Cardiotoxicity in oncology and coronary microcirculation: future challenges in theranostics

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1. ABSTRACT

Many of the patients undergoing chemotherapy or radiotherapy for cancer are at increased risk of developing cardiovascular diseases. Recent evidence suggests that cardiac dysfunction and subsequent heart failure are mainly due to vascular toxicity rather than only to due to myocyte toxicity. However, not all of the vascular toxicity of cancer therapies can be explained by epicardial coronary artery disease. In fact, in the last decades, it has been found that myocardial ischemia may occur as a consequence of structural or functional dysfunction of the complex network of vessels, which cannot be seen by a coronary angiography: the coronary microcirculation. Nowadays many diagnostic and therapeutic options are available both in coronary microvascular dysfunction and cardio-oncology. Aim of

this review is to suggest future theranostic implications of the relationship between cardiotoxicity in oncology and coronary microvascular dysfunction, showing common pathophysiologic mechanisms, proposing new diagnostic approaches and therapeutic options for cardioprotection.

2. INTRODUCTION

In recent years, cardiotoxicity of cancer treatments is a topic of growing interest. However, many aspects of both radiation-induced and cancer drug-induced cardiovascular diseases remain not fully elucidated. A careful review of the literature suggests that the main pathophysiological mechanisms

implicated in cardiotoxicity are due to vascular toxicity of cancer treatments. Either in the presence or in the absence of epicardial coronary artery disease, coronary microcirculation plays a key role in determining of myocardial ischemia. Aim of this review is to suggest potential theranostic implications of the relationship between cardiotoxicity in oncology and coronary microvascular dysfunction (CMD).

3. THE VASCULAR BRANCH OF CARDIOTOXICITY IN ONCOLOGY

As defined by the National Cancer Institute, cardiotoxicity is a general term indicating "toxicity affecting the heart" (1). In fact, regardless of whether the anticancer therapy is physical (ionizing radiation), chemical, hormonal or biological (the so called "targeted therapy"), it may have negative effects on the cardiovascular system (2). Given that targeted signalling cascades that promote cancer cell proliferation conversely protect vascular cells and cardiomyocytes (3), cancer therapies may be considered as a double-edged sword. More specifically, as expressed in a recent paper on cancer treatments and cardiovascular toxicity published by the European Society of Cardiology (4), about 50% of the cardiovascular toxicity in oncologic therapy is characterized by either functional or structural vascular damage leading to the worsening or the developing of coronary artery disease (CAD), peripheral vascular disease, thromboembolic disease, arterial hypertension and pulmonary hypertension. This observation suggests that a significant amount of cardiotoxicity from either chemotherapy (CT) or radiotherapy (RT) primarily involves vessels rather than cardiomyocytes. In fact, myocardial ischemia has been described as a very common side effect of several cancer therapies (4). For these reasons, patients with pre-existing cardiovascular diseases or with high cardiovascular risk should be considered at higher risk of developing vascular damage due to anti-cancer treatments. The most known risk factors associated with the development of vascular toxicity in patients underwent cancer therapies are arterial hypertension, diabetes mellitus, smoking, previous left ventricle dysfunction, heart failure (HF), previous CAD, increasing age, female gender and postmenopausal status (5). Moreover, genetic polymorphisms may predispose to cardiotoxicity (6), suggesting that genetic features might play a role in modulating the risk of cardiovascular toxicity after cancer treatment. Finally, therapy-related risk factors may include treatment type, drug class, drug dose, duration of treatment and use of combined therapy (7). However, there are gaps in evidence about the factors related to the development of CAD in patients treated with anti-cancer therapy. In this context, CMD precedes the development of coronary macrovascular disease, and it could offer a promising field of research in cardiotoxicity related to cancer treatment. Surprisingly, a great number of pathophysiological correlations may be found between these apparently unrelated topics. Moreover, CMD treatment may share similar therapeutic targets with the current best-evidence cardioprotective strategies.

4. CORONARY MICROVASCULAR DYSFUNCTION

Among patients undergoing coronary angiography because of angina, up to 40% are found to have normal-appearing epicardial coronary arteries (8). In fact, epicardial arteries, often referred to as the conductance vessels, represent the "visible" segment of the coronary circulation and give rise to the pre-arterioles and arterioles that constitute the microcirculation, the "invisible" segments of coronary circulation. In contrast with epicardial coronary arteries, the coronary microcirculation cannot be directly imaged in vivo neither with coronary angiography nor by intracoronary imaging techniques. Indeed, small coronary arteries are below the spatial resolution of coronary angiography (about 0.5. mm). Visual assessment of small coronary arteries might be possible by endomyocardial biopsy (11), but this invasive approach is not justified in the majority patients with CMD and does not allow assessment of functional alterations. Several methods have been proposed to investigate the functional state of coronary microcirculation (12), even though their application in the clinical setting is not always simple. In the past 20 years, a large number of studies using both invasive and non-invasive techniques for the assessment of coronary physiology, have produced a large wealth of data leading to a better understanding of CMD. Specifically, studies using positron emission tomography (PET) have permitted to establish the normal range of absolute myocardial blood flow (MBF, mL/min/g) and of coronary flow reserve (CFR) - which represents the ratio of MBF during near maximal coronary vasodilatation to baseline MBF (9). Patients with CMD show a reduced CFR that is usually identified by values lower than 2.0., which are unlikely to be detectable in apparently healthy subjects (10). In patients with normal coronary angiogram, CMD can represent an additional mechanism of subendocardial ischaemia manifesting typical chest pain and ST-segment depression during exercise, a condition commonly known as microvascular angina (MVA) (9). Even in the absence of significant epicardial CAD, CMD can cause myocardial ischemia, which has been shown to bear an independent prognostic value (8). In 2007, Camici and Crea (8) classified CMD into four main types on the basis of the clinical setting in which it occurs: CMD in the absence of myocardial diseases and obstructive CAD (type A), CMD in myocardial diseases (type B), CMD in obstructive CAD (type C) and iatrogenic CMD (type D). Similarly, in vascular toxicity of cancer treatments, CMD could present in different settings.

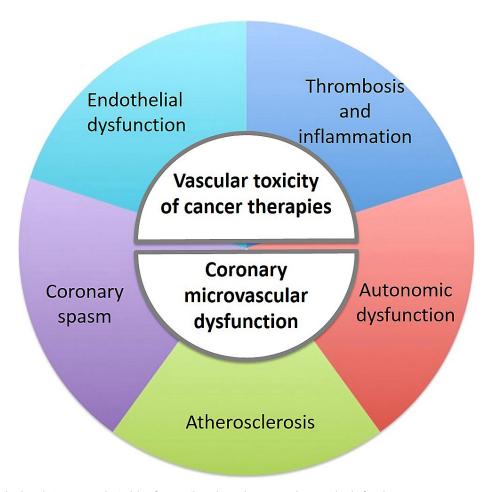


Figure 1. Shared mechanisms between vascular toxicity of cancer therapies and coronary microvascular dysfunction.

5. CMD AND VASCULAR CARDIOTOXICITY: SHARED PATHOPHYSIOLOGICAL MECHANISMS

The mechanisms by which cancer therapies cause myocardial ischemia are diverse and include accelerated atherosclerosis, thrombosis, vasospasm, and coronary microvascular impairment. We will show that many of them are shared with CMD, thus representing suitable targets for cardioprotection in cancer patients (figure 1).

5.1. Endothelial dysfunction

Endothelial cells have the key function of participating in the maintenance of patent and functional capillaries. Endothelial dysfunction is characterized by a shift down in the actions of the endothelium toward reduced vasodilation and prothrombic properties, as well as exacerbated pro-inflammatory state. Moreover, endothelial dysfunction has been associated with the majority of cardiovascular and peripheral vascular diseases (13).

5.1.1. Nitrix oxide (NO) production

Production and release of nitric oxide (NO) are the most important mechanisms of endotheliummediated vasodilation, and also the first to be lost in case of endothelial dysfunction (14). Among chemotherapeutics, vascular endothelial growth factor (VEGF) inhibitors like bevacizumab, sunitinib and sorafenibare known to starve cancer by inhibiting neo-angiogenesis (4). Notably, this class of drug is related with reduced NO production, resulting in vasoconstriction (15). In CMD, endothelium-mediated vasodilation has been found to be impaired, mainly due to either reduced activity of NO synthase, the enzyme that catalyzes NO synthesis from the aminoacid L-arginine, or increased serum levels of asymmetric dimethylarginine (ADMA), a major endogenous inhibitor of NO synthesis (16).

5.1.2. Reactive oxygen species (ROS)

Oxidative stress and subsequent release of reactive oxygen species (ROS) seems to play a

key role in endothelial dysfunction pathophysiology (17). Accordingly, mechanisms dependent on ROS were among the first to be linked to endothelial toxicity of cancer therapies (18). This is the case of anthracycline- and cisplatin-induced endothelial toxicity (19). Also 5-fluorouracil (5-FU) was found to induce ROS-induced endothelial damage (20). Similarly, vinca alkaloids (21), anti-Her2 target therapy (22) and almost every chemotherapeutic compound display significant detrimental effects on endothelial function due to oxidative stress (23). A parallelism with CMD may exist, since excessive generation of ROS is a common feature in several conditions that have been associated with a pathological impairment of endothelium-dependent coronary microvascular dilatation, as was demonstrated in diabetes, obesity, smoking, and other cardiovascular risk factors (24). Accordingly, administration of antioxidant substances. which prevent superoxide anion formation, including glutathione and antioxidant vitamins, improves or normalizes endothelium-dependent coronary microvascular dilatation both in experimental and in clinical conditions (25,26).

5.1.3. Endothelin-1 and angiotensin II

Mechanisms endothelial leading to dysfunction by bevacizumab and the others VEGF inhibitors are mainly related to increased endothelin-1 (ET-1) and angiotensin II release and production (27). In CMD, it has been demonstrated an imbalance between locally-released vasodilating agents and vasoconstrictors, thereby increasing the vasoconstrictor susceptibility of the endothelium. ET-1, in particular, is the most powerful vasoconstrictor substance produced in the body (28) and its production or release is often increased in the presence of CMD (29). In animal models, the intracoronary injection of ET-1 or angiotensin II resulted in myocardial ischemia due to vasoconstriction (29). Similar results can be observed in human by intracoronary injection of high doses of acetylcholine (30), which may cause chest pain and objective evidence of myocardial ischemia, in the absence of epicardial coronary arteries disease.

5.2. Atherosclerosis

Atherosclerosis is a chronic disease of the arterial wall, leading to the development of atheromatous plaques in the inner lining of the arteries (17). From a pathophysiological viewpoint, atherosclerosis is no more considered merely as a storage disease, but as an inflammatory disease (31). In cardio-oncology, the main risk factor for accelerated atherosclerosis is RT (32), which may lead to severe CAD, complicated by plaque rupture and thrombosis (33). Radiation-related CAD is usually a late complication, especially in survivors of breast cancer

or Hodgkin lymphoma (34), and it may not be detected until at least 10 years after thoracic RT exposure. CT also may lead to accelerated atherosclerosis and its dangerous consequences, as described for instance in cisplatin-treated survivors of testicular cancer (35). A significant contribution to atherosclerosis development in both cancer treatment toxicity and macro/microvascular CAD seems to be related to the activation of tissue renin angiotensin aldosterone systems (RAAS). In fact, RAAS exacerbation promotes significant pro inflammatory background by activating the transcription factor NF-kB, stimulating the expression of cell adhesion molecules and the release of pro-inflammatory cytokines like IL-1, IL-6 and TNFalpha (17). Moreover, angiotensin converting enzyme (ACE) expression is increased by the activation of macrophages by oxidized LDL (36) while angiotensin Il directly stimulate the activation of growth factors and the release of matrix metallo-proteinases (37), thereby making atherosclerotic plaque more prone to rupture and thrombosis (38).

5.3. Thrombosis

Vessel thrombosis is a critical event associated with myocardial infarction, stroke, and venous thromboembolic disorders, accounting for considerable morbidity and mortality. Moreover, venous thrombosis is the second leading cause of death in patients with cancer (39). Cancer therapy itself can induce blood clotting, thrombosis and thromboembolic events (34). This is particularly true for cisplatin, which may lead to arterial thrombosis with subsequent myocardial and cerebrovascular ischemia (40). The pathophysiology is multifactorial, including pro-coagulant and direct endothelial toxic effects, resulting in platelets aggregation and thromboxane formation (34). Among the immune and targeted therapeutics, those inhibiting the VEGF signalling pathway have an increased risk for coronary thrombosis, as shown in patients treated for breast cancer (41) or metastatic diseases (42). In the field of CMD, intravascular plugging of coronary microcirculation has been extensively described, and can be caused by atherosclerotic debris, microemboli and neutrophil-platelet aggregates (29). The main evidence comes from type 4 CMD, typically occurring during percutaneous coronary interventions and related to intracoronary manipulation of friable plagues (43). In these cases, microvascular plugging often causes "infarctlets", as indicated by a modest raise of markers of myocardial necrosis, and has a negative prognostic impact (44). Microvascular occlusion (MVO) has been described also in the setting of transmural myocardial infarction, resulting from a complex interplay of ischemia-reperfusion damage, endothelial dysfunction, platelet activation and vasoconstriction (45): all these mechanisms are largely shared with those from vascular toxicity resulting from cancer therapy described above.

5.4. Coronary spasm

Epicardial coronary spasm is defined as a condition in which a relatively large coronary artery running on the surface of the heart transiently exhibits abnormal contraction, leading to a transient complete or incomplete occlusion of the this artery (46). Abnormal vasoreactivity may be triggered by multiple stimuli acting on different cellular pathways involving both endothelium and underlying vascular smooth muscle cells (VSMCs). VSMCs in particular, are the main regulators of vascular tone and vessel patency (17), having protein-kinase C (PKC), the intracellular enzyme Rho-kinase 5 and ATP-dependent K+ channels as key mediators (46). After CT or RT as well as any kind of intimal injury, VSMCs change from the quiescent "contractile" phenotype state to the active "synthetic" state, that can migrate and proliferate from media to the intima (19). Among anti-cancer drugs in particular, the proteasome inhibitor bortezomib. approved for the treatment of multiple myeloma and non-Hodgkin's lymphoma, interferes with the degradation of cell cycle proteins in VSMCs, causing apoptosis and leading to coronary vasospasm (47). Conversely, Fluoropyrimidines, like 5-FU, have both direct toxicity on vascular endothelium and an indirect vasospastic effect via PKC activation in VSMCs (19). Similar effects have been described also for the modern "targeted therapy", since sorafenib too has been reported to induce epicardial vasospasm (48). Finally, coronary artery spasm may be triggered by RT in some cases (32), even though the predominant clinical manifestation of radiation-related heart disease today is atherosclerotic CAD (49). Similarly, an impaired VSMCs response to vasodilator stimuli have been described in the presence of CMD (50). Notably, endothelium-independent abnormalities in coronary microvascular dilatation may involve impaired opening of ATP-dependent K+ channels (51) and abnormal Rho-kinase 5 activity (52).

5.5. Hormonal effects

A confounding aspect of identifying the mechanisms of cardiotoxicity is that not all patients receiving these agents develop cardiotoxicity. The lack of uniform effect in males and females, for example, suggests possible hormonal interactions that modulate the cardiotoxic effect of some drugs. It is the case, for instance, of the tyrosine-kinase inhibitor sunitinib, which has shown more toxicities in multiple organ systems in females compared to men (53); this seems to be related to inhibition of the positive effects from estradiol in endothelial cells (54).

Estrogen deficiency has been demonstrated also in primary CMD and may, at least in part, explain the high prevalence of this disease in females often in the pre- and post-menopausal state, as accurately

described in a recent editorial (55). In fact, estrogen deficiency is associated with vasomotor abnormalities, including an impairment of endothelial function (56). Accordingly, estrogen administration has been demonstrated to improve endothelial function (57).

5.6. Autonomic dysfunction

Autonomic innervation plays a key role in regulating heart rate, myocardial function and MBF (58). Its impairment is associated with the development and the progression of cardiovascular diseases in cancer patients (59). Indeed, anti-cancer CT directly affects the function of the autonomic system. In this context, a reduced heart rate variability has been reported in patients treated with vincristine (60), doxorubicin (61) and paclitaxel (62). Moreover, aberrant blood pressure variability and maladaptive orthostatic responses are frequently observed in patients taking paclitaxel, taxanes, vinca alkaloids and cisplatin (58). Finally, damage to the cardiac nervous system by CT or thoracic RT may lead to sympathetic-vagal imbalance leading to sinus tachycardia that, reducing diastolic time and enhancing cardiac oxygen consumption, progressively induces myocardial ischemia (4). However, a significant quote of silent ischemia has been reported in cancer treated patients, since sensitivity fibers may be damaged in turn (63).

Moreover, it is well known that coronary microcirculation is under control of the autonomic nervous system (29). In particular, microvascular vasodilation is regulated by beta-2 adrenoceptor in small arterioles (64), while vasoconstriction is mediated by both alpha1- and alpha2-adrenoceptors (65). It has been demonstrated that either impaired beta-2 vasodilatation or augmented alpha-adrenergic constriction during exercise may be powerful enough to induce myocardial ischemia and MVA (66). Finally, parasympathetic activity has shown to cause coronary vasodilatation through a NO mediated mechanism (67).

6. CARDIOTOXICITY AND CMD: THERANOSTIC IMPLICATIONS

Theranostics combine the therapeutic ("thera") and diagnostic ("nostic") potentials of a certain compound (68). Prior to therapy, the compound is used in a diagnostic test to determine whether the drug will (potentially) exert a therapeutic effect, making it a powerful tool for personalizing disease treatment. Clinical applications of theranostics in oncology range from tissue-specific biomarkers to the most modern nanoparticles technologies. As extensively shown above, given that cardiotoxicity of cancer treatments and CMD shares several pathophysiological mechanisms, future research in theranostics should be addressed regarding the utility to diagnose and

treat CMD in patients receiving cancer therapies, especially because many effective drugs used to treat CMD already represent the current best-evidence cardioprotective strategies. Recently, dysregulation of microRNAs (miRNAs) involved in microvascular remodelling has been found in several cardiovascular diseases (69). In parallel, strong evidence has been found that some miRNAs can act as oncogenes or tumor suppressor genes, dysregulating neoangiogenesis and being involved in the initiation and progression of several human cancers (70).

6.1. Common diagnostic workup

The most common biomarkers employed in cardiotoxicity monitoring are troponin I and natriuretic peptides because of their higher cardiac specificity. However, they are not a reliable indicator of vascular toxicity of cancer therapy (7, 71). On the other hand, CMD can be non-invasively assessed through measurements of MBF and/or CFR in response to appropriate vasodilating stimuli (e.g. adenosine, dipyridamole) (29). Among non-invasive methods for the assessment of CMD, transthoracic stress-echocardiography allows the measurement of CFR through the quantification of diastolic flow velocity in the left anterior descendent artery at baseline and during maximal vasodilator stimulation (72). This non-invasive technique seems to be a promising diagnostic approach due to its widespread availability and safety, especially since 2-dimensional echocardiography is routinely used for monitoring patients with cancer. Moreover recent techniques. including 3d-echocardiogram, strain and speckle tracking, may allow the earlier detection of more subtle changes in myocardial function (71). In particular, the value of echocardiographic myocardial deformation parameters such as peak systolic global longitudinal strain for the early detection of myocardial changes and prediction of cardiotoxicity in patients receiving cancer therapy is well described in a recent review (73). In addition, diastolic dysfunction can be considered an early biomarker, since often it precedes systolic dysfunction in patients receiving chemotherapy (71). This is of particular interest since, according to a novel pathophysiologic paradigm, preserved ejection fraction HF and CMD are two facets of the same coin (74). Among radionuclide techniques, PET offers unrivalled sensitivity and specificity for the non-invasive study of coronary microcirculation in humans (75). Reliable assessment of CFR and MBF have been obtained by two tracers in particular, oxygen-15 labeled water (76) and nitrogen-13 labeled ammonia (77). However, because of its expensive and time-consuming features, imaging with PET can only be performed in highly specialized centres (29). Moreover, cardiac magnetic resonance (CMR) affords the opportunity for non-invasive tissue characterization including myocardial oedema, inflammation, and fibrosis,

playing an important role in identification of both early and late cardiotoxicity in patients with cancer (78). At the same time, CMR perfusion sequences have shown to allow the accurate investigation of CMD (79). Finally, a complete assessment of CMD may also include, at least in some patients, the assessment of coronary microvascular response to constrictor stimuli (acetylcholine) (30). These tests, however, need to be carried out during invasive procedures, identifying a significant coronary vasoconstriction during coronary angiography in the absence of established coronary artery disease (29). A proposed algorithm that can be considered in the assessment of cardiac toxicity of cancer therapies including the identification of myocardial ischemia is summarized in figure 2.

6.2. Rationale for cardioprotection

Prevention from cardiotoxicity may be primary, extended to all patients already treated with potentially cardiotoxic therapies, or secondary in selected high-risk patients showing pre-clinical signs of cardiotoxicity (71). A tailored prevention strategy based on the cardiac risk stratification according to patient-related and therapy-related risk factors bears further investigation. Given the wide sharing of pathophysiological mechanisms between cardiotoxicity and CMD, several common therapeutic approaches have shown positive effects in both clinical conditions. While waiting for the advent of modern molecular therapostics, there is a rational of using best-evidence therapies for CMD to treat cardiotoxicity in oncology.

6.2.1. Beta-blockers

Beta-blockers have shown several beneficial effects in patients with CMD and stable MVA, by reducing myocardial oxygen consumption and improving coronary perfusion (29). Consistently, an improvement of ischemic and angina threshold was reported in some studies (80, 81). On the other hand, there is growing evidence suggesting a cardioprotective role of beta-blockers in prevention of CT-induced cardiotoxicity. Carvedilol in particular. which has also antioxidant properties and the ability to chelate iron, was reported as able to prevent cardiac histopathological damage caused by doxorubicin (82). However, there is no definite evidence for a class-effect benefit of these compounds in terms of cardioprotection. In fact, metoprolol showed a neutral effect (83), while nonselective beta-blockers such as propranolol resulted dangerous because of potential enhanced cardiotoxicity (84).

6.2.2. RAAS-inhibitors

ACE-inhibitors have been proposed as therapeutic agents in MVA due to their lowering effects on serum and tissue angiotensin II levels (85). In

RISK OF CARDIOTOXICITY Age, gender, cardiovascular risk factors, known CAD, pre-existent cardiac disease and co-morbidities Screening, diagnosis and development of new BASELINE EVALUATION CLINICAL EVALUATION therapeutic options Usual care New proposed alahorithm Identify Identify Identify myocyte Myocadial cardiac dysfuction damage and Ischemia overload **ECHOCARDIOGRAPHY** ASSESMENT OF CORONARY LABORATORY MARKERS Systolic dsyfunction FLOW RESERVE Troponin Diastolic dysfunction Stress-Echo NT-proBNP LV Hypertrophy PET/SPECT LV Remodelling Stress-CMR Cardioprotective drugs

Proposed algorithm to assess cardiotoxicity of cancer therapies

Figure 2. Flow diagram summarizing hypothetical algorithm in the assessment of cardiac toxicity of cancer therapies including the identification of myocardial ischemia. LV, left ventricular; CMR, cardiac magnetic resonance; SPECT, single-photonemission computed tomography; PET, positron emission tomography.

particular, enalapril has been found to improve CMD through the increase of NO availability and reduction of oxidative stress in MVA patients (86). In cardiotoxicity, animal studies suggest that enalapril and other ACE-inhibitors may be cardioprotective in anthracycline toxicity by preserving mitochondrial function and down regulating ROS generation (87, 88). In particular, thanks to its ROS scavenger role, the ACE-inhibitor zofenopril exerts protective properties through off-target mechanisms in endothelial cells, as increased acidic sulphide group availability (89). Similar benefits seem to apply to angiotensin receptor blockers in reducing the formation of ROS, thus attenuating the development of myocardial dysfunction in cancer patients treated by CT (90).

6.2.3. Statins

Statins might have beneficial effects on CMD by improving endothelial function through several effects such as antioxidant, anti-inflammatory and cholesterol-lowering effects. They were able to improve exercise stress test performance in MVA patients (91) and to significantly reduce oxidative

stress and endothelial function after 6 months of treatment (92). In cardiotoxicity, studies suggest the benefit of statins in reducing anthracycline-mediated cardiomyocyte death (93) and subsequent HF (94). However, to date no prospective trials have addressed the role of statins in the prevention of cancer therapy-related cardiotoxicity.

6.2.4. Antioxidants

Benefits from antioxidant therapy have been clearly observed only in type A CMD, in which short-term administration of the antioxidant vitamin C restored coronary microcirculatory responsiveness and normalized CFR in smokers without significant CAD nor structural heart disease (95). However, clinical use of antioxidants to protect the heart during chemotherapy is controversial due to the potentially reduced cytotoxic efficacy toward cancer cells (19). Nevertheless, recent evidences suggest the protective role of mitochondrial aldehyde dehydrogenase-2 (ALDH-2) in endothelial cells exposed to stress insult (96). Thus, since dysfunction of this molecule has been associated with both ischemic heart disease

(97) and doxorubicin-mediated cardiotoxicity (98), ALDH-2 targeting seems to be a promising therapeutic challenge to modulate mitochondrial functions and possibly neo-angiogenesis (96).

6.2.5. Other drugs

Non-dihydropyridine calcium antagonists, nitrates, adenosine, nicorandil and alpha-antagonists constitute other therapeutic options for MVA (29). However, their possible role in cardioprotection has still to be elucidated. Finally, it would be of particular interest to investigate the cardioprotective role of ranolazine and ivabradin, which have shown their positive effects on CMD by improving diastolic dysfunction and prolonging diastolic MBF time, respectively. Ranolazine and ivabradin have been tested as cardioprotective agents in both animal models and small series of patients with CT-related reduced ejection fraction HF (29). Given the recently described parallelism between CMD and diastolic dysfunction (74), studies are required to prove their efficacy in patients with CT/RT-related preserved ejection fraction HF.

7. CONCLUSIONS: KNOWLEDGE GAPS AND FUTURE DIRECTIONS

The specialty of cardio-oncology has gained significant momentum, with increasing awareness and interest in advancing this field. Although many important progress has been reached in this field, not all the cardiac toxicity of cancer treatment can be prevented or justified only assessing myocardial function and structure. After the evaluation of cardiotoxicity risk, current guidelines recommend to assess cardiotoxicity mainly using laboratory markers, electrocardiogram and echocardiogram. However, the study of cardiotoxicity revealed that the majority of early myocardial damage from CT/RT primary involve vascular toxicity rather that direct myocyte toxicity. For this reason, we propose myocardial ischemia as new theranostic target in the field of cardiooncology. In this view, in patients undergoing anti-cancer treatment, especially in the presence of high risk of ischemic heart disease, the evaluation of myocardial ischemia, using PET/SPET or stress-echocardiography, allow to non-invasively identify patients with CMD treatable with well-known anti-ischemic drugs (figure 2). Moreover, shared molecular pathways between CMD and cardiotoxicity, such as oxidative stress response. VSMCs tone, inflammation and thrombosis, represent the basis for the development of future research on new strategies for tailored cardioprotection.

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9. REFERENCES

- I. Brana, J. Tabernero: Cardiotoxicity. *Ann Oncol* 21, 173–179 (2010)
 DOI: 10.1093/annonc/mdq295
- M.S. Ewer, S.M. Swain, D. Cardinale, A. Fadol, T.M. Suter: Cardiac dysfunction after cancer treatment. *Tex Heart Inst J* 38, 148–252 (2011)
- G. Minotti, P. Menna, E. Salvatorelli, G. Cairo, L. Gianni: Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. Pharmacol Rev 56, 185–229 (2004) DOI: 10.1124/pr.56.2.6
- 4. J.L. Zamorano, P. Lancellotti, D. Rodriguez Muñoz, V. Abonyas, R. Asteggiano, M. Galderisi, G. Habib, D.J. Lenihan, G.Y. Lip, A.R. Lyon, T. Lopez Fernandez, D. Mothy, M.F. Piepoli, J. Tamargo, A. Torbicki, T.M. Suter, Authors/Task Force Members; ESC Committee for Practice Guidelines (CPG): 2016 ESC Position Paper on cancer treatments and cardiovascular toxicity developed under the auspices of the ESC Committee for Practice Guidelines: The Task Force for cancer treatments and cardiovascular toxicity of the European Society of Cardiology (ESC) Eur Heart J 37, 2768–2801 (2016)

DOI: 10.1093/eurhearti/ehw211

A. Albini, G. Pennesi, F. Donatelli, R. Cammarota, S. De Flora, D.M. Noonan: Cardiotoxicity of anticancer drugs: the need for cardio-oncology and cardio-oncological prevention. *J Natl Cancer Inst* 102, 14–25 (2010)

DOI: 10.1093/jnci/djp440

- J.G. Blanco, C.L. Sun, W. Landier, L. Chen, D. Esparza-Duran, W. Leisenring, A. Mays, D.L. Friedman, J.P. Ginsberg, M.M. Hudson, J.P. Neglia, K.C. Oeffinger, A.K. Ritchey, D. Villaluna, M.V. Relling, S. Bhatia: Anthracycline related cardiomyopathy after childhood cancer: role of polymorphisms in carbonyl reductase genes a report from the Children's Oncology Group. *J Clin Oncol* 30, 1415–1421 (2012)
 DOI: 10.1200/JCO.2011.34.8987
- M.W. Bloom, C.E. Hamo, D. Cardinale, B. Ky, A. Nohria, L. Baer, H. Skopicki, D.J. Lenihan, M. Gheorghiade, A.R. Lyon, J. Butler: Cancer therapy-related cardiac

- dysfunction and heart failure Part 1: definitions, pathophysiology, risk factors, and imaging. *Circ Heart Fail* 9, e002661 (2016)
- DOI:10.1161/CIRCHEARTFAILURE.115.002661
- P.G. Camici, F. Crea: Coronary microvascular dysfunction. N Engl J Med 356, 830–840 (2007)
 DOI: 10.1056/NEJMra061889
- 9. F. Crea, P.G. Camici, C.N. Bairey Merz: Coronary microvascular dysfunction: an update. *Eur Heart J* 35,1101–1111 (2014) DOI: 10.1093/eurheartj/eht513
- A. Sestito, G.A. Lanza, A. Di Monaco, P. Lamendola, G. Careri, P. Tarzia, G. Pinnacchio, I. Battipaglia, F. Crea: Relation between cardiovascular risk factors and coronary microvascular dysfunction in cardiac syndrome X. *J Cardiovasc Med* 12, 322–327 (2011) DOI: 10.2459/JCM.0b013e3283406479
- J. Escaned, A. Flores, P. García-Pavía, J. Segovia, J. Jimenez, P. Aragoncillo, C. Salas, F. Alfonso, R. Hernàndez, D.J. Angiolillo, P. Jiménez-Quevedo, C. Bañuelos, L. Alonso-Pulpón, C. Macaya: Assessment of microcirculatory remodeling with intracoronary flow velocity and pressure measurements: validation with endomyocardial sampling in cardiac allografts. Circulation 120, 1561–1568 (2009) DOI:10.1161/CIRCULATIONAHA.108.834739
- D.Y. Leung, M. Leung M. Non-invasive/ invasive imaging: significance and assessment of coronary microvascular dysfunction. *Heart* 97, 587–595 (2011) DOI: 10.1136/hrt.2009.183327
- P. Rajendran, T. Rengarjan, J. Thangavel, Y. Nishigaki, D. Sakthisekaran, G. Sethi, I. Nishigaki: The vascular endothelium and human diseases. *Int J Biol Sci* 9, 1057–1069 (2013) DOI: 10.7150/ijbs.7502
- 14. N.D. Roe, J. Ren: Nitric oxide synthase uncoupling: a therapeutic target in cardiovascular diseases. *Vascul Pharmacol* 57, 168–172 (2012)
 DOI: 10.1016/j.vph.2012.02.004
- D.C. Sane, L. Anton, K.B. Brosnihan: Angiogenic growth factor and hypertension. Angiogenesis 7, 193–201 (2004) DOI: 10.1007/s10456–004-2699–3

- F. Perticone, A. Sciacqua, R. Maio, M. Perticone, R. Maas, R.H. Boger, G. Tripepi, G. Sesti, C. Zoccali: Asymmetric dimethylarginine, L-arginine, and endothelial dysfunction in essential hypertension. *J Am Coll Cardiol* 46, 518–523 (2005) DOI: 10.1016/j.jacc.2005.04.040
- A. Durante, G. Peretto, A. Laricchia, F. Ancona, M. Spartera, A. Mangieri, D. Cianflone: Role of the renin-angiotensin-aldosterone system in the pathogenesis of atherosclerosis. *Curr Pharm Des* 18, 981–1004 (2012)
 DOI: 10.2174/138161212799436467
- M.B. Wolf, J.W. Baynes. The anti-cancer drug, doxorubicin, causes oxidant stress induced endothelial dysfunction. Biochim Biophys Acta 1760, 267–271 (2006) DOI: 10.1016/j.bbagen.2005.10.012
- L. Morbidelli, S. Donnini, M. Ziche: Targeting endothelial cell metabolism for cardioprotection from the toxicity of antitumor agents. *Cardiooncology*, 2:3 (2016) DOI: 10.1186/s40959–016-0010–6
- S. Kinhult, M. Albertsson, J. Eskilsson, M. Cwikici: Effects on probucol on endothelial damage by 5-fluorouracil. *Acta Oncol* 42, 304–308 (2003)
 DOI: 10.1080/02841860310004409
- B.L. Samuels, N.J. Vogelzang, B.J. Kennedy. Severe vascular toxicity associated with vinblastine, bleomycin, and cisplatin chemotherapy. Cancer Chemother Pharmacol 19, 253–256 (1987)
 DOI: 10.1007/BF00252982
- H. Hurwitz, L. Fehrenbacher. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med 350, 2335–2342 (2004) DOI: 10.1056/NEJMoa032691
- 23. A. Soultati, G. Mountzios, C. Avgerinou, G. Papaxoinis, D. Pectasides, M.A. Dimopoulos, C. Papadimitriou: Endothelial vascular toxicity from chemotherapeutic agents: preclinical evidence and clinical implications. *Cancer Treat Rev* 38, 473–483 (2012)
 DOI: 10.1016/j.ctrv.2011.09.002
- 24. Harrison DG: Cellular and molecular mechanisms of endothelial cell dysfunction. *J Clin Invest* 100, 2153–2157 (1997) DOI: 10.1172/JCI119751

- 25. A. Mügge, J.H. Elwell, T.E. Peterson, T.G. Hofmeyer, D.D. Heistad, D.G. Harrison: Chronic treatment with polyethylene glycolated superoxide dismutase partially restores endotheliumdependent vascular relaxations in cholesterol-fed rabbits. *Circ Res* 69, 1293–1300 (1991) DOI: 10.1161/01.RES.69.5.1293
- T. Heitzer, H. Just, T. Munzel: Antioxidant vitamin C improves endothelial dysfunction in chronic smokers. *Circulation* 94, 6–9 (1996)
 DOI: 10.1161/01.CIR.94.1.6
- T. Kamba, D.M. McDonald: Mechanisms of adverse effects of anti-VEGF therapy for cancer. *Br J Cancer* 96, 1788–1795 (2007)
 DOI: 10.1038/sj.bjc.6603813
- 28. J.R. Vane, E.E. Anggard, R.M. Botting: Regulatory functions of the vascular endothelium. *N Engl J Med* 323, 27–36 (1990)
 DOI: 10.1056/NEJM199007053230106
- P.G. Camici, G. d'Amati, O.E. Rimoldi: Coronary microvascular dysfunction: mechanisms and functional assessment. *Nat Rev Cardiol* 12, 48–62 (2015)
 DOI: 10.1038/nrcardio.2014.160
- 30. Chayama: Response of the left anterior descending coronary artery to acetylcholine in patients with chest pain and angiographically normal coronary arteries. *Am J Cardiol* 92, 1394–1398 (2003) DOI: 10.1016/j.amjcard.2003.08.043
- 31. P. Libby, P.M. Ridker, G.K. Hansson: Inflammation in atherosclerosis: from pathophysiology to practice. *J Am Coll Cardiol* 54, 2129–2138 (2009) DOI: 10.1016/j.jacc.2009.09.009
- P. McGale, S.C. Darby, P. Hall, J. Adolfsson, N.O. Bengtsson, A.M. Bennet, T. Fornander, B. Gigante, M.B. Jensen, R. Peto, K. Rahimi, C.W. Taylor, M. Ewertz: Incidence of heart disease in 35,000 women treated with radiotherapy for breast cancer in Denmark and Sweden. Radiother Oncol 100, 167– 175 (2011) DOI: 10.1016/j.radonc.2011.06.016
- F.C. Brosius 3rd, B.F. Waller, W.C. Roberts: Radiation heart disease. Analysis of 16 young (aged 15 to 33 years) necropsy

- patients who received over 3,500 rads to the heart. *Am J Med* 70, 519–530 (1981) DOI: 10.1016/0002–9343(81)90574-X
- 34. M. Florescu, M. Cinteza, D. Vinereanu: Chemotherapy-induced cardiotoxicity. Maedica 8, 59–67 (2013)
- H.S. Haugnes, T. Wethal, N. Aass, O. Dahl, O. Klepp, C.W. Langberg, T. Wilsgaard, R.M. Bremnes, S.D. Fossa. Cardiovascular risk factors and morbidity in long-term survivors of testicular cancer: a 20-year follow-up study. *J Clin Oncol* 28, 4649–4657 (2010) DOI: 10.1200/JCO.2010.29.9362
- P. Rouet-Benzineb, B. Gontero, P. Dreyfus, C. Lafuma: Angiotensin II induces nuclear factor- kappa B activation in cultured neonatal rat cardiomyocytes through protein kinase C signaling pathway. J Mol Cell Cardiol 32, 1767–1778 (2000) DOI: 10.1006/jmcc.2000.1211
- A.J. Naftilan, R.E. Pratt, V.J. Dzau VJ: Induction of platelet-derived growth factor A-chain and c-myc gene expressions by angiotensin II in cultured rat vascular smooth muscle cells. *J Clin Invest* 83, 1419–1424 (1989)
 DOI: 10.1172/JCI114032
- R.T. Lee, F.J. Schoen, H.M. Loree, M.W. Lark, P. Libby: Circumferential stress and matrix metalloproteinase 1 in human coronary atherosclerosis: implications for plaque rupture. *Arterioscler Thromb Vasc Biol* 16, 1070–1073 (1996)
 DOI: 10.1161/01.ATV.16.8.1070
- 39. B. Furie, B.C. Furie: Mechanisms of thrombus formation. *N Engl J Med* 359, 938–949 (2008)
 DOI: 10.1056/NEJMra0801082
- R.A. Moore, N. Adel, E. Riedel, M. Bhutani, D.R. Feldman, N.E. Tabbara, G. Soff, R. Parameswaran, H. Hassoun: High incidence of thromboembolic events in patients treated with cisplatin-based chemotherapy: a large retrospective analysis. *J Clin Oncol* 29, 3466–3473 (2011) DOI: 10.1200/JCO.2011.35.5669
- D. Cameron, J. Brown, R. Dent, C. Jackisch, J. Mackey, X. Pivot, G.G. Steger, T.M. Suter, M. Toi, M. Parmar, R. Laeufle, Y.H. Im, G. Romieu, V. Harvey, O. Lipatov, T. Pienkowski T, P. Cottu, A. Chan A, S.A. Im, P.S. Hall, L. Bubuteishvili-Pacaud, V. Henschel, R.J.

- Deurloo, C. Pallaud, R. Bell: Adjuvant bevacizumab-containing therapy in triplenegative breast cancer (BEATRICE): primary results of a randomised, phase 3 trial. *Lancet Oncol* 14, 933–942 (2013) DOI: 10.1016/S1470–2045(13)70335–8
- 42. F.A. Scappaticci, J.R. Skillings, S.N. Holden, H.P. Gerber, K. Miller, F. Kabbinavar, E. Bergsland, J. Ngai, E. Holmgren, J. Wang, H. Hurwitz. Arterial thromboembolic events in patients with metastatic carcinoma treated with chemotherapy and bevacizumab. *J Natl Cancer Inst* 2007, 99, 1232–1239 (2007) DOI: 10.1093/jnci/djm086
- I. Porto, F. Belloni, G. Niccoli, C. Larosa, A.M. Leone, F. Burzotta, C. Trani, G.L. De Maria, C. Hamilton-Craig, F. Crea: Filter no-reflow during percutaneous coronary intervention of saphenous vein grafts: incidence, predictors and effect of the type of protection device. *EuroIntervention* 7, 955–961 (2011) DOI: 10.4244/EIJV7I8A151
- 44. R. Corbalán, G. Larrain, C. Nazzal, P.F. Castro, M. Acevedo, J.M. Dominiquez, F. Bellolio, M.W. Krucoff: Association of noninvasive markers of coronary artery reperfusion to assess microvascular obstruction in patients with acute myocardial infarction treated with primary angioplasty. Am J Cardiol 88, 342–346 (2001) DOI: 10.1016/S0002–9149(01)01676–9
- G. Niccoli, F. Burzotta, L. Galiuto, F. Crea F: Myocardial no-reflow in humans. *J Am Coll Cardiol* 54, 281–292 (2009)
 DOI: 10.1016/j.jacc.2009.03.054
- H. Ogawa, T. Akasaka, R. Hattori, Japanese Circulation Society (JCS) Joint Working Group: Guidelines for diagnosis and treatment of patients with vasospastic angina (coronary spastic angina) (JCS 2008): digest version. Circ J 74, 1745–1762 (2010) DOI: 10.1253/circi.CJ-10-74-0802
- 47. C.E. Hamo, M.W. Bloom: Getting to the heart of the matter: An overview of cardiac toxicity related to cancer therapy. *Clin Med Insights Cardiol* 9, 47–51 (2015)
- Y. Arima, S. Oshima, K. Noda, H. Fukushima, I. Taniguchi, S. Nakamura, M. Shono, H. Ogawa: Sorafenib-induced acute myocardial infarction due to coronary artery spasm. *J Cardiol* 54, 512–515 (2009) DOI: 10.1016/j.jjcc.2009.03.009

- 49. R. Virmani, A. Farb, A.J. Carter, R.M. Jones: Comparative pathology: radiation-induced coronary artery disease in man and animals. Semin Interv Cardiol 3, 163–172 (1998)
- M.A. Kurz, K.G. Lamping, J.N. Bates, C.L. Eastham, M.L. Marcus, D.G. Harrison: Mechanisms responsible for the heterogeneous coronary microvascular response to nitroglycerin. Circ Res 68, 847–855 (1991) DOI: 10.1161/01.RES.68.3.847
- 51. W.F. Jackson: Potassium channels in the peripheral microcirculation. *Microcirculation* 12, 113–127 (2005)
 DOI: 10.1080/10739680590896072
- A. Masumoto, M. Mohri M, H. Shimokawa, L. Urakami, M. Usui, A. Takeshita: Suppression of coronary artery spasm by the Rho-kinase inhibitor fasudil in patients with vasospastic angina. *Circulation* 105, 1545–1547 (2002) DOI: 10.1161/hc1002.105938
- 53. A.A. van der Veldt, E. Boven, H.H. Helgason, M. van Wouwe M, J. Berkhof, G. de Gast, H. Mallo, C.N. Tillier, A.J. van den Eertwegh, J.B. Haanen: Predictive factors for severe toxicity of sunitinib in unselected patients with advanced renal cell cancer. *Br J Cancer* 99, 259–265 (2008) DOI: 10.1038/sj.bjc.6604456
- 54. Pamela Ann Harvey and Leslie Anne Leinwand. Oestrogen enhances cardiotoxicity induced by Sunitinib by regulation of drug transport and metabolism. *Cardiovascular Research*, 107, 66–77(2015) DOI: 10.1093/cvr/cvv152
- 55. P.G. Camici, F. Crea: Microvascular angina: a women's affair? *Circ Cardiovasc Imaging* 8, pii: e003252 (2015)
 DOI: 10.1161/CIRCIMAGING.115.003252
- P. Collins P: Role of endothelial dysfunction and oestrogens in syndrome X. Coron Artery Dis 3, 593–598 (1992)
 DOI: 10.1097/00019501–199207000-00008
- 57. E.H. Lieberman, M.D. Gerhard, A. Uehata, B.W. Walsh, A.P. Selwyn, P. Ganz, A.C. Yeung, M.A. Creager: Estrogen improves endotheliumdependent, flow-mediated vasodilation in postmenopausal women. *Ann Intern Med* 121, 936–941 (1994) DOI: 10.7326/0003–4819-121–12-199412150–00005

- 58. S.C. Adams, R. Schondorf, J. Benoit, R.D. Kilgour: Impact of cancer and chemotherapy on autonomic nervous system function and cardiovascular reactivity in young adults with cancer: a case-controlled feasibility study. *BMC Cancer* 15, 414 (2015) DOI: 10.1186/s12885–015-1418–3
- R.D. Brook, S. Julius: Autonomic imbalance, hypertension, and cardiovascular risk. *Am J Hypertens* 13, 112–122 (2000)
 DOI: 10.1016/S0895–7061(00)00228–4
- H.E. Hirvonen, T.T. Salmi, E. Heinonen, K.J. Antila, I.A. Valimaki: Vincristine treatment of acute lymphoblastic leukemia induces transient autonomic cardioneuropathy. *Cancer* 64, 801–805 (1989)
 DOI: 10.1002/1097–0142(19890815)64:4<801:: AID-CNCR2820640406>3.0.CO;2-E
- 61. W.J. Hrushesky, D.J. Fader, J.S. Berestka, M. Sommer, J. Hayes, F.O. Cope. Diminishment of respiratory sinus arrhythmia foreshadows doxorubicin-induced cardiomyopathy. *Circulation*, 84, 697–707 (1991) DOI: 10.1161/01.CIR.84.2.697
- E. Ekholm, E. Salminen, H. Huikuri, J. Jalonen, K. Antila, T.A. Salmi, V.T. Rantanen: Impairment of heart rate variability during paclitaxel therapy. *Cancer* 88, 2149–53 (2000)
 DOI: 10.1002/(SICI)1097–0142(20000501) 88:9<2149::AID-CNCR22>3.0.CO;2-Z
- 63. K.K. Ness, G.T. Armstrong GT: Screening for cardiac autonomic dysfunction among Hodgkin lymphoma survivors treated with thoracic radiation. *J Am Coll Cardiol* 65, 584–585 (2015)
 DOI: 10.1016/j.jacc.2014.11.036
- 64. J. Quillen, F. Sellke, P. Banitt, D. Harrison D: The effect of norepinephrine on the coronary microcirculation. *J Vasc Res* 29, 2–7 (1992)
- 65. E.O. Feigl: The paradox of adrenergic coronary vasoconstriction. *Circulation* 76, 737–745 (1987)
 DOI: 10.1161/01.CIR.76.4.737
- A. Maseri: Coronary vasoconstriction: visible and invisible. N Engl J Med 325, 1579–1580 (1991)
 DOI: 10.1056/NEJM199111283252210

- 67. T.P. Broten, J.K. Miyashiro, S. Moncada, E.O. Feigl EO: Role of endothelium-derived relaxing factor in parasympathetic coronary vasodilatation. *Am J Physiol* 262, H1579–H1584 (1992)
- E.D.G. Fleurena, Y.M.H. Versleijen-Jonkersa, S. Heskampb, C.M.L. van Herpena, W.J.G. Oyenb, W.T.A. van der Graafa, O.C. Boermanb: Theranostic applications of antibodies in oncology. Mol Oncol 8, 799–812 (2014) DOI: 10.1016/j.molonc.2014.03.010
- 69. A. Busch, S.M. Eken, L. Maegdefessel: Prospective and therapeutic screening value of non-coding RNA as biomarkers in cardiovascular disease. *Ann Transl Med* 4, 236 (2016)
 DOI: 10.21037/atm.2016.06.06
- G. Bertoli, C. Cava, I. Castiglioni: MicroRNAs: new biomarkers for diagnosis, prognosis, therapy prediction and therapeutic tools for breast cancer. *Theranostics* 5, 1122–1143 (2015)
 DOI: 10.7150/thno.11543
- M.W. Bloom, C.E. Hamo, D. Cardinale, B. Ky, A. Nohria, L. Baer, H. Skopicki, D.J. Lenihan, M. Gheorghiade, A.R. Lyon, J. Butler: Cancer therapy-related cardiac dysfunction and heart failure Part 2: prevention, treatment, guidelines, and future directions. Circ Heart Fail 9, e002843 (2016) DOI: 10.1161/CIRCHEARTFAILURE.115. 002661
 DOI: 10.1161/CIRCHEARTFAILURE.115. 002843
- 72. T. Hozumi, K. Yoshida, Y. Ogata, T. Akasaka, Y. Asami, T. Takagi, S. Morioka: Noninvasive assessment of coronary flow velocity and coronary flow velocity reserve in the left anterior descending coronary artery by Doppler echocardiography: comparison with invasive technique. *J Am Coll Cardiol* 32, 1251–1259 (1998)
 DOI: 10.1016/S0735–1097(98)00389–1
- 73. P. Thavendiranathan, F. Poulin, K.D. Lim, J.C. Plana, A. Woo, T.H. Marwick: Use of Myocardial Strain Imaging by Echocardiography for the Early Detection of Cardiotoxicity in Patients During and After Cancer Chemotherapy: A Systematic Review. *J Am Coll Cardiol* 65, 2751–2768 (2015)

- K.S. Shah, A.S. Maisel: Novel biomarkers in heart failure with preserved ejection fraction. Heart Fail Clin 10,471–479 (2014)
 DOI: 10.1016/j.hfc.2014.04.005
 DOI: 10.1016/j.hfc.2014.04.008
- 75. P.G. Camici, O.E. Rimoldi: The clinical value of myocardial blood flow measurement. *J Nucl Med* 50, 1076–1087 (2009) DOI: 10.2967/jnumed.108.054478
- 76. P.A. Kaufmann, T. Gnecchi-Ruscone, J.T. Yap, O.E. Rimoldi, P.G. Camici: Assessment of the reproducibility of baseline and hyperemic myocardial blood flow measurements with 15O-labeled water and PET. *J Nucl Med* 40, 1848–1856 (1999)
- T.H. Schindler, H.R. Schelbert, A. Quercioli,
 V. Dilsizian: Cardiac PET Imaging for the detection and monitoring of coronary artery disease and microvascular health. *JACC Cardiovas Imaging* 3, 623–640 (2010)
 DOI: 10.1016/j.jcmg.2010.04.007
- P. Thavendiranathan, B.J. Wintersperger, S.D. Flamm, T.H. Marwick. Cardiac MRI in the assessment of cardiac injury and toxicity from cancer chemotherapy: a systematic review. *Circ Cardiovasc Imaging* 6,1080– 1091 (2013) DOI: 10.1161/CIRCIMAGING.113.000899
- 79. R.J. Kim RJ, D.S. Fieno, T.B. Parrish, K. Harris, E.L. Chen, O. Simonetti, J. Bundy, J.P. Finn, F.J. Klocke, R.M. Judd: Relationship of MRI delayed contrast enhancement to irreversible injury, infarct age, and contractile function. *Circulation* 100, 1992–2002 (1999) DOI: 10.1161/01.CIR.100.19.1992
- 80. G. Fragasso, S.L. Chierchia, G. Pizzetti, E. Rossetti, M. Carlino, S. Gerosa, O. Carandente, A. Fedele, N. Cattaneo: Impaired left ventricular filling dynamics in patients with angina and angiographically normal coronary arteries: effect of beta adrenergic blockade. *Heart* 77, 32–39 (1997)
 DOI: 10.1136/hrt.77.1.32
- 81. F. Leonardo, G. Fragasso, E. Rossetti, P. Dabrowski, P. Pagnotta, G.M. Rosano, S.L. Chierchia: comparison of trimetazidine with atenolol in patients with syndrome X: effects on diastolic function and exercise tolerance. *Cardiologia* 44, 1065–1069 (1999)

- 82. P.J. Oliveira, J.A. Bjork, M.S. Santos, R.L. Leino, M.K. Froberg, A.J. Moreno, K.B. Wallace: Carvedilol-mediated antioxidant protection against doxorubicin-induced cardiac mitochondrial toxicity. *Toxicol Appl Pharmacol* 200, 159–168 (2004) DOI: 10.1016/j.taap.2004.04.005
- 83. P.Georgakopoulos, P.Roussou, E.Matsakas, A. Karavidas, N. Anagnostopoulos, T. Marinakis, A. Galanopoulos, F. Georgiakodis, S. Zimeras, M. Kyriakidis, A. Ahimastos: Cardioprotective effect of metoprolol and enalapril in doxorubicin-treated lymphoma patients: a prospective, parallel-group, randomized, controlled study with 36-month follow-up. *Am J Hematol* 85, 894–896 (2010) DOI: 10.1002/ajh.21840
- 84. J.Y Choe, A.B. Combs, K. Folkers: Potentiation of the toxicity of Adriamycin by propranolol. *Res Commun Chem Pathol Pharmacol.* 21, 577–580 (1978)
- 85. J.C. Kaski, G. Rosano, S. Gavrielides, L. Chen: Effects of angiotensin-converting enzyme inhibition on exercise-induced angina and ST segment depression in patients with microvascular angina. *J Am Coll Cardiol* 23, 652–657 (1994) DOI: 10.1016/0735–1097(94)90750–1
- 86. P.G. Camici, P. Marracini, R. Gistri, P.A. Salvadori, O. Sorace, A. L'Abbate: Adrenergically mediated coronary vasoconstriction in patients with syndrome X. Cardiovasc Drugs Ther 8, 221–226 (1994) DOI: 10.1007/BF00877330
- 87. M.A. Abd El-Aziz, A.I. Othman, M. Amer, M.A. El-Missiry: Potential protective role of angiotensin-converting enzyme inhibitors captopril and enalapril against adriamycin-induced acute cardiac and hepatic toxicity in rats. *J Appl Toxicol* 21, 469–73 (2001) DOI: 10.1002/jat.782
- 88. A. Hiona, A.S. Lee, J. Nagendran, X. Xie, A.J. Connolly, R.C. Robbins, J.C. Wu: Pretreatment with angiotensin-converting enzyme inhibitor improves doxorubicininduced cardiomyopathy via preservation of mitochondrial function. *J Thorac Cardiovasc Surg* 142, 396–403 (2011)
 DOI: 10.1016/j.jtcvs.2010.07.097
- 89. H. Buikema, S.H. Monmink, R.A. Tio, H.J. Crijns, D. de Zeeuw, W.H. Gilst: Comparison

of zofenopril and lisinopril to study the role of the sulfhydryl-group in improvement of endothelial dysfunction with ACE-inhibitors in experimental heart failure. Br J Pharmacol, 1999–2007 (2000)

DOI: 10.1038/sj.bjp.0703498

- 90. C. Cadeddu, A. Piras, G. Mantovani, M. Deidda, M. Dessì, C. Madeddu, E. Massa, G. Mercuro: Protective effects of the angiotensin II receptor blocker telmisartan on epirubicin-induced inflammation, oxidative stress, and early ventricular impairment. *Am Heart J* 160, 487 (2010) DOI: 10.1016/j.ahj.2010.05.037
- E. Fábián, A. Varga, E. Picano, Z. Vajo, A. Rónaszéki, M. Csanády: Effect of simvastatin on endothelial function in cardiac syndrome X patients. *Am J Cardiol* 94, 652–655 (2004)
 DOI: 10.1016/j.amjcard.2004.05.035
- C. Pizzi, O. Manfrini, F. Fontana, R. Bugiardini: Angiotensin-converting enzyme inhibitors and 3-hydroxy-3-methylglutaryl coenzyme A reductase in cardiac syndrome X: role of superoxide dismutase activity. *Circulation* 109, 53–58 (2004) DOI: 10.1161/01.CIR.0000100722.34034.E4
- J. Huelsenbeck, C. Henninger, A. Schad, K.J. Lackner, B. Kaina, G. Fritz: Inhibition of Rac1 signaling by lovastatin protects against anthracyclineinduced cardiac toxicity. *Cell Death Dis* 2, e190 (2011) DOI: 10.1038/cddis.2011.65
- 94. S. Seicean, A. Seicean, J.C. Plana, G.T. Budd, T.H. Marwick TH: Effect of statin therapy on the risk for incident heart failure in patients with breast cancer receiving anthracycline chemotherapy: an observational clinical cohort study. *J Am Coll Cardiol* 60, 2384–2390 (2012) DOI: 10.1016/j.jacc.2012.07.067
- P.A. Kaufmann, T. Gnecchi-Ruscone, M. di Terlizzi, K.P. Schäfers, T.F. Lüscher, P.G. Camici Coronary heart disease in smokers: vitamin C restores coronary microcirculatory function. *Circulation* 102, 1233–1238 (2000)
 DOI: 10.1161/01.CIR.102.11.1233
- 96. R. Solito, F. Corti, C.H. Chen, D. Mochly-Rosen, A. Giachetti, M. Ziche, S. Donnini: Mitochondrial aldehyde dehydrogenase-2 activation prevents β-amyloidinduced endothelial cell dysfunction

and restores angiogenesis. *J Cell Sci* 126, 1952–1961 (2013) DOI: 10.1242/jcs.117184

- 97. C.H. Chen, G.R. Budas, E.N. Disatnik, T.D. Hurley D. Mochly-Rosen: Activation of aldehyde dehydrogenase-2 reduces ischemic damage to the heart. *Science* 21, 1493–1495 (2008) DOI: 10.1126/science.1158554
- 98. W. Ge, M. Yuan, A.F. Ceylan, X. Wang, J. Ren: Mitochondrial aldehyde dehydrogenase protects against doxorubicin cardiotoxicity through a transient receptor potential channel vanilloid 1-mediated mechanism. *Biochim Biophys Acta*, 1862(4):622–34 (2016)

DOI: 10.1016/j.bbadis.2015.12.014

Abbreviations: ALDH-2: aldehyde dehydrogenase-2; CAD: coronary artery disease; CFR = coronary flow reserve; CMD: coronary microvascular dysfunction; CMR: cardiac magnetic resonance; CT: chemotherapy; ET-1: endothelin-1; 5-FU: 5-fluorouracil; HF: heart failure; MBF: myocardial blood flow; miRNA: microRNA; MVA: microvascular angina: MVO: microvascular occlusion: NO: nitric oxide; PET: positron emission tomography; PKC: protein-kinase C; RAAS: renin angiotensin aldosterone system; ROS: reactive oxygen species; RT: radiotherapy; VEGF: vascular endothelial growth factor; VSMCs: vascular smooth muscle cells

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