

Transcription factors in the development of inner ear hair cells

Shuna Li¹, Wei Qian¹, Guochang Jiang¹, Yongming Ma¹

¹Department of Otolaryngology and Head-Neck Surgery, Zhenjiang First People's Hospital, The Affiliated People's Hospital of Jiangsu University, Zhenjiang, Jiangsu, China

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Transcription factors in inner ear development
4. Transcription factors in hair cells development
 - 4.1. Math1 in hair cells development
 - 4.2. Sox2 in hair cells development
 - 4.3. Atoh1 in hair cells development
 - 4.4. Gata3 in hair cells development
5. Conclusions
6. Acknowledgements
7. References

1. ABSTRACT

Inner ear hair cells are the sensory receptors that detect and convert sound vibrations and head movements into neural signals. However, in humans, these cells are unable to regenerate if they are damaged or lost. Over the past decade, there has been an exponential increase in interest and progress in understanding of the development of the inner ear and of hair cells, aiming to gain insights into hair cell repair or even regeneration. In hair cell development, various transcription factors have been found to be involved in the processes of hair cell proliferation, differentiation and survival. Among these transcription factors, Math1, Gata3, Sox2 and Atoh1 have been highlighted for their crucial role in the fate of hair cells. In this article, we will summarize the current understanding of the role of transcription factors in hair cell development, focusing on the role and possible mechanisms of Math1, Gata3, Sox2 and Atoh1.

2. INTRODUCTION

The inner ear is a complex structure with functions of sound detection and balance. Impaired development of the inner ear leads to hearing or balance disorders. A transcription factor is a key protein in the transformation of genetic information, binding to DNA and then controlling its transcription to messenger RNA, and thus participating in various important cellular processes (1). Several transcription factors participate in regulating the development of the inner ear, even from the embryonic stage (2-7). Among the various types of cells in the inner ear, hair cells are the sensory receptors that detect and convert sound vibrations and head movements into neural signals (8,9). However, hair

cells lack the capacity to regenerate in humans if they are damaged or lost (10). Reviewing the key transcription factors that participate in the survival, differentiation or proliferation of hair cells may provide insights into hair cell regeneration.

3. TRANSCRIPTION FACTORS IN INNER EAR DEVELOPMENT

The inner ear is derived from the otic placode, an epithelial structure which forms part of the surface non-neural ectoderm adjacent to the caudal part of the hindbrain (11,12). In the otic placode, transcription factors Fgf3, Fgf8 and Pax2 are essential for its specification (13,14). Moreover, Wnt signaling can specify otic placode by upregulating otic genes or by upregulating the Notch signaling pathway, which then provides positive feedback to Wnt signaling (15). Another example is that Sox9, the high mobility group (HMG)-domain-containing transcription factor, controls adhesive properties and invagination of placodal cells in a cell-autonomous manner, and maintains the progenitors in the otic epithelium (16,17). When these factors were abnormal, the inner ear may develop abnormally during the early stages.

In different parts of the inner ear, different transcription factors are involved in development. In the semicircular canals, the nuclear receptors Nor-1 and Gbx2 are essential for proliferation or differentiation (18,19), whereas in the vestibulum, BETA2/NeuroD1, Fkh10 and Pax2 transcription factors are involved in its development (20-22). Pax2 not only

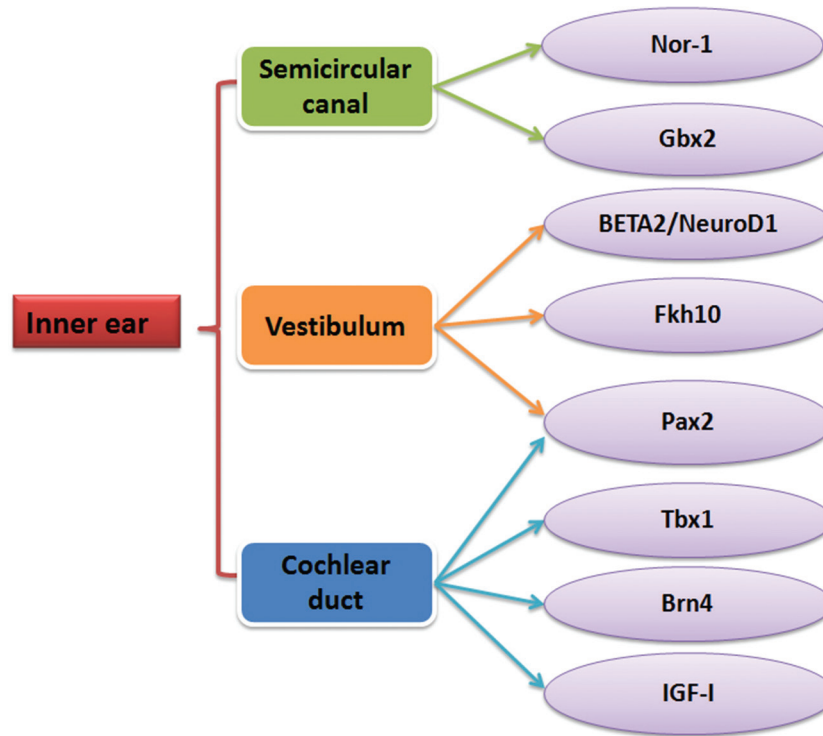


Figure 1. Transcription factors in different aspects of inner ear development. Nor-1 and Gbx2 are essential for semicircular canal development. Meanwhile BETA2/NeuroD1, Fkh10 and Pax2 transcription factors are involved in development of the vestibulum. Pax2 not only participates in development of the vestibulum, but also in cochlear duct development. The development of the cochlear duct also involves factors such as Tbx1, Brn4 and insulin-like growth factor (IGF)-I.

participates in development of the vestibulum, but is also essential in regulating differentiation during cochlear duct development (23,24). In contrast, development of the cochlear duct involves factors such as Tbx1, Brn4 and insulin-like growth factor (IGF)-I (25,26) (Figure 1). IGF-I also participates in the survival and differentiation of sensory cells and otic neuroblasts (26,27). Moreover, sensory cells can also be regulated by Rbpj, Hedgehog, Foxi2 and Foxi3 (28-30). Thus, the survival, differentiation and proliferation of cells in the inner ear involve several transcription factors. In addition to its role in sensory cells, STOX1 regulates proliferation of epithelial cells via the AKT pathway (31), and Tfp2a promotes specification and maturation of neurons (32). Therefore, transcription factors can participate in development of the inner ear by regulating survival, differentiation and proliferation of different types of cells.

4. TRANSCRIPTION FACTORS IN HAIR CELL DEVELOPMENT

In the development of hair cells, the POU-domain factor Brn-3c is essential for hair cells to acquire auditory and vestibular functions during the late embryonic and early postnatal period (33). On the other hand, the development of hair cells can be inhibited by the transcription factor RY-box containing gene 2, whereas

the motor protein myosin II can regulate extension of the organ of Corti and the alignment of hair cells (34).

Furthermore, several transcription factors regulate the differentiation of hair cells, such as repressor element-1 silencing transcription factor, six1 (35,36). Among the factors involved in cell differentiation, Pax2 is an early marker of hair cells (37). As an architectural transcription factor, Hmga2 is also tissue-specifically expressed in early development of the cochlea, and may have a dual role in regulating hair cell differentiation and maintenance that is predominantly expressed in embryonic hair cells, but is downregulated in the cochlear epithelium of postnatal ears (38). On the other hand, the survival of hair cells can be promoted by the anti-apoptotic factor z-Val-Ala-Asp-fluoromethylketone in a mouse model of deafness (39). Similarly, the LIM-homeodomain transcription factor Isl1 was also found to play a role of maintaining hair cell survival (40,41). Interestingly, the zinc finger transcription factor Gfi1, a target of the Pou4f3 deafness gene, is essential not only for differentiation of hair cells but also for their survival (42,43), suggesting a crucial role of Gfi1 in hair cell development and degeneration. In addition to Gfi1, Nr2f2 is another target of Pou4f3 and is also related to hair cell development and survival (44).

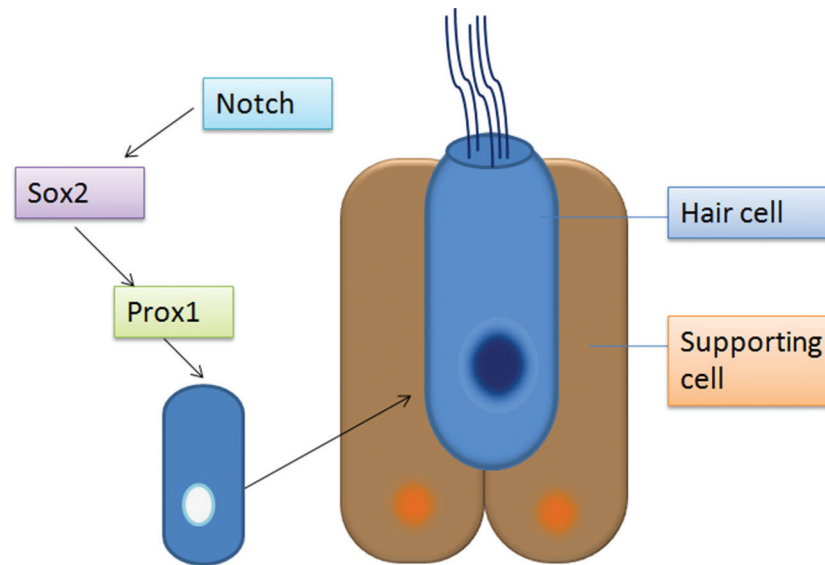


Figure 2. Sox2 in hair cell differentiation. Sox2 expression is positively regulated by Notch signaling and then targets Prox1. Decreased expression of Sox2 leads to precocious hair cell differentiation and over- production of hair cells in the inner ear.

These findings illustrate the crucial role transcription factors play during hair cell development, from differentiation to survival. Among the various transcription factors, Math1, Gata3, Sox2 and atonal homolog 1 (Atoh1) have attracted much attention for their crucial roles in hair cell development. Here, we will discuss recent findings on their role and possible mechanisms in hair cell development.

4.1. Math1 in hair cell development

Math1, the basic helix-loop-helix transcription factor, plays an essential role in controlling the generation of all elements of the proprioceptive pathway, including hair cell development (45,46). It regulates formation of the sensory epithelia by initiating both inductive and inhibitory signaling in the development of the cochlea (47). Furthermore, Math1 is a key factor in inducing differentiation of hair cells from embryonic stem cells (48). Recently, a study on neonatal mice showed that Math1 can induce hair cell-like cells to proliferate and differentiate in the lesser epithelial ridge (49). Thus, Math1 may be crucial in promoting the proliferation and differentiation of hair cells even from the embryonic stage. Although the upstream regulator of Math1 is known to be Sox2 (50), better understanding of the upstream and downstream mediators of Math1 is needed to further elucidate the mechanism of Math1 regulation in hair cell development.

4.2. Sox2 in hair cell development

Sox2 is a member of the Sox family of transcription factors and is similar to Sox9 in that it contains an HMG-domain. It is essential in maintenance of embryonic and neural stem cells and is responsible for progenitor self-renewal and commitment. The impaired function of Sox2 leads to defects of ear sensory epithelia. As mentioned

above, Sox2 has been found to act upstream of Math1 in regulating the development of sensory organs in the inner ear (50). In a study of chick inner ear, Sox2 protein was found to be expressed in a spatially- and temporally-restricted manner throughout ear development, and is transiently expressed in embryonic hair cells (51). In development of the inner ear sensory cell, Sox2 can specify sensory progenitors and drive development of the sensory cell progenitors in the inner ear (52). Furthermore, Dabdoub and colleagues identified diverse roles of Sox2 in the development, specification, and maintenance of sensory cells (53). They observed that mutations in Sox2 could lead to sensorineural hearing loss. Meanwhile, decreased expression of Sox2 can cause precocious hair cell differentiation and an over-production of inner hair cells. Sox2 expression was positively regulated by Notch signaling, and subsequently targeted Prox1, a homeobox transcription factor (53). In addition, a surprising finding was that Sox2 could trigger an incoherent feed-forward loop in hair cell differentiation. It directly activates Atoh1 through its transcriptional activator function, whereas it also promotes the expression of Atoh1 negative regulators (54). This finding suggests a complex role of Sox2 in hair cell differentiation, which may be a key factor connecting different signaling pathways during the process of cell differentiation.

4.3. Atoh1 in hair cell development

Atoh1 is a member of the basic helix-loop-helix family of transcription factors. Atoh1 is expressed in progenitors that may differentiate into hair cells and participates in hair cell development (55,54). Deletion of Atoh1 leads to impaired differentiation of hair cells and loss of most of the organ of Corti, suggesting the essential role of Atoh1 in inner ear hair cell formation (56).

Atoh1 mutation, namely Atoh1(trhl), leads to hair cell loss in the inner ear (57). Atoh1 is also essential for development of hair cell mechanotransduction, viability, and maintenance such that reduction or deletion of Atoh1 leads to progressive loss of almost all the inner ear hair cells, as well as the majority of the outer hair cells (58). Lin and colleagues reported an exciting finding that Atoh1 can induce mesenchymal stem cells to transform into hair cell-like cells when co-cultured with spiral ganglion neurons (59), which may provide insights into hair cell regeneration and deafness therapy.

4.4. Gata3 in hair cell development

Gata3 belongs to the Gata family of transcription factors, which usually is essential in the differentiation of epithelial cells and T cells (60,61). In inner ear development, Gata3 plays a crucial role and haploinsufficiency leads to hearing loss even in early postnatal development, which is maintained through to adulthood (62). Milo and colleagues subsequently found that the function of Gata3 during inner ear development may be associated with IGF signaling and AKT signaling. GATA3 deficiency leads to decreased expression of IGF1, IGF2, and several IGF-binding proteins as well as the serine-threonine kinase Akt2/PKB β , but to increased expression of Akt1/PKB α protein (63) (Figure 2). In cochlear development, Gata3 is thought to be essential for neurosensory specification and differentiation. Gata3 deficiency leads to impaired differentiation of hair cells, with decreased hair cell differentiation in the mouse cochlea (64,65). However, the way in which Gata3 participates in hair cell differentiation is largely unknown. Further study on this mechanism will provide a better understanding of hair cell differentiation.

5. CONCLUSIONS

Hair cells are essential for the functions of hearing and balance in the inner ear. The past decade has seen exponential increases in interest and progress in the field of hair cell development. In this field, various transcription factors have been found to be involved in the processes of hair cell proliferation, differentiation and survival. As we have discussed, Math1, Gata3, Sox2 and Atoh1 have been better investigated than other transcription factors. These factors are crucial in determining the fate of hair cells. Atoh1 can even induce hair cells to differentiate from mesenchymal stem cells. These findings provide insight into hair cell regeneration. The challenge in future is how to intervene to regulate or even control the fate of hair cells. The future is exciting in terms of the benefits for deafness patients that may emerge as a result of hair cell regeneration.

6. ACKNOWLEDGEMENTS

This work was supported by the Natural Science Foundation of Jiangsu Province for Youths (BK2012280).

7. REFERENCES

1. Latchman DS: Transcription factors: an overview. *Int J Biochem Cell Biol* 29 (12),1305-12 (1997)
DOI: 10.1016/S1357-2725(97)00085-X
2. Bagheri-Fam S, Barrionuevo F, Dohrmann U, Günther T, Schüle R, Kemler R, *et al*: Long-range upstream and downstream enhancers control distinct subsets of the complex spatiotemporal Sox9 expression pattern. *Dev Biol* 291 (2),382-97 (2006)
DOI: 10.1016/j.ydbio.2005.11.013
3. Sajjan SA, Rubenstein JL, Warchol ME, Lovett M: Identification of direct downstream targets of Dlx5 during early inner ear development. *Hum Mol Genet* 20 (7),1262-73 (2011)
DOI: 10.1093/hmg/ddq567
4. Trowe MO, Maier H, Schweizer M, Kispert A: Deafness in mice lacking the T-box transcription factor Tbx18 in otic fibrocytes. *Development* 135 (9),1725-34 (2008)
DOI: 10.1242/dev.014043
5. Monks DC, Morrow BE: Identification of putative retinoic acid target genes downstream of mesenchymal Tbx1 during inner ear development. *Dev Dyn* 241 (3),563-73 (2012)
DOI: 10.1002/dvdy.23731
6. Engelen E, Akinci U, Bryne JC, Hou J, Gontan C, Moen M, *et al*: Sox2 cooperates with Chd7 to regulate genes that are mutated in human syndromes. *Nat Genet* 43 (6),607-11 (2011)
DOI: 10.1038/ng.825
7. Pilipenko VV, Reece A, Choo DI, Greinwald JH Jr: Genomic organization and expression analysis of the murine Fam3c gene. *Gene* 335,159-68 (2004)
DOI: 10.1016/j.gene.2004.03.026
8. Hudspeth AJ, Choe Y, Mehta AD, Martin P: Putting ion channels to work: mechanoelectrical transduction, adaptation, and amplification by hair cells. *Proc Natl Acad Sci U S A* 97 (22),11765-72 (2000)
DOI: 10.1073/pnas.97.22.11765
9. Hackney CM, Furness DN: The composition and role of cross links in mechanoelectrical transduction in vertebrate sensory hair cells. *J Cell Sci* 126 (Pt 8),1721-31 (2013)
DOI: 10.1242/jcs.106120
10. Rubel EW, Furrer SA, Stone JS: A brief

- history of hair cell regeneration research and speculations on the future. *Hear Res* 297,42-51 (2013)
DOI: 10.1016/j.heares.2012.12.014
11. Barald KF, Kelley MW: From placode to polarization: new tunes in inner ear development. *Development* 131 (17),4119-30 (2004)
DOI: 10.1242/dev.01339
12. Groves AK, Fekete DM: Shaping sound in space: the regulation of inner ear patterning. *Development* 139 (2),245-57 (2012)
DOI: 10.1242/dev.067074
13. Liu D, Chu H, Maves L, Yan YL, Morcos PA, Postlethwait JH, Westerfield M: Fgf3 and Fgf8 dependent and independent transcription factors are required for otic placode specification. *Development* 130 (10),2213-24 (2003)
DOI: 10.1242/dev.00445
14. Freter S, Muta Y, O'Neill P, Vassilev VS, Kuraku S, Ladher RK: Pax2 modulates proliferation during specification of the otic and epibranchial placodes. *Dev Dyn* 241 (11),1716-28 (2012)
DOI: 10.1002/dvdy.23856
15. Jayasena CS, Ohshima T, Segil N, Groves AK: Notch signaling augments the canonical Wnt pathway to specify the size of the otic placode. *Development* 135 (13),2251-61 (2008)
DOI: 10.1242/dev.017905
16. Barrionuevo F, Naumann A, Bagheri-Fam S, Speth V, Taketo MM, Scherer G, Neubüser A: Sox9 is required for invagination of the otic placode in mice. *Dev Biol* 317 (1),213-24 (2008)
DOI: 10.1016/j.ydbio.2008.02.011
17. Park BY, Saint-Jeannet JP: Long-term consequences of Sox9 depletion on inner ear development. *Dev Dyn* 239 (4),1102-12 (2010)
DOI: 10.1002/dvdy.22259
18. Ponnio T, Burton Q, Pereira FA, Wu DK, Conneely OM: The nuclear receptor Nor-1 is essential for proliferation of the semicircular canals of the mouse inner ear. *Mol Cell Biol* 22 (3),935-45 (2002)
DOI: 10.1128/MCB.22.3.935-945.2002
19. Lin Z, Cantos R, Patente M, Wu DK: Gbx2 is required for the morphogenesis of the mouse inner ear: a downstream candidate of hindbrain signaling. *Development* 132 (10),2309-18 (2005)
DOI: 10.1242/dev.01804
20. Liu M, Pereira FA, Price SD, Chu MJ, Shope C, Himes D, et al. Essential role of BETA2/NeuroD1 in development of the vestibular and auditory systems. *Genes Dev* 14 (22),2839-54 (2000)
DOI: 10.1101/gad.840500
21. Hulander M, Wurst W, Carlsson P, Enerbäck S: The winged helix transcription factor Fkh10 is required for normal development of the inner ear. *Nat Genet* 20 (4),374-6 (1998)
DOI: 10.1038/3850
22. Warchol ME, Richardson GP: Expression of the Pax2 transcription factor is associated with vestibular phenotype in the avian inner ear. *Dev Neurobiol* 69 (2-3),191-202 (2009)
DOI: 10.1002/dneu.20684
23. Hutson MR, Lewis JE, Nguyen-Luu D, Lindberg KH, Barald KF: Expression of Pax2 and patterning of the chick inner ear. *J Neurocytol* 28 (10-11),795-807 (1999)
DOI: 10.1023/A:1007057719025
24. Burton Q, Cole LK, Mulheisen M, Chang W, Wu DK: The role of Pax2 in mouse inner ear development. *Dev Biol* 272 (1),161-75 (2004)
DOI: 10.1016/j.ydbio.2004.04.024
25. Braunstein EM, Crenshaw EB 3rd, Morrow BE, Adams JC: Cooperative function of Tbx1 and Brn4 in the periotic mesenchyme is necessary for cochlea formation. *J Assoc Res Otolaryngol* 9 (1),33-43 (2008)
DOI: 10.1007/s10162-008-0110-6
26. Sanchez-Calderon H, Rodriguez-de la Rosa L, Milo M, Pichel JG, Holley M, Varela-Nieto I: RNA microarray analysis in prenatal mouse cochlea reveals novel IGF-I target genes: implication of MEF2 and FOXM1 transcription factors. *PLoS One* 5 (1),e8699 (2010)
DOI: 10.1371/journal.pone.0008699
27. Aburto MR, Magariños M, Leon Y, Varela-Nieto I, Sanchez-Calderon H: AKT signaling mediates IGF-I survival actions on otic neural progenitors. *PLoS One* 7 (1),e30790 (2012)
DOI: 10.1371/journal.pone.0030790
28. Yamamoto N, Chang W, Kelley MW: Rbpj regulates development of prosensory cells in the mammalian inner ear. *Dev Biol* 353 (2),367-79 (2011)

- DOI: 10.1016/j.ydbio.2011.03.016
29. Driver EC, Pryor SP, Hill P, Turner J, Rüther U, Biesecker LG, *et al*: Hedgehog signaling regulates sensory cell formation and auditory function in mice and humans. *J Neurosci* 28 (29),7350-8 (2008)
DOI: 10.1523/JNEUROSCI.0312-08.2008
 30. Khatri SB, Groves AK: Expression of the Foxi2 and Foxi3 transcription factors during development of chicken sensory placodes and pharyngeal arches. *Gene Expr Patterns* 13 (1-2),38-42 (2013)
DOI: 10.1016/j.gep.2012.10.001
 31. Nie X, Zhang K, Wang L, Ou G, Zhu H, Gao WQ: Transcription factor STOX1 regulates proliferation of inner ear epithelial cells via the AKT pathway. *Cell Prolif* 48 (2),209-20 (2015)
DOI: 10.1111/cpr.12174
 32. Kantarci H, Edlund RK, Groves AK, Riley BB: Tfap2a promotes specification and maturation of neurons in the inner ear through modulation of Bmp, Fgf and notch signaling. *PLoS Genet* 11 (3),e1005037 (2015)
DOI: 10.1371/journal.pgen.1005037
 33. Xiang M, Gan L, Li D, Chen ZY, Zhou L, O'Malley BW Jr, *et al*: Essential role of POU-domain factor Brn-3c in auditory and vestibular hair cell development. *Proc Natl Acad Sci U S A* 94 (17),9445-50 (1997)
DOI: 10.1073/pnas.94.17.9445
 34. Kelley MW, Driver EC, Puligilla C: Regulation of cell fate and patterning in the developing mammalian cochlea. *Curr Opin Otolaryngol Head Neck Surg* 17 (5),381-7 (2009)
DOI: 10.1097/MOO.0b013e3283303347
 35. Roberson DW, Alosi JA, Mercola M, Cotanche DA: REST mRNA expression in normal and regenerating avian auditory epithelium. *Hear Res* 172 (1-2),62-72 (2002)
DOI: 10.1016/S0378-5955(02)00512-9
 36. Bricaud O, Collazo A: The transcription factor six1 inhibits neuronal and promotes hair cell fate in the developing zebrafish (*Danio rerio*) inner ear. *J Neurosci* 26 (41),10438-51 (2006)
DOI: 10.1523/JNEUROSCI.1025-06.2006
 37. Pechriggl EJ, Bitsche M, Glueckert R, Rask-Andersen H, Blumer MJ, Schrott-Fischer A, Fritsch H: Development of the innervation of the human inner ear. *Dev Neurobiol* 75 (7),683-702 (2015)
 38. Smeti I, Watabe I, Savary E, Fontbonne A, Zine A: HMGA2, the architectural transcription factor high mobility group, is expressed in the developing and mature mouse cochlea. *PLoS One* 9 (2),e88757 (2014)
DOI: 10.1371/journal.pone.0088757
 39. Atar O, Avraham KB: Anti-apoptotic factor z-Val-Ala-Asp-fluoromethylketone promotes the survival of cochlear hair cells in a mouse model for human deafness. *Neuroscience* 168 (3),851-7 (2010)
DOI: 10.1016/j.neuroscience.2010.04.011
 40. Zhuang S, Zhang Q, Zhuang T, Evans SM, Liang X, Sun Y: Expression of Isl1 during mouse development. *Gene Expr Patterns* 13 (8),407-12 (2013)
DOI: 10.1016/j.gep.2013.07.001
 41. Huang M, Kantardzhieva A, Scheffer D, Liberman MC, Chen ZY: Hair cell overexpression of Isl1 reduces age-related and noise-induced hearing loss. *J Neurosci* 33 (38),15086-94 (2013)
DOI: 10.1523/JNEUROSCI.1489-13.2013
 42. Wallis D, Hamblen M, Zhou Y, Venken KJ, Schumacher A, Grimes HL, *et al*: The zinc finger transcription factor Gfi1, implicated in lymphomagenesis, is required for inner ear hair cell differentiation and survival. *Development* 130 (1),221-32 (2003)
DOI: 10.1242/dev.00190
 43. Hertzano R, Montcouquiol M, Rashi-Elkeles S, Elkon R, Yücel R, Frankel WN, *et al*: Transcription profiling of inner ears from Pou4f3 (ddl/ddl) identifies Gfi1 as a target of the Pou4f3 deafness gene. *Hum Mol Genet* 13 (18),2143-53 (2004)
DOI: 10.1093/hmg/ddh218
 44. Tornari C, Towers ER, Gale JE, Dawson SJ: Regulation of the orphan nuclear receptor Nr2f2 by the DFNA15 deafness gene Pou4f3. *PLoS One* 9 (11),e112247 (2014)
DOI: 10.1371/journal.pone.0112247
 45. Bermingham NA, Hassan BA, Price SD, Vollrath MA, Ben-Arie N, Eatock RA, *et al*: Math1: an essential gene for the generation of inner ear hair cells. *Science* 284 (5421),1837-41 (1999)
DOI: 10.1126/science.284.5421.1837
 46. Bermingham NA, Hassan BA, Wang VY,

- Fernandez M, Banfi S, Bellen HJ, *et al*: Proprioceptor pathway development is dependent on Math1. *Neuron* 30 (2), 411-22 (2001)
DOI: 10.1016/S0896-6273(01)00305-1
47. Woods C, Montcouquiol M, Kelley MW: Math1 regulates development of the sensory epithelium in the mammalian cochlea. *Nat Neurosci* 7 (12),1310-8 (2004)
DOI: 10.1038/nn1349
48. Ouji Y, Ishizaka S, Nakamura-Uchiyama F, Wanaka A, Yoshikawa M: Induction of inner ear hair cell-like cells from Math1-transfected mouse ES cells. *Cell Death Dis* 4,e700 (2013)
DOI: 10.1038/cddis.2013.230
49. Yang XY, Jin K, Ma R, Yang JM, Luo WW, Han Z, *et al*: Role of the planar cell polarity pathway in regulating ectopic hair cell-like cells induced by Math1 and testosterone treatment. *Brain Res* 1615,22-30 (2015)
DOI: 10.1016/j.brainres.2015.04.017
50. Kiernan AE, Pelling AL, Leung KK, Tang AS, Bell DM, Tease C, *et al*: Sox2 is required for sensory organ development in the mammalian inner ear. *Nature* 434 (7036),1031-5 (2005)
DOI: 10.1038/nature03487
51. Neves J, Kamaid A, Alsina B, Giraldez F: Differential expression of Sox2 and Sox3 in neuronal and sensory progenitors of the developing inner ear of the chick. *J Comp Neurol* 503 (4),487-500 (2007)
DOI: 10.1002/cne.21299
52. Pan W, Jin Y, Chen J, Rottier RJ, Steel KP, Kiernan AE: Ectopic expression of activated notch or SOX2 reveals similar and unique roles in the development of the sensory cell progenitors in the mammalian inner ear. *J Neurosci* 33 (41),16146-57 (2013)
DOI: 10.1523/JNEUROSCI.3150-12.2013
53. Dabdoub A, Puligilla C, Jones JM, Fritzsche B, Cheah KS, Pevny LH, *et al*: Sox2 signaling in prosensory domain specification and subsequent hair cell differentiation in the developing cochlea. *Proc Natl Acad Sci U S A* 105 (47),18396-401 (2008)
DOI: 10.1073/pnas.0808175105
54. Neves J, Uchikawa M, Bigas A, Giraldez F: The prosensory function of Sox2 in the chicken inner ear relies on the direct regulation of Atoh1. *PLoS One* 7 (1),e30871 (2012)
DOI: 10.1371/journal.pone.0030871
55. Yang H, Xie X, Deng M, Chen X, Gan L: Generation and characterization of Atoh1-Cre knock-in mouse line. *Genesis* 48 (6),407-13 (2010)
DOI: 10.1002/dvg.20633
56. Pan N, Jahan I, Kersigo J, Kopecky B, Santi P, Johnson S, *et al*: Conditional deletion of Atoh1 using Pax2-Cre results in viable mice without differentiated cochlear hair cells that have lost most of the organ of Corti. *Hear Res* 275 (1-2),66-80 (2011)
DOI: 10.1016/j.heares.2010.12.002
57. Sheykhosslami K, Thimmappa V, Nava C, Bai X, Yu H, Zheng T, *et al*: A new mutation of the Atoh1 gene in mice with normal life span allows analysis of inner ear and cerebellar phenotype in aging. *PLoS One* 8 (11),e79791 (2013)
DOI: 10.1371/journal.pone.0079791
58. Pan N, Jahan I, Kersigo J, Duncan JS, Kopecky B, Fritzsche B: A novel Atoh1 "self-terminating" mouse model reveals the necessity of proper Atoh1 level and duration for hair cell differentiation and viability. *PLoS One* 7 (1),e30358 (2012)
DOI: 10.1371/journal.pone.0030358
59. Lin Z, Perez P, Sun Z, Liu JJ, Shin JH, Hyrc KL, *et al*: Reprogramming of single-cell-derived mesenchymal stem cells into hair cell-like cells. *Otol Neurotol* 33 (9),1648-55 (2012)
DOI: 10.1097/MAO.0b013e3182713680
60. Kouros-Mehr H, Slorach EM, Sternlicht MD, Werb Z: GATA-3 maintains the differentiation of the luminal cell fate in the mammary gland. *Cell* 127 (5),1041-55 (2006)
DOI: 10.1016/j.cell.2006.09.048
61. Yagi R, Zhu J, Paul WE: An updated view on transcription factor GATA3-mediated regulation of Th1 and Th2 cell differentiation. *Int Immunol* 23 (7),415-20 (2011)
DOI: 10.1093/intimm/dxr029
62. van der Wees J, van Looij MA, de Ruiter MM, Elias H, van der Burg H, *et al*: Hearing loss following Gata3 haploinsufficiency is caused by cochlear disorder. *Neurobiol Dis* 16 (1),169-78 (2004)
DOI: 10.1016/j.nbd.2004.02.004
63. Milo M, Cacciabue-Rivolta D, Kneebone A, Van Doorninck H, Johnson C, Lawoko-Kerali G,

et al: Genomic analysis of the function of the transcription factor gata3 during development of the mammalian inner ear. *PLoS One* 4 (9):e7144 (2009)
DOI: 10.1371/journal.pone.0007144

64. Duncan JS, Fritzsche B: Continued expression of GATA3 is necessary for cochlear neurosensory development. *PLoS One* 8 (4):e62046 (2013)
DOI: 10.1371/journal.pone.0062046
65. Rivolta MN, Holley MC: GATA3 is downregulated during hair cell differentiation in the mouse cochlea. *J Neurocytol* 27 (9):637-47 (1998)
DOI: 10.1023/A:1006951813063

Abbreviations: HMG: high mobility group; IGF: Insulin-like growth factor

Key Words: Inner Ear, Hair Cell, Transcription Factor, Review

Send correspondence to: Shuna Li, Department of Otolaryngology and Head-Neck Surgery, Zhenjiang First People's Hospital, The Affiliated People's Hospital of Jiangsu University, 8 Dianli Road, Zhenjiang, Jiangsu, China, 212002, Tel: 86-511-88915351, Fax: 86-511-88915351, E-mail: lishuna01@yeah.net