Clinical implications and pathological associations of circulating mitochondrial DNA

Eszter Tuboly¹, Daniel McIlroy^{1,2}, Gabrielle Briggs², Natalie Lott¹, Zsolt J Balogh^{1,2}

¹Department of Traumatology, John Hunter Hospital, Lookout Road, New Lambton Height, NSW 2305 Newcastle, NSW 2310, Australia, ²Department of Traumatology, University of Newcastle, University Drive, Callaghan NSW 2308, Australia

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
 - 2.1. Conditions when mitochondrial DNA release is accompanied by mechanical and chemical stress
 - 2.1.1. Mechanical stress related mtDNA release
 - 2.1.2. Stress causes oxidative damage and a subsequent DNA release in mitochondria
 - 2.1.3. Cell death pathways culminating in lysis
 - 2.1.4. Oxidative cellular damage pathways initiated by mitochondria
- 3. Circulating mitochondrial DNA in innate immune responses
 - 3.1. MtDNA and TLR interaction
 - 3.2. MtDNA-Nod-like receptor 3 relationship
 - 3.3. MtDNA-STING pathway relationship
 - 3.4. MtDNA-neutrophil extracellular trap formation
- 4. Clinical implications of cell-free mtDNA
 - 4.1. Diagnostic application of mtDNA
 - 4.2. Therapeutical implications of mtDNA
- 5. Future directions
- 6. Acknowledgement
- 7. References

1. ABSTRACT

Mitochondria are membrane-enclosed organelles, the energy-producing centers in almost all eukaryotic cells. The evolutionary emergence of mitochondria is a result of the endocytosis of a-proteobacteria. There are several characteristic features which refer to its prokaryotic ancestors including its independent sets of double-stranded mitochondrial DNA, which is uniquely circular in form and contains a significant amount of unmethylated DNA as CpG islands. Resent research has proven that free mitochondrial DNA found in blood was associated with innate immunomodulation in a broad-range of clinical conditions. Upon release, mitochondrial DNA acts as a danger-associated molecular pattern in the circulation, it is recognized by pattern recognition receptors and it facilitates inflammatory responses. Besides its high receptor activation potential, mitochondrial DNA is likely to perform direct crosstalk with activated leukocytes and to be contributed to other anti-microbial activities. Here we highlight the pathological conditions where cell free mtDNA is involved, describe the potential sources and mechanisms of extracellular mtDNA release and explore evidence for its mechanism of action after being excreted and potential therapeutic strategies.

2. INTRODUCTION

Due to its high mutation rate, mitochondrial DNA (mtDNA) has been studied in the context of ageingrelated and multifactorial diseases where mitochondrial dysfunction is thought to play a role (1, 2). However, in the last few years, a novel and significant role of mtDNA has emerged, involving its ability to trigger innate immune system responses and drive inflammation when released from mechanically injured cells (3). MtDNA, along with other host molecules released upon cell damage, falls into the category of damage-associated molecular patterns (DAMPs). According to the endosymbiotic theory, the evolutionary emergence of mitochondria is a result of the endocytosis of a-proteobacteria (4, 5). Phylogenetic investigations support this assumption, as mitochondria possess several features characteristic of their prokaryotic ancestors, such as their barrel-shape and diameter of 0.2. to 1.0. µm. Mitochondria have their own independent sets of double-stranded mitochondrial DNA (mtDNA), which is not linear like nuclear DNA, but circular in form and consists of a high number of unmethylated CpG islands. as typically found in bacteria (6). The genome size of mtDNA is significantly smaller than the nuclear (16.5.69 bp vs. 3.2. billion bp) in humans, the number of encoding mitochondrial genes is only 37, which encode no more

than 16 proteins, all belonging to the electron transportchain (7). The release of mtDNA and its presence in the circulation were described to play a significant role in various clinical conditions; in particular, inflammatory diseases, several types of cancer and in conditions leading to critical illness requiring intensive care unit admission (8-10). Currently, several conditions are being explored where mechanical and chemical stress can lead to cellular necrosis accompanied by mtDNA release. This review will provide a summary of the pathological conditions where cell free mtDNA is involved, describe the potential sources and mechanisms of extracellular mtDNA release and explore evidence for its mechanism of action after being excreted and potential therapeutic strategies.

2.1. Conditions when mitochondrial DNA release is accompanied by mechanical and chemical stress

The release of mtDNA has been described in diverse inflammation and cell necrosis-related clinical conditions, where the loss of cell membrane integrity leads to the release of intracellular content (11). However, it is important to note that in these conditions, the stress destroying the mitochondria can be of either extracellular or intra-mitochondrial origin, with the latter case initiated by oxidative burst inside the mitochondrion, resulting in mitochondrial and cellular disintegration (12). As such, mechanisms of mtDNA release may vary depending on the origin and the type of insult.

2.1.1. Mechanical stress related mtDNA release

Any type of cell death that culminates in lysis and subsequent release of intracellular content into the extracellular environment could conceivably result in mtDNA being released into the circulation (11). MtDNA and other particles are not exposed to the innate immune system following normal apoptosis, but cell death due to mechanical stress and subsequent lysis can mediate their entry into the systemic circulation to provoke immune response (13). Tissue injury is one such example of mechanical stress leading to release of mtDNA (14). In major trauma patients, the initial cell death due to the injury is an important but nonmodifiable factor for post-injury inflammation-associated complications (15). The recognition of mtDNA by innate immune cells (primarily neutrophils) plays a pivotal role in the pathophysiology of sterile inflammation after major trauma (16). Our group has hypothesized that mtDNA may have a primary inflammatory source following major trauma rather than as a result of direct tissue injury and subsequent cell necrosis (17). It is supported by our recent findings, where elevated and increasing concentration of mtDNA was shown to be present in the sera of major trauma patients. We also identified sustained high concentrations of cell free mtDNA in the sera of postoperative trauma patients undergoing major orthopedic trauma surgery without association with wellestablished markers of tissue necrosis (17).

2.1.2. Stress causes oxidative damage and a subsequent DNA release in mitochondria

In addition to its presence in major trauma patients, alterations in mtDNA content of the blood have been measured in a number of diseases relating to oxidative stress. Besides the measurable oxidative damage, the involvement of the mitochondrial genome was described in response to focal or acute myocardial ischemia/reperfusion in animal models and in human cells (18-20). Elevated blood mtDNA content was described to be in association with higher cardiovascular risk or development of coronary heart disease (21, 22). This latter study was conducted on patients with diabetes mellitus and others found hyperglycemia-induced elevation in mtDNA of the peripheral blood in early diabetes (23). Likewise, neurodegenerative diseases such as Alzheimer's or Huntington's disease are also seemed to be associated with alterations in mtDNA concentration (24-26). Also, different types of cancer or critical conditions like sepsis or hemorrhagic shock are considered to be connected with mtDNA copy number changes (16, 27-30). In the context of highaltitude oxygen deprivation, blood mtDNA content was observed to be increased in lowlanders as compared to highlanders, regardless of age and gender, suggesting that the changes in mtDNA concentration might be due to the ROS-stress adaptation mechanisms (31, 32). Both acute and chronic inflammatory diseases have been associated with increased cell-free mtDNA and correlate with elevated free radical production that may have originated from mitochondria (33), but the question of the source of ROS remains open, as the ROS productive capacity of immune cells is also well-known in these conditions. Liu and co-workers published in 2003, that the copy number of mtDNA in human leukocytes was highly affected by alterations in plasma antioxidants/ pro-oxidants (34). In addition to its role in diseases itself, mtDNA is also affected by diagnostic and therapeutic tools. Mitochondria are highly susceptible to ionizing radiation at the clinically relevant dosages and oxidative stress resulting from irradiation was found to be accompanied by a rise in extracellular mtDNA release (35, 36). In fact, the natural process of aging is relevant here, since oxidative stress within aging mitochondria can lead to a vicious cycle in which damaged mitochondria produce increased amounts of reactive oxygen species. This could explain the significant increase in the mtDNA mutation rate also found in human clinical studies in healthy older people's plasma, as compared to young volunteers (37, 38). The cut-off for clinically relevant rise in these mutations is likely to be close to the 6th decade of life in humans, highlighting the importance of age-matching in mtDNA concentration-based human studies (39).

Paradoxically, oxidative injury to mitochondria takes place during "reductive stress": when electron acceptors are expected to be mostly reduced, some redox proteins can donate electrons to O_2 instead,

1012 © 1996-2017

which increases the NADH/NAD+ ratio of mitochondria. Conditions such as high-intensity exercise training, alcohol intake or chronic fatigue syndrome and other forms of hyperglycemia-induced diseases all result in reductive stress and were reported to be accompanied by cellular damage and therefore mtDNA excretion (40-42).

2.1.3. Cell death pathways culminating in lysis

If a cell is directly injured through a physical insult or severely stress it may become necrotic. Necrosis is characterized morphologically by cell rounding, swelling (oncosis), and expansion of organelles and de-condensation of nuclear chromatin (43). This process culminates in cell lysis. Recent evidence suggests that complex signal transduction mechanisms can control necrosis (43). Further cell death essentially occurs as a result of the physiological stress caused by the mass release of cytokines and other cell signaling molecules from injured tissues and the innate immune cells involved in the acute inflammatory response. "Necrosis" in the postinjury state can be triggered through subsequent complex tightly regulated intracellular signaling cascades, not just through the initial mechanical tissue injury. Necrosis can be triggered by exogenous molecules such as TNFa and Fas ligand binding to cell surface receptors (44). The activation of such receptors can lead to a tightly regulated and controlled form of necrosis. This process is mediated through caspase-8 (anti-cell death enzyme) and receptor interacting protein kinases (RIPK family) and the term "necropoptosis" coined for it (11). The role of DAMPs in triggering necropoptosis has also been explored through their activation of pathogen recognition receptors (PRRs) which then trigger intracellular signaling cascades through RIPK1 and RIPK3 (11). Whilst RIPK1 and RIPK3 can play a role, they are not essential in necrosis following ischemia reperfusion (IR) injury (44). This is characterized by exposure to high levels of hydrogen peroxide (H2O2) and is dependent on the activity of a different enzyme poly (ADP-ribose) polymerase (44). Free intracellular iron redox reactions with H₂O₂ appear to play a pivotal role in this modality of cell death by inducing lysosomal permeability (43). Intracellular chelation of free iron was demonstrated to be cell-protective in such conditions (43). Regardless of the initiating stimulus, loss of membrane continuity and lysis leads to the extravasation of the intracellular contents, including mtDNA and associated mtDAMPs into the extra-cellular environment.

2.1.4. Oxidative cellular damage pathways initiated by mitochondria

The majority of ROS are products of mitochondrial respiration, as the electrontransport-chain contains several redox centers that may leak electrons to O_2 , serving as the primary source of O_2 production in most tissues (12). A major threat to this controlled equilibrium is hypoxia, since the absence of the electron acceptor O_2 leads to a shift in reducing potential to a

higher than normal reducing power, which results in progressive structural and functional cell damage. It is therefore widely accepted that several disease states are linked to "oxidative stress" and a subsequent mtDNA release (45). The effect of abnormal Ca²⁺ is also inevitable in response to stress, but the mechanisms of the harmful effect of Ca2+ on mitochondria is not well characterized (46). ROS and Ca2+ constitute important mediators of the propagation of the necrotic signal from the mitochondrial matrix towards the outside and can cause damage to all of the major classes of biological macromolecules, including nucleic acids, proteins, carbohydrates, and lipids (47). Mitochondrial calcium has been described to stimulate oxidative phosphorylation, thereby promoting more ROS generation (48). In addition calcium-mediated activation of calpain can lead to cleavage and inactivation of caspases (49) whereas ROS can target the active site of caspases and render them inactive, promoting necrosis (50). Likewise, mitochondrial H_oO_o can cause the release of cytochrome c from mitochondria into the cytosol and H2O2 may also activate nuclear transcription factors, like NF-κB, AP-1, and p53, which may upregulate death proteins or produce inhibitors of survival proteins (51).

3. CIRCULATING MITOCHONDRIAL DNA IN INNATE IMMUNE RESPONSES

Despite their seemingly independent existence within the cell, mitochondrial transcription and replication are co-dependent on nuclear encoded factors transported into mitochondria (52). The aforementioned similarity to bacterial DNA makes mtDNA highly immunostimulatory to cells of the innate immune system (6). The latest evidence suggests that it does not only facilitate antibacterial immune responses, but significantly contributes to further adverse effects and may have important roles in inflammatory diseases and complicated outcomes following cellular damage or oxido-reductive stress (8). MtDNA has been shown to bind PRRs, namely to the Tolllike receptor (TLR) superfamily members or nucleotide oligomerization domain (NOD)-like receptors (NLRs) and more recently it has been shown to be linked with the stimulator of interferon genes (STING) pathway (53, 54).

3.1. MtDNA and TLR interaction

MtDNA has been demonstrated to induce neutrophil activation and facilitates adverse immune reactions through activation of TLR 4 (55) and TLR 9 (56), mediated through MAP kinases p38 (16, 57) and p44/42 (58). MtDNA triggers activation of the nuclear factor kappa B pathway (NFkB) via TLR9, resulting in upregulation of pro-inflammatory cytokine production including TNF- α (59), IL-1 β (60) and IL-6 (61). MtDAMPs have been shown to potentiate inflammatory lung injury when introduced into healthy rats in a landmark paper by Zhang and colleagues (9). One possible contributory factor is that mtDNA triggers increased neutrophil

expression of matrix metalloprotease 8 (MM8) through p38 activation, which is a collagen cleavage enzyme that potentiates tissue degradation (57).

3.2. MtDNA-Nod-like receptor 3 relationship

Of the NLR receptors NLR pyrin domain 3 (NLRP3) inflammasome is the most widely studied mainly due to its affinity for a wide variety of ligands (62). Mitochondria have been implicated in the recruitment of NLRP3 in a variety of different ways including through direct activation with mtDNA (63). The assembly of the NLRP3 inflammasome in complexes containing caspase-1 has now been directly implicated in triggering a novel form of cell death termed "pyroptosis" (64). Interestingly when cells lack mtDNA (induced by treatment with ethidium bromide) NLRP3 inflammasome formation was completely inhibited (65). Conversely, NLRP3 inflammasome formation releases mtDNA (65). This indicates a possible positive feedback loop where mtDNA potentiates its own release by stimulating further NLRP3 inflammasome formation.

3.3. MtDNA-STING pathway relationship

MtDNA has the ability to stimulate the innate immune system through stimulation of interferon genes (STING) pathway, resulting in interferon release. The STING pathway was recently mechanistically dissected to reveal an intricate relationship demonstrating how mtDNA triggers interferon release (54). The study showed that through depletion of mitochondrial transcription factor A (TFAM) during a herpes viral infection, mtDNA stability was disturbed, causing enlargement of the mitochondrial nucleoid. Subsequently, fragmented mtDNA was released, activating peri-mitochondrial cyclic GMP-AMP synthase (cGAS) causing increased cGAMP formation. The second messenger cGAMP then activates the endoplasmic reticulum bound STING pathway which ultimately upregulates type I interferon (IFN I) expression which inhibits viral propagation. Interestingly, proapoptotic caspase activation inhibits this response and suppresses downstream interferon production (66).

3.4. MtDNA-neutrophil extracellular trap formation

Neutrophil extracellular trap (NET) formation or "NETosis" was first described by Brinkmann and colleagues in 2004 (67). It is characterized by smooth extracellular filaments-17nm in diameter- which are composed of stacked and probably modified nucleosomes (68). This filamentous chromatin backbone is adorned with globular domains of approximately 50nm diameter containing neutrophilic granular proteins. The principle function of the NET is believed to be to entrap and kill circulating pathogens and this function has been directly shown in both Gram- and Gram+ bacteria, viruses and fungi (68, 69).

The composition of NETs was initially widely believed to be predominantly nuclear DNA (nDNA),

however under specific stimulatory conditions NETs composed exclusively of mtDNA were demonstrated (70). Our group described that NETs formed after trauma and subsequent surgery were predominantly composed of mtDNA (71). More recently NETs rich in oxidized mtDNA have been discovered in systemic lupus erythematous (72). The emerging body of evidence suggesting NETs can indeed be composed exclusively or predominantly of mtDNA means NETosis may represent a significant source of circulating mtDNA in certain inflammatory conditions.

In addition to the role of intracellular mtDNA in NET composition, mtDNA may also trigger NET formation as a DAMP. NETosis has widely been considered as a NADPH oxidase (PHOX) dependent process, reliant on mitochondrial release of reactive oxygen species (73). However mtDNA as a trigger for NETosis is a much more recent concept and there is growing evidence that extracellular trap formation takes place independently from pro-oxidant activity (74, 75). MtDNA has been demonstrated to be a trigger for NETosis after major trauma and with signaling mediated through a TLR9dependent pathway, independent of PHOX (38). The concept of mtDNA as a signaling molecule involved in NETosis suggests it may have a more diverse role in regulating certain inflammatory processes in a novel and previously unstudied way.

4. CLINICAL IMPLICATIONS OF CELL-FREE mtDNA

4.1. Diagnostic application of mtDNA

The number of studies investigating the concentration of mtDNA as a potential biomarker in different human body fluids has grown significantly in recent years. Real-time PCR allows simultaneous detection and quantification of mtDNA using a small amount of sample and a downstream real-time PCR analysis give an accurate reproducible result within 2 hrs. The detection from blood, saliva, urine or sperm is a minimally or non-invasive process for diagnosis and was proven to be valuable for the prognosis of various clinical conditions, such as different types of cancer, type 2 diabetes, sepsis, multiple organ failure, fertility impairment or neurodegenerative disorders (28, 76-81). However, the exact cellular mechanisms and cell-type of origin which cause mtDNA concentration to fluctuate in many conditions remain unclear.

Elevated mtDNA content in peripheral blood has been demonstrated as a diagnostic factor in various types of cancer, including non-Hodgkin lymphoma, lung cancer, pancreatic cancer, breast cancer, colorectal cancer, or glioma (81-86). In contrast, an increased risk of renal cancer or hepatocellular carcinoma was observed to be associated with decreased circulating mtDNA concentrations within the tumor tissues of

cancer patients (87, 88). In other human studies, mtDNA quantity measured in the blood of patients with sepsis, pulmonary embolism or out-of-hospital cardiac arrest was proven to be a more powerful prognostic marker than those conventionally used, including nuclear DNA or other existing semiquantitative score systems (89, 90).

The rapidly elevated concentrations of circulating mtDNA levels that are observed in trauma patients with severe injury suggests that extracellular DNA originates from direct tissue injury and subsequent necrosis. It was described to be a trustworthy prognostic marker either in blood or in cerebrospinal fluid with good prediction for unfavorable outcome, or even mortality (65, 91-93). Although, in some studies, nuclear DNA concentration and well-established markers of tissue necrosis were not found to correlate with mtDNA levels, or mtDNA concentration was observed to have no contribution in the pathophysiology of critical illness (17, 29, 94). These findings together with the abovementioned dichotomies, raise some concerns regarding the nucleic acid-based diagnosis. The reduced clearance of DNA over time caused by impaired organ function during systemic inflammation may also be a contributing factor (92) and similarly, the limited capacity of inflammatory cells for taking up dying cells, thereby DNA (95). In the near future, investigation of the mtDNA methylation pattern rather than its concentration might be used for the diagnostic purpose for identifying tissue specific origin, as it was successfully performed to predict cardiovascular problems or amyotrophic lateral sclerosis (96, 97) and suggests a promising approach to diagnose health problems caused by environmental pollution exposure, aging, drug treatment, and oxidative stress (98). Moreover, since mitochondria do not contain histones, it is likely that the mtDNA methylation/hydroxymethylation ratio rather than histone modification is important for mitochondrial genome-based diagnostics.

4.2. Therapeutical implications of mtDNA

Major trauma patients often require lifesaving allogenic blood products in which cellular remnants, such as mitochondria and extracellular mtDNA are described to be present and to mediate adverse inflammatory processes, as neutrophil, eosinophil and basophil leukocyte activation (99, 100). It is important to take into account, that platelet units represent a potential reservoir of mtDNA, since unlike leukodepleted red blood cell units, stored platelets contain mitochondria.

Moreover, other therapies might also cause cytolysis and may be accompanied by circulatory mtDNA release. Plasma mtDNA content was proven to be elevated and observed to mediate pro-inflammatory effects in maintained haemodialysis patients (101) and

in another study, increased mtDNA amount in the plasma of patients was considered to be related to the overall procedure of artificial kidney therapy and probably was due to the death of leukocytes (102).

The fact that mtDNA has such potent immunostimulatory effects makes it an exciting target for immunomodulation therapy attenuating some of the potentially deleterious effects of excessive innate immune activation. Whether mtDNA is free or conjugated in NETs it is readily digestible with DNAse. There is certainly good evidence to suggest that focally targeting NETs with DNAse has yielded a reduction in associated inflammatory lung damage in a mouse model of transfusion related acute lung injury (TRALI) (103). Human recombinant DNAse therapy has been used to good effect when nebulized in cystic fibrosis patients by enhancing sputum solubilisation (104), however no studies have been performed in humans to treat acute inflammatory conditions. With such an emergent role of mtDNA in NETs associated with trauma (71) and more recently in SLE (105) the investigation of DNAse therapy in different inflammatory conditions would be very reasonable.

Targeting mtDNA receptors may also yield ways to modify its proinflammatory properties. It has a diverse role as a signalling molecule in various inflammatory pathways as a ligand of multiple receptors including mtDNA stimulation of the NLRP3 inflammasome (63), cGAS in mtDNA-STING pathway (54) and TLR9 in mtDNA mediated NETosis (38). Modulation of these receptors may convey benefit in a variety of clinical conditions and attenuate the immunostimulatory effects of mtDNA.

5. FUTURE DIRECTIONS

It is essential to understand the tissue specific origin of circulating mtDNA for both diagnostic and therapeutic considerations. The natural history of free polynucleotides in the circulation in inflammatory conditions is largely unknown. The available active DNAse concentration in physiological and pathological conditions could indicate the potential need for enzyme supplementation as a therapeutic strategy. We believe that our current knowledge on cell free circulating mtDNA is in a rather exploratory phase with a potential for the future to rewrite the pathology of the leading causes of morbidity and mortality such as inflammatory conditions, autoimmune disorders, cancer, heart disease, stroke and injury.

6. ACKNOWLEDGEMENT

Eszter Tuboly and Daniel McIlroy contributed equally to this paper. The authors have no conflicts of interest to disclose.

7. REFERENCES

Johns DR: The other human genome: mitochondrial DNA and disease. Nat Med 2,1065-8 (1996)

DOI: 10.1038/nm1096-1065

- 2. Khan N: Recent advancements in diagnostic tools in mitochondrial energy metabolism diseases. Adv Med Sci 61,244-248. (Epub ahead of print) (2016)
- Krysko DV, Agostinis P, Krysko O, Garg AD, Bachert C, Lambrecht BN, Vandenabeele P: Emerging role of damage-associated molecular patterns derived from mitochondria inflammation. Trends Immunol 32,157-64 (2011)

DOI: 10.1016/j.it.2011.01.005

- Gray MW, Burger G, Lang BF: Mitochondrial evolution. Science 283,1476-81 (1999) DOI: 10.1126/science.283.5407.1476
- Zimmer C: Origins. On the origin of eukaryotes. Science 325.666-8 (2009) DOI: 10.1126/science.325 666
- 6. Hochhauser D: Relevance of mitochondrial DNA in cancer. *Lancet* 356,181-2 (2000) DOI: 10.1016/S0140-6736(00)02475-2
- Anderson S, Bankier AT, Barrell BG, de Bruijn MH, Coulson AR, Drouin J, Eperon IC, Nierlich DP, Roe BA, Sanger F, Schreier PH, Smith AJ, Staden R, Young IG: Sequence and organization of the human mitochondrial genome. *Nature* 290,457-65 (1981) DOI: 10.1038/290457a0
- Collins LV, Hajizadeh S, Holme E, Jonsson IM, Tarkowski A: Endogenously oxidized mitochondrial DNA induces in vivo and in vitro inflammatory responses. J Leukoc Biol 75,995-1000 (2004)

DOI: 10.1189/jlb.0703328

- Zhang Q, Raoof M, Chen Y, Sumi Y, Sursal T, Junger W, Brohi K, Itagaki K, Hauser CJ: Circulating mitochondrial DAMPs cause inflammatory responses to injury. Nature 464,104-7 (2010) DOI: 10.1038/nature08780
- 10. Kepp O, Senovilla L, Vitale I, Vacchelli E, Adjemian S, Agostinis P, Apetoh L, Aranda F. Barnaba V. Bloy N. Bracci L. Breckpot K. Brough D, Buqué A, Castro MG, Cirone M, Colombo MI, Cremer I, Demaria S, Dini L,

- Eliopoulos AG, Faggioni A, Formenti SC, Galluzzi L and co-workers: Consensus guidelines for the detection of immunogenic cell death. Oncoimmunology 3(9):e955691. eCollection (2014)
- P. 11. Kaczmarek Vandenabeele Krysko DV: Necroptosis: the release of damage-associated molecular patterns and its physiological relevance. Immunity 38,209-23 (2013) DOI: 10.1016/j.immuni.2013.02.003
- Dröse S, Brandt U: Molecular mechanisms of superoxide production by the mitochondrial respiratory chain. Adv Exp Med Biol 748,145-69 (2012) DOI: 10.1007/978-1-4614-3573-0 6
- 13. Murakami Y, Matsumoto H, Roh M, Giani A, Kataoka K, Morizane Y, Kayama M, Thanos A, Nakatake S, Notomi S, Hisatomi T, Ikeda Y, Ishibashi T, Connor KM, Miller JW, Vavvas DG: Programmed necrosis, not apoptosis, is a key mediator of cell loss and DAMP-mediated inflammation in dsRNA-induced retinal degeneration. Cell Death Differ 21,270-7 (2014) DOI: 10.1038/cdd.2013.109
- 14. Simmons JD, Lee YL, Mulekar S, Kuck JL, Brevard SB. Gonzalez RP. Gillespie MN. Richards WO: Elevated levels of plasma mitochondrial DNA DAMPs are linked to clinical outcome in severely injured human subjects. Ann Surg 258,591-6; discussion 596-8 (2013)
- Balogh ZJ, Reumann MK, Gruen RL, Mayer-Kuckuk P, Schuetz MA, Harris IA, Gabbe BJ, Bhandari M: Advances and future directions for management of trauma patients with musculoskeletal injuries. Lancet 380,1109-19 (2012) DOI: 10.1016/S0140-6736(12)60991-X
- 16. Zhang Q, Itagaki K, Hauser CJ: Mitochondrial DNA is released by shock and activates neutrophils via p38 map kinase. Shock 34,55-9 (2010) DOI: 10.1097/SHK.0b013e3181cd8c08
- 17. McIlroy DJ, Bigland M, White AE, Hardy BM, Lott N, Smith DW, Balogh ZJ: Cell necrosisindependent sustained mitochondrial and nuclear DNA release following trauma surgery. J Trauma Acute Care Surg 78,282-8 (2015) DOI: 10.1097/TA.0000000000000519
- 18. Chen H, Hu CJ, He YY, Yang DI, Xu J, Hsu CY:

- Reduction and restoration of mitochondrial dna content after focal cerebral ischemia/reperfusion. *Stroke* 32,2382-7 (2001) DOI: 10.1161/hs1001.097099
- Corbucci GC, Lettieri B, Luongo C, Orrù A, Musu M, Marchi A: Mitochondrial genome involvement in ischemia/reperfusion-induced adaptive changes in human myocardial cells. *Minerva Anestesiol* 72,337-47 (2006)
- Bliksøen M, Baysa A, Eide L, Bjørås M, Suganthan R, Vaage J, Stensløkken KO, Valen G: Mitochondrial DNA damage and repair during ischemia-reperfusion injury of the heart. J Mol Cell Cardiol 78,9-22 (2015) DOI: 10.1016/j.yjmcc.2014.11.010
- BerezinAE: Circulating Cell-Free Mitochondrial DNA as Biomarker of Cardiovascular risk: New Challenges of Old Findings. *Angiol* 3,161 (2015)
- Liu J, Cai X, Xie L, Tang Y, Cheng J, Wang J, Wang L, Gong J: Circulating Cell Free Mitochondrial DNA is a Biomarker in the Development of Coronary Heart Disease in the Patients with Type 2 Diabetes. Clin Lab 61,661-7 (2015)
- Malik AN, Parsade CK, Ajaz S, Crosby-Nwaobi R, Gnudi L, Czajka A, Sivaprasad S: Altered circulating mitochondrial DNA and increased inflammation in patients with diabetic retinopathy. *Diabetes Res Clin Pract* 110,257-65 (2015) DOI: 10.1016/j.diabres.2015.10.006
- 24. Lin MT, Cantuti-Castelvetri I, Zheng K, Jackson KE, Tan YB, Arzberger T, Lees AJ, Betensky RA, Beal MF, Simon DK: Somatic mitochondrial DNA mutations in early Parkinson and incidental Lewy body disease. Ann Neurol 71,850-4 (2012) DOI: 10.1002/ana.23568
- Wang HC, Lin YJ, Lin WC, Ho JT, Chen WF, Chang WN, Tsai NW, Lu CH: The value of serial plasma nuclear and mitochondrial DNA levels in acute spontaneous intra-cerebral haemorrhage. *Eur J Neurol* 19,1532-8 (2012) DOI: 10.1111/j.1468-1331.2012.03761.x
- 26. Petersen MH, Budtz-Jørgensen E, Sørensen SA, Nielsen JE, Hjermind LE, Vinther-Jensen T, Nielsen SM, Nørremølle A: Reduction in mitochondrial DNA copy number in peripheral leukocytes after onset of Huntington's disease. *Mitochondrion* 17,14-21 (2014)

- DOI: 10.1016/j.mito.2014.05.001
- Masayesva BG, Mambo E, Taylor RJ, Goloubeva OG, Zhou S, Cohen Y, Minhas K, Koch W, Sciubba J, Alberg AJ, Sidransky D, Califano J: Mitochondrial DNA content increase in response to cigarette smoking. Cancer Epidemiol Biomarkers Prev 15,19-24 (2006) DOI: 10.1158/1055-9965.EPI-05-0210
- 28. Yu M. Generation, function and diagnostic value of mitochondrial DNA copy number alterations in human cancers. *Life Sci* 89,65-71 (2011)
 DOI: 10.1016/j.lfs.2011.05.010
- Puskarich MA, Shapiro NI, Trzeciak S, Kline JA, Jones AE: Plasma levels of mitochondrial DNA in patients presenting to the emergency department with sepsis. Shock 38,337-40 (2012)
 DOI: 10.1097/SHK.0b013e318266a169
- Kosaka J, Morimatsu H, Takahashi T, Shimizu H, Kawanishi S, Omori E, Endo Y, Tamaki N, Morita M, Morita K: Effects of biliverdin administration on acute lung injury induced by hemorrhagic shock and resuscitation in rats. *PLoS One* 8,e63606 (2013)
- 31. Luo Y, Yang X, Gao Y: Mitochondrial DNA response to high altitude: a new perspective on high-altitude adaptation. *Mitochondrial DNA* 24,313-9 (2013)
 DOI: 10.3109/19401736.2012.760558
- 32. Li Y, Huang W, Yu Q, Cheng YT1, Kong QP: Lower mitochondrial DNA content relates to high-altitude adaptation in Tibetans. *Mitochondrial DNA* 27,753-7 (2016) DOI: 10.3109/19401736.2014.915526
- Escames G, López LC, García JA, García-Corzo L, Ortiz F, Acuña-Castroviejo D: Mitochondrial DNA and inflammatory diseases. *Hum Genet* 131,161-73 (2012) DOI: 10.1007/s00439-011-1057-y
- Liu CS, Tsai CS, Kuo CL, Chen HW, Lii CK, Ma YS, Wei YH: Oxidative stress-related alteration of the copy number of mitochondrial DNA in human leukocytes. *Free Radic Res* 37,1307-17 (2003) DOI: 10.1080/10715760310001621342
- Prithivirajsingh S, Story MD, Bergh SA, Geara FB, Ang KK, Ismail SM, Stevens CW, Buchholz TA, Brock WA: Accumulation of the common mitochondrial DNA deletion

- induced by ionizing radiation. FEBS Lett 571,227-32 (2004)
- DOI: 10.1016/j.febslet.2004.06.078
- Evdokimovsky EV, Ushakova TE, Kudriavtcev AA, Gaziev AI: Alteration of mtDNA copy number, mitochondrial gene expression and extracellular DNA content in mice after irradiation at lethal dose. *Radiat Environ Biophys* 50,181-8 (2011)
 DOI: 10.1007/s00411-010-0329-6
- Pinti M, Cevenini E, Nasi M, De Biasi S, Salvioli S, Monti D, Benatti S, Gibellini L, Cotichini R, Stazi MA, Trenti T, Franceschi C, Cossarizza A: Circulating mitochondrial DNA increases with age and is a familiar trait: Implications for "inflamm-aging". Eur *J Immunol* 44,1552-62 (2014) DOI: 10.1002/eji.201343921
- Itagaki K, Kaczmarek E, Lee YT, Tang IT, Isal B, Adibnia Y, Sandler N, Grimm MJ, Segal BH, Otterbein LE, Hauser CJ: Mitochondrial DNA released by trauma induces neutrophil extracellular traps. *PLoS One* 10,e0120549 (2015)
- 39. Trifunovic A: Mitochondrial DNA and ageing. *Biochim Biophys Acta* 1757,611-7 (2006) DOI: 10.1016/j.bbabio.2006.03.003
- Menshikova EV, Ritov VB, Fairfull L, Ferrell RE, Kelley DE, Goodpaster BH: Effects of exercise on mitochondrial content and function in aging human skeletal muscle. *J Gerontol A Biol Sci Med Sci* 61,534-40 (2006) DOI: 10.1093/gerona/61.6.534
- von Wurmb-Schwark N, Ringleb A, Schwark T, Broese T, Weirich S, Schlaefke D, Wegener R, Oehmichen M: The effect of chronic alcohol consumption on mitochondrial DNA mutagenesis in human blood. *Mutat Res* 637,73-9 (2008)
 - DOI: 10.1016/j.mrfmmm.2007.07.003
- Billing-Ross P, Germain A, Ye K, Keinan A, Gu Z, Hanson MR: Mitochondrial DNA variants correlate with symptoms in myalgic encephalomyelitis/chronic fatigue syndrome. J Transl Med 14,19 (2016)
- 43. Vanden Berghe T, Vanlangenakker N, Parthoens E, Deckers W, Devos M, Festjens N, Guerin CJ, Brunk UT, Declercq W, Vandenabeele P: Necroptosis, necrosis and secondary necrosis converge on similar cellular disintegration features. *Cell Death*

- Differ 17,922-30 (2010) DOI: 10.1038/cdd.2009.184
- 44. Vanlangenakker N, Vanden Berghe T, Bogaert P, Laukens B, Zobel K, Deshayes K, Vucic D, Fulda S, Vandenabeele P, Bertrand MJ: cIAP1 and TAK1 protect cells from TNF-induced necrosis by preventing RIP1/RIP3-dependent reactive oxygen species production. *Cell Death Differ* 18,656-65 (2011) DOI: 10.1038/cdd.2010.138
- 45. Gutteridge JM: Does redox regulation of cell function explain why antioxidants perform so poorly as therapeutic agents? *Redox Rep* 4,129-31 (1999)
- 46. Adam-Vizi V, Starkov AA: Calcium and mitochondrial reactive oxygen species generation: how to read the facts. *J Alzheimers Dis* 20 Suppl 2,S413-26 (2010)
- 47. Brookes PS, Yoon Y, Robotham JL, Anders MW, Sheu SS: Calcium, ATP, and ROS: a mitochondrial love-hate triangle. *Am J Physiol Cell Physiol* 287,C817-33 (2004)
- 48. Feissner RF, Skalska J, Gaum WE, Sheu SS: Crosstalk signaling between mitochondrial Ca2+ and ROS. *Front Biosci (Landmark Ed)* 14,1197-218 (2009) DOI: 10.2741/3303
- Chua BT, Guo K, Li P: Direct cleavage by the calcium-activated protease calpain can lead to inactivation of caspases. *J Biol Chem* 275,5131-5 (2000)
 DOI: 10.1074/jbc.275.7.5131
- 50. Samali A, Orrenius S: Heat shock proteins: regulators of stress response and apoptosis. *Cell Stress Chaperones* 3,228-36 (1998) DOI: 10.1379/1466-1268(1998)003<0228 :HSPROS>2.3.CO;2
- Dumont A, Hehner SP, Hofmann TG, Ueffing M, Dröge W, Schmitz ML: Hydrogen peroxideinduced apoptosis is CD95-independent, requires the release of mitochondria-derived reactive oxygen species and the activation of NF-kappaB. Oncogene 18,747-57 (1999) DOI: 10.1038/sj.onc.1202325
- 52. Leigh-Brown S, Enriquez JA, Odom DT: Nuclear transcription factors in mammalian mitochondria. *Genome Biol* 11,215 (2010)
- 53. Palm NW, Medzhitov R: Pattern recognition receptors and control of adaptive immunity. *Immunol Rev* 227,221-33 (2009)

- DOI: 10.1111/j.1600-065X.2008.00731.x
- 54. West AP, Khoury-Hanold W, Staron M, Tal MC, Pineda CM, Lang SM, Bestwick M, Duguay BA, Raimundo N, MacDuff DA, Kaech SM, Smiley JR, Means RE, Iwasaki A, Shadel GS: Mitochondrial DNA stress primes the antiviral innate immune response. Nature 520.553-7 (2015)

DOI: 10.1038/nature14156

- Suliman HB, Welty-Wolf KE, Carraway MS, Schwartz DA, Hollingsworth JW, Piantadosi CA: Toll-like receptor 4 mediates mitochondrial DNA damage and biogenic responses after heat-inactivated E. coli. FASEB J 19,1531-3 (2015)
- 56. Krychtiuk KA, Ruhittel S, Hohensinner PJ, Koller L, Kaun C, Lenz M, Bauer B, Wutzlhofer L, Draxler DF, Maurer G, Huber K, Woita J, Heinz G, Niessner A, Speidl WS: Mitochondrial DNA and Toll-Like Receptor-9 Are Associated With Mortality in Critically III Patients. Crit Care Med 43,2633-41 (2015) DOI: 10.1097/CCM.0000000000001311
- 57. Wei X, Shao B, He Z, Ye T, Luo M, Sang Y, Liang X, Wang W, Luo S, Yang S, Zhang S, Gong C, Gou M, Deng H, Zhao Y, Yang H, Deng S, Zhao C, Yang L, Qian Z, Li J, Sun X, Han J, Jiang C, Wu M, Zhang Z: Cationic nanocarriers induce cell necrosis through impairment of Na(+)/K(+)-ATPase and cause subsequent inflammatory response. Cell Res 25,237-53 (2015) DOI: 10.1038/cr.2015.9
- 58. Hauser CJ, Sursal T, Rodriguez EK, Appleton PT, Zhang Q, Itagaki K: Mitochondrial damage associated molecular patterns from femoral reamings activate neutrophils through formyl peptide receptors and P44/42 MAP kinase. J Orthop Trauma 24,534-8 (2010) DOI: 10.1097/BOT.0b013e3181ec4991
- 59. Julian MW, Shao G, Vangundy ZC, Papenfuss TL, Crouser ED: Mitochondrial transcription factor A, an endogenous danger signal, promotes TNFα release via RAGE- and TLR9responsive plasmacytoid dendritic cells. PLoS One 8,e72354 (2013)
- 60. Yu EP, Bennett MR: Mitochondrial DNA damage and atherosclerosis. Trends Endocrinol Metab 25,481-7 (2014) DOI: 10.1016/j.tem.2014.06.008
- 61. Zhang JZ, Liu Z, Liu J, Ren JX, Sun TS:

- Mitochondrial DNA induces inflammation and increases TLR9/NF-kB expression in lung tissue. Int J Mol Med 33,817-24 (2014) DOI: 10.3892/ijmm.2014.1650
- 62. Gurung P, Lukens JR, Kanneganti TD: Mitochondria: diversity in the regulation of the NLRP3 inflammasome. Trends Mol Med 21,193-201 (2015) DOI: 10.1016/j.molmed.2014.11.008
- 63. Shimada K, Crother TR, Karlin J, Dagvadori J. Chiba N. Chen S. Ramanujan VK, Wolf AJ, Vergnes L, Ojcius DM, Rentsendorj A, Vargas M, Guerrero C, Wang Y, Fitzgerald KA, Underhill DM, Town T, Arditi M: Oxidized mitochondrial DNA activates the NLRP3 inflammasome during apoptosis. Immunity 36,401-14 (2012)

DOI: 10.1016/j.immuni.2012.01.009

- 64. Bergsbaken T, Fink SL, Cookson BT: Pyroptosis: host cell death and inflammation. Nat Rev Microbiol 7,99-109 (2009) DOI: 10.1038/nrmicro2070
- 65. Nakahira K, Haspel JA, Rathinam VA, Lee SJ, Dolinay T, Lam HC, Englert JA, Rabinovitch M, Cernadas M, Kim HP, Fitzgerald KA, Ryter SW, Choi AM: Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. Nat Immunol. 12,222-30 (2011)

DOI: 10.1038/ni.1980

- 66. White MJ. Kile BT: Stressed mitochondria sound the alarm. Immunol Cell Biol 93,427-8 (2015) DOI: 10.1038/icb.2015.31
- 67. Brinkmann V, Reichard U, Goosmann C. Fauler B, Uhlemann Y, Weiss DS, Weinrauch Y, Zychlinsky A: Neutrophil extracellular traps kill bacteria. Science 303,1532-5 (2004) DOI: 10.1126/science.1092385
- 68. Brinkmann V, Zychlinsky A. Neutrophil extracellular traps: is immunity second function of chromatin? J Cell Biol 198,773-83 (2012)
- 69. Urban CF, Reichard U, Brinkmann V, Zychlinsky A: Neutrophil extracellular traps capture and kill Candida albicans yeast and hyphal forms. Cell Microbiol 8,668-76 (2006) DOI: 10.1111/j.1462-5822.2005.00659.x
- 70. Yousefi S, Gold JA, Andina N, Lee JJ, Kelly

- AM, Kozlowski E, Schmid I, Straumann A, Reichenbach J, Gleich GJ, Simon HU: Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. *Nat Med* 14,949-53 (2008) DOI: 10.1038/nm.1855
- 71. McIlroy DJ, Jarnicki AG, Au GG, Lott N, Smith DW, Hansbro PM, Balogh ZJ: Mitochondrial DNA neutrophil extracellular traps are formed after trauma and subsequent surgery. *J Crit Care* 29,1133.e1-5 (2014)
- Muller S, Radic M: Oxidation and mitochondrial origin of NET DNA in the pathogenesis of lupus. *Nat Med* 22,126-7 (2016) DOI: 10.1038/nm.4044
- Hakkim A, Fuchs TA, Martinez NE, Hess S, Prinz H, Zychlinsky A, Waldmann H: Activation of the Raf-MEK-ERK pathway is required for neutrophil extracellular trap formation. *Nat Chem Biol* 7,75-7 (2011) DOI: 10.1038/nchembio.496
- 74. Marcos V, Zhou Z, Yildirim AO, Bohla A, Hector A, Vitkov L, Wiedenbauer EM, Krautgartner WD, Stoiber W, Belohradsky BH, Rieber N, Kormann M, Koller B, Roscher A, Roos D, Griese M, Eickelberg O, Döring G, Mall MA, Hartl D: CXCR2 mediates NADPH oxidase-independent neutrophil extracellular trap formation in cystic fibrosis airway inflammation. *Nat Med* 16,1018-23 (2010) DOI: 10.1038/nm.2209
- 75. Morshed M, Hlushchuk R, Simon D, Walls AF, Obata-Ninomiya K, Karasuyama H, Djonov V, Eggel A7, Kaufmann T, Simon HU, Yousefi S: NADPH oxidase-independent formation of extracellular DNA traps by basophils. *J Immunol* 192,5314-23 (2014) DOI: 10.4049/jimmunol.1303418
- Kao SH, Chao HT, Liu HW, Liao TL, Wei YH: Sperm mitochondrial DNA depletion in men with asthenospermia. Fertil Steril 82,66-73 (2004)
 DOI: 10.1016/j.fertnstert.2003.11.056
- Liu CS, Cheng WL, Kuo SJ, Li JY, Soong BW, Wei YH: Depletion of mitochondrial DNA in leukocytes of patients with poly-Q diseases. *J Neurol Sci* 264,18-21 (2008)
 DOI: 10.1016/j.jns.2007.07.016
- Garrabou G, Morén C, López S, Tobías E, Cardellach F, Miró O, Casademont J: The effects of sepsis on mitochondria. J Infect Dis

- 205,392-400 (2012) DOI: 10.1093/infdis/jir764
- 79. Xu FX, Zhou X, Shen F, Pang R, Liu SM: Decreased peripheral blood mitochondrial DNA content is related to HbA1c, fasting plasma glucose level and age of onset in type 2 diabetes mellitus. *Diabet Med* 29,e47-54 (2012)
- 80. Pyle A, Anugrha H, Kurzawa-Akanbi M, Yarnall A, Burn D, Hudson G: Reduced mitochondrial DNA copy number is a biomarker of Parkinson's disease. *Neurobiol Aging* 38,216. e7-10 (2016)
- 81. Zhang Y, Qu Y, Gao K, Yang Q, Shi B, Hou P, Ji M: High copy number of mitochondrial DNA (mtDNA) predicts good prognosis in glioma patients. *Am J Cancer Res* 5,1207-16. eCollection (2015)
- 82. Lan Q, Lim U, Liu CS, Weinstein SJ, Chanock S, Bonner MR, Virtamo J, Albanes D, Rothman N: A prospective study of mitochondrial DNA copy number and risk of non-Hodgkin lymphoma. *Blood* 112,4247-9 (2008) DOI: 10.1182/blood-2008-05-157974
- Shen J, Platek M, Mahasneh A, Ambrosone CB, Zhao H: Mitochondrial copy number and risk of breast cancer: a pilot study. Mitochondrion 10,62-8 (2010)
 DOI: 10.1016/j.mito.2009.09.004
- 84. Hosgood HD, Liu CS, Rothman N, Weinstein SJ, Bonner MR, Shen M, Lim U, Virtamo J, Cheng WL, Albanes D, Lan Q: Mitochondrial DNA copy number and lung cancer risk in a prospective cohort study. *Carcinogenesis* 31,847-9 (2010)
 DOI: 10.1093/carcin/bgg045
- Chen T, He J, Shen L, Fang H, Nie H, Jin T, Wei X, Xin Y, Jiang Y, Li H, Chen G, Lu J, Bai Y: The mitochondrial DNA 4,977-bp deletion and its implication in copy number alteration in colorectal cancer. *BMC Med Genet* 12,8 (2011)
- 86. Lynch SM, Weinstein SJ, Virtamo J, Lan Q, Liu CS, Cheng WL, Rothman N, Albanes D, Stolzenberg-Solomon RZ: Mitochondrial DNA copy number and pancreatic cancer in the alpha-tocopherol beta-carotene cancer prevention study. *Cancer Prev Res (Phila)* 4,1912-9 (2011)

DOI: 10.1158/1940-6207.CAPR-11-0002

- Yamada S, Nomoto S, Fujii T, Kaneko T, Takeda S, Inoue S, Kanazumi N, Nakao A: Correlation between copy number of mitochondrial DNA and clinico-pathologic parameters of hepatocellular carcinoma. *Eur J Surg Oncol* 32,303-7 (2006)
 DOI: 10.1016/j.ejso.2006.01.002
- 88. Xing J, Chen M, Wood CG, Lin J, Spitz MR, Ma J, Amos CI, Shields PG, Benowitz NL, Gu J, de Andrade M, Swan GE, Wu X: Mitochondrial DNA content: its genetic heritability and association with renal cell carcinoma. *J Natl Cancer Inst* 100,1104-12 (2008) DOI: 10.1093/jnci/djn213
- Arnalich F, Codoceo R, López-Collazo E, Montiel C: Circulating cell-free mitochondrial DNA: a better early prognostic marker in patients with out-of-hospital cardiac arrest. *Resuscitation* 83,e162-3 (2012)
 DOI: 10.1016/j.resuscitation.2012.03.032
- 90. Arnalich F, Maldifassi MC, Ciria E, Codoceo R, Renart J, Fernández-Capitán C, Herruzo R, Garcia-Rio F, López-Collazo E, Montiel C: Plasma levels of mitochondrial and nuclear DNA in patients with massive pulmonary embolism in the emergency department: a prospective cohort study. *Crit Care* 17,R90 (2013)
- 91. Nakahira K, Kyung SY, Rogers AJ, Gazourian L, Youn S, Massaro AF, Quintana C, Osorio JC, Wang Z, Zhao Y, Lawler LA, Christie JD, Meyer NJ, Mc Causland FR, Waikar SS, Waxman AB, Chung RT, Bueno R, Rosas IO, Fredenburgh LE, Baron RM, Christiani DC10, Hunninghake GM, Choi AM: Circulating mitochondrial DNA in patients in the ICU as a marker of mortality: derivation and validation. *PLoS Med* 10,e1001577; discussion e1001577 (2013)
- Lam NY, Rainer TH, Chiu RW, Joynt GM, Lo YM: Plasma mitochondrial DNA concentrations after trauma. *Clin Chem* 50,213-6 (2004) DOI: 10.1373/clinchem.2003.025783
- 93. Walko TD, Bola RA, Hong JD, Au AK, Bell MJ, Kochanek PM, Clark RS, Aneja RK: Cerebrospinal fluid mitochondrial DNA: a novel DAMP in pediatric traumatic brain injury. Shock 41,499-503 (2014) DOI: 10.1097/SHK.0000000000000160
- 94. Timmermans K, Kox M, Vaneker M, van den Berg M, John A, van Laarhoven A, van der

- Hoeven H, Scheffer GJ, Pickkers P: Plasma levels of danger-associated molecular patterns are associated with immune suppression in trauma patients. *Intensive Care Med* 42,551-61 (2016) DOI: 10.1007/s00134-015-4205-3
- Kawane K, Ohtani M, Miwa K, Kizawa T, Kanbara Y, Yoshioka Y, Yoshikawa H, Nagata S: Chronic polyarthritis caused by mammalian DNA that escapes from degradation in macrophages. *Nature* 443,998-1002 (2006) DOI: 10.1038/nature05245
- Wong M, Gertz B, Chestnut BA, Martin LJ: Mitochondrial DNMT3A and DNA methylation in skeletal muscle and CNS of transgenic mouse models of ALS. Front Cell Neurosci 7,279 (2013)
- 97. Baccarelli AA, Byun HM: Platelet mitochondrial DNA methylation: a potential new marker of cardiovascular disease. *Clin Epigenetics* 7,44 (2015)
- 98. Iacobazzi V, Castegna A, Infantino V, Andria G: Mitochondrial DNA methylation as a next-generation biomarker and diagnostic tool. *Mol Genet Metab* 110,25-34 (2013) DOI: 10.1016/j.ymgme.2013.07.012
- 99. Lee YL, King MB, Gonzalez RP, Brevard SB, Frotan MA, Gillespie MN, Simmons JD: Blood transfusion products contain mitochondrial DNA damage-associated molecular patterns: a potential effector of transfusion-related acute lung injury. *J Surg Res* 191,286-9 (2014) DOI: 10.1016/j.iss.2014.06.003
- 100. Yasui K, Matsuyama N, Kuroishi A, Tani Y, Furuta RA, Hirayama F: Mitochondrial damage-associated molecular patterns as potential proinflammatory mediators in post-platelet transfusion adverse effects. *Transfusion* 56,1201-12 (2016) DOI: 10.1111/trf.13535
- 101. Cao H, Ye H, Sun Z, Shen X, Song Z, Wu X, He W, Dai C, Yang J: Circulatory mitochondrial DNA is a pro-inflammatory agent in maintenance hemodialysis patients. *PLoS One* 9,e113179 (2014)
- 102. Fournié GJ, Lulé J, Dueymes JM, Laval F, Delobbe I, Vernier I, Pourrat JP: Plasma DNA in patients undergoing hemodialysis or hemofiltration: cytolysis in artificial kidney is responsible for the release of DNA in circulation. Am J Nephrol 9,384-91 (1989)

DOI: 10.1159/000168000

- 103. Caudrillier A, Kessenbrock K, Gilliss BM, Nguyen JX, Marques MB, Monestier M, Toy P, Werb Z, Looney MR: Platelets induce neutrophil extracellular traps in transfusionrelated acute lung injury. J Clin Invest 122,2661-71 (2012) DOI: 10.1172/JCI61303
- 104. Fuchs HJ, Borowitz DS, Christiansen DH, Morris EM, Nash ML, Ramsey BW, Rosenstein BJ, Smith AL, Wohl ME: Effect of aerosolized recombinant human DNase on exacerbations of respiratory symptoms and on pulmonary function in patients with cystic fibrosis. The Pulmozyme Study Group. N Engl J Med 331,637-42 (1994)

DOI: 10.1056/NEJM199409083311003

105. Lood C, Blanco LP, Purmalek MM, Carmona-Rivera C, De Ravin SS, Smith CK, Malech HL, Ledbetter JA, Elkon KB, Kaplan MJ: Neutrophil extracellular traps enriched in oxidized mitochondrial DNA are interferogenic and contribute to lupus-like disease. *Nat Med* 22,146-53 (2016)

DOI: 10.1038/nm.4027

Key Words: Mitochondria, Mitochondrial DNA, mtDNA, Inflammation, Organ Failure, Review

Send correspondence to: Zsolt J Balogh, Department of Traumatology, John Hunter Hospital and University of Newcastle, Newcastle, NSW 2310, Australia, and Department of Traumatology, University of Newcastle, NSW 2310, Australia, Tel: 61249214259, Fax: 61249214274, E-mail: zsolt.balogh@hnehealth.nsw.gov.au