oxidative damage, suggesting that soy isoflavone is a potent antioxidant and could be used for cancer prevention.

# 4.2. Indole-3-carbinol and 3,3'-diindolylmethane

Indole-3-carbinol (I3C) is produced from cruciferous vegetables. Under the acidic conditions of the stomach, I3C undergoes rapid self-condensation reactions to form several derivatives. 3,3'-diindolylmethane (DIM) is the major derivative and condensation product of I3C. Epidemiological and dietary studies have shown an association between high dietary intake of cruciferous vegetables and a decreased risk of cancer (38). The formation of DIM from I3C has been believed to be a likely

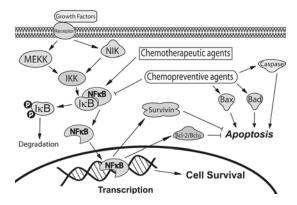
prerequisite for I3C-induced anti-carcinogenesis. I3C and DIM have shown their ability to reduce oxidative stress and stimulate antioxidant response element-driven gene expression as antioxidants (39, 40). We and other investigators have found that I3C and DIM inhibit oncogenesis and cancer cell growth, and induce apoptosis in various cancer cells (21, 23, 41, 42), suggesting that I3C and DIM may serve as potent agents for the prevention of cancers. We have also found that I3C and DIM significantly inhibited NF-kappaB DNA binding activity in prostate and breast cancer cells, corresponding with the inhibition of cell proliferation and the induction of apoptosis by I3C and DIM (21, 23). These results suggest that inhibition of NF-kappaB activity by I3C and DIM may reduce the oxidative stress induced by ROS or TNF-alpha and that I3C and DIM could be used for cancer prevention and treatment.

### 4.3. Curcumin

Curcumin is a compound from Curcuma longa (tumeric). Curcumin has received much attention due to its pronounced anti-inflammatory, anti-oxidative. immunomodulating, anti-atherogenic, and anticarcinogenic activities (43, 44). It has been reported that curcumin is a potent scavenger of oxygen free radicals such as hydroxyl radical and nitrogen dioxide radical (45). Curcumin inhibits lipid peroxidation in rat brain, liver, and lens, suggesting its antioxidant properties (46-48). Chuang have shown that curcumin diethylnitrosamine-induced inflammation liver activation of NF-kappaB in rats (49). It has been reported that curcumin inhibited IKK, suppressed both constitutive and inducible NF-kappaB activation, and potentiated TNFalpha induced apoptosis (20). Curcumin also showed strong antioxidant and anticancer properties mediated through the regulation of the expression of genes that require the activation of activator protein 1 (AP1) and NF-kappaB (50). Moreover, curcumin inhibits the growth of cancer cells, induces apoptosis, inactivates Akt, and regulates the expression of genes related to anti-invasion (51, 52), suggesting its role in cancer prevention and treatment.

# 4.4. Epigallocatechin-3-gallate

Epigallocatechin-3-gallate (EGCG) is the most potent catechins from green tea. Epidemiologic evidence has shown that consumption of green tea significantly decreased overall cancer incidence (53). EGCG has shown its strong antioxidant activity. It has been reported that EGCG treatment resulted in a significant inhibition of NFkappaB activation and translocation to the nucleus by suppressing the degradation of IkappaBalpha in the cytoplasm (19, 54). EGCG also inhibited the activation of IKK and phosphorylation of IkappaBalpha (55, 56), corresponding with the inhibition of NF-kappaB activation. Growing evidence shows that EGCG inhibits the proliferation of various cancer cells and induces apoptotic processes in cancer cells through inhibition of NF-kappaB (53, 57). It has also been found that EGCG had a concurrent effect on two important transcription factors, p53 (stabilization of p53) and NF-kappaB (negative regulation of NF-kappaB activity) (58). Moreover, EGCG has been found to reduce the levels of matrix



**Figure 2.** Targeting NF-kappaB in the combination treatment with chemopreventive agents and chemotherapeutics.

metalloproteinases, suppress angiogenesis, and inhibit invasion and metastasis (59, 60). These findings suggest that EGCG is a potent antioxidant which is likely to be useful for cancer prevention and treatment.

#### 4.5. Resveratrol

Resveratrol is present in a wide variety of plants including grapes, mulberries, and peanuts. Relatively high quantities of resveratrol are also found in grapes. Resveratrol has beneficial effects on the reduction of oxidative stress and the prevention of cancers (61, 62). It has been found that resveratrol reduces DNA damage and the formation of A2E-epoxidation, which is implicated in the degenerative disease (63). Experimental studies have shown that resveratrol inhibits the growth of various cancer cells and induces apoptotic cell death (64-66). Resveratrol also shows its inhibitory effects on the activity of NF-kappaB (67), suggesting that its role as antioxidant may contribute to the prevention of cancers.

### 4.6. Lycopene

Tomatoes are rich in lycopene. The consumption of tomatoes and tomato products containing lycopene has been associated with a decreased risk of cancers (68). Lycopene is a potent antioxidant. It has been found that lycopene exhibits a high physical quenching rate constant with singlet oxygen, suggesting its high activity as antioxidant (69). Lycopene also shows beneficial effects on cancer prevention. Giovannucci et al. have reported that frequent consumption of tomato products is associated with a lower risk of prostate cancer (70). The inverse associations between plasma lycopene and prostate cancer have also been reported (71). Experimental studies have shown that lycopene inhibits cell growth in breast, prostate and endometrial cancer cells through the regulation of cell cycle-related genes (72, 73). Lycopene also inhibits the activity of NF-kappaB (74). A clinical trial has revealed that lycopene supplementation could reduce tumor size and PSA level in localized prostate cancers (75), suggesting its promising effect on prostate cancer treatment.

# 4.7. Vitamins

Vitamin E is a lipid-soluble antioxidant distributed in green leaf vegetables, nuts, seeds, sunflower,

and plant oils. Vitamin E exerts a potent antioxidant effect. It has been reported that vitamin E inhibits NF-kappaB activation and NF-kappaB-dependent transcription, and induces differentiation through reduction of NF-kappaB (76, 77), suggesting that vitamin E may exert its antioxidant effect through modulation of NF-kappaB. Vitamin C is a water-soluble antioxidant. Sources of vitamin C include fruits and vegetables. It has been reported that vitamin C inhibits NF-kappaB activation by the inhibition of IkappaBalpha phosphorylation or activation of p38 mitogen-activated protein kinase (78, 79). These results suggest that vitamin E and C are beneficial for cancer prevention.

### 4.8. COX-2 inhibitors

Recently, aspirin has been considered as a cancer prevention drug because of its ability to down-regulate COX signaling. Experimental studies have shown that aspirin inhibits constitutively activated NF-kappaB in cell culture and, in turn, decreases the expression of the NF-kappaB downstream gene, especially COX-2, in pancreatic cancer cells (80). Another COX-2 inhibitor, SC236, has been shown to suppress NF-kappaB DNA binding activity and NF-kappaB mediated gene transcription (81). This effect was regulated through a mechanism independent of cyclooxygenase activity and prostaglandin synthesis. We and others have found that celecoxib, one of the COX-2 inhibitors, significantly inhibited tumor growth and induced apoptosis through the inhibition of NF-kappaB activation and COX-2 expression (82, 83).

# 5. TARGETING NF-kappaB IN THE COMBINATION TREATMENT WITH NF-kappaB INHIBITOR AND CANCER THERAPEUTICS

NF-kappaB is constitutively activated in most human cancers, suggesting that the activation of NFkappaB is involved in the processes of carcinogenesis. Current evidence has also shown that NF-κB participates in the processes of angiogenesis, invasion, and metastasis (10). Moreover, it has been known that several chemotherapeutics such as cisplatin and docetaxel can activate NF-kappaB (84-86). The activation of NFkappaB by chemotherapeutics has been found to cause acquired resistance to chemotherapeutic agents, resulting in treatment failure in cancer therapy. Experimental studies also demonstrated that other molecules (i.e. IL-1, E3-ubiquitin ligase receptor subunit betaTRCP1, etc) could induce NF-kappaB activation and lead to chemoresistance in human cancer cell lines (87, 88), suggesting that NF-kappaB plays important roles in the development of chemoresistance in cancers. Therefore, strategies targeting the NF-kappaB pathway could be a novel approach for improved cancer cell killing. Recently, more natural chemopreventive agents with anti-NF-kappaB activity and synthetic NF-kappaB inhibitors have been used in combination treatment with chemotherapeutics. Chemotherapy combined with these NF-kappaB inhibitors have shown enhanced anti-tumor activity synergistic through action, chemoresistance, or compensation of inverse properties (Figure 2).

### 5.1. Genistein

The in vitro and in vivo studies from our laboratory and others have demonstrated that the anti-tumor effects of chemotherapeutic agents could be enhanced by combination treatment with genistein through inhibition of NF-kappaB. We have reported that genistein in vitro and in vivo potentiated growth inhibition and apoptotic cell death caused by cisplatin, docetaxel, doxorubicin, gemcitabine, and CHOP (cyclophosphamide, doxorubicin, vincristine, prednisone) in lymphoma and cancers of the prostate, breast, pancreas, and lung (32, 89-91). By in vitro and in vivo studies, we also found that NF-kappaB activity was significantly increased by cisplatin, docetaxel, doxorubicin, and gemcitabine treatment and that the NF-kappaB inducing activity of these agents was completely abrogated by genistein pre-treatment (32, 89-91), suggesting that genistein pre-treatment inactivates NF-kappaB and leads to increased growth inhibition and apoptosis induced by these agents. Recently, we found that anti-tumor and antimetastatic activities of docetaxel could be enhanced by genistein through the regulation of osteoprotegerin/receptor activator of NF-kappaB (RANK)/RANK ligand/MMP-9 signaling in vitro and in vivo (92). These results suggest that genistein could potentiate the anti-tumor activities of chemotherapeutic agents in vitro and in vivo, resulting in more cancer cell killing and apoptotic cell death through regulation of NF-kappaB pathways in various cancers.

### 5.2. I3C and DIM

We and other investigators have found that I3C and DIM inhibit oncogenesis and cancer cell growth, and induce apoptosis through the inhibition of NF-kappaB activity in various cancer cells. Moreover, we and others have also found that combinations of I3C and cisplatin or tamoxifen cooperate to inhibit the growth of PC-3 prostate and MCF-7 breast cancer cells more effectively than either agent alone (42, 93). These results suggest that inhibition of NF-kappaB activity by I3C and DIM may contribute to the enhanced anti-tumor activity of chemotherapeutic agents.

# 5.3. Curcumin

It has been known that curcumin could enhance cancer therapeutic efficacy by inhibition of NF-kappaB. Curcumin has been found to significantly inhibit doxorubicin and paclitaxel induced NF-kappaB activation (85, 94), suggesting that curcumin could reduce drug resistance and sensitize cancer cell to chemotherapeutic agents. Curcumin and celecoxib also synergistically inhibited cell growth and induced apoptosis though the inhibition of NF-kappaB activity and the down-regulation of the expression of NF-kappaB-regulated gene products including COX-2, prostaglandin E2, and IL-8 in pancreatic cancers (13, 95). Curcumin also enhanced the anti-tumor activities of cisplatin, doxorubicin, and Taxol through inhibition of NF-kappaB in different type of cancers (96-98). A recent report showed that curcumin potentiated antitumor activity of gemcitabine in an orthotopic model of pancreatic cancer through suppression of proliferation, angiogenesis, and NF-kappaB-regulated gene products (99). These results suggest that curcumin is a potent agent in enhancing therapeutic efficacy of multiple chemotherapeutic agents in various cancers.

### 5.4. EGCG

It has been reported that EGCG and tamoxifen synergistically induced apoptosis and growth inhibition in MDA-MB-231 human breast cancer cells through inhibition of NF-kappaB (100). EGCG could also chemosensitize resistant tumor cells to doxorubicin through an increase in the accumulation of doxorubicin in the tumors of human carcinoma xenograft model (101). EGCG was also shown to inhibit activation of IKK, phosphorylation of IkappaBalpha, and activation of NF-kappaB, suggesting its novel effects on cancer therapy (54).

### 5.5. Resveratrol

Resveratrol shows its inhibitory effects on the activation of NF-kappaB, suggesting its role as a NF-kappaB inhibitor contributing to the chemosensitization of cancer cells to chemotherapeutic agents. It has been found that resveratrol from grapes could sensitize non-Hodgkin's lymphoma and multiple myeloma cells to paclitaxel-mediated apoptosis (102). Resveratrol could also sensitize TRAIL-induced apoptosis in pancreatic cancer cells (103). The chemotherapeutic strategy combined with resveratrol may be a novel approach to enhance the efficacy of chemotherapy for pancreatic cancers.

# 5.6. Dehydroxymethylepoxyquinomicin

Dehydroxymethylepoxyquinomicin (DHMEQ) is a synthetic NF-kappaB inhibitor. It has been found that DHMEQ could inhibit constitutively activated NF-kappaB and exhibit a synergistic inhibitory effect on cell growth with cisplatin in HA22T/VGH hepatic cancer cells (104). In HA22T/VGH cells, DHMEQ decreased the level of activated nuclear NF-kappaB in a dose-dependent manner and attenuated NF-kappaB activation induced by cisplatin. The combination of DHMEQ with cisplatin also decreased the levels of IL-6 and Bcl-xL mRNA, suggesting that DHMEO could inhibit the activation of NF-kappaB and the expression of NF-kappaB downstream target genes. In another study, the combination of DHMEO and IFNgamma synergistically inhibited cell proliferation with a pronounced attenuation of survivin expression, which is a known NF-kappaB downstream target gene (105). The combination of DHMEQ and TNF-alpha were also evaluated. NF-kappaB was activated by TNF-alpha; however, the administration of DHMEQ abrogated NFkappaB transcriptional activity (106). The addition of DHMEO to TNF-alpha markedly induced apoptosis with down-regulation of anti-apoptotic c-FLIP and survivin in PK-8 cells (106). These findings suggest that DHMEQ in combination with chemotherapeutic agents may be a promising strategy for the treatment of cancers.

### 5.7. COX-2 inhibitors

It has been known that NF-kappaB up-regulates COX-2 which activates EGFR signaling, suggesting that the inhibition of NF-kappaB, COX-2, and EGFR could be synergistic in killing cancer cells. We and others have found that celecoxib, a COX-2 inhibitor, combined with erlotinib (EGFR blocker) or curcumin synergistically potentiate the growth inhibitory and pro-apoptotic effects in cancer cells through the inhibition of NF-kappaB (82, 95),

suggesting that celecoxib could be a potent agent for combination treatment of cancers together with inhibition of NF-kappaB and EGFR. More recently, Mukhtar *et al.* have shown that lower doses of COX-2 inhibitors (NS-398 and celecoxib) combined with EGCG resulted in the enhanced cell growth inhibition, apoptosis induction, and NF-kappaB inhibition *in vitro* and *in vivo*, suggesting their potential role in cancer treatment (107).

### 5.8. Parthenolide

Parthenolide is a synthesized small molecule which suppresses NF-kappaB activation and sensitizes cancer cells to TNF-alpha-induced apoptosis in human cancer cells. The combination treatment with parthenolide and NSAID sulindac synergistically inhibited cell growth. induced apoptosis, increased levels of IkappaBalpha, and decreased NF-kappaB DNA binding activity (108), suggesting that a chemotherapeutic approach including NFkappaB inhibitors and NSAIDs could be a novel strategy for the treatment of cancer. In addition, parthenolide also enhanced ICI 182,780 (Faslodex; fulvestrant)-induced apoptosis and cell growth inhibition in antiestrogenresistant breast cancer cells (109), which have constitutively activated NF-kappaB, suggesting the protective role of NF-kappaB in antiestrogen-resistant breast cancer cells. The effect of parthenolide on the inhibition of NF-kappaB activation leads to growth inhibition of cancer cells.

### 5.9. Sulfasalazine

Sulfasalazine is commonly used as an anti inflammatory agent and is known as a potent inhibitor of NF-kappaB. Muerkoster et al. have investigated whether blockade of NF-kappaB activity with sulfasalazine could overcome NF-kappaB-induced chemoresistance in vivo in a mouse model of pancreatic cancer (110). They found that treatment with the chemotherapeutic agent etoposide alone moderately reduced tumor size (32-35% reduction), as compared to untreated tumors. Sulfasalazine alone only temporarily decreased the tumor size. However, sulfasalazine in combination with etoposide significantly reduced tumor size (80% reduction) in all experiments. TUNEL-staining showed higher numbers of apoptotic cells from the combination tumors Immunohistochemical staining of the activated p65 subunit showed that sulfasalazine treatment abolished the basal NFkappaB activity in tumor xenografts, suggesting that the anti-inflammatory drug sulfasalazine sensitizes cancer cells to chemotherapeutic agents by inhibition of NF-kappaB (110).

## 5.10. Proteasome inhibitor

Proteasome inhibitors PS-341 and MG-132 have been found to block intracellular degradation of IkappaBalpha proteins and, in turn, inhibit activation of NF-kappaB. It has been reported that PS-341 sensitized cancer cells to apoptosis induced by Taxol (111). Sensitization of cancer cells to Taxol could be mediated through the inhibition of IkappaBalpha degradation by PS-341. MG-132 has also been found to sensitize cancer cells to Fas-mediated apoptosis and radiation therapy (112, 113). These findings suggest that combination treatment with

proteasome inhibitors could enhance the efficacy of cancer therapy.

### 6. SUMMARY AND PERSPECTIVE

The *in vitro* and *in vivo* studies reviewed above all suggest that NF-kappaB is an important target for cancer prevention and treatment. The strategies to reduce the activity of NF-κB by natural or synthetic NF-κB inhibitors could become a novel and potent approach for enhancing the therapeutic efficacy of conventional chemotherapeutic agents in the treatment of human cancers. However, further in-depth mechanistic studies, *in vivo* animal experiments, and clinical trials are needed for investigating the effects of NF-kappaB inhibitors in combination with conventional cancer therapeutics.

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### 8. REFERENCES

- 1. Chen F., V.Castranova & X.Shi: New insights into the role of nuclear factor-kappaB in cell growth regulation. *Am J Pathol* 159,387-397 (2001)
- 2. Lenardo M.J. & D.Baltimore: NF-kappa B: a pleiotropic mediator of inducible and tissue-specific gene control. *Cell* 58,227-229 (1989)
- 3. Karin M.: Nuclear factor-kappaB in cancer development and progression. *Nature* 441,431-436 (2006)
- 4. Karin M.: NF-kappaB and cancer: mechanisms and targets. *Mol Carcinog* 45,355-361 (2006)
- 5. Ghosh G., G.van Duyne, S.Ghosh & P.B.Sigler: Structure of NF-kappa B p50 homodimer bound to a kappa B site. *Nature* 373,303-310 (1995)
- 6. Karin M. & F.R.Greten: NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol* 5,749-759 (2005)
- 7. Bowie A. & L.A.O'Neill: Oxidative stress and nuclear factor-kappaB activation: a reassessment of the evidence in the light of recent discoveries. *Biochem Pharmacol* 59,13-23 (2000)
- 8. Toledano M.B. & W.J.Leonard: Modulation of transcription factor NF-kappa B binding activity by oxidation-reduction *in vitro*. *Proc Natl Acad Sci U S A* 88,4328-4332 (1991)
- 9. Galaris D. & A.Evangelou: The role of oxidative stress in mechanisms of metal-induced carcinogenesis. *Crit Rev Oncol Hematol* 42.93-103 (2002)
- 10. Bharti A.C. & B.B.Aggarwal: Nuclear factor-kappa B and cancer: its role in prevention and therapy. *Biochem Pharmacol* 64,883-888 (2002)

- 11. Bargou R.C., F.Emmerich, D.Krappmann, K.Bommert, M.Y.Mapara, W.Arnold, H.D.Royer, E.Grinstein, A.Greiner, C.Scheidereit & B.Dorken: Constitutive nuclear factor-kappaB-RelA activation is required for proliferation and survival of Hodgkin's disease tumor cells. *J Clin Invest* 100,2961-2969 (1997)
- 12. Hideshima T., D.Chauhan, P.Richardson, C.Mitsiades, N.Mitsiades, T.Hayashi, N.Munshi, L.Dang, A.Castro, V.Palombella, J.Adams & K.C.Anderson: NF-kappa B as a therapeutic target in multiple myeloma. *J Biol Chem* 277,16639-16647 (2002)
- 13. Li L., B.B.Aggarwal, S.Shishodia, J.Abbruzzese & R.Kurzrock: Nuclear factor-kappaB and IkappaB kinase are constitutively active in human pancreatic cells, and their down-regulation by curcumin (diferuloylmethane) is associated with the suppression of proliferation and the induction of apoptosis. *Cancer* 101,2351-2362 (2004)
- 14. Wang W., J.L.Abbruzzese, D.B.Evans, L.Larry, K.R.Cleary & P.J.Chiao: The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. *Clin Cancer Res* 5,119-127 (1999)
- 15. Nakshatri H., P.Bhat-Nakshatri, D.A.Martin, R.J.Goulet, Jr. & G.W.Sledge, Jr.: Constitutive activation of NF-kappaB during progression of breast cancer to hormone-independent growth. *Mol Cell Biol* 17,3629-3639 (1997)
- 16. Shukla S., G.T.Maclennan, P.Fu, J.Patel, S.R.Marengo, M.I.Resnick & S.Gupta: Nuclear factor-kappaB/p65 (Rel A) is constitutively activated in human prostate adenocarcinoma and correlates with disease progression. *Neoplasia* 6,390-400 (2004)
- 17. Levidou G., P.Korkolopoulou, N.Nikiteas, N.Tzanakis, I.Thymara, A.A.Saetta, C.Tsigris, G.Rallis, K.Vlasis & E.Patsouris: Expression of nuclear factor kappaB in human gastric carcinoma: relationship with IkappaBa and prognostic significance. *Virchows Arch* (2007)
- 18. Allen C.T., J.L.Ricker, Z.Chen & W.C.Van: Role of activated nuclear factor-kappaB in the pathogenesis and therapy of squamous cell carcinoma of the head and neck. *Head Neck* (2007)
- 19. Ahmad N., S.Gupta & H.Mukhtar: Green tea polyphenol epigallocatechin-3-gallate differentially modulates nuclear factor kappaB in cancer cells versus normal cells. *Arch Biochem Biophys* 376,338-346 (2000)
- 20. Bharti A.C., N.Donato, S.Singh & B.B.Aggarwal: Curcumin (diferuloylmethane) down-regulates the constitutive activation of nuclear factor-kappa B and IkappaBalpha kinase in human multiple myeloma cells, leading to suppression of proliferation and induction of apoptosis. *Blood* 101,1053-1062 (2003)
- 21. Chinni S.R., Y.Li, S.Upadhyay, P.K.Koppolu & F.H.Sarkar: Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest and apoptosis in prostate cancer cells. *Oncogene* 20,2927-2936 (2001)
- 22. Li Y. & F.H.Sarkar: Inhibition of nuclear factor kappaB activation in PC3 cells by genistein is mediated via Akt signaling pathway. *Clin Cancer Res* 8,2369-2377 (2002)
- 23. Li Y., S.R.Chinni & F.H.Sarkar: Selective growth regulatory and pro-apoptotic effects of DIM is mediated by AKT and NF-kappaB pathways in prostate cancer cells. *Front Biosci* 10,236-243 (2005)

- 24. Kurahashi N., M.Iwasaki, S.Sasazuki, T.Otani, M.Inoue & S.Tsugane: Soy product and isoflavone consumption in relation to prostate cancer in Japanese men. *Cancer Epidemiol Biomarkers Prev* 16,538-545 (2007)
- 25. Zhang M., X.Xie, A.H.Lee & C.W.Binns: Soy and isoflavone intake are associated with reduced risk of ovarian cancer in southeast china. *Nutr Cancer* 49,125-130 (2004)
- 26. Wu A.H., M.C.Yu, C.C.Tseng, N.C.Twaddle & D.R.Doerge: Plasma isoflavone levels versus self-reported soy isoflavone levels in Asian-American women in Los Angeles County. *Carcinogenesis* 25,77-81 (2004)
- 27. Ruiz-Larrea M.B., A.R.Mohan, G.Paganga, N.J.Miller, G.P.Bolwell & C.A.Rice-Evans: Antioxidant activity of phytoestrogenic isoflavones. *Free Radic Res* 26,63-70 (1997)
- 28. Zhou Y. & A.S.Lee: Mechanism for the suppression of the mammalian stress response by genistein, an anticancer phytoestrogen from soy. *J Natl Cancer Inst* 90,381-388 (1998)
- 29. Kameoka S., P.Leavitt, C.Chang & S.M.Kuo: Expression of antioxidant proteins in human intestinal Caco-2 cells treated with dietary flavonoids. *Cancer Lett* 146,161-167 (1999)
- 30. Sierens J., J.A.Hartley, M.J.Campbell, A.J.Leathem & J.V.Woodside: *In vitro* isoflavone supplementation reduces hydrogen peroxide-induced DNA damage in sperm. *Teratog Carcinog Mutagen* 22,227-234 (2002)
- 31. Wei H., L.Wei, K.Frenkel, R.Bowen & S.Barnes: Inhibition of tumor promoter-induced hydrogen peroxide formation *in vitro* and *in vivo* by genistein. *Nutr Cancer* 20.1-12 (1993)
- 32. Banerjee S., Y.Zhang, S.Ali, M.Bhuiyan, Z.Wang, P.J.Chiao, P.A.Philip, J.Abbruzzese & F.H.Sarkar: Molecular evidence for increased antitumor activity of gemcitabine by genistein *in vitro* and *in vivo* using an orthotopic model of pancreatic cancer. *Cancer Res* 65,9064-9072 (2005)
- 33. Davis J.N., O.Kucuk & F.H.Sarkar: Genistein inhibits NF-kappa B activation in prostate cancer cells. *Nutr Cancer* 35,167-174 (1999)
- 34. Rahman K.W., Y.Li & F.H.Sarkar: Inactivation of Akt and NF-kappaB Play Important Roles During Indole-3-Carbinol-Induced Apoptosis in Breast Cancer Cells. *Nutr Cancer* 48,84-94 (2004)
- 35. Baxa D.M. & F.K. Yoshimura: Genistein reduces NF-kappa B in T lymphoma cells via a caspase-mediated cleavage of I kappa B alpha. *Biochem Pharmacol* 66,1009-1018 (2003)
- 36. Tabary O., S.Escotte, J.P.Couetil, D.Hubert, D.Dusser, E.Puchelle & J.Jacquot: Genistein inhibits constitutive and inducible NFkappaB activation and decreases IL-8 production by human cystic fibrosis bronchial gland cells. *Am J Pathol* 155,473-481 (1999)
- 37. Davis J.N., O.Kucuk, Z.Djuric & F.H.Sarkar: Soy isoflavone supplementation in healthy men prevents NF-kappa B activation by TNF-alpha in blood lymphocytes. *Free Radic Biol Med* 30,1293-1302 (2001)
- 38. Steinmetz K.A. & J.D.Potter: Vegetables, fruit, and cancer prevention: a review. *J Am Diet Assoc* 96,1027-1039 (1996)

- 39. Benabadji S.H., R.Wen, J.B.Zheng, X.C.Dong & S.G.Yuan: Anticarcinogenic and antioxidant activity of diindolylmethane derivatives. *Acta Pharmacol Sin* 25,666-671 (2004)
- 40. Nho C.W. & E.Jeffery: Crambene, a bioactive nitrile derived from glucosinolate hydrolysis, acts via the antioxidant response element to upregulate quinone reductase alone or synergistically with indole-3-carbinol. *Toxicol Appl Pharmacol* 198,40-48 (2004)
- 41. Firestone G.L. & L.F.Bjeldanes: Indole-3-carbinol and 3-3'-diindolylmethane antiproliferative signaling pathways control cell-cycle gene transcription in human breast cancer cells by regulating promoter-Sp1 transcription factor interactions. *J Nutr* 133,2448S-2455S (2003)
- 42. Sarkar F.H. & Y.Li: Indole-3-carbinol and prostate cancer. *J Nutr* 134,3493S-3498S (2004)
- 43. Banerjee M., L.M.Tripathi, V.M.Srivastava, A.Puri & R.Shukla: Modulation of inflammatory mediators by ibuprofen and curcumin treatment during chronic inflammation in rat. *Immunopharmacol Immunotoxicol* 25,213-224 (2003)
- 44. Miquel J., A.Bernd, J.M.Sempere, J.Diaz-Alperi & A.Ramirez: The curcuma antioxidants: pharmacological effects and prospects for future clinical use. A review. *Arch Gerontol Geriatr* 34,37-46 (2002)
- 45. Tonnesen H.H. & J.V.Greenhill: Studies on curcumin and curcuminoids. XXII: Curcumin as a reducing agent and as a radical scavenger. *Int J Pharm* 87,79-87 (1992)
- 46. Awasthi S., S.K.Srivatava, J.T.Piper, S.S.Singhal, M.Chaubey & Y.C.Awasthi: Curcumin protects against 4-hydroxy-2-trans-nonenal-induced cataract formation in rat lenses. *Am J Clin Nutr* 64,761-766 (1996)
- 47. Reddy A.C. & B.R.Lokesh: Effect of curcumin and eugenol on iron-induced hepatic toxicity in rats. *Toxicology* 107,39-45 (1996)
- 48. Shukla P.K., V.K.Khanna, M.Y.Khan & R.C.Srimal: Protective effect of curcumin against lead neurotoxicity in rat. *Hum Exp Toxicol* 22,653-658 (2003)
- 49. Chuang S.E., A.L.Cheng, J.K.Lin & M.L.Kuo: Inhibition by curcumin of diethylnitrosamine-induced hepatic hyperplasia, inflammation, cellular gene products and cell-cycle-related proteins in rats. *Food Chem Toxicol* 38,991-995 (2000)
- 50. Duvoix A., F.Morceau, S.Delhalle, M.Schmitz, M.Schnekenburger, M.M.Galteau, M.Dicato & M.Diederich: Induction of apoptosis by curcumin: mediation by glutathione S-transferase P1-1 inhibition. *Biochem Pharmacol* 66,1475-1483 (2003)
- 51. Aggarwal B.B., A.Kumar & A.C.Bharti: Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 23,363-398 (2003)
- 52. Radhakrishna P.G., A.S.Srivastava, T.I.Hassanein, D.P.Chauhan & E.Carrier: Induction of apoptosis in human lung cancer cells by curcumin. *Cancer Lett* 208,163-170 (2004)
- 53. Mukhtar H. & N.Ahmad: Green tea in chemoprevention of cancer. *Toxicol Sci* 52,111-117 (1999)
- 54. Afaq F., V.M.Adhami, N.Ahmad & H.Mukhtar: Inhibition of ultraviolet B-mediated activation of nuclear factor kappaB in normal human epidermal keratinocytes by green tea Constituent (-)-epigallocatechin-3-gallate. *Oncogene* 22,1035-1044 (2003)

- 55. Chen P.C., D.S.Wheeler, V.Malhotra, K.Odoms, A.G.Denenberg & H.R.Wong: A green tea-derived polyphenol, epigallocatechin-3-gallate, inhibits IkappaB kinase activation and IL-8 gene expression in respiratory epithelium. *Inflammation* 26,233-241 (2002)
- 56. Yang F., H.S.Oz, S.Barve, W.J.de Villiers, C.J.McClain & G.W.Varilek: The green tea polyphenol (-)-epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting I kappa B kinase activity in the intestinal epithelial cell line IEC-6. *Mol Pharmacol* 60,528-533 (2001)
- 57. Lambert J.D. & C.S.Yang: Mechanisms of cancer prevention by tea constituents. *J Nutr* 133,3262S-3267S (2003)
- 58. Hastak K., S.Gupta, N.Ahmad, M.K.Agarwal, M.L.Agarwal & H.Mukhtar: Role of p53 and NF-kappaB in epigallocatechin-3-gallate-induced apoptosis of LNCaP cells. *Oncogene* 22,4851-4859 (2003)
- 59. Jung Y.D. & L.M.Ellis: Inhibition of tumour invasion and angiogenesis by epigallocatechin gallate (EGCG), a major component of green tea. *Int J Exp Pathol* 82,309-316 (2001)
- 60. Kim H.S., M.H.Kim, M.Jeong, Y.S.Hwang, S.H.Lim, B.A.Shin, B.W.Ahn & Y.D.Jung: EGCG blocks tumor promoter-induced MMP-9 expression via suppression of MAPK and AP-1 activation in human gastric AGS cells. *Anticancer Res* 24,747-753 (2004)
- 61. Fremont L.: Biological effects of resveratrol. *Life Sci* 66,663-673 (2000)
- 62. Dong Z.: Molecular mechanism of the chemopreventive effect of resveratrol. *Mutat Res* 523-524,145-150 (2003)
- 63. Sparrow J.R., H.R.Vollmer-Snarr, J.Zhou, Y.P.Jang, S.Jockusch, Y.Itagaki & K.Nakanishi: A2E-epoxides damage DNA in retinal pigment epithelial cells. Vitamin E and other antioxidants inhibit A2E-epoxide formation. *J Biol Chem* 278,18207-18213 (2003)
- 64. Delmas D., C.Rebe, S.Lacour, R.Filomenko, A.Athias, P.Gambert, M.Cherkaoui-Malki, B.Jannin, L.Dubrez-Daloz, N.Latruffe & E.Solary: Resveratrol-induced apoptosis is associated with Fas redistribution in the rafts and the formation of a death-inducing signaling complex in colon cancer cells. *J Biol Chem* 278,41482-41490 (2003)
- 65. Laux M.T., M.Aregullin, J.P.Berry, J.A.Flanders & E.Rodriguez: Identification of a p53-dependent pathway in the induction of apoptosis of human breast cancer cells by the natural product, resveratrol. *J Altern Complement Med* 10,235-239 (2004)
- 66. Scarlatti F., G.Sala, G.Somenzi, P.Signorelli, N.Sacchi & R.Ghidoni: Resveratrol induces growth inhibition and apoptosis in metastatic breast cancer cells via de novo ceramide signaling. *FASEB J* 17,2339-2341 (2003)
- 67. Estrov Z., S.Shishodia, S.Faderl, D.Harris, Q.Van, H.M.Kantarjian, M.Talpaz & B.B.Aggarwal: Resveratrol blocks interleukin-1beta-induced activation of the nuclear transcription factor NF-kappaB, inhibits proliferation, causes S-phase arrest, and induces apoptosis of acute myeloid leukemia cells. *Blood* 102,987-995 (2003)
- 68. Heber D. & Q.Y.Lu: Overview of mechanisms of action of lycopene. *Exp Biol Med (Maywood )* 227,920-923 (2002)

- 69. Di Mascio P., S.Kaiser & H.Sies: Lycopene as the most efficient biological carotenoid singlet oxygen quencher. *Arch Biochem Biophys* 274,532-538 (1989)
- 70. Giovannucci E., E.B.Rimm, Y.Liu, M.J.Stampfer & W.C.Willett: A prospective study of tomato products, lycopene, and prostate cancer risk. *J Natl Cancer Inst* 94,391-398 (2002)
- 71. Lu Q.Y., J.C.Hung, D.Heber, V.L.Go, V.E.Reuter, C.Cordon-Cardo, H.I.Scher, J.R.Marshall & Z.F.Zhang: Inverse associations between plasma lycopene and other carotenoids and prostate cancer. *Cancer Epidemiol Biomarkers Prev* 10,749-756 (2001)
- 72. Kim L., A.V.Rao & L.G.Rao: Effect of lycopene on prostate LNCaP cancer cells in culture. *J Med Food* 5,181-187 (2002)
- 73. Nahum A., K.Hirsch, M.Danilenko, C.K.Watts, O.W.Prall, J.Levy & Y.Sharoni: Lycopene inhibition of cell cycle progression in breast and endometrial cancer cells is associated with reduction in cyclin D levels and retention of p27(Kip1) in the cyclin E-cdk2 complexes. *Oncogene* 20,3428-3436 (2001)
- 74. Kim G.Y., J.H.Kim, S.C.Ahn, H.J.Lee, D.O.Moon, C.M.Lee & Y.M.Park: Lycopene suppresses the lipopolysaccharide-induced phenotypic and functional maturation of murine dendritic cells through inhibition of mitogen-activated protein kinases and nuclear factor-kappaB. *Immunology* 113,203-211 (2004)
- 75. Kucuk O., F.H.Sarkar, Z.Djuric, W.Sakr, M.N.Pollak, F.Khachik, M.Banerjee, J.S.Bertram & D.P.Wood, Jr.: Effects of lycopene supplementation in patients with localized prostate cancer. *Exp Biol Med (Maywood)* 227,881-885 (2002)
- 76. Calfee-Mason K.G., B.T.Spear & H.P.Glauert: Vitamin E inhibits hepatic NF-kappaB activation in rats administered the hepatic tumor promoter, phenobarbital. *J Nutr* 132,3178-3185 (2002)
- 77. Sokoloski J.A., W.F.Hodnick, S.T.Mayne, C.Cinquina, C.S.Kim & A.C.Sartorelli: Induction of the differentiation of HL-60 promyelocytic leukemia cells by vitamin E and other antioxidants in combination with low levels of vitamin D3: possible relationship to NF-kappaB. *Leukemia* 11,1546-1553 (1997)
- 78. Bowie A.G. & L.A.O'Neill: Vitamin C inhibits NF-kappa B activation by TNF via the activation of p38 mitogen-activated protein kinase. *J Immunol* 165,7180-7188 (2000)
- 79. Carcamo J.M., A.Pedraza, O.Borquez-Ojeda & D.W.Golde: Vitamin C suppresses TNF alpha-induced NF kappa B activation by inhibiting I kappa B alpha phosphorylation. *Biochemistry* 41,12995-13002 (2002)
- 80. Sclabas G.M., T.Uwagawa, C.Schmidt, K.R.Hess, D.B.Evans, J.L.Abbruzzese & P.J.Chiao: Nuclear factor kappa B activation is a potential target for preventing pancreatic carcinoma by aspirin. *Cancer* 103,2485-2490 (2005)
- 81. Wong B.C., X.Jiang, X.M.Fan, M.C.Lin, S.H.Jiang, S.K.Lam & H.F.Kung: Suppression of RelA/p65 nuclear translocation independent of IkappaB-alpha degradation by cyclooxygenase-2 inhibitor in gastric cancer. *Oncogene* 22,1189-1197 (2003)
- 82. Ali S., B.F.El-Rayes, F.H.Sarkar & P.A.Philip: Simultaneous targeting of the epidermal growth factor

- receptor and cyclooxygenase-2 pathways for pancreatic cancer therapy. *Mol Cancer Ther* 4,1943-1951 (2005)
- 83. El-Rayes B.F., S.Ali, F.H.Sarkar & P.A.Philip: Cyclooxygenase-2-dependent and -independent effects of celecoxib in pancreatic cancer cell lines. *Mol Cancer Ther* 3,1421-1426 (2004)
- 84. Bian X., L.M.McAllister-Lucas, F.Shao, K.R.Schumacher, Z.Feng, A.G.Porter, V.P.Castle & A.W.Opipari, Jr.: NF-kappa B activation mediates doxorubicin-induced cell death in N-type neuroblastoma cells. *J Biol Chem* 276,48921-48929 (2001)
- 85. Chuang S.E., P.Y.Yeh, Y.S.Lu, G.M.Lai, C.M.Liao, M.Gao & A.L.Cheng: Basal levels and patterns of anticancer drug-induced activation of nuclear factor-kappaB (NF-kappaB), and its attenuation by tamoxifen, dexamethasone, and curcumin in carcinoma cells. *Biochem Pharmacol* 63,1709-1716 (2002)
- 86. Yeh P.Y., S.E.Chuang, K.H.Yeh, Y.C.Song, C.K.Ea & A.L.Cheng: Increase of the resistance of human cervical carcinoma cells to cisplatin by inhibition of the MEK to ERK signaling pathway partly via enhancement of anticancer drug-induced NF kappa B activation. *Biochem Pharmacol* 63,1423-1430 (2002)
- 87. Arlt A., J.Vorndamm, S.Muerkoster, H.Yu, W.E.Schmidt, U.R.Folsch & H.Schafer: Autocrine production of interleukin 1beta confers constitutive nuclear factor kappaB activity and chemoresistance in pancreatic carcinoma cell lines. *Cancer Res* 62,910-916 (2002)
- 88. Muerkoster S., A.Arlt, B.Sipos, M.Witt, M.Grossmann, G.Kloppel, H.Kalthoff, U.R.Folsch & H.Schafer: Increased expression of the E3-ubiquitin ligase receptor subunit betaTRCP1 relates to constitutive nuclear factor-kappaB activation and chemoresistance in pancreatic carcinoma cells. *Cancer Res* 65,1316-1324 (2005)
- K.L.Ellis, 89 Li Y., S.Ali, B.F.El-Rayes, O.Kucuk, P.A.Philip A.Nedeljkovic-Kurepa, Apoptosis-inducing effect F.H.Sarkar: of chemotherapeutic agents is potentiated by isoflavone genistein, a natural inhibitor of NF-kappaB in BxPC-3 pancreatic cancer cell line. Pancreas 28,e90e95 (2004)
- 90. Li Y., F.Ahmed, S.Ali, P.A.Philip, O.Kucuk & F.H.Sarkar: Inactivation of nuclear factor kappaB by soy isoflavone genistein contributes to increased apoptosis induced by chemotherapeutic agents in human cancer cells. *Cancer Res* 65,6934-6942 (2005)
- 91. Mohammad R.M., A.Al-Katib, A.Aboukameel, D.R.Doerge, F.Sarkar & O.Kucuk: Genistein sensitizes diffuse large cell lymphoma to CHOP (cyclophosphamide, doxorubicin, vincristine, prednisone) chemotherapy. *Mol Cancer Ther* 2,1361-1368 (2003)
- 92. Li Y., O.Kucuk, M.Hussain, J.Abrams, M.L.Cher & F.H.Sarkar: Antitumor and antimetastatic activities of docetaxel are enhanced by genistein through regulation of osteoprotegerin/receptor activator of nuclear factor-kappaB (RANK)/RANK ligand/MMP-9 signaling in prostate cancer. *Cancer Res* 66,4816-4825 (2006)
- 93. Cover C.M., S.J.Hsieh, E.J.Cram, C.Hong, J.E.Riby, L.F.Bjeldanes & G.L.Firestone: Indole-3-carbinol and

- tamoxifen cooperate to arrest the cell cycle of MCF-7 human breast cancer cells. *Cancer Res* 59,1244-1251 (1999)
- 94. Aggarwal B.B., S.Shishodia, Y.Takada, S.Banerjee, R.A.Newman, C.E.Bueso-Ramos & J.E.Price: Curcumin suppresses the paclitaxel-induced nuclear factor-kappaB pathway in breast cancer cells and inhibits lung metastasis of human breast cancer in nude mice. *Clin Cancer Res* 11,7490-7498 (2005)
- 95. Lev-Ari S., H.Zinger, D.Kazanov, D.Yona, R.Ben-Yosef, A.Starr, A.Figer & N.Arber: Curcumin synergistically potentiates the growth inhibitory and proapoptotic effects of celecoxib in pancreatic adenocarcinoma cells. *Biomed Pharmacother* 59 Suppl 2,S276-S280 (2005) 96. Bava S.V., V.T.Puliappadamba, A.Deepti, A.Nair, D.Karunagaran & R.J.Anto: Sensitization of taxol-induced apoptosis by curcumin involves down-regulation of nuclear factor-kappaB and the serine/threonine kinase Akt and is independent of tubulin polymerization. *J Biol Chem* 280,6301-6308 (2005)
- 97. Notarbartolo M., P.Poma, D.Perri, L.Dusonchet, M.Cervello & N.D'Alessandro: Antitumor effects of curcumin, alone or in combination with cisplatin or doxorubicin, on human hepatic cancer cells. Analysis of their possible relationship to changes in NF-kB activation levels and in IAP gene expression. *Cancer Lett* 224,53-65 (2005)
- 98. Venkatraman M., R.J.Anto, A.Nair, M.Varghese & D.Karunagaran: Biological and chemical inhibitors of NF-kappaB sensitize SiHa cells to cisplatin-induced apoptosis. *Mol Carcinog* 44,51-59 (2005)
- 99. Kunnumakkara A.B., S.Guha, S.Krishnan, P.Diagaradjane, J.Gelovani & B.B.Aggarwal: Curcumin Potentiates Antitumor Activity of Gemcitabine in an Orthotopic Model of Pancreatic Cancer through Suppression of Proliferation, Angiogenesis, and Inhibition of Nuclear Factor-{kappa}B-Regulated Gene Products. Cancer Res 67,3853-3861 (2007)
- 100. Chisholm K., B.J.Bray & R.J.Rosengren: Tamoxifen and epigallocatechin gallate are synergistically cytotoxic to MDA-MB-231 human breast cancer cells. *Anticancer Drugs* 15,889-897 (2004)
- 101. Zhang Q., D.Wei & J.Liu: *In vivo* reversal of doxorubicin resistance by (-)-epigallocatechin gallate in a solid human carcinoma xenograft. *Cancer Lett* 208,179-186 (2004)
- 102. Jazirehi A.R. & B.Bonavida: Resveratrol modifies the expression of apoptotic regulatory proteins and sensitizes non-Hodgkin's lymphoma and multiple myeloma cell lines to paclitaxel-induced apoptosis. *Mol Cancer Ther* 3,71-84 (2004)
- 103. Fulda S. & K.M.Debatin: Sensitization for tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis by the chemopreventive agent resveratrol. *Cancer Res* 64,337-346 (2004)
- 104. Poma P., M.Notarbartolo, M.Labbozzetta, R.Sanguedolce, A.Alaimo, V.Carina, A.Maurici, A.Cusimano, M.Cervello & N.D'Alessandro: Antitumor effects of the novel NF-kappaB inhibitor dehydroxymethylepoxyquinomicin on human hepatic cancer cells: analysis of synergy with cisplatin and of possible correlation with

- inhibition of pro-survival genes and IL-6 production. *Int J Oncol* 28,923-930 (2006)
- 105. Sato A., M.Oya, K.Ito, R.Mizuno, Y.Horiguchi, K.Umezawa, M.Hayakawa & M.Murai: Survivin associates with cell proliferation in renal cancer cells: regulation of survivin expression by insulin-like growth factor-1, interferon-gamma and a novel NF-kappaB inhibitor. *Int J Oncol* 28,841-846 (2006)
- 106. Matsumoto G., M.Muta, K.Umezawa, T.Suzuki, K.Misumi, K.Tsuruta, A.Okamoto & M.Toi: Enhancement of the caspase-independent apoptotic sensitivity of pancreatic cancer cells by DHMEQ, an NF-kappaB inhibitor. *Int J Oncol* 27,1247-1255 (2005)
- 107. Adhami V.M., A.Malik, N.Zaman, S.Sarfaraz, I.A.Siddiqui, D.N.Syed, F.Afaq, F.S.Pasha, M.Saleem & H.Mukhtar: Combined Inhibitory Effects of Green Tea Polyphenols and Selective Cyclooxygenase-2 Inhibitors on the Growth of Human Prostate Cancer Cells Both *In vitro* and *In vivo. Clin Cancer Res* 13,1611-1619 (2007)
- 108. Yip-Schneider M.T., H.Nakshatri, C.J.Sweeney, M.S.Marshall, E.A.Wiebke & C.M.Schmidt: Parthenolide and sulindac cooperate to mediate growth suppression and inhibit the nuclear factor-kappa B pathway in pancreatic carcinoma cells. *Mol Cancer Ther* 4,587-594 (2005)
- 109. Riggins R.B., A.Zwart, R.Nehra & R.Clarke: The nuclear factor kappa B inhibitor parthenolide restores ICI 182,780 (Faslodex; fulvestrant)-induced apoptosis in antiestrogen-resistant breast cancer cells. *Mol Cancer Ther* 4,33-41 (2005)
- 110. Muerkoster S., A.Arlt, M.Witt, A.Gehrz, S.Haye, C.March, F.Grohmann, K.Wegehenkel, H.Kalthoff, U.R.Folsch & H.Schafer: Usage of the NF-kappaB inhibitor sulfasalazine as sensitizing agent in combined chemotherapy of pancreatic cancer. *Int J Cancer* 104,469-476 (2003)
- 111. Dong Q.G., G.M.Sclabas, S.Fujioka, C.Schmidt, B.Peng, T.Wu, M.S.Tsao, D.B.Evans, J.L.Abbruzzese, T.J.McDonnell & P.J.Chiao: The function of multiple IkappaB: NF-kappaB complexes in the resistance of cancer cells to Taxol-induced apoptosis. *Oncogene* 21,6510-6519 (2002)
- 112. Meli M., N.D'Alessandro, M.Tolomeo, L.Rausa, M.Notarbartolo & L.Dusonchet: NF-kappaB inhibition restores sensitivity to Fas-mediated apoptosis in lymphoma cell lines. *Ann N Y Acad Sci* 1010,232-236 (2003)
- 113. Munshi A., J.F.Kurland, T.Nishikawa, P.J.Chiao, M.Andreeff & R.E.Meyn: Inhibition of constitutively activated nuclear factor-kappaB radiosensitizes human melanoma cells. *Mol Cancer Ther* 3,985-992 (2004)
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