

Molecular basis for prostate cancer racial disparities

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1. ABSTRACT

Prostate cancer (PCa) remains the most common cancer in American men. African-American (AA) men continue to have higher PCa prevalence and mortality rates compared to men in other populations. In addition to socioeconomic factors and lifestyle differences, molecular alterations contribute to this discrepancy. We summarize molecular genetics research results interrelated with the biology of PCa racial disparity. Androgen and androgen receptor (AR) pathways have long been associated with prostate growth. Racial differences have also been found among variants of genes of the enzymes involved in androgen biosynthesis and metabolism. Growth

factors and their receptors are a potential cause of the disparity in PCa. Recent molecular and biotechnological approaches in the field of proteomics and genomics will greatly aid the advancement of translational research on racial disparity in PCa, which may help, in finding new prognostic markers and novel therapeutic approaches for the treatment of PCa in AA.

2. INTRODUCTION

Prostate cancer (PCa) is the second most common cause of death from malignancy in American

men. The incidence of PCa varies widely between ethnic populations and countries. In 2015, approximately 220,800 cases of prostate cancer were newly diagnosed; 27,540 PCa deaths were estimated in the United States. PCa is the most commonly diagnosed cancer among African American (AA) men. In AA, there is a higher incidence of PCa at a younger age with poor prognosis than men of other ethnicities (1-3). It is estimated that one in five AA men will be diagnosed with prostate cancer in their lifetime (4). Along with positive family history and older age, African ancestry has long been recognized as an important risk factor for PCa (5, 6). The underlying reasons for this disparity are not well understood, although existing evidence implicates important genetic components. While it has been argued that racial variation may be largely due to lifestyle, dietary, socioeconomic (7, 8), or clinical factors, these cannot fully explain the discrepancy (1-3, 9) or the results of migration studies, and consequently, genetic parameters may be important. Studies of the pathology and recurrence of tumors in AA and Caucasian American (CA) men have suggested that racial differences in the biology of PCa tumors may explain observed differences in outcome (10, 11).

Although there are some effective treatment approaches for clinically localized PCa through surgery and radiotherapy, however, the metastatic PCa remains incurable. The metastatic potential of tumor cells and its possible dissemination to secondary sites are critical factors related to its mortality rates. In spite of the high incidence and mortality rates, the molecular mechanisms involved in oncogenesis and progression to PCa are still poorly understood, especially related to the progression to the metastatic form. PCa etiology remains obscure and its tumors vary from indolent forms with low progression rates to extremely aggressive with rapid growth rates. Several methods have been used for the study of oncogenes expression, characterization, chromosomal aberrations and heredity loci in this neoplastic disease (12, 13). In the current review, we are highlighting recent research on differential androgen levels in prostate cancer development in ethnic population.

3. DIFFERENTIAL ANDROGEN LEVELS IN PROSTATE CANCER DEVELOPMENT IN ETHNIC POPULATION

Androgens and the androgen receptor pathway constitute the most intensely studied field in PCa. Several aspects of the pathway are related to the racial disparity of PCa.

3.1. Androgen receptors (AR)

Prostate cells require androgen stimulation in the form of testosterone and 5 α -dihydrotestosterone (DHT-A) for normal growth and maintenance. The AR is the key receiver of androgen signal. After formation of the complex AR- androgen, it translocated to the nucleus and

acts in the transcription of androgen response genes (14). AR- androgen complex regulates the expression of genes necessary for the growth and development of both normal and malignant prostate tissue. Immunohistochemical studies of malignant and benign prostate tissue from AA and Caucasian American (CA) men who underwent radical prostatectomy for PCa, showed expression of AR protein was 22% higher in the benign prostate and 81% higher in PCa of AA patients (15). This suggests that differences in androgenic stimulation may play an important role in racial disparity.

The AR gene is over 90 kb in length, located within chromosome Xq11-12 and has eight exons. Exon 1 encodes the N-terminal (transactivation) domain, which controls transcriptional activation of the receptor as well as two polymorphic trinucleotide repeats (CAG and GGC), which code for polyglutamine and polyglycine tracts, respectively, in the N-terminal domain. Studies have shown that CAG repeat varies in length from 11 to 31 repeats in normal men (16), and an inverse relationship has been demonstrated between CAG repeat length and AR transcriptional activation ability (17). Short CAG and GGC repeat lengths are associated with an increased risk of developing PCa (18-20), specifically individuals with CAG repeat length less than 20 and GGC repeat length less than 16 (18, 20-22). AA men populations have significantly shorter repeat length than CA men (17, 23, 24).

Prostate cancer may occur at a younger age and progress more rapidly in AA than CA because of racial differences in androgenic stimulation of the prostate. CWR-22 xenograft model of androgen regulated genes include CDK1 and CDK2, cyclin A and B1 (25); α -enolase, α -tubulin, Ikb α , IGFBP-5 (26), PSA, hK-2, Nkx3.1., ARA-70 (27); EF-1 α , tomoregulin, TRX-R1, and Mxi-1 are expressed at higher levels in recurrent and androgen-dependent tumors with higher proliferation rate in PCa (27, 28).

3.2. Serum androgen levels

Although there is no clear relationship between circulating androgen levels and PCa (29-31), high levels of androgens have long been considered as risk factor (29, 32). Racial differences in maternal androgens during gestation, serum androgens and time of puberty during adolescence may imprint the AA prostate to respond more to similar levels of androgenic stimulation in adulthood. AA men (aged 31 to 50) have higher mean serum testosterone level (15%) (33) than CA men (34). However, elevated testosterone and dihydrotestosterone (DHT) have not been persuasively shown to increase the risk of PCa (35).

3.3. Tissue androgens

Testosterone is the major circulating androgen, and DHT is 1/10 the concentration of testosterone in

serum. DHT is the major intra-prostatic androgen. In prostate tissues 5 α -reductase converts testosterone to DHT. DHT is a preferred ligand for the AR, since AR–DHT complex is more stable than the AR–testosterone complex. Also androgen levels are measured for radical prostatectomy from clinically localized PCa (37).

Steroid hormones analysis from snap frozen tissue obtained intra-operatively from radical prostatectomy specimens from AA and CA showed similar levels of testosterone, DHT, dehydroepiandrosterone, dehydroepiandrosterone sulfate, and PSA. However, AA had higher androstenedione (ASD) and sex hormone-binding globulin (SHBG) levels. Age, BMI, PSA, and pathologic gleason sum and stage (nonparametric rank analysis) make the racial differences between ASD and SHBG. A higher ASD tissue level in AA is not the result of higher levels of testosterone whereas higher serum levels of SHBG have been postulated to be protective against prostate cancer. SHBG decreases bioavailable testosterone for androgen receptor ligand activation. Although AA has been reported to have higher serum levels of SHBG than CA (36), but SHBG serum levels have not correlated significantly with risk of prostate cancer (37, 38).

More recently, SHBG has been demonstrated to be produced in the human PCa cell lines (LNCaP, DU-145, and PC3), and in cultured human prostate epithelial and stromal cells (27). SHBG binds to a membrane receptor in prostate (39), which was reported to initiate an intracellular signal that increased cAMP levels and modulated androgen action in the prostate (40). Immunostaining revealed heterogeneous expression of SHBG protein, primarily in the epithelium of both benign prostate (27) and prostate cancer (41). Also *in situ* hybridization of adjacent sections confirmed local synthesis of SHBG (27). Higher tissue levels of SHBG may enhance androgen action in prostate tissue (42) of AA through cAMP-dependent pathways.

4. GENES INVOLVED IN ANDROGEN BIOSYNTHESIS

Variants in the genes involved in androgen biosynthesis and metabolism are compelling candidates for susceptibility factors in PCa pathogenesis. Many of these genes have been found to harbor genetic polymorphisms. These polymorphisms can potentially change androgen levels in prostate tissue. Genetic polymorphisms in these pathways are summarized in Figure 1.

4.1. CYP11A1

CYP11A1 gene is the first rate-limiting step for biosynthesis of both testosterone and estrogen, located on 15q23–q24 chromosome encodes for the enzyme P450_{scc}, and has pentanucleotide (TAAAA) *n* repeat range (4 to 10-repeat sequences) at 5'-untranslated region (UTR) (60), which catalyzes cholesterol to

pregnenolone. Population based studies have found higher prevalence of a 6-repeat allele in Japanese populations compared with the higher prevalence of a 4-repeat allele in European and African populations (43, 44). Interestingly, Japanese PCa patients without the 4-repeat allele have an increased risk of metastatic tiPCa compared to those with the 4-repeat allele (44). However, a positive association with PCa risk was not identified in the European populations (45).

4.2. CYP17

CYP17 gene is located on chromosome 10, encodes the cytochrome P450_{c17a} enzyme (46), which mediates both 17 α -hydroxylase and 17,20-lyase activities in testosterone biosynthesis in the gonads and adrenals (46). The 5'-untranslated promoter region of *CYP17* contains a polymorphic T-to-C substitution that gives rise to A1 (T) and A2 (C) alleles (47). Previous studies reported that the A2 allele may be associated with an increased risk of PCa (48–53); however, other results have either been inconclusive (54, 55) or showed a possible increased risk from the A1 allele (56, 57). The results of a meta-analysis suggest that *CYP17* polymorphisms may have a role in PCa susceptibility in AA but not CA men (58). The A2 allele was slightly less frequent in AA versus CA men, but a different study had the opposite finding (59). Ultimately, there may be little difference in A2 frequency and a null effect of the *CYP17* polymorphism on androgen levels.

4.3. SRD5A2

Intra-prostatic DHT levels may be integral to racial variations in risk (60). Steroid 5 α -reductase irreversibly converts testosterone into DHT. Two forms of steroid 5 α -reductase exist, steroid 5 α -reductase type 1 (*SRD5A1*) and steroid 5 α -reductase type 2 (*SRD5A2*). *SRD5A1* is expressed more abundantly in extra-prostatic tissues (e.g., skin), whereas *SRD5A2* is exclusively expressed in the prostate (61). Activity of 5 α -reductase was reported lower in Asian than in white and black men (60, 62). This gene is more polymorphic in AA men than previously assumed (63), though *SRD5A2* TA repeat alleles are only present in high-risk AA men and not in lower risk CA and Asian men (60, 62). Thus, it has been proposed that certain steroid 5 α -reductase enzyme variants encoded by *SRD5A2* genes marked by particular TA repeat alleles may result in an elevation of enzyme activity, leading to an increased prostatic level of DHT, which may increase the risk for developing PCa. Furthermore, the V89L and A49T variants of *SRD5A2* gene have been shown to alter the conversion of testosterone to DHT (64, 65). While the V89L polymorphism is decreased the production of DHT (66) and A49T variant increases its production, particularly in AA and Hispanic men (67).

4.4. HSD3B family

The *HSD3B1* and *HSD3B2* genes are located on 1p13.1., and encode 3 β -hydroxysteroid dehydrogenase/



Figure 1. Gene involved in Androgen biosynthesis and metabolism in prostate cancer. Genetic polymorphisms in genes associated with androgen biosynthesis/metabolism and AR, which show differential racial frequencies and potential association with prostate cancer. Red star indicates the genes, which has racial frequency difference.

$\Delta 5$ - $\Delta 4$ isomerase 1 and 2 isoenzymes (3β -HSD types 1 and 2). The proteins are bifunctional enzymes that catalyze androstenedione production in steroidogenic tissues and convert the active DHT into inactive metabolites in steroid target tissues (68). A N367T (AAC>ACC, rs1047303) polymorphism in *HSD3B1* has been reported to present at a high frequency in Caucasian (31%), inter-medium frequency in AA (11.7.%) and low frequency in Asian men (8.5.%), although the variant has a similar activity to the wild type (69). A complex (TG)_n (TA)_n(CA)_n dinucleotide repeat polymorphism is found in intron 3 of the *HSD3B2* gene (70). The common alleles occurred at variable frequencies in different racial populations. The longer allele formed more stable hairpin structures with faster degradation rate of DHT. Longer alleles commonly found in Asian men (71, 72), whereas short alleles have been found to be associated with an increased PCa risk in Caucasian but not in AA men (71). There are two SNPs in *HSD3B2*,

rs1819698 and rs1538989 reported which is more common in AA than CA and increased the risk of PCa in AA but not CA (77). The interaction between *HSD3B1* and *HSD3B2* polymorphisms has also been investigated. Although the N367T polymorphism in *HSD3B1* is weakly associated with PCa risk, but the combination with *HSD3B2* rs1819698 is greatly enhanced the association (73).

4.5. CYP19A1

The *CYP19A1* gene is located on chromosome 15q21.1., and encodes the enzyme aromatase, which catalyzes the irreversible conversion of C19 androgens, androstenedione and testosterone, to the C18 estrogens, estrone and estradiol respectively. More than 30 SNPs have been detected in different populations. Several SNPs (rs2470152, rs749292, rs727479) are confirmed to be associated with serum estradiol of men (74, 75). The polymorphisms of *CYP19* gene in Caucasian and AA men

Table 1. *CYP3A* family genes and their allelic functions

Gene	Characteristic	Reference
<i>CYP3A4</i>	• Cytochrome P4503A4 (<i>CYP3A4</i>) protein facilitates the oxidative deactivation of testosterone to biologically less active metabolites, and the inhibition of <i>CYP3A4</i> causes increased levels of testosterone. • <i>CYP3A4</i> supports oxidative metabolism of finasteride and may be a effective molecule in PCa treatment. • Germline genetic variation in the 5' regulatory region of the <i>CYP3A4</i> gene (A to G transition) on chromosome 7 has been reported. This variant G allele (referred to as <i>CYP3A4</i> G variant) is more common among AA men (gene frequency>50%) than CA (<10%), Hispanic, or Asian men. • The G variant is inversely associated with less aggressive PCa. ., and the <i>CYP3A4</i> variant is strongly associated with AA population.	(167-169) (170, 171) (172)
<i>CYP3A5</i>	• <i>CYP3A5</i> expressed at high levels in the non-tumoral prostate tissue, specifically in the basolateral cells and catalyzes 6β-hydroxylation of testosterone. An A to G transition (A6986G) within intron 3 leads to a variant in the <i>CYP3A5</i> mRNA expression in human prostatic tissue. Allele <i>CYP3A5*1</i> (A allele) produces a correctly spliced transcript leading to high levels of full-length <i>CYP3A5</i> mRNA and protein, however allele <i>CYP3A5*3</i> (rs776746, G allele) creates a cryptic splice site leading to the inclusion of a novel exon, and ultimately a premature stop codon. • <i>CYP3A5*1</i> has a higher frequency in AA individuals than Caucasian or Asian men. • <i>CYP3A5*3*3</i> decreases <i>CYP3A5</i> mRNA content 13-fold compared to <i>CYP3A5*1*3</i>. <i>CYP3A5*1</i> and show linkage disequilibrium with <i>CYP3A4*1B</i> in Caucasian and African men, • <i>CYP3A4*1B/CYP3A5*1</i> haplotype is inversely associated with risk among Caucasian men with less aggressive disease. • In Japanese population, <i>CYP3A5*1*1</i> men have lower risk of developing a low-grade localized PCa than <i>CYP3A5*3*3</i> carried men. <i>CYP3A5*3</i> is not associate with PCa in either white or African men, the <i>CYP3A4*1B/CYP3A5*3</i> haplotype is significantly associated with increasing PCa risk in European American but not in AA men. • <i>CYP3A5</i> interacts with <i>SRD5A2</i> or <i>KLK3</i>, which influenced the development of PCa.	(173) (170, 174) (175, 176)
<i>CYP3A43</i>	• <i>CYP3A43</i> is predominantly expressed in the prostate. The allelic frequency of <i>CYP3A43*3</i> (rs680055) is higher in AA than CA men. • There is a 2.6.-fold increase in PCa risk among individuals with the <i>CYP3A43*3</i> homozygous genotype compared with those with the <i>CYP3A43*1</i> homozygous genotype in AA, but not in CA men.	(177, 178) (179)

(rs2470152, rs12439137, rs3751592, rs2470164) are associated with PCa risk (77). Particularly, the rs2470164 was reported to increase PCa risk in Caucasian men with different frequency among healthy Caucasian (50%) and AA men (5.6.%). The tetranucleotide repeat (TTTA) n is located in intron 4 of *CYP19A1*. The TTTA repeat numbers range from 7 to 13 and are designated as A1 to A7 according to the repeat number. Most studies for this polymorphism are among Asian men. In Asian men, A1 is found more frequently (~50% of the population) than all other alleles (76, 77). Several studies have shown that TTTA repeat length is associated with PCa risk (57, 80, 81, 82), however, TTTA repeat length is not associated with the American population (80). The polymorphism Arg264Cys substitution (rs700519) is found at a higher frequency in Indian men (27%) (78) in comparison to AA (16.8.%) and Caucasian men (4-8.1.%) (79-81). Studies among Caucasian and Indian men showed a tendency for this polymorphism to increase risk of PCa (78, 81), but large scale population studies failed to confirmed the results in Caucasian men (75, 79, 80).

4.6. *CYP3A* family

Cytochrome P450 3A (*CYP3A*) enzymes hydroxylate testosterone and dehydroepiandrosterone to less active metabolites. The *CYP3A* locus consists

of four genes in humans, *CYP3A4*, *CYP3A5*, *CYP3A7* and *CYP3A43*, all of which reside in a 231 kb region of chromosome 7q21-22.1 (82) (see details in Table 1).

4.7. *HSD17B* family

The 17 β -hydroxysteroid dehydrogenases (17 β -HSDs) are involved in regulation of estrogens and androgens by catalyzing the reduction of 17-ketosteroids or the oxidation of 17 β -hydroxysteroids. 17 β -HSD1 is encoded by *HSD17B1* gene located on 17q21 and involved in estrogen and testosterone biosynthesis. The polymorphism of *HSD17B1* (Ser313Gly, rs605059) is detected in Caucasian men (40%), and associated with either familial or sporadic cases of PCa (83). In studies from multi-ethnic groups (The Breast and Prostate Cancer Cohort Consortium, BPC3), four common SNPs (rs676387, rs605059, rs598126, rs2010750) were detected, although none were found to be associated with PCa risk, however, only varying frequencies of haplotypes between different races. The haplotype CAAC is commonly found in AA men and CAGC is more prevalent in white and black than Asian men, whereas inverse association with PCa risk in Latino and Japanese American but not in AA, Native Hawaiian, or white men (83).

17 β -HSD2 is encoded by *HSD17B2* and located on 16q24. 17 β -HSD2 is involved in the conversion of

Table 2: Growth receptors and their characteristics in the prostate cancer health disparities

Growth receptors	Characteristics	References
EGFR (Epidermal growth factor receptor)	- EGFR helps in cellular proliferation, progression, tumor cell invasion and also a target of anticancer agents for androgen-independent PCa. - Over-expresses of EGFR in PCa is more common in African American (AA) than Caucasian American (CA). - EGFR inhibitors hinder the growth of both androgen dependent and independent PCa xenografts. - Androgen independent, metastatic PCa and androgen ablation increases the expression of EGFR Intronic dinucleotide repeats (CA) n (range from 14 to 21 correlated with transcriptional activity) polymorphism. - Longer allele of EGFR is associated with 80% reduction in EGFR protein expression than shorter allele. - Three out of four missense mutations in EGFR TK domain identified as oncogenic in nature. These mutations are reported in 3 Koreans, 1 in CA but none in AA.	(180-186)
EPHB2 (Ephrin type-B receptor 2)	- Located on 1p36, and link with hereditary PCa with racially diverse family. - Studies in DU145 PCa cell line suggested that it may be tumor suppressor gene. - Screening of the EphB2 gene for germline polymorphisms in AA PCa identified ten sequence variants in the gene, including a common nonsense mutation and K1019X. - The risk for PCa increased 3-fold among AA men who carried at least one copy of the K1019X allele and had a family history of PCa.	(187-189)
VDR (Vitamin D receptor)	- European American men have high plasma levels of the active form of vitamin D, 25-hydroxyvitamin D, which is associated with decreased risk of lethal PCa. - SNP sets linked to 7 vitamin D pathway-related genes, including VDR, which are associated with PCa risk. - A single VDR polymorphism, the <i>BsmI</i> B allele, is protective against recurrence of PCa in European American men but not in AA men. - The decrease VDR signaling contributes to the greater risk of advanced or lethal PCa in AA population.	(190-192)

active androgens into their less active forms. SNPs in HSD17B2 (rs1424151) are significantly associated with plasma testosterone level in Caucasian men (84), but no association with PCa was detected (84, 85).

17 β -HSD3 is encoded by *HSD17B3* and located on 9q22, catalyzes androstenedione to testosterone. The frequency of the G289S polymorphism (rs2066479) of *HSD17B3*, was 4.3-7.3.% in Caucasian men and was reported to significantly increase PCa risk in Italian men (86), but studies in Finnish and Swedish men found no positive associations (87).

HSD17B4 gene is located on 5q21 and encodes androgen/estrogen inactivating enzyme 17 β -HSD4. It was reported to be associated with the outcome of PCa patients (88, 89). 17 β -HSD5 belongs to the aldo-keto reductase (AKR) superfamily and is formally known as AKR1C3 encoded by the *AKR1C3* gene located on 10p14-p15. It catalyzes the conversion of androstenedione to testosterone and DHT to androstenediol. An A to G substitution was identified in exon 2 that confers a Glu77 Gly (rs41306308) change, and occurred in 4.8.% of Caucasian men but was completely absent in Asian men, and the Glu77Gly polymorphism was associated with lower testosterone levels in serum (90). Moreover, promoter

polymorphism (A to G, rs3763676) of *AKR1C3* is more prevalent in Caucasian than Asian men (90), whereas men with the A allele have significantly lower risk of PCa (91).

4.8. UGT2B15

UGT2B15 is a member of UDP glucuronosyltransferases (UGTs) family, which glucuronidate steroids and other endogenous molecules, encoded by the *UGT2B15* gene and located on 14q13-q21.1. It has a high capacity to glucuronidate 3 α - androstenediol and a moderate capacity for DHT. A nonsense mutation in codon 85 (aspartate>tyrosine, D85Y, Asp85Tyr) has been identified in the *UGT2B15* gene. The 85Y variant associates with a 2-fold increase in activity for 3 α - androstenediol and DHT, it is likely to lead to lower androgen exposure compared with 85D. A study found that Asians have higher 85D allele frequency than Caucasians (92), however, other studies are inconclusive (93, 94).

5. OTHER FACTORS INVOLVE IN PROSTATE CANCER DEVELOPMENT

5.1. Sex hormone binding globulin

Sex hormone-binding globulin (*SHBG*) gene is located on 17p12-p13 and encodes a steroid binding

protein, and act as regulator of free plasma androgens. It also mediates androgen and estrogen signaling at the cell membrane via cyclic adenosine monophosphate. Most of the studies are reported that black men have higher PCa risk with higher plasma SHBG levels in prostate tissue than white and Asian men (95). Interestingly, the higher risk population have a higher SHBG level. A collaborative analysis of 18 prospective studies found the fifth highest serum SHBG levels had a relative PCa risk reduction of 14% when compared with the fifth lowest (96). A common polymorphism in the *SHBG*, and *D356N* gene encodes for an additional N-glycosylation consensus site, which may reduce its clearance from circulation and alter its binding to membrane receptors (97). In the multicenter study the *SHBG-D356N* heterozygotic polymorphism had a higher frequency in white men (17%) than black men (7.8%) (129). The *D356N* heterozygote is associated with increasing PCa risk in non-Hispanic white but not in black men. Studies carried out in British and US men reported no association found between PCa and *SHBG* polymorphisms (98).

5.2. Growth factors and receptors

In most studies growth factor receptors concerning the racial disparity of PCa are EGFR and EPHB2 (Table 2). In addition, AA men have been found to have higher IGF-1 and lower IGFB-3 levels, which may cause higher tumor growth with lower anti-tumor activity (99).

5.3. Differences in apoptotic genes in relation to prostate cancer racial disparity

5.3.1. Anti-apoptotic *BCL-2*

Studies show that altered expression of the *BCL-2* gene may be an important factor underlying the greater aggressiveness of PCa in AA men (100). *BCL-2* has a central role in preventing cancer cells from death, via its anti-apoptotic effect, and its up-regulation in AA men may be responsible for PCa cell survival and resistance to therapies. Thus, a positive connection between over-expression of *BCL-2* and increased in prostate tumors proliferation in AA but not in CA men, and suggest *BCL-2* dependent aggressive behavior of PCa in AA men (100).

5.3.2. *MDM2*

In response to stress, cells activate a complex pathway involving tumor suppressor gene p53 that is responsible for cell cycle arrest, DNA repair, and apoptosis as protection from the deleterious effects of mutation (101). *MDM2* is a key negative regulator of tumor suppressor p53, by targeting p53 for proteasomal degradation (102-104). Several reports suggested that *MDM2* overexpression was significantly associated with advanced stage PCa (105-107). Another studies have also shown that inhibiting *MDM2* expression enhances the effects of radiation and chemotherapy on PCa cells (108-110). A single nucleotide polymorphism in the *MDM2* promoter, SNP309, enhances transcriptional

activation of *MDM2* and has been associated with early onset of several types of cancer (111-115). Interestingly, *MDM2*, protein expression was significantly higher in CA (78%) than AA (45%) patients (116), and also *MDM2* and AA ethnicity have both been associated with poor prognosis. However, the relationship between these two variables was neither causative nor correlative.

5.4. Genetics variations between AA and CA prostate cancer

PCa is also caused by multiple genes through complex interaction including environmental factors (85, 117-119). There may be ethnic variation in the frequency of alleles that may be associated with PCa risk and/or progression. Although the incidence and mortality for PCa may differ among different racial groups, the increased risk for PCa attributed to family history of this disease is consistent across different racial backgrounds, supporting the possibility of a common genetic basis of disease (120). The analysis of genetic alterations in PCa is challenging because PCa often has genetic and morphological heterogeneity, multifocality and the presence of more than one lesion of independent origin (121-124). Linkage studies, determine the susceptibility loci for PCa on several chromosomes and several candidate genes if the tumors in AA men are different from those in CA men (125).

5.4.1. Chromosome 8

The short arm of chromosome 8 (8p22-23) has been proposed as a potential location for one or more genes important in the development of PCa (126, 127). The short arm of chromosome 8 is frequently deleted in both adenocarcinomas and PINs (128, 129), which has lead to the assumption that the inactivation of an unidentified tumor suppressor gene on 8p is involved in prostate tumor initiation (130-132). Other studies on the loss of chromosome 8p in AA and CA men have generated conflict findings (133), however, the racial differences in the association between PCa recurrence and several prognostic factors of cancer progression, including Gleason score, surgical margin, and TNM stage are positively correlated.

5.4.2. miRNA

MicroRNAs (miRNAs) are a class of small, endogenous, non-coding RNAs that regulate gene expression at the levels of transcription and post-transcription (134, 135). miRNA inhibits translation of target genes involved in a variety of fundamental cellular processes including organ development, differentiation, and cancer formation (136-139). Functional studies of individual miRNAs have shown that miRNAs can act as oncogenes or tumor suppressor genes (140-144). miRNAs are differently dysregulated in uterine leiomyomas in both AA and CA women, indicating that miRNAs expression are associated with the racial disparity in cancer (145). In the prostate, the expression of 5 miRNAs, miR- 30c,

miR-301, miR-219, miR-261, and miR-1b1, were reported racially different in benign prostate tissue (146). Expression of commonly dysregulated miRNAs in PCA revealed racial differences in the expression of let-7c and miR30c in AA prostate tissue (147). Thus, distinct gene expression and genome-wide copy number variation between AA and CA prostate cancer might play a role in the racial disparity of PCA.

5.5. PSA levels in AA and CA prostate cancer patients

FDA approved the PSA test in the year 1986 for monitoring the status of disease and diagnosis (1992). The test is performed on symptomatic and asymptomatic men in an effort to diagnosed PCA early and to monitor disease recurrence and progression (148). Past surveys of urologists revealed significant variation in the use of the PSA test (149), including racial disparities in PSA surveillance, with AA men half as likely as CA men to receive annual monitoring (150). A recent study concluded that PSA testing is probably not able to explain current racial differences in PCA mortality rates (151). Interestingly, recently a relationship was reported between serum PSA levels and polymorphisms in the PSA and AR genes (152). Specifically, serum PSA levels increased by 7% with each decreasing AR CAG repeats allele size among individuals homozygous for a single nucleotide polymorphism in the PSA gene promoter. A recent study of the ERDA1 locus revealed that large CAG repeats are more common among Asian populations, less common in populations of European ancestry, and least common in African populations (153). This pattern showed similarity with the AR trinucleotide repeats studies.

5.6. Familial prostate cancer susceptibility genes

Variation in genetic susceptibility also play a key role in the incidence of, and mortality from, prostate cancer in AA and CA men (154). Although the etiology of PCA is complex, the study on family and twin cases suggested that the inherited genetic susceptibility is a critical risk factor in PCA. PCA susceptibility genes have been mapped to 1q24–25 (155), Xq27–28 (156), and 8p22–23 (126), and other loci including PCaP and CaPB (157), HPC20 (158), and HPC2/ELAC (159), have been examined. The frequency of these genes and an understanding of their importance in hereditary prostate cancer cases, sporadic white American cases, and unaffected controls continues to evolve, but almost all reported studies contain few AA. Recently, more groups have begun to examine affected AA families. Familial aggregation rates were similar in AA and white Americans men (85). Race-specific penetrance estimates for the carriers of deleterious genotypes, which were similar in AA and white Americans, but low in Asian Americans (160). The higher incidences of prostate cancer in AA may not be caused by a higher prevalence of germline mutations predisposing to the disease in AA. Men with more than six

family members with prostate cancer had a higher chance of presenting with lymph node metastatic or widely metastatic prostate cancer than men with only four to six family members affected (161). Linkage analysis of 33 AA prostate cancer families from two independent research groups provided some evidence for clustering at HPC1. Increased evidence of linkage was found in families with prostate cancer diagnosis younger than age 65 years, and male-to-male transmission (162). These studies, though very preliminary, suggest that genetics could contribute to the increased aggressiveness of prostate cancer in AA.

5.7. Other genes in prostate cancer

5.7.1. *MSR1*

Recently, the macrophage scavenger receptor 1 (*MSR1*) gene has been proposed as a link between germline alterations in 8p and PCA (163, 164). Both common sequence variants and rare germline mutations have been suggested as potential PCA susceptibility factors. Several rare germline mutations of the *MSR1* gene were found to co-segregate with PCA, and one of the germline mutations was associated with an increased risk of PCA among AA men (163). In a subsequent study of CA men, the same authors examined five common sequence variants of *MSR1* and reported significantly different allele frequencies for each of the variants among men with PCA compared with unaffected men (164), with each, except INDEL7, associated with an elevated risk for PCA. An ensuing study examined each of these five common *MSR1* sequence variants in AA men (118). They found that the Asp174Tyr mutation is nearly twice as common among PCA patients compared with controls; however, after adjusting for age, none of the sequence variants were associated with a significantly increased risk of PCA, providing limited support for an association in AA men.

5.7.2. *Caveolin-1*

Caveolin-1 is a structural protein found in caveolae that support cholesterol transport and signal transduction. Caveolin-1 suppresses c-myc-mediated apoptosis, and is overexpressed in murine and human PCA. Caveolin-1 is expressed higher levels in PCA in AA than CA (165), and PCA grows more rapidly in AA due to reduced rates of apoptosis (165). Therefore, radiation and androgen-deprivation therapy are less effective for AA PCA patients (166).

6. CONCLUSIONS

Due to the complex nature of the AR signaling pathway, there are different ways that genetic polymorphisms contribute to the deregulation of this pathway and increase PCA risk. Future detailed studies need to include an integrated analysis of the combined effect of these polymorphisms on the AR pathway as well as androgen metabolism/biosynthesis in the development of PCA. Analysis of these complex polymorphisms in androgen synthesis and AR signaling pathway with

racial disparities are causing difficult situation at clinical front to achieve functional confirmation to establish their contribution to PCa development. Recent molecular and biotechnological approaches in the field of proteomics and genomics will greatly aid the advancement of translational research on racial disparity in PCa, which may help, in finding new prognostic markers and novel therapeutic approaches for the treatment of PCa in AA.

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8. REFERENCES

1. M. P. Cotter, R. W. Gern, G. Y. Ho, R. Y. Chang and R. D. Burk: Role of family history and ethnicity on the mode and age of prostate cancer presentation. *Prostate*, 50(4), 216-21 (2002)
DOI: 10.1002/pros.10051
PMid:11870799
2. R. M. Hoffman, F. D. Gilliland, J. W. Eley, L. C. Harlan, R. A. Stephenson, J. L. Stanford, P. C. Albertson, A. S. Hamilton, W. C. Hunt and A. L. Potosky: Racial and ethnic differences in advanced-stage prostate cancer: the Prostate Cancer Outcomes Study. *J Natl Cancer Inst*, 93(5), 388-95 (2001)
DOI: 10.1093/jnci/93.5.388
PMid:11238701
3. E. A. Platz, E. B. Rimm, W. C. Willett, P. W. Kantoff and E. Giovannucci: Racial variation in prostate cancer incidence and in hormonal system markers among male health professionals. *J Natl Cancer Inst*, 92(24), 2009-17 (2000)
DOI: 10.1093/jnci/92.24.2009
PMid:11121463
4. C. Desantis, D. Naishadham and A. Jemal: Cancer statistics for African Americans, 2013. *CA Cancer J Clin* (2013)
5. H. Gronberg: Prostate cancer epidemiology. *Lancet*, 361(9360), 859-64 (2003)
DOI: 10.1016/S0140-6736(03)12713-4
6. D. J. Schaid: The complex genetic epidemiology of prostate cancer. *Hum Mol Genet*, 13 Spec No 1, R103-21 (2004)
7. X. L. Du, S. Fang, A. L. Coker, M. Sanderson, C. Aragaki, J. N. Cormier, Y. Xing, B. J. Gor and W. Chan: Racial disparity and socioeconomic status in association with survival in older men with local/regional stage prostate carcinoma: findings from a large community-based cohort. *Cancer*, 106(6), 1276-85 (2006)
DOI: 10.1002/cncr.21732
PMid:16475208
8. S. Reddy, M. Shapiro, R. Morton, Jr. and O. W. Brawley: Prostate cancer in black and white Americans. *Cancer Metastasis Rev*, 22(1), 83-86 (2003)
DOI: 10.1023/A:1022216119066
PMid:12716039
9. A. S. Robbins, A. S. Whittemore and D. H. Thom: Differences in socioeconomic status and survival among white and black men with prostate cancer. *Am J Epidemiol*, 151(4), 409-16 (2000)
DOI: 10.1093/oxfordjournals.aje.a010221
PMid:10695600
10. I. J. Powell: Prostate cancer in the African American: is this a different disease? *Semin Urol Oncol*, 16(4), 221-6 (1998)
11. A. S. Robbins, A. S. Whittemore and S. K. Van Den Eeden: Race, prostate cancer survival, and membership in a large health maintenance organization. *J Natl Cancer Inst*, 90(13), 986-90 (1998)
DOI: 10.1093/jnci/90.13.986
PMid:9665146
12. P. E. Li and P. S. Nelson: Prostate cancer genomics. *Curr Urol Rep*, 2(1), 70-8 (2001)
DOI: 10.1007/s11934-001-0028-6
PMid:12084298
13. D. Karan, M. F. Lin, S. L. Johansson and S. K. Batra: Current status of the molecular genetics of human prostatic adenocarcinomas. *Int J Cancer*, 103(3), 285-93 (2003)
DOI: 10.1002/ijc.10813
PMid:12471610
14. C. A. Heinlein and C. Chang: Androgen receptor in prostate cancer. *Endocr Rev*, 25(2), 276-308 (2004)
DOI: 10.1210/er.2002-0032
PMid:15082523
15. K. E. Gaston, D. Kim, S. Singh, O. H. Ford, 3rd and J. L. Mohler: Racial differences in androgen receptor protein expression in men with clinically localized prostate cancer. *J Urol*, 170(3), 990-3 (2003)
DOI: 10.1097/01.ju.0000079761.56154.e5
PMid:12913756

16. A. Edwards, H. A. Hammond, L. Jin, C. T. Caskey and R. Chakraborty: Genetic variation at five trimeric and tetrameric tandem repeat loci in four human population groups. *Genomics*, 12(2), 241-53 (1992)
DOI: 10.1016/0888-7543(92)90371-X
17. N. L. Chamberlain, E. D. Driver and R. L. Miesfeld: The length and location of CAG trinucleotide repeats in the androgen receptor N-terminal domain affect transactivation function. *Nucleic Acids Res*, 22(15), 3181-6 (1994)
DOI: 10.1093/nar/22.15.3181
PMid:8065934 PMCID: PMC310294
18. E. Giovannucci, M. J. Stampfer, K. Krithivas, M. Brown, D. Dahl, A. Brufsky, J. Talcott, C. H. Hennekens and P. W. Kantoff: The CAG repeat within the androgen receptor gene and its relationship to prostate cancer. *Proc Natl Acad Sci U S A*, 94(7), 3320-3 (1997)
DOI: 10.1073/pnas.94.7.3320
PMid:9096391 PMCID: PMC20367
19. D. O. Hardy, H. I. Scher, T. Bogenreider, P. Sabbatini, Z. F. Zhang, D. M. Nanus and J. F. Catterall: Androgen receptor CAG repeat lengths in prostate cancer: correlation with age of onset. *J Clin Endocrinol Metab*, 81(12), 4400-5 (1996)
20. E. A. Platz, E. Giovannucci, D. M. Dahl, K. Krithivas, C. H. Hennekens, M. Brown, M. J. Stampfer and P. W. Kantoff: The androgen receptor gene GGN microsatellite and prostate cancer risk. *Cancer Epidemiol Biomarkers Prev*, 7(5), 379-84 (1998)
21. S. A. Ingles, R. K. Ross, M. C. Yu, R. A. Irvine, G. La Pera, R. W. Haile and G. A. Coetzee: Association of prostate cancer risk with genetic polymorphisms in vitamin D receptor and androgen receptor. *J Natl Cancer Inst*, 89(2), 166-70 (1997)
DOI: 10.1093/jnci/89.2.166
PMid:8998186
22. J. L. Stanford, J. J. Just, M. Gibbs, K. G. Wicklund, C. L. Neal, B. A. Blumenstein and E. A. Ostrander: Polymorphic repeats in the androgen receptor gene: molecular markers of prostate cancer risk. *Cancer Res*, 57(6), 1194-8 (1997)
23. O. Sartor, Q. Zheng and J. A. Eastham: Androgen receptor gene CAG repeat length varies in a race-specific fashion in men without prostate cancer. *Urology*, 53(2), 378-80 (1999)
DOI: 10.1016/S0090-4295(98)00481-6
24. R. A. Irvine, M. C. Yu, R. K. Ross and G. A. Coetzee: The CAG and GGC microsatellites of the androgen receptor gene are in linkage disequilibrium in men with prostate cancer. *Cancer Res*, 55(9), 1937-40 (1995)
25. C. W. Gregory, R. T. Johnson, Jr., S. C. Presnell, J. L. Mohler and F. S. French: Androgen receptor regulation of G1 cyclin and cyclin-dependent kinase function in the CWR22 human prostate cancer xenograft. *J Androl*, 22(4), 537-48 (2001)
26. C. W. Gregory, D. Kim, P. Ye, A. J. D'Ercole, T. G. Pretlow, J. L. Mohler and F. S. French: Androgen receptor up-regulates insulin-like growth factor binding protein-5 (IGFBP-5) expression in a human prostate cancer xenograft. *Endocrinology*, 140(5), 2372-81 (1999)
DOI: 10.1210/en.140.5.2372
27. D. J. Hryb, A. M. Nakhla, S. M. Kahn, J. St George, N. C. Levy, N. A. Romas and W. Rosner: Sex hormone-binding globulin in the human prostate is locally synthesized and may act as an autocrine/paracrine effector. *J Biol Chem*, 277(29), 26618-22 (2002)
DOI: 10.1074/jbc.M202495200
PMid:12015315
28. J. L. Mohler, T. L. Morris, O. H. Ford, 3rd, R. F. Alvey, C. Sakamoto and C. W. Gregory: Identification of differentially expressed genes associated with androgen-independent growth of prostate cancer. *Prostate*, 51(4), 247-55 (2002)
DOI: 10.1002/pros.10086
PMid:11987153
29. A. Wolk, S. O. Andersson and R. Bergstrom: Prospective study of sex hormone levels and risk of prostate cancer. *J Natl Cancer Inst*, 89(11), 820 (1997)
DOI: 10.1093/jnci/89.11.820
PMid:9182984
30. K. J. Pienta, J. A. Goodson and P. S. Esper: Epidemiology of prostate cancer: molecular and environmental clues. *Urology*, 48(5), 676-83 (1996)
DOI: 10.1016/S0090-4295(96)00219-1
31. A. W. Meikle and J. A. Smith, Jr.: Epidemiology of prostate cancer. *Urol Clin North Am*, 17(4),

- 709-18 (1990)
32. N. M. Makridakis and J. K. Reichardt: Molecular epidemiology of hormone-metabolic loci in prostate cancer. *Epidemiol Rev*, 23(1), 24-9 (2001)
DOI: 10.1093/oxfordjournals.epirev.a000791
PMid:11588850
33. L. Ellis and H. Nyborg: Racial/ethnic variations in male testosterone levels: a probable contributor to group differences in health. *Steroids*, 57(2), 72-5 (1992)
DOI: 10.1016/0039-128X(92)90032-5
34. R. Ross, L. Bernstein, H. Judd, R. Hanisch, M. Pike and B. Henderson: Serum testosterone levels in healthy young black and white men. *J Natl Cancer Inst*, 76(1), 45-8 (1986)
35. E. D. Crawford: Epidemiology of prostate cancer. *Urology*, 62(6 Suppl 1), 3-12 (2003)
36. A. H. Wu, A. S. Whittemore, L. N. Kolonel, E. M. John, R. P. Gallagher, D. W. West, J. Hankin, C. Z. Teh, D. M. Dreon and R. S. Paffenbarger, Jr.: Serum androgens and sex hormone-binding globulins in relation to lifestyle factors in older African-American, white, and Asian men in the United States and Canada. *Cancer Epidemiol Biomarkers Prev*, 4(7), 735-41 (1995)
37. P. H. Gann, C. H. Hennekens, J. Ma, C. Longcope and M. J. Stampfer: Prospective study of sex hormone levels and risk of prostate cancer. *J Natl Cancer Inst*, 88(16), 1118-26 (1996)
DOI: 10.1093/jnci/88.16.1118
PMid:8757191
38. R. Heikkila, K. Aho, M. Heliovaara, M. Hakama, J. Marniemi, A. Reunanen and P. Knekt: Serum testosterone and sex hormone-binding globulin concentrations and the risk of prostate carcinoma: a longitudinal study. *Cancer*, 86(2), 312-5 (1999)
DOI: 10.1002/(SICI)1097-0142(19990715)86:2<312::AID-CNCR15>3.0.CO;2-7
39. V. D. Ding, D. E. Moller, W. P. Feeney, V. Didolkar, A. M. Nakhla, L. Rhodes, W. Rosner and R. G. Smith: Sex hormone-binding globulin mediates prostate androgen receptor action via a novel signaling pathway. *Endocrinology*, 139(1), 213-8 (1998)
DOI: 10.1210/endo.139.1.5681
PMid:9421417
40. D. J. Hryb, M. S. Khan, N. A. Romas and W. Rosner: The control of the interaction of sex hormone-binding globulin with its receptor by steroid hormones. *J Biol Chem*, 265(11), 6048-54 (1990)
41. T. Usui, T. Ishibe, H. Nakatsu, M. Nakahara, H. Nihira and Y. Miyachi: Immunohistochemical localization of testosterone-estradiol-binding globulin in a clinically hormone-independent prostatic carcinoma. *Urol Int*, 38(1), 16-8 (1983)
DOI: 10.1159/000280854
PMid:6340306
42. S. M. Kahn, D. J. Hryb, A. M. Nakhla, N. A. Romas and W. Rosner: Sex hormone-binding globulin is synthesized in target cells. *J Endocrinol*, 175(1), 113-20 (2002)
DOI: 10.1677/joe.0.1750113
PMid:12379495
43. I. Piras, A. Falchi, P. Morai, A. Melis, L. Giovannoni, G. Paoli, C. Calo, G. Vona and L. Varesi: Frequencies of promoter pentanucleotide (TTTAA)n of CYP11A gene in European and North African populations. *Genet Test*, 12(1), 93-6 (2008)
DOI: 10.1089/gte.2007.0060
PMid:18307388
44. T. Kumazawa, N. Tsuchiya, L. Wang, K. Sato, T. Kamoto, O. Ogawa, A. Nakamura, T. Kato and T. Habuchi: Microsatellite polymorphism of steroid hormone synthesis gene CYP11A1 is associated with advanced prostate cancer. *Int J Cancer*, 110(1), 140-4 (2004)
DOI: 10.1002/ijc.20070
PMid:15054879
45. L. Tang, S. Yao, C. Till, P. J. Goodman, C. M. Tangen, Y. Wu, A. R. Kristal, E. A. Platz, M. L. Neuhaus, F. Z. Stanczyk, J. K. Reichardt, R. M. Santella, A. Hsing, A. Hoque, S. M. Lippman, I. M. Thompson and C. B. Ambrosone: Repeat polymorphisms in estrogen metabolism genes and prostate cancer risk: results from the Prostate Cancer Prevention Trial. *Carcinogenesis*, 32(10), 1500-6 (2011)
DOI: 10.1093/carcin/bgr139
PMid:21771722 PMCID: PMC3179424
46. J. Picado-Leonard and W. L. Miller: Cloning and sequence of the human gene for P450c17 (steroid 17 α -hydroxylase/17,20 lyase): similarity with the gene for P450c21. *DNA*,

- 6(5), 439-48 (1987)
DOI: 10.1089/dna.1987.6.439
PMid:3500022
47. A. H. Carey, D. Waterworth, K. Patel, D. White, J. Little, P. Novelli, S. Franks and R. Williamson: Polycystic ovaries and premature male pattern baldness are associated with one allele of the steroid metabolism gene CYP17. *Hum Mol Genet*, 3(10), 1873-6 (1994)
DOI: 10.1093/hmg/3.10.1873
PMid:7849715
 48. R. M. Lunn, D. A. Bell, J. L. Mohler and J. A. Taylor: Prostate cancer risk and polymorphism in 17 hydroxylase (CYP17) and steroid reductase (SRD5A2). *Carcinogenesis*, 20(9), 1727-31 (1999)
DOI: 10.1093/carcin/20.9.1727
PMid:10469617
 49. A. Gsur, G. Bernhofer, S. Hinteregger, G. Haidinger, G. Schatzl, S. Madersbacher, M. Marberger, C. Vutuc and M. Micksche: A polymorphism in the CYP17 gene is associated with prostate cancer risk. *Int J Cancer*, 87(3), 434-7 (2000)
DOI: 10.1002/1097-0215 (20000801)87:3<434:AID-IJC19>3.0.CO;2-G
 50. Y. Yamada, M. Watanabe, M. Murata, M. Yamanaka, Y. Kubota, H. Ito, T. Katoh, J. Kawamura, R. Yatani and T. Shiraishi: Impact of genetic polymorphisms of 17-hydroxylase cytochrome P-450 (CYP17) and steroid 5alpha-reductase type II (SRD5A2) genes on prostate-cancer risk among the Japanese population. *Int J Cancer*, 92(5), 683-6 (2001)
DOI: 10.1002/1097-0215(20010601)92:5<683:AID-IJC1255>3.0.CO;2-4
 51. C. A. Haiman, M. J. Stampfer, E. Giovannucci, J. Ma, N. E. Decalo, P. W. Kantoff and D. J. Hunter: The relationship between a polymorphism in CYP17 with plasma hormone levels and prostate cancer. *Cancer Epidemiol Biomarkers Prev*, 10(7), 743-8 (2001)
 52. R. A. Kittles, R. K. Panguluri, W. Chen, A. Massac, C. Ahaghotu, A. Jackson, F. Ukoli, L. Adams-Campbell, W. Isaacs and G. M. Dunston: Cyp17 promoter variant associated with prostate cancer aggressiveness in African Americans. *Cancer Epidemiol Biomarkers Prev*, 10(9), 943-7 (2001)
 53. J. L. Stanford, E. A. Noonan, L. Iwasaki, S. Kolb, R. B. Chadwick, Z. Feng and E. A. Ostrander: A polymorphism in the CYP17 gene and risk of prostate cancer. *Cancer Epidemiol Biomarkers Prev*, 11(3), 243-7 (2002)
 54. B. Chang, S. L. Zheng, S. D. Isaacs, K. E. Wiley, J. D. Carpten, G. A. Hawkins, E. R. Bleecker, P. C. Walsh, J. M. Trent, D. A. Meyers, W. B. Isaacs and J. Xu: Linkage and association of CYP17 gene in hereditary and sporadic prostate cancer. *Int J Cancer*, 95(6), 354-9 (2001)
DOI: 10.1002/1097-0215(20011120)95:6<354:AID-IJC1062>3.0.CO;2-3
 55. A. G. Latil, R. Azzouzi, G. S. Cancel, E. C. Guillaume, B. Cochran-Priollet, P. L. Berthon and O. Cussenot: Prostate carcinoma risk and allelic variants of genes involved in androgen biosynthesis and metabolism pathways. *Cancer*, 92(5), 1130-7 (2001)
DOI: 10.1002/1097-0142(20010901)92:5<1130::AID-CNCR 1430>3.0.CO;2-B
 56. M. Wadelius, A. O. Andersson, J. E. Johansson, C. Wadelius and E. Rane: Prostate cancer associated with CYP17 genotype. *Pharmacogenetics*, 9(5), 635-9 (1999)
DOI: 10.1097/00008571-199910000-00010
PMid:10591544
 57. T. Habuchi, Z. Liqing, T. Suzuki, R. Sasaki, N. Tsuchiya, H. Tachiki, N. Shimoda, S. Satoh, K. Sato, Y. Kakehi, T. Kamoto, O. Ogawa and T. Kato: Increased risk of prostate cancer and benign prostatic hyperplasia associated with a CYP17 gene polymorphism with a gene dosage effect. *Cancer Res*, 60(20), 5710-3 (2000)
 58. C. Ntais, A. Polycarpou and J. P. Ioannidis: Association of the CYP17 gene polymorphism with the risk of prostate cancer: a meta-analysis. *Cancer Epidemiol Biomarkers Prev*, 12(2), 120-6 (2003)
 59. H. S. Feigelson, R. McKean-Cowdin, M. C. Pike, G. A. Coetzee, L. N. Kolonel, A. M. Nomura, L. Le Marchand and B. E. Henderson: Cytochrome P450c17alpha gene (CYP17) polymorphism predicts use of hormone replacement therapy. *Cancer Res*, 59(16), 3908-10 (1999)
 60. R. K. Ross, L. Bernstein, R. A. Lobo, H. Shimizu, F. Z. Stanczyk, M. C. Pike and B. E. Henderson: 5-alpha-reductase activity and risk of prostate cancer among Japanese and US white and black males. *Lancet*, 339(8798), 887-9 (1992)

- DOI: 10.1016/0140-6736(92)90927-U
61. A. E. Thigpen, R. I. Silver, J. M. Guileyardo, M. L. Casey, J. D. McConnell and D. W. Russell: Tissue distribution and ontogeny of steroid 5 alpha-reductase isozyme expression. *J Clin Invest*, 92(2), 903-10 (1993)
DOI: 10.1172/JCI116665
PMid:7688765 PMCID: PMC294929
 62. D. P. Lookingbill, L. M. Demers, C. Wang, A. Leung, R. S. Rittmaster and R. J. Santen: Clinical and biochemical parameters of androgen action in normal healthy Caucasian versus Chinese subjects. *J Clin Endocrinol Metab*, 72(6), 1242-8 (1991)
DOI: 10.1210/jcem-72-6-1242
PMid:1827450
 63. J. K. Reichardt, N. Makridakis, B. E. Henderson, M. C. Yu, M. C. Pike and R. K. Ross: Genetic variability of the human SRD5A2 gene: implications for prostate cancer risk. *Cancer Res*, 55(18), 3973-5 (1995)
 64. J. E. Montie and K. J. Pienta: Review of the role of androgenic hormones in the epidemiology of benign prostatic hyperplasia and prostate cancer. *Urology*, 43(6), 892-9 (1994)
DOI: 10.1016/0090-4295(94)90163-5
 65. R. Anwar, S. G. Gilbey, J. P. New and A. F. Markham: Male pseudohermaphroditism resulting from a novel mutation in the human steroid 5 alpha-reductase type 2 gene (SRD5A2). *Mol Pathol*, 50(1), 51-2 (1997)
DOI: 10.1136/mp.50.1.51
PMid:9208814 PMCID: PMC379579
 66. N. Makridakis, R. K. Ross, M. C. Pike, L. Chang, F. Z. Stanczyk, L. N. Kolonel, C. Y. Shi, M. C. Yu, B. E. Henderson and J. K. Reichardt: A prevalent missense substitution that modulates activity of prostatic steroid 5alpha-reductase. *Cancer Res*, 57(6), 1020-2 (1997)
 67. N. M. Makridakis, R. K. Ross, M. C. Pike, L. E. Crocitto, L. N. Kolonel, C. L. Pearce, B. E. Henderson and J. K. Reichardt: Association of mis-sense substitution in SRD5A2 gene with prostate cancer in African-American and Hispanic men in Los Angeles, USA. *Lancet*, 354(9183), 975-8 (1999)
DOI: 10.1016/S0140-6736(98)11282-5
 68. J. Simard, F. Durocher, F. Mebarki, C. Turgeon, R. Sanchez, Y. Labrie, J. Couet, C. Trudel, E. Rheaume, Y. Morel, V. Luu-The and F. Labrie: Molecular biology and genetics of the 3 beta-hydroxysteroid dehydrogenase/delta5-delta4 isomerase gene family. *J Endocrinol*, 150 Suppl, S189-207 (1996)
 69. L. Wang, E. Salavaggione, L. Pellemounter, B. Eckloff, E. Wieben and R. Weinshilbom: Human 3beta-hydroxysteroid dehydrogenase types 1 and 2: Gene sequence variation and functional genomics. *J Steroid Biochem Mol Biol*, 107(1-2), 88-99 (2007)
 70. H. Verreault, I. Dufort, J. Simard, F. Labrie and V. Luu-The: Dinucleotide repeat polymorphisms in the HSD3B2 gene. *Hum Mol Genet*, 3(2), 384 (1994)
DOI: 10.1093/hmg/3.2.384
PMid:8004119
 71. C. Neslund-Dudas, C. H. Bock, K. Monaghan, N. L. Nock, J. J. Yang, A. Rundle, D. Tang and B. A. Rybicki: SRD5A2 and HSD3B2 polymorphisms are associated with prostate cancer risk and aggressiveness. *Prostate*, 67(15), 1654-63 (2007)
DOI: 10.1002/pros.20625
PMid:17823934 PMCID: PMC2132439
 72. S. A. Devgan, B. E. Henderson, M. C. Yu, C. Y. Shi, M. C. Pike, R. K. Ross and J. K. Reichardt: Genetic variation of 3 beta-hydroxysteroid dehydrogenase type II in three racial/ethnic groups: implications for prostate cancer risk. *Prostate*, 33(1), 9-12 (1997)
DOI: 10.1002/(SICI)1097-0045(19970915)33:1<9::AID-PROS2>3.0.CO;2-H
 73. B. L. Chang, S. L. Zheng, G. A. Hawkins, S. D. Isaacs, K. E. Wiley, A. Turner, J. D. Carpten, E. R. Bleecker, P. C. Walsh, J. M. Trent, D. A. Meyers, W. B. Isaacs and J. Xu: Joint effect of HSD3B1 and HSD3B2 genes is associated with hereditary and sporadic prostate cancer susceptibility. *Cancer Res*, 62(6), 1784-9 (2002)
 74. A. L. Eriksson, M. Lorentzon, L. Vandenput, F. Labrie, M. Lindersson, A. C. Syvanen, E. S. Orwoll, S. R. Cummings, J. M. Zmuda, O. Ljunggren, M. K. Karlsson, D. Mellstrom and C. Ohlsson: Genetic variations in sex steroid-related genes as predictors of serum estrogen levels in men. *J Clin Endocrinol Metab*, 94(3), 1033-41 (2009)
DOI: 10.1210/jc.2008-1283

- PMid:19116238 PMCID: PMC2681277
75. R. C. Travis, F. Schumacher, J. N. Hirschhorn, P. Kraft, N. E. Allen, D. Albanes, G. Berglund, S. I. Berndt, H. Boeing, H. B. Bueno-de-Mesquita, E. E. Calle, S. Chanock, A. M. Dunning, R. Hayes, H. S. Feigelson, J. M. Gaziano, E. Giovannucci, C. A. Haiman, B. E. Henderson, R. Kaaks, L. N. Kolonel, J. Ma, L. Rodriguez, E. Riboli, M. Stampfer, D. O. Stram, M. J. Thun, A. Tjonneland, D. Trichopoulos, P. Vineis, J. Virtamo, L. Le Marchand and D. J. Hunter: CYP19A1 genetic variation in relation to prostate cancer risk and circulating sex hormone concentrations in men from the Breast and Prostate Cancer Cohort Consortium. *Cancer Epidemiol Biomarkers Prev*, 18(10), 2734-44 (2009)
DOI: 10.1158/1055-9965.EPI-09-0496
PMid:19789370 PMCID: PMC2812905
 76. T. Sonoda, H. Suzuki, M. Mori, T. Tsukamoto, A. Yokomizo, S. Naito, K. Fujimoto, Y. Hirao, N. Miyanaga and H. Akaza: Polymorphisms in estrogen related genes may modify the protective effect of isoflavones against prostate cancer risk in Japanese men. *Eur J Cancer Prev*, 19(2), 131-7 (2010)
DOI: 10.1097/CEJ.0b013e328333f3be2
PMid:19952760
 77. Y. C. Huang, M. Chen, M. W. Lin, M. Y. Chung, Y. H. Chang, W. J. Huang, T. T. Wu, J. M. Hsu, S. Yang and Y. M. Chen: CYP19 TCT trinucleotide Del/Del genotype is a susceptibility marker for prostate cancer in a Taiwanese population. *Urology*, 69(5), 996-1000 (2007)
DOI: 10.1016/j.urology.2007.02.014
PMid:17482958
 78. K. Onsory, R. C. Sobti, A. I. Al-Badran, M. Watanabe, T. Shiraishi, A. Krishan, H. Mohan and P. Kaur: Hormone receptor-related gene polymorphisms and prostate cancer risk in North Indian population. *Mol Cell Biochem*, 314(1-2), 25-35 (2008)
 79. N. Mononen, E. H. Seppala, P. Duggal, V. Autio, T. Ikonen, P. Ellonen, J. Saharinen, J. Saarela, M. Vihinen, T. L. Tammela, O. Kallioniemi, J. E. Bailey-Wilson and J. Schleutker: Profiling genetic variation along the androgen biosynthesis and metabolism pathways implicates several single nucleotide polymorphisms and their combinations as prostate cancer risk factors. *Cancer Res*, 66(2), 743-7 (2006)
DOI: 10.1158/0008-5472.CAN-05-1723
PMid:16424004
 80. J. Beuten, J. A. Gelfond, J. L. Franke, K. S. Weldon, A. C. Crandall, T. L. Johnson-Pais, I. M. Thompson and R. J. Leach: Single and multigenic analysis of the association between variants in 12 steroid hormone metabolism genes and risk of prostate cancer. *Cancer Epidemiol Biomarkers Prev*, 18(6), 1869-80 (2009)
DOI: 10.1158/1055-9965.EPI-09-0076
PMid:19505920
 81. F. Modugno, J. L. Weissfeld, D. L. Trump, J. M. Zmuda, P. Shea, J. A. Cauley and R. E. Ferrell: Allelic variants of aromatase and the androgen and estrogen receptors: toward a multigenic model of prostate cancer risk. *Clin Cancer Res*, 7(10), 3092-6 (2001)
 82. K. Gellner, R. Eiselt, E. Hustert, H. Arnold, I. Koch, M. Haberl, C. J. Deglmann, O. Burk, D. Buntfuss, S. Escher, C. Bishop, H. G. Koebe, U. Brinkmann, H. P. Klenk, K. Kleine, U. A. Meyer and L. Wojnowski: Genomic organization of the human CYP3A locus: identification of a new, inducible CYP3A gene. *Pharmacogenetics*, 11(2), 111-21 (2001)
DOI: 10.1097/00008571-200103000-00002
PMid:11266076
 83. P. Kraft, P. Pharoah, S. J. Chanock, D. Albanes, L. N. Kolonel, R. B. Hayes, D. Altshuler, G. Andriole, C. Berg, H. Boeing, N. P. Burt, B. Bueno-de-Mesquita, E. E. Calle, H. Cann, F. Canzian, Y. C. Chen, D. E. Crawford, A. M. Dunning, H. S. Feigelson, M. L. Freedman, J. M. Gaziano, E. Giovannucci, C. A. Gonzalez, C. A. Haiman, G. Hallmans, B. E. Henderson, J. N. Hirschhorn, D. J. Hunter, R. Kaaks, T. Key, L. Le Marchand, J. Ma, K. Overvad, D. Palli, M. C. Pike, E. Riboli, C. Rodriguez, W. V. Setiawan, M. J. Stampfer, D. O. Stram, G. Thomas, M. J. Thun, R. Travis, A. Trichopoulos, J. Virtamo and S. Wacholder: Genetic variation in the HSD17B1 gene and risk of prostate cancer. *PLoS Genet*, 1(5), e68 (2005)
DOI: 10.1371/journal.pgen.0010068
PMid:16311626 PMCID: PMC1287955
 84. T. Sun, W. K. Oh, S. Jacobus, M. Regan, M. Pomerantz, M. L. Freedman, G. S. Lee and P. W. Kantoff: The impact of common genetic

- variations in genes of the sex hormone metabolic pathways on steroid hormone levels and prostate cancer aggressiveness. *Cancer Prev Res (Phila)*, 4(12), 2044-50 (2011)
DOI: 10.1158/1940-6207.CAPR-11-0283
PMid:21900597 PMCID: PMC3773969
85. G. R. Cunningham, C. M. Ashton, J. F. Annegers, J. Soucek, M. Klima and B. Miles: Familial aggregation of prostate cancer in African-Americans and white Americans. *Prostate*, 56(4), 256-62 (2003)
DOI: 10.1002/pros.10252
PMid:12858353
 86. K. Margiotti, E. Kim, C. L. Pearce, E. Spera, G. Novelli and J. K. Reichardt: Association of the G289S single nucleotide polymorphism in the HSD17B3 gene with prostate cancer in Italian men. *Prostate*, 53(1), 65-8 (2002)
DOI: 10.1002/pros.10134
PMid:12210481
 87. S. Lindstrom, S. L. Zheng, F. Wiklund, B. A. Jonsson, H. O. Adami, K. A. Balter, A. J. Brookes, J. Sun, B. L. Chang, W. Liu, G. Li, W. B. Isaacs, J. Adolfsson, H. Gronberg and J. Xu: Systematic replication study of reported genetic associations in prostate cancer: Strong support for genetic variation in the androgen pathway. *Prostate*, 66(16), 1729-43 (2006)
DOI: 10.1002/pros.20489
PMid:16998812
 88. K. K. Rasiah, M. Gardiner-Garden, E. J. Padilla, G. Moller, J. G. Kench, M. C. Alles, S. A. Eggleton, P. D. Stricker, J. Adamski, R. L. Sutherland, S. M. Henshall and V. M. Hayes: HSD17B4 overexpression, an independent biomarker of poor patient outcome in prostate cancer. *Mol Cell Endocrinol*, 301(1-2), 89-96 (2009)
 89. R. W. Ross, W. K. Oh, W. Xie, M. Pomerantz, M. Nakabayashi, O. Sartor, M. E. Taplin, M. M. Regan, P. W. Kantoff and M. Freedman: Inherited variation in the androgen pathway is associated with the efficacy of androgen-deprivation therapy in men with prostate cancer. *J Clin Oncol*, 26(6), 842-7 (2008)
DOI: 10.1200/JCO.2007.13.6804
PMid:18281655
 90. J. Jakobsson, E. Palonek, M. Lorentzon, C. Ohlsson, A. Rane and L. Ekstrom: A novel polymorphism in the 17beta-hydroxysteroid dehydrogenase type 5 (aldo-keto reductase 1C3) gene is associated with lower serum testosterone levels in caucasian men. *Pharmacogenomics J*, 7(4), 282-9 (2007)
DOI: 10.1038/sj.tpj.6500419
PMid:16983398
 91. J. J. Schulze, H. Karypidis and L. Ekstrom: Basal and Regulatory Promoter Studies of the AKR1C3 Gene in Relation to Prostate Cancer. *Front Pharmacol*, 3, 151 (2012)
 92. J. W. Lampe, J. Bigler, A. C. Bush and J. D. Potter: Prevalence of polymorphisms in the human UDP-glucuronosyltransferase 2B family: UGT2B4(D458E), UGT2B7(H268Y), and UGT2B15(D85Y). *Cancer Epidemiol Biomarkers Prev*, 9(3), 329-33 (2000)
 93. H. Okugi, H. Nakazato, H. Matsui, N. Ohtake, S. Nakata and K. Suzuki: Association of the polymorphisms of genes involved in androgen metabolism and signaling pathways with familial prostate cancer risk in a Japanese population. *Cancer Detect Prev*, 30(3), 262-8 (2006)
DOI: 10.1016/j.cdp.2006.04.004
PMid:16859836
 94. A. Gsur, M. Preyer, G. Haidinger, G. Schatzl, S. Madersbacher, M. Marberger, C. Vutuc and M. Micksche: A polymorphism in the UDP-Glucuronosyltransferase 2B15 gene (D85Y) is not associated with prostate cancer risk. *Cancer Epidemiol Biomarkers Prev*, 11(5), 497-8 (2002)
 95. A. H. Heald, F. Ivison, S. G. Anderson, K. Cruickshank, I. Laing and J. M. Gibson: Significant ethnic variation in total and free testosterone concentration. *Clin Endocrinol (Oxf)*, 58(3), 262-6 (2003)
DOI: 10.1046/j.1365-2265.2003.01653.x
 96. A. W. Roddam, N. E. Allen, P. Appleby and T. J. Key: Endogenous sex hormones and prostate cancer: a collaborative analysis of 18 prospective studies. *J Natl Cancer Inst*, 100(3), 170-83 (2008)
DOI: 10.1093/jnci/djm323
PMid:18230794
 97. S. G. Power, W. P. Bocchinfuso, M. Pallesen, S. Warmels-Rodenhiser, H. Van Baelen and G. L. Hammond: Molecular analyses of a human sex hormone-binding globulin variant: evidence for an additional carbohydrate

- chain. *J Clin Endocrinol Metab*, 75(4), 1066-70 (1992)
DOI: 10.1210/jc.75.4.1066
98. Y. L. Low, J. I. Taylor, P. B. Grace, A. A. Mulligan, A. A. Welch, S. Scollen, A. M. Dunning, R. N. Luben, K. T. Khaw, N. E. Day, N. J. Wareham and S. A. Bingham: Phytoestrogen exposure, polymorphisms in COMT, CYP19, ESR1, and SHBG genes, and their associations with prostate cancer risk. *Nutr Cancer*, 56(1), 31-9 (2006)
DOI: 10.1207/s15327914nc5601_5
PMid:17176215
 99. D. L. Winter, A. L. Hanlon, S. L. Raysor, D. Watkins-Bruner, W. H. Pinover, G. E. Hanks and J. V. Tricoli: Plasma levels of IGF-1, IGF-2, and IGFBP-3 in white and African-American men at increased risk of prostate cancer. *Urology*, 58(4), 614-8 (2001)
DOI: 10.1016/S0090-4295(01)01273-0
 100. R. W. de Vere White, A. D. Deitch, A. G. Jackson, R. Gandour-Edwards, J. Marshall, S. E. Soares, S. N. Toscano, J. M. Lunetta and S. L. Stewart: Racial differences in clinically localized prostate cancers of black and white men. *J Urol*, 159(6), 1979-82; discussion 1982-3 (1998)
 101. E. C. Pietsch, O. Humbey and M. E. Murphy: Polymorphisms in the p53 pathway. *Oncogene*, 25(11), 1602-11 (2006)
DOI: 10.1038/sj.onc.1209367
PMid:16550160
 102. Y. Haupt, R. Maya, A. Kazaz and M. Oren: Mdm2 promotes the rapid degradation of p53. *Nature*, 387(6630), 296-9 (1997)
DOI: 10.1038/387296a0
PMid:9153395
 103. M. H. Kubbutat, S. N. Jones and K. H. Vousden: Regulation of p53 stability by Mdm2. *Nature*, 387(6630), 299-303 (1997)
DOI: 10.1038/387299a0
PMid:9153396
 104. G. L. Bond, W. Hu and A. J. Levine: MDM2 is a central node in the p53 pathway: 12 years and counting. *Curr Cancer Drug Targets*, 5(1), 3-8 (2005)
DOI: 10.2174/1568009053332627
PMid:15720184
 105. I. Osman, M. Drobnjak, M. Fazzari, J. Ferrara, H. I. Scher and C. Cordon-Cardo: Inactivation of the p53 pathway in prostate cancer: impact on tumor progression. *Clin Cancer Res*, 5(8), 2082-8 (1999)
 106. K. R. Leite, M. F. Franco, M. Srougi, L. J. Nesrallah, A. Nesrallah, R. G. Bevilacqua, E. Darini, C. M. Carvalho, M. I. Meirelles, I. Santana and L. H. Camara-Lopes: Abnormal expression of MDM2 in prostate carcinoma. *Mod Pathol*, 14(5), 428-36 (2001)
DOI: 10.1038/modpathol.3880330
PMid:11353053
 107. L. Y. Khor, M. Desilvio, T. Al-Saleem, M. E. Hammond, D. J. Grignon, W. Sause, M. Pilepich, P. Okunieff, H. Sandler and A. Pollack: MDM2 as a predictor of prostate carcinoma outcome: an analysis of Radiation Therapy Oncology Group Protocol 8610. *Cancer*, 104(5), 962-7 (2005)
DOI: 10.1002/cncr.21261
PMid:16007688
 108. R. Bianco, R. Caputo, R. Caputo, V. Damiano, S. De Placido, C. Ficorella, S. Agrawal, A. R. Bianco, F. Ciardiello and G. Tortora: Combined targeting of epidermal growth factor receptor and MDM2 by gefitinib and antisense MDM2 cooperatively inhibit hormone-independent prostate cancer. *Clin Cancer Res*, 10(14), 4858-64 (2004)
DOI: 10.1158/1078-0432.CCR-03-0497
PMid:15269162
 109. H. Wang, D. Yu, S. Agrawal and R. Zhang: Experimental therapy of human prostate cancer by inhibiting MDM2 expression with novel mixed-backbone antisense oligonucleotides: *in vitro* and *in vivo* activities and mechanisms. *Prostate*, 54(3), 194-205 (2003)
DOI: 10.1002/pros.10187
PMid:12518324
 110. Z. Zhang, M. Li, H. Wang, S. Agrawal and R. Zhang: Antisense therapy targeting MDM2 oncogene in prostate cancer: Effects on proliferation, apoptosis, multiple gene expression, and chemotherapy. *Proc Natl Acad Sci U S A*, 100(20), 11636-41 (2003)
DOI: 10.1073/pnas.1934692100
PMid:13130078 PMCID: PMC208810
 111. G. L. Bond, C. Menin, R. Bertorelle, P. Alhopuro, L. A. Aaltonen and A. J. Levine: MDM2 SNP309 accelerates colorectal tumour formation in women. *J Med Genet*, 43(12), 950-2 (2006)
DOI: 10.1136/jmg.2006.043539

- PMid:16825430 PMCID: PMC2563203
112. H. Lind, S. Zienolddiny, P. O. Ekstrom, V. Skaug and A. Haugen: Association of a functional polymorphism in the promoter of the MDM2 gene with risk of nonsmall cell lung cancer. *Int J Cancer*, 119(3), 718-21 (2006)
DOI: 10.1002/ijc.21872
PMid:16496380
 113. C. Menin, M. C. Scaini, G. L. De Salvo, M. Biscuola, M. Quaggio, G. Esposito, C. Belluco, M. Montagna, S. Agata, E. D'Andrea, D. Nitti, A. Amadori and R. Bertorelle: Association between MDM2-SNP309 and age at colorectal cancer diagnosis according to p53 mutation status. *J Natl Cancer Inst*, 98(4), 285-8 (2006)
DOI: 10.1093/jnci/djj054
PMid:16478747
 114. P. Alhopuro, S. K. Ylisaukko-Oja, W. J. Koskinen, P. Bono, J. Arola, H. J. Jarvinen, J. P. Mecklin, T. Atula, R. Kontio, A. A. Makitie, S. Suominen, I. Leivo, P. Vahteristo, L. M. Aaltonen and L. A. Aaltonen: The MDM2 promoter polymorphism SNP309T-->G and the risk of uterine leiomyosarcoma, colorectal cancer, and squamous cell carcinoma of the head and neck. *J Med Genet*, 42(9), 694-8 (2005)
DOI: 10.1136/jmg.2005.031260
PMid:16141004 PMCID: PMC1736129
 115. R. I. Yarden, E. Friedman, S. Metsuyanin, T. Olender, E. Ben-Asher and M. Z. Papa: MDM2 SNP309 accelerates breast and ovarian carcinogenesis in BRCA1 and BRCA2 carriers of Jewish-Ashkenazi descent. *Breast Cancer Res Treat*, 111(3), 497-504 (2008)
DOI: 10.1007/s10549-007-9797-z
PMid:18026875
 116. G. Wang, E. F. Firoz, A. Rose, E. Blochin, P. Christos, D. Pollens, M. Mazumdar, W. Gerald, C. Oddoux, P. Lee and I. Osman: MDM2 expression and regulation in prostate cancer racial disparity. *Int J Clin Exp Pathol*, 2(4), 353-60 (2009)
 117. C. M. Zeigler-Johnson, A. H. Walker, B. Mancke, E. Spangler, M. Jalloh, S. McBride, A. Deitz, S. B. Malkowicz, D. Ofori-Adjei, S. M. Gueye and T. R. Rebbeck: Ethnic differences in the frequency of prostate cancer susceptibility alleles at SRD5A2 and CYP3A4. *Hum Hered*, 54(1), 13-21 (2002)
DOI: 10.1159/000066695
 118. D. C. Miller, S. L. Zheng, R. L. Dunn, A. V. Sarma, J. E. Montie, E. M. Lange, D. A. Meyers, J. Xu and K. A. Cooney: Germ-line mutations of the macrophage scavenger receptor 1 gene: association with prostate cancer risk in African-American men. *Cancer Res*, 63(13), 3486-9 (2003)
 119. I. Oakley-Girvan, D. Feldman, T. R. Eccleshall, R. P. Gallagher, A. H. Wu, L. N. Kolonel, J. Halpern, R. R. Balise, D. W. West, R. S. Paffenbarger, Jr. and A. S. Whittemore: Risk of early-onset prostate cancer in relation to germ line polymorphisms of the vitamin D receptor. *Cancer Epidemiol Biomarkers Prev*, 13(8), 1325-30 (2004)
 120. K. R. Monroe, M. C. Yu, L. N. Kolonel, G. A. Coetzee, L. R. Wilkens, R. K. Ross and B. E. Henderson: Evidence of an X-linked or recessive genetic component to prostate cancer risk. *Nat Med*, 1(8), 827-9 (1995)
DOI: 10.1038/nm0895-827
PMid:7585188
 121. S. I. Bastacky, K. J. Wojno, P. C. Walsh, M. J. Carmichael and J. I. Epstein: Pathological features of hereditary prostate cancer. *J Urol*, 153(3 Pt 2), 987-92 (1995)
 122. D. Mirchandani, J. Zheng, G. J. Miller, A. K. Ghosh, D. K. Shibata, R. J. Cote and P. Roy-Burman: Heterogeneity in intratumor distribution of p53 mutations in human prostate cancer. *Am J Pathol*, 147(1), 92-101 (1995)
 123. J. Qian, D. G. Bostwick, S. Takahashi, T. J. Borell, J. F. Herath, M. M. Lieber and R. B. Jenkins: Chromosomal anomalies in prostatic intraepithelial neoplasia and carcinoma detected by fluorescence *in situ* hybridization. *Cancer Res*, 55(22), 5408-14 (1995)
 124. W. A. Sakr, D. J. Grignon, J. D. Crissman, L. K. Heilbrun, B. J. Cassin, J. J. Pontes and G. P. Haas: High grade prostatic intraepithelial neoplasia (HGPIN) and prostatic adenocarcinoma between the ages of 20-69: an autopsy study of 249 cases. *In vivo*, 8(3), 439-43 (1994)
 125. T. R. Rebbeck: Genetics, disparities, and prostate cancer. *LDI Issue Brief*, 10(7), 1-4 (2005)
 126. J. Xu, S. L. Zheng, G. A. Hawkins, D. A. Faith, B. Kelly, S. D. Isaacs, K. E. Wiley, B.

- Chang, C. M. Ewing, P. Bujnovszky, J. D. Carpten, E. R. Bleecker, P. C. Walsh, J. M. Trent, D. A. Meyers and W. B. Isaacs: Linkage and association studies of prostate cancer susceptibility: evidence for linkage at 8p22-23. *Am J Hum Genet*, 69(2), 341-50 (2001)
DOI: 10.1086/321967
PMid:11443539 PMCID: PMC1235306
127. M. Gibbs, J. L. Stanford, G. P. Jarvik, M. Janer, M. Badzioch, M. A. Peters, E. L. Goode, S. Kolb, L. Chakrabarti, M. Shook, R. Basom, E. A. Ostrander and L. Hood: A genomic scan of families with prostate cancer identifies multiple regions of interest. *Am J Hum Genet*, 67(1), 100-9 (2000)
DOI: 10.1086/302969
PMid:10820127 PMCID: PMC1287067
 128. M. R. Emmert-Buck, C. D. Vocke, R. O. Pozzatti, P. H. Duray, S. B. Jennings, C. D. Florence, Z. Zhuang, D. G. Bostwick, L. A. Liotta and W. M. Linehan: Allelic loss on chromosome 8p12-21 in microdissected prostatic intraepithelial neoplasia. *Cancer Res*, 55(14), 2959-62 (1995)
 129. C. D. Vocke, R. O. Pozzatti, D. G. Bostwick, C. D. Florence, S. B. Jennings, S. E. Strup, P. H. Duray, L. A. Liotta, M. R. Emmert-Buck and W. M. Linehan: Analysis of 99 microdissected prostate carcinomas reveals a high frequency of allelic loss on chromosome 8p12-21. *Cancer Res*, 56(10), 2411-6 (1996)
 130. H. Zitzelsberger, D. Engert, A. Walch, U. Kulka, M. Aubele, H. Hofler, M. Bauchinger and M. Werner: Chromosomal changes during development and progression of prostate adenocarcinomas. *Br J Cancer*, 84(2), 202-8 (2001)
DOI: 10.1054/bjoc.2000.1533
PMid:11161378 PMCID: PMC2363712
 131. R. S. Verma, M. Manikal, R. A. Conte and C. J. Godec: Chromosomal basis of adenocarcinoma of the prostate. *Cancer Invest*, 17(6), 441-7 (1999)
DOI: 10.3109/07357909909021436
PMid:10434955
 132. R. Bookstein: Tumor suppressor genes in prostatic oncogenesis. *J Cell Biochem Suppl*, 19, 217-23 (1994)
 133. J. A. Kalapurakal, A. N. Jacob, P. Y. Kim, D. D. Najjar, Y. C. Hsieh, P. Ginsberg, I. Daskal, S. O. Asbell and R. P. Kandpal: Racial differences in prostate cancer related to loss of heterozygosity on chromosome 8p12-23. *Int J Radiat Oncol Biol Phys*, 45(4), 835-40 (1999)
DOI: 10.1016/S0360-3016(99)00283-7
 134. D. P. BarTel: MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell*, 116(2), 281-97 (2004)
DOI: 10.1016/S0092-8674(04)00045-5
 135. A. M. Denli, B. B. Tops, R. H. Plasterk, R. F. Ketting and G. J. Hannon: Processing of primary microRNAs by the Microprocessor complex. *Nature*, 432(7014), 231-5 (2004)
DOI: 10.1038/nature03049
PMid:15531879
 136. G. A. Calin, C. Sevignani, C. D. Dumitru, T. Hyslop, E. Noch, S. Yendamuri, M. Shimizu, S. Rattan, F. Bullrich, M. Negrini and C. M. Croce: Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci U S A*, 101(9), 2999-3004 (2004)
DOI: 10.1073/pnas.0307323101
PMid:14973191 PMCID: PMC365734
 137. J. Lu, G. Getz, E. A. Miska, E. Alvarez-Saavedra, J. Lamb, D. Peck, A. Sweet-Cordero, B. L. Ebert, R. H. Mak, A. A. Ferrando, J. R. Downing, T. Jacks, H. R. Horvitz and T. R. Golub: MicroRNA expression profiles classify human cancers. *Nature*, 435(7043), 834-8 (2005)
DOI: 10.1038/nature03702
PMid:15944708
 138. M. V. Iorio, M. Ferracin, C. G. Liu, A. Veronese, R. Spizzo, S. Sabbioni, E. Magri, M. Pedriali, M. Fabbri, M. Campiglio, S. Menard, J. P. Palazzo, A. Rosenberg, P. Musiani, S. Volinia, I. Nenci, G. A. Calin, P. Querzoli, M. Negrini and C. M. Croce: MicroRNA gene expression deregulation in human breast cancer. *Cancer Res*, 65(16), 7065-70 (2005)
DOI: 10.1158/0008-5472.CAN-05-1783
PMid:16103053
 139. C. G. Liu, G. A. Calin, B. Meloon, N. Gamliel, C. Sevignani, M. Ferracin, C. D. Dumitru, M. Shimizu, S. Zupo, M. Dono, H. Alder, F. Bullrich, M. Negrini and C. M. Croce: An oligonucleotide microchip for genome-wide microRNA profiling in human and mouse tissues. *Proc Natl Acad Sci U S A*, 101(26), 9740-4 (2004)
DOI: 10.1073/pnas.0403293101

- PMid:15210942 PMCID: PMC470744
140. Y. Akao, Y. Nakagawa and T. Naoe: MicroRNAs 143 and 145 are possible common onco-microRNAs in human cancers. *Oncol Rep*, 16(4), 845-50 (2006)
DOI: 10.3892/or.16.4.845
PMid:16266980
 141. Y. Hayashita, H. Osada, Y. Tatematsu, H. Yamada, K. Yanagisawa, S. Tomida, Y. Yatabe, K. Kawahara, Y. Sekido and T. Takahashi: A polycistronic microRNA cluster, miR-17-92, is overexpressed in human lung cancers and enhances cell proliferation. *Cancer Res*, 65(21), 9628-32 (2005)
DOI: 10.1158/0008-5472.CAN-05-2352
PMid:16266980
 142. L. He, J. M. Thomson, M. T. Hemann, E. Hernando-Monge, D. Mu, S. Goodson, S. Powers, C. Cordon-Cardo, S. W. Lowe, G. J. Hannon and S. M. Hammond: A microRNA polycistron as a potential human oncogene. *Nature*, 435(7043), 828-33 (2005)
DOI: 10.1038/nature03552
PMid:15944707 PMCID: PMC4599349
 143. P. M. Voorhoeve, C. le Sage, M. Schrier, A. J. Gillis, H. Stoop, R. Nagel, Y. P. Liu, J. van Duijse, J. Drost, A. Griekspoor, E. Zlotorynski, N. Yabuta, G. De Vita, H. Nojima, L. H. Looijenga and R. Agami: A genetic screen implicates miRNA-372 and miRNA-373 as oncogenes in testicular germ cell tumors. *Cell*, 124(6), 1169-81 (2006)
DOI: 10.1016/j.cell.2006.02.037
PMid:16564011
 144. M. L. Si, S. Zhu, H. Wu, Z. Lu, F. Wu and Y. Y. Mo: miR-21-mediated tumor growth. *Oncogene*, 26(19), 2799-803 (2007)
DOI: 10.1038/sj.onc.1210083
PMid:17072344
 145. T. Wang, X. Zhang, L. Obijuru, J. Laser, V. Aris, P. Lee, K. Mittal, P. Soteropoulos and J. J. Wei: A micro-RNA signature associated with race, tumor size, and target gene activity in human uterine leiomyomas. *Genes Chromosomes Cancer*, 46(4), 336-47 (2007)
DOI: 10.1002/gcc.20415
PMid:17243163
 146. G. A. Calin and C. M. Croce: MicroRNA signatures in human cancers. *Nat Rev Cancer*, 6(11), 857-66 (2006)
DOI: 10.1038/nrc1997
PMid:17060945
 147. K. M. Rahman and W. A. Sakr: The therapeutic value of natural agents to treat miRNA targeted breast cancer in African-American and Caucasian-American women. *Curr Drug Targets*, 13(14), 1917-25 (2012)
DOI: 10.2174/138945012804545461
PMid:23140300
 148. T. J. Polascik, J. E. Oesterling and A. W. Partin: Prostate specific antigen: a decade of discovery--what we have learned and where we are going. *J Urol*, 162(2), 293-306 (1999)
DOI: 10.1016/S0022-5347(05)68543-6
 149. J. Oh, J. W. Colberg, D. K. Ornstein, E. T. Johnson, D. Chan, K. S. Virgo and F. E. Johnson: Current followup strategies after radical prostatectomy: a survey of American Urological Association urologists. *J Urol*, 161(2), 520-3 (1999)
DOI: 10.1016/S0022-5347(01)61939-6
 150. S. B. Zeliadt, D. F. Penson, P. C. Albertsen, J. Concato and R. D. Etzioni: Race independently predicts prostate specific antigen testing frequency following a prostate carcinoma diagnosis. *Cancer*, 98(3), 496-503 (2003)
DOI: 10.1002/cncr.11492
PMid:12879465
 151. A. B. Mariotto, R. Etzioni, M. Krapcho and E. J. Feuer: Reconstructing PSA testing patterns between black and white men in the US from Medicare claims and the National Health Interview Survey. *Cancer*, 109(9), 1877-86 (2007)
DOI: 10.1002/cncr.22607
PMid:17372918
 152. W. M. Xue, G. A. Coetzee, R. K. Ross, R. Irvine, L. Kolonel, B. E. Henderson and S. A. Ingles: Genetic determinants of serum prostate-specific antigen levels in healthy men from a multiethnic cohort. *Cancer Epidemiol Biomarkers Prev*, 10(6), 575-9 (2001)
 153. R. Deka, S. Guangyun, J. Wiest, D. Smelser, S. Chunhua, Y. Zhong and R. Chakraborty: Patterns of instability of expanded CAG repeats at the ERDA1 locus in general populations. *Am J Hum Genet*, 65(1), 192-8 (1999)
DOI: 10.1086/302453
PMid:10364532 PMCID: PMC1378090
 154. A. Shibata and A. S. Whittemore: Genetic predisposition to prostate cancer: possible explanations for ethnic differences in risk.

- Prostate*, 32(1), 65-72 (1997)
DOI: 10.1002/(SICI)1097-0045(19970615)32:1<65::AID-PROS9>3.0.CO;2-B
155. J. R. Smith, D. Freije, J. D. Carpten, H. Gronberg, J. Xu, S. D. Isaacs, M. J. Brownstein, G. S. Bova, H. Guo, P. Bujnovszky, D. R. Nusskern, J. E. Damber, A. Bergh, M. Emanuelsson, O. P. Kallioniemi, J. Walker-Daniels, J. E. Bailey-Wilson, T. H. Beaty, D. A. Meyers, P. C. Walsh, F. S. Collins, J. M. Trent and W. B. Isaacs: Major susceptibility locus for prostate cancer on chromosome 1 suggested by a genome-wide search. *Science*, 274(5291), 1371-4 (1996)
DOI: 10.1126/science.274.5291.1371
PMid:8910276
156. J. Xu, D. Meyers, D. Freije, S. Isaacs, K. Wiley, D. Nusskern, C. Ewing, E. Wilkens, P. Bujnovszky, G. S. Bova, P. Walsh, W. Isaacs, J. Schleutker, M. Matikainen, T. Tammela, T. Visakorpi, O. P. Kallioniemi, R. Berry, D. Schaid, A. French, S. McDonnell, J. Schroeder, M. Blute, S. Thibodeau, H. Gronberg, M. Emanuelsson, J. E. Damber, A. Bergh, B. A. Jonsson, J. Smith, J. Bailey-Wilson, J. Carpten, D. Stephan, E. Gillanders, I. Amundson, T. Kainu, D. Freas-Lutz, A. Baffoe-Bonnie, A. Van Aucken, R. Sood, F. Collins, M. Brownstein and J. Trent: Evidence for a prostate cancer susceptibility locus on the X chromosome. *Nat Genet*, 20(2), 175-9 (1998)
DOI: 10.1038/2477
PMid:9771711
157. J. Xu, S. L. Zheng, B. Chang, J. R. Smith, J. D. Carpten, O. C. Stine, S. D. Isaacs, K. E. Wiley, L. Henning, C. Ewing, P. Bujnovszky, E. R. Bleeker, P. C. Walsh, J. M. Trent, D. A. Meyers and W. B. Isaacs: Linkage of prostate cancer susceptibility loci to chromosome 1. *Hum Genet*, 108(4), 335-45 (2001)
DOI: 10.1007/s004390100488
PMid:11379880
158. S. L. Zheng, J. Xu, S. D. Isaacs, K. Wiley, B. Chang, E. R. Bleeker, P. C. Walsh, J. M. Trent, D. A. Meyers and W. B. Isaacs: Evidence for a prostate cancer linkage to chromosome 20 in 159 hereditary prostate cancer families. *Hum Genet*, 108(5), 430-5 (2001)
DOI: 10.1007/s004390100513
PMid:11409871
159. J. Xu, S. L. Zheng, J. D. Carpten, N. N. Nupponen, C. M. Robbins, J. Mestre, T. Y. Moses, D. A. Faith, B. D. Kelly, S. D. Isaacs, K. E. Wiley, C. M. Ewing, P. Bujnovszky, B. Chang, J. Bailey-Wilson, E. R. Bleeker, P. C. Walsh, J. M. Trent, D. A. Meyers and W. B. Isaacs: Evaluation of linkage and association of HPC2/ELAC2 in patients with familial or sporadic prostate cancer. *Am J Hum Genet*, 68(4), 901-11 (2001)
DOI: 10.1086/319513
PMid:11254448 PMCID: PMC1275644
160. G. Gong, I. Oakley-Girvan, A. H. Wu, L. N. Kolonel, E. M. John, D. W. West, A. Felberg, R. P. Gallagher and A. S. Whittemore: Segregation analysis of prostate cancer in 1,719 white, African-American and Asian-American families in the United States and Canada. *Cancer Causes Control*, 13(5), 471-82 (2002)
DOI: 10.1023/A:1015755219674
PMid:12146852
161. C. Ahaghotu, A. Baffoe-Bonnie, R. Kittles, C. Pettaway, I. Powell, C. Royal, H. Wang, S. Vijayakumar, J. Bennett, G. Hoke, T. Mason, J. Bailey-Wilson, W. Boykin, K. Berg, J. Carpten, S. Weinrich, J. Trent, G. Dunston and F. Collins: Clinical characteristics of African-American men with hereditary prostate cancer: the AAHPC Study. *Prostate Cancer Prostatic Dis*, 7(2), 165-9 (2004)
DOI: 10.1038/sj.pcan.4500719
PMid:15175665
162. W. M. Brown, E. M. Lange, H. Chen, S. L. Zheng, B. Chang, K. E. Wiley, S. D. Isaacs, P. C. Walsh, W. B. Isaacs, J. Xu and K. A. Cooney: Hereditary prostate cancer in African American families: linkage analysis using markers that map to five candidate susceptibility loci. *Br J Cancer*, 90(2), 510-4 (2004)
DOI: 10.1038/sj.bjc.6601417
PMid:14735201 PMCID: PMC2410149
163. J. Xu, S. L. Zheng, A. Komiya, J. C. Mychaleckyj, S. D. Isaacs, J. J. Hu, D. Sterling, E. M. Lange, G. A. Hawkins, A. Turner, C. M. Ewing, D. A. Faith, J. R. Johnson, H. Suzuki, P. Bujnovszky, K. E. Wiley, A. M. DeMarzo, G. S. Bova, B. Chang, M. C. Hall, D. L. McCullough, A. W. Partin, V. S. Kassabian, J. D. Carpten, J. E. Bailey-Wilson, J. M. Trent, J. Ohar, E. R. Bleeker, P. C. Walsh, W. B. Isaacs and D. A. Meyers: Germline mutations and sequence variants of the macrophage scavenger receptor 1 gene are associated

- with prostate cancer risk. *Nat Genet*, 32(2), 321-5 (2002)
DOI: 10.1038/ng994
PMid:12244320
164. J. Xu, S. L. Zheng, A. Komiya, J. C. Mychaleckyj, S. D. Isaacs, B. Chang, A. R. Turner, C. M. Ewing, K. E. Wiley, G. A. Hawkins, E. R. Bleeker, P. C. Walsh, D. A. Meyers and W. B. Isaacs: Common sequence variants of the macrophage scavenger receptor 1 gene are associated with prostate cancer risk. *Am J Hum Genet*, 72(1), 208-12 (2003)
DOI: 10.1086/345802
PMid:12471593 PMCID: PMC378627
 165. G. Yang, J. Addai, M. Ittmann, T. M. Wheeler and T. C. Thompson: Elevated caveolin-1 levels in African-American versus white-American prostate cancer. *Clin Cancer Res*, 6(9), 3430-3 (2000)
 166. V. Mouraviev, L. Li, S. A. Tahir, G. Yang, T. M. Timme, A. Goltsov, C. Ren, T. Satoh, T. M. Wheeler, M. M. Ittmann, B. J. Miles, R. J. Amato, D. Kadmon and T. C. Thompson: The role of caveolin-1 in androgen insensitive prostate cancer. *J Urol*, 168(4 Pt 1), 1589-96 (2002)
 167. D. J. Waxman, C. Attisano, F. P. Guengerich and D. P. Lapenson: Human liver microsomal steroid metabolism: identification of the major microsomal steroid hormone 6 beta-hydroxylase cytochrome P-450 enzyme. *Arch Biochem Biophys*, 263(2), 424-36 (1988)
DOI: 10.1016/0003-9861(88)90655-8
 168. D. J. Waxman, D. P. Lapenson, T. Aoyama, H. V. Gelboin, F. J. Gonzalez and K. Korzekwa: Steroid hormone hydroxylase specificities of eleven cDNA-expressed human cytochrome P450s. *Arch Biochem Biophys*, 290(1), 160-6 (1991)
DOI: 10.1016/0003-9861(91)90602-F
 169. V. Ozdemir, W. Kalow, B. K. Tang, A. D. Paterson, S. E. Walker, L. Endrenyi and A. D. Kashuba: Evaluation of the genetic component of variability in CYP3A4 activity: a repeated drug administration method. *Pharmacogenetics*, 10(5), 373-88 (2000)
DOI: 10.1097/00008571-200007000-00001
PMid:10898107
 170. S. J. Plummer, D. V. Conti, P. L. Paris, A. P. Curran, G. Casey and J. S. Witte: CYP3A4 and CYP3A5 genotypes, haplotypes, and risk of prostate cancer. *Cancer Epidemiol Biomarkers Prev*, 12(9), 928-32 (2003)
 171. A. Loukola, M. Chadha, S. G. Penn, D. Rank, D. V. Conti, D. Thompson, M. Cicek, B. Love, V. Bivolarevic, Q. Yang, Y. Jiang, D. K. Hanzel, K. Dains, P. L. Paris, G. Casey and J. S. Witte: Comprehensive evaluation of the association between prostate cancer and genotypes/haplotypes in CYP17A1, CYP3A4, and SRD5A2. *Eur J Hum Genet*, 12(4), 321-32 (2004)
DOI: 10.1038/sj.ejhg.5201101
PMid:14560315
 172. I. J. Powell, J. Zhou, Y. Sun, W. A. Sakr, N. P. Patel, L. K. Heilbrun and R. B. Everson: CYP3A4 genetic variant and disease-free survival among white and black men after radical prostatectomy. *J Urol*, 172(5 Pt 1), 1848-52 (2004)
 173. S. Leskela, E. Honrado, C. Montero-Conde, I. Landa, A. Cascon, R. Leton, P. Talavera, J. M. Cozar, A. Concha, M. Robledo and C. Rodriguez-Antona: Cytochrome P450 3A5 is highly expressed in normal prostate cells but absent in prostate cancer. *Endocr Relat Cancer*, 14(3), 645-54 (2007)
DOI: 10.1677/ERC-07-0078
PMid:17914095
 174. P. Kuehl, J. Zhang, Y. Lin, J. Lamba, M. Assem, J. Schuetz, P. B. Watkins, A. Daly, S. A. Wrighton, S. D. Hall, P. Maurel, M. Relling, C. Brimer, K. Yasuda, R. Venkataramanan, S. Strom, K. Thummel, M. S. Boguski and E. Schuetz: Sequence diversity in CYP3A promoters and characterization of the genetic basis of polymorphic CYP3A5 expression. *Nat Genet*, 27(4), 383-91 (2001)
DOI: 10.1038/86882
PMid:11279519
 175. L. Zhenhua, N. Tsuchiya, S. Narita, T. Inoue, Y. Horikawa, H. Kakinuma, T. Kato, O. Ogawa and T. Habuchi: CYP3A5 gene polymorphism and risk of prostate cancer in a Japanese population. *Cancer Lett*, 225(2), 237-43 (2005)
DOI: 10.1016/j.canlet.2005.03.009
PMid:15876487
 176. M. H. Vaarala, H. Mattila, P. Ohtonen, T. L. Tammela, T. K. Paavonen and J. Schleutker: The interaction of CYP3A5 polymorphisms

- along the androgen metabolism pathway in prostate cancer. *Int J Cancer*, 122(11), 2511-6 (2008)
DOI: 10.1002/ijc.23425
PMid:18306354
177. T. L. Domanski, C. Finta, J. R. Halpert and P. G. Zaphiropoulos: cDNA cloning and initial characterization of CYP3A43, a novel human cytochrome P450. *Mol Pharmacol*, 59(2), 386-92 (2001)
178. A. Stone, L. D. Ratnasinghe, G. L. Emerson, R. Modali, T. Lehman, G. Runnells, A. Carroll, W. Carter, S. Barnhart, A. A. Rasheed, G. Greene, D. E. Johnson, C. B. Ambrosone, F. F. Kadlubar and N. P. Lang: CYP3A43 Pro(340) Ala polymorphism and prostate cancer risk in African Americans and Caucasians. *Cancer Epidemiol Biomarkers Prev*, 14(5), 1257-61 (2005)
DOI: 10.1158/1055-9965.EPI-04-0534
PMid:15894682
179. C. Siemes, L. E. Visser, F. H. de Jong, J. W. Coebergh, A. G. Uitterlinden, A. Hofman, B. H. Stricker and R. H. van Schaik: Cytochrome P450 3A gene variation, steroid hormone serum levels and prostate cancer-The Rotterdam Study. *Steroids*, 75(12), 1024-32 (2010)
DOI: 10.1016/j.steroids.2010.07.001
PMid:20621111
180. H. L. Ratan, A. Gescher, W. P. Steward and J. K. Mellon: ErbB receptors: possible therapeutic targets in prostate cancer? *BJU Int*, 92(9), 890-5 (2003)
181. F. M. Sirotnak, M. F. Zakowski, V. A. Miller, H. I. Scher and M. G. Kris: Efficacy of cytotoxic agents against human tumor xenografts is markedly enhanced by coadministration of ZD1839 (Iressa), an inhibitor of EGFR tyrosine kinase. *Clin Cancer Res*, 6(12), 4885-92 (2000)
182. F. M. Sirotnak, Y. She, F. Lee, J. Chen and H. I. Scher: Studies with CWR22 xenografts in nude mice suggest that ZD1839 may have a role in the treatment of both androgen-dependent and androgen-independent human prostate cancer. *Clin Cancer Res*, 8(12), 3870-6 (2002)
183. D. B. Agus, R. W. Akita, W. D. Fox, G. D. Lewis, B. Higgins, P. I. Pisacane, J. A. Lofgren, C. Tindell, D. P. Evans, K. Maiese, H. I. Scher and M. X. Sliwkowski: Targeting ligand-activated ErbB2 signaling inhibits breast and prostate tumor growth. *Cancer Cell*, 2(2), 127-37 (2002)
DOI: 10.1016/S1535-6108(02)00097-1
184. G. Di Lorenzo, G. Tortora, F. P. D'Armiento, G. De Rosa, S. Staibano, R. Autorino, M. D'Armiento, M. De Laurentiis, S. De Placido, G. Catalano, A. R. Bianco and F. Ciardiello: Expression of epidermal growth factor receptor correlates with disease relapse and progression to androgen-independence in human prostate cancer. *Clin Cancer Res*, 8(11), 3438-44 (2002)
185. B. Shuch, M. Mikhail, J. Satagopan, P. Lee, H. Yee, C. Chang, C. Cordon-Cardo, S. S. Taneja and I. Osman: Racial disparity of epidermal growth factor receptor expression in prostate cancer. *J Clin Oncol*, 22(23), 4725-9 (2004)
DOI: 10.1200/JCO.2004.06.134
PMid:15570072
186. D. A. Douglas, H. Zhong, J. Y. Ro, C. Oddoux, A. D. Berger, M. R. Pincus, J. M. Satagopan, W. L. Gerald, H. I. Scher, P. Lee and I. Osman: Novel mutations of epidermal growth factor receptor in localized prostate cancer. *Front Biosci*, 11, 2518-25 (2006)
DOI: 10.2741/1986
PMid:16720329
187. M. Gibbs, J. L. Stanford, R. A. McIndoe, G. P. Jarvik, S. Kolb, E. L. Goode, L. Chakrabarti, E. F. Schuster, V. A. Buckley, E. L. Miller, S. Brandzel, S. Li, L. Hood and E. A. Ostrander: Evidence for a rare prostate cancer-susceptibility locus at chromosome 1p36. *Am J Hum Genet*, 64(3), 776-87 (1999)
DOI: 10.1086/302287
PMid:10053012 PMCID: PMC1377795
188. H. Matsui, K. Suzuki, N. Ohtake, S. Nakata, T. Takeuchi, H. Yamanaka and I. Inoue: Genomewide linkage analysis of familial prostate cancer in the Japanese population. *J Hum Genet*, 49(1), 9-15 (2004)
DOI: 10.1007/s10038-003-0099-y
PMid:14666403
189. R. A. Kittles, A. B. Baffoe-Bonnie, T. Y. Moses, C. M. Robbins, C. Ahaghotu, P. Huusko, C. Pettaway, S. Vijayakumar, J. Bennett, G. Hoke, T. Mason, S. Weinrich, J. M. Trent, F. S. Collins, S. Mousses, J. Bailey-Wilson, P. Furbert-Harris, G. Dunston, I. J. Powell and J.

- D. Carpten: A common nonsense mutation in EphB2 is associated with prostate cancer risk in African American men with a positive family history. *J Med Genet*, 43(6), 507-11 (2006)
DOI: 10.1136/jmg.2005.035790
PMid:16155194 PMCID: PMC2564535
190. H. Williams, I. J. Powell, S. J. Land, W. A. Sakr, M. R. Hughes, N. P. Patel, L. K. Heilbrun and R. B. Everson: Vitamin D receptor gene polymorphisms and disease free survival after radical prostatectomy. *Prostate*, 61(3), 267-75 (2004)
DOI: 10.1002/pros.20103
PMid:15368470
191. I. M. Shui, L. A. Mucci, P. Kraft, R. M. Tamimi, S. Lindstrom, K. L. Penney, K. Nimptsch, B. W. Hollis, N. Dupre, E. A. Platz, M. J. Stampfer and E. Giovannucci: Vitamin D-related genetic variation, plasma vitamin D, and risk of lethal prostate cancer: a prospective nested case-control study. *J Natl Cancer Inst*, 104(9), 690-9 (2012)
DOI: 10.1093/jnci/djs189
PMid:22499501 PMCID: PMC3341310
192. L. B. Signorello, S. M. Williams, W. Zheng, J. R. Smith, J. Long, Q. Cai, M. K. Hargreaves, B. W. Hollis and W. J. Blot: Blood vitamin d levels in relation to genetic estimation of African ancestry. *Cancer Epidemiol Biomarkers Prev*, 19(9), 2325-31 (2010)
DOI: 10.1158/1055-9965.EPI-10-0482
PMid:20647395 PMCID: PMC2938736

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