TRPM2: a potential drug target to retard oxidative stress

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TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. The gene location, tissues distribution of TRPM2
- 4. The basic structure of TRPM2
- 5. Variants of TRPM2
- 6. Activators for TRPM2
- 7. Inhibitors for TRPM2
- 8. TRPM2 will be activated during oxidative stress
- 9. Function and potential application in drug discovery of TRPM2
- 10. Discussion
- 11. Acknowledgement
- 12. References

1. ABSTRACT

The Transient Receptor Potential Melastatin 2 (TRPM2) is a member of G protein coupled receptor superfamily and a novel dual-function protein that possesses both ion channel and Adenosine 5'-Diphos-Phatase Ribose (ADPR) hydrolase function. TRPM2 is involved in Ca2+ signaling in various cells as an endogenous redox sensor for oxidative stress and reactive oxygen species, and contributes to cytokine production, insulin release, motility, Ca2+ entry and Ca2+-dependent cellular reactions such as endothelial hyper-permeability and apoptosis. The wide expression of TRPM2 might render it as a potentially significant therapeutic target in pathological settings including cardiovascular and neurodegenerative diseases and of great relevance in drug design, feed additives and other industries. Here, we discuss the TRPM2 gene structure, function, its variants, as well as its activators and inhibitors and provide a peptide drug design for modulation of oxidative stress.

2. INTRODUCTION

Oxidative Stress (OS) represents an imbalance between the generation of Reactive Oxygen Species (ROS) and the functions of antioxidant systems (1). Disturbances in the normal redox state of tissues will induce toxic effects through ROS that could damage proteins, lipids, DNA and other molecules (2). Suitable concentrations of ROS are beneficial and, in fact, necessary in lives due to their signaling roles in metabolism (3). However, when under various stresses, like cold, heat, age, disease, and other conditions, the balance between ROS production and quenching is disturbed, OS is induced and bring damages to organisms, and further induce sickle cell disease, atherosclerosis, parkinson's disease, Alzheimer's Disease (AD), heart failure, myocardial infarction, Schizophrenia, Bipolar Disorder (BD), fragile X syndrome and chronic fatigue syndrome (4-6).

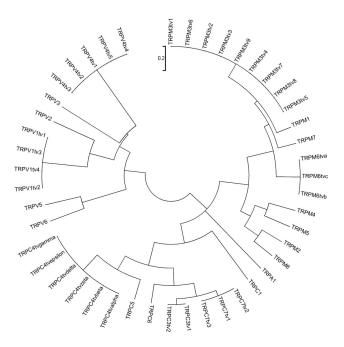


Figure 1. Phylogenetic tree for Homo sapiens (human) TRP channels built by using MEGA 4.1, tv means transcript variant.

The Transient Receptor Potential Melastatin 2 (TRPM2) is a member of G protein coupled receptor superfamily and a novel dual-function protein that possesses both ion channel and Adenosine 5'-Diphos-Phatase Ribose (ADPR) hydrolase function, and the two functions are uncoupled (7-9). It plays a pivotal role in temperature sensing, cancer, numerous metabolic and neuronal diseases such as diabetes mellitus, BD and AD (10-15). TRPM2 also acts as an endogenous redox sensor for mediating OS / ROS-induced Ca2+ entry and the subsequent specific Ca2+-dependent cellular reactions. As for the nearly half of commercial medicines target to G protein coupled receptor superfamily, TRPM2 is also a potentially significant therapeutic target for diseases, and researches on it can be of great relevance in drug designing, feed additives and other industries (16).

3. THE GENE LOCATION, TISSUES DISTRIBUTION OF TRPM2

The TRPM2 gene was initially reported as transient receptor potential related channel 7 by the genomic DNA sequence analysis (Figure 1). This gene consists of 32 exons and spans approx. 90 kb (17). In human, it locates on disease-rich chromosome 21q22.3, pitches in the region between two markers, D21S400 and D21S171 (18, 19). Chromosome 21q22.3. (Figure 2) is near to the SOD1 locus (21q22.1.–22.2.), which is associated with familial amyotrophic lateral sclerosis (20).

TRPM2 is predominantly expressed in brain (17), and to be highest in the hippocampus (21),

cerebral cortex, thalamus, and midbrain, specifically in microglia and neuronal cell (22). TRPM2 is also detected in many other tissues, including bone, gastrointestinal tract, marrow, spleen, heart, liver, pancreas, placenta, ovary, lung, vascular smooth muscle, hematopoietic cells, monocytes, endocrine cells and endothelial cells (23).

4. THE BASIC STRUCTURE OF TRPM2

TRPM2 comprise an ion channel core domain and cytoplasmic NH2 and COOH termini. Similar to other six Trans-Membrane (6TM) channels including cyclic nucleotide-gated channels, its ion channel domain (residues 762-1048) is composed of 6TM segments (S1-S6) and a pore loop between S5 and S6 (Figure 3). The distal part of S6 is important for the gating function for charge discrimination (24). No functional role be attributed to any of the structural motifs within the N-stretch that may be a spacer segment for other functional sites in the N terminus (25). Glu-960, Gln-981, Asp-987, and Glu-1022 residues are engaged in determining divalent cationic permeation properties of the channel. The residues Glu-960, Gln-981, and Asp-987 contribute significantly to defining Ca2+ permeation while Glu-1022 to Mg2+ (26). The N-terminal cytosolic tail of about 700 amino acids contains calmodulin binding domains that mediate Ca²⁺ regulation of TRPM2 activity (25). The N-terminal part also has a high affinity binding site for Protein Kinase C-alpha (PKCα) and two PxxP motifs implicated in protein-protein interactions, so this part likely to be crucial for the proper interaction with essential regulatory cytosolic components, assembly of the

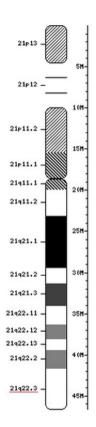


Figure 2. The master map of Homo sapiens (human) Chromosome 21. Region located TRPM2 was point out with red underline.

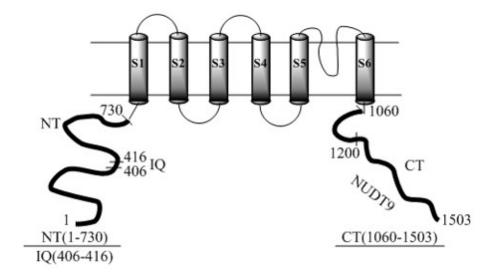


Figure 3. Diagram of TRPM2. The N- and C-terminal fragments (dark lines), the IQ-like motif (residues 406-416), the NUDT9 ADPR hydrolase domain (residues 1236-1503), and the six transmembrane domains (S1-S6) are shown.

channel units and membrane trafficking (25). There are two coiled-coil domains contained in TRPM2. The one in N-terminal part (655-679) is required for protein expression and function, but not subunit interaction, and the Ile-658 residue in this domain is required for normal channel function (27), while the other one in the C-terminus may play a role in ion channel subunit

multi-mediation or in recruitment of regulatory proteins (28). The whole TRPM subfamily contains a pair of cysteine residues (positions 996 and 1008 in the human TRPM2 subunit), and the first is completely conserved (Figure 4) while the second is present at slightly different positions. The two cysteine residues and Lys¹¹¹⁰ are obligatory for TRPM2 channel function

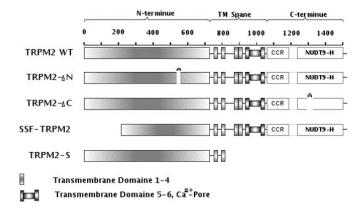


Figure 4. Schematic representation of TRPM2 isoforms. Membrane spanning domains 1-4 and the putative pore region including transmembrane domains 5-6 are indicated. CCR represents the coiled coil region which may mediate protein/protein interactions. NUDT9-H represents the NUDT9 ADPR hydrolase domain. TRPM2-DN has a deletion of amino acids 538-557 in the N-terminus. TRPM2-DC has a deletion of AA 1292-1325 in the C-terminus. TRPM2-S is missing four of six transmembrane domains and the putative calcium pore.

(29, 30). Functional TRPM2 molecules are tetramers and subunit composition is a factor in regulation of the channel opening (31).

5. VARIANTS OF TRPM2

Until now, some TRPM2 variants have been found, each of them has unique construction and characters, which was showed in Figure 4. TRPM2-S was cloned from human bone marrow and consists of only the N terminus and the first two trans-membrane domains. The four C-terminal trans-membrane domains, the putative calcium-permeable pore region, and the entire C terminus were deleted because of a stop codon (TAG) alternative splicing which locates at the splice junction between exons 16 and 17. Interaction between TRPM2-S and TRPM2-WT is an important mechanism for channel activity regulation and cellular response to OS (32). Expression of TRPM2-S inhibited susceptibility to cell death and onset of apoptosis induced by H2O2 in cells expressing TRPM2-WT.

The striatum short form (SSF-TRPM2), detected only in striatum (caudate nucleus and putamen), is a 5.5 kb shorter transcript transcribed from the intron 4 of the TRPM2 gene with 1289 amino acid residues. As compared to the long form protein, the N-terminal 214 amino acid residues are removed in SSF-TRPM2. The SSF-TRPM2 still maintained $\rm H_2O_2$ -induced $\rm Ca^{2^+}$ influx activity (19).

TRPM2ΔNΔC is a new TRPM2 isoform by two deletions, from 538 to 557 and from 1292 to 1325.

Additionally, two amino acid residues are exchanged (S1088N and D1291E). One deletion is located in the cytosolic N terminus and the other one in the cytosolic C terminus of TRPM2. This splicing variant lost channel activity to respond to ADPR (33).

TRPM2P1018L produces a missense change whereby proline 1018 (Pro1018) is replaced by leucine (Leu1018), and inactive the channel function. The ability of TRPM2 to maintain sustained ion influx is a physiologically important function and that its disruption may, under certain conditions, contribute to disease states. TRPM2P1018L may confer susceptibility to western pacific amyotrophic lateral sclerosis and parkinsonism-dementia, which shared a unique mineral environment characterized by the presence of severely low levels of Ca2+ and Mg2+, coupled with high levels of bioavailable transition metals in the soil and drinking water (34). TRPM2 genetic variants also have significant association with BD. TRPM2 SNP rs1556314 in exon 11 was significantly associated with BD-I but not BD-II (35).

6. ACTIVATORS FOR TRPM2

ADPR is the intracellular TRPM2 agonist (36). However, ADPR alone is insufficient to gate TRPM2 channel rather channel activation is contingent on the binding of Ca²⁺ to an intracellular channel domain via calmodulin (37, 38). ADPR most likely binds to C-terminal Nudix hydrolase domain NUDT9H for TRPM2 activation (Figure 5), but there also may be other mechanism (39). TRPM2 channel is intimately coupled to N-methyl-d-aspartate receptor s via Ca²⁺ influx (40)

Figure 5. Molecular structures of ADPR and mainly related adenine nucleotides that can activate TRPM2 channels.

Figure 6. Molecular structures of mainly TRPM2 channel inhibitors.

and are necessary for the induction of N-methyl-daspartate receptor -dependent long term depression (41). Additionally, nicotinic acid adenine dinucleotide phosphate, cyclic ADPR, 2'-O-acetyl-ADPR, ADPR-2'-phosphate and H₂O₂ facilitate activation of TRPM2 channel (33, 42-45). The mechanism of how H₂O₂ activate TRPM2 is still unclear. Hara et al reported that TRPM2 activation by H₂O₂ is mediated by increase of NAD+ levels and binding of NAD+ to the nudix domain, but Kühn and Lückhoff thought the channels are not gated by H₂O₂, but NAD+ (46, 47). Others thought H₂O₂ activates TRPM2 through the release of ADPR from the mitochondria or the ectoenzyme CD38 (42, 48). Conversely, the direct activation of TRPM2 by cyclic ADPR and nicotinic acid adenine dinucleotide phosphate, albeit limited, cannot be suppressed by the specific ADPR antagonist AMP (42). The intracellular Ca2+ appears to be an important modulator and cofactor of TRPM2, as elevated intracellular Ca2+ can significantly increase the sensitivity of TRPM2 toward ADPR (46). TRPM2 channel activation is sensitive to intracellular CI⁻ concentration (49).

7. INHIBITORS FOR TRPM2

Generally, there are less specific TRPM2 inhibitors which have a potentially clinically useful effect on the activation are found now. But several compounds show inhibit ability toward TRPM2, they

are probably divided into 2 different classes, namely the fenamates, such as flufenamic acid (FFA), and anti-fungal imidazoles (miconazole, econazol and clotrimazole) (50, 51) (Figure 6). FFA, a nonsteroidal anti-inflammatory drug, is a pH-dependent antagonist of TRPM2 channels. Decreasing extracellular pH accelerated FFA inhibition of TRPM2. FFA is not easily dissolved in the aqueous solution, which inhibit its application (50). N- (p-amylcinnamoyl) anthranilic acid (ACA), usually functions as a phospholipase A_a inhibitor, is another inhibitor of TRPM2 (52), it modulates different TRP channels most probably by a direct interaction. ACA induces a complete and rapid decline of TRPM2 currents in cells overexpressing this channel, most likely by interfering with the pore and in a manner independent of the channel activator (24, 52). Owing to its high potency and efficacy on TRPM2, ACA can serve, in combination with other blockers, as a pharmacological tool for studying H₂O₂induced Ca2+ signal and biological functions of TRPM2 channels in native cells. Chlorpromazine, Curcumin and N-Acetylcysteine also inhibits activation of TRPM2 channels (53-55). 8- phenyl-2'-deoxy-ADPR can highly specific binding to NUDT9H and inhibit TRPM2 (56).

Although FFA, miconazole and clotrimazole, and ACA inhibit TRPM2, their inhibition was either gradual or irreversible. For example imidazole derivatives, miconazole and clotrimazole, inhibit

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TRPM2 in an irreversible manner (51). 2-aminoethyl diphenylborinate (2-APB) can reversibly and completely inhibit TRPM2 (57).

TRPM2 is also inhibited by both intracellular and extracellular acidic pH. Extracellular protons inhibit TRPM2 by decreasing single-channel conductance. H958, D964, and E994, at the outer vestibule of the channel pore are responsible for this characteristic. Titration of H958, D964, and E994 by external protons inhibits TRPM2 gating by causing conformation change of the channel, and/or by decreasing local Ca²⁺ concentration at the outer vestibule, therefore reducing Ca2+ permeation and inhibiting Ca2+ -mediated TRPM2 gating. Intracellular protons inhibit TRPM2 by inducing channel closure without changing channel conductance. D933 located at the C terminus of the S4-S5 linker is responsible for intracellular pH sensitivity. D933 is not only essential for intracellular pH sensitivity, but it is also crucial for TRPM2 channel gating (58).

8. TRPM2 WILL BE ACTIVATED DURING OXI-DATIVE STRESS

TRPM2 is shown to gate in response to OS via second-messenger production (59), which can be inhibited by pharmacological reagents. Oxidants, applied externally or produced in the cytosol during OS (33), stimulate ADPR formation in the nucleus and mitochondria (48). The free radicals intermediates produced in the cytosol during OS include superoxide anion (O2-), H2O2, nitric oxide, and a more damaging compound, hydroxyl radical (OH.). These radicals contribute to DNA oxidation and damage, which initiate PARP-mediated ADPR generation. PARP act as a potential source of ADPR for the activation of TRPM2 in OS. PARP binds to single-stranded and double-stranded DNA breaks and catalyses the breakdown of NAD into nicotinamide and poly (ADPR) to initiate DNA repair mechanisms (48). Free ADPR is then generated from poly (ADPR) degradation by poly (ADPR) glycohydrolase. It has been suggested that OS-induced TRPM2 activation is triggered via the production of ADPR from mitochondria (48). TRPM2 confers susceptibility to cell death through the activation of caspases and PARP (46). Alternative supply of ADPR may results from the direct hydrolysis of NAD+ into nicotinamide and ADPR or indirectly through cyclic ADPR formation. This reaction is catalyzed by NADases that are not only located at cell surface, but also in mitochondria and the nucleus (60). Although most investigators demonstrated an indirect action of oxidants on TRPM2 through ADPR generation, direct action of oxidants on TRPM2 has been also proposed for neutrophil granulocytes, as the TRPM2-ΔC splice variant was stimulated by H2O2 but did not respond to ADPR (33). H₂O₂-mediated Ca²⁺ entry through TRPM2 in pulmonary artery endothelial cells was reduced by at least 65 % in cells treated with a PARP inhibitor (either 3, 4-dihydro-5- (4- (1-piperidinyl)butoxyl)-1 (2H)-isoquinolinone or 3-aminobenzamide) to prevent ADPR agonist formation (61). Phosphotyrosine phosphatase L1 and PKC α can regulate oxidant-induced TRPM2 activation (62). The H_2O_2 induced apoptotic changes involve TRPM2 opening, which results in mitochondrial (Na⁺)_m (and (Ca²⁺)_m) overload, followed by mitochondrial membrane disruption, cytC release, and caspase 3-dependent nuclear condensation /fragmentation, while the necrotic changes are caspase-3-independent, but PARP dependent. Inhibition of both TRPM2 and PARP activities totally abolishes H_2O_2 -induced myocyte death (23).

9. FUNCTION AND POTENTIAL APPLICATION IN DRUG DISCOVERY OF TRPM2

TRPM2 regulates endothelial barrier function, plays a critical role in the mechanism of endothelial barrier disruption following OS (63), inhibition of TRPM2 may provide a useful therapeutic strategy for the treatment of endothelial barrier dysfunction and vascular inflammation (61). TRPM2 is abundant in pancreatic islet cells (64) and plays a critical role in insulin secretion by pancreatic β-cells. TRPM2 also functions as a Ca2+- release channel activated by intracellular ADPR in a lysosomal compartment. TRPM2-mediated chemokine production in monocytes/ macrophages is an important mechanism in the progressive severity of DSS-induced ulcerative colitis. Ca2+ influx via native TRPM2 plays an important role in the H₂O₂-induced CXCL8 production inmonocytes. Erk and NF-kB are involved in CXCL8 production inmonocytes and other cell types (65). Erk activation amplified by Ca2+ influx through TRPM2 is mediated via Ras (66). TRPM2 are potential therapeutic targets for oxytocin release in psychiatric diseases caused by social stress (67).

10. DISCUSSION

TRPM2 is implicated in endothelial dysfunction and many pathological states, which has emerged as an important Ca²⁺ signal molecular in various cells, and be involved in cytokine production, insulin release, motility and death. It is a potentially significant therapeutic target in pathological settings including cardiovascular and neurodegenerative diseases (68).

Chemical drug occupied most of modern drugs, which are susceptibility to resistance and usually have long-term toxicity. The use of these drugs should be used under serious supervision. So, new safely and effectively therapeutic approaches were developed in clinic. Peptides have attractive features compared to small molecule and protein, such as high

structural compatibility with target protein, disrupted protein-protein interfaces, less susceptibility to drug resistance, small size, lower costs, improved organ or tumor penetration and higher activity per mass when compared to antibodies or large proteins (69, 70). Novel delivery and chemical modification technologies have sparked much interest in peptide therapeutics (71-73). More than 50 peptide-based products have been approved for clinical use (e.g. Fuzeon (Roche); Byetta (Amylin/Eli Lilly); Sandostatin (Novartis); Zoladex (Astra- Zeneca); Copaxone (Teva)) (74). Peptideprotein interactions have important roles in mediating in protein-protein interactions, predominantly in signaling and regulatory networks (75), which are also attractive drug targets for designed inhibitory peptides (76). As for peptide drug design, there been some strategies, such as peptide phage display (77). Researchers described computational framework to identify peptide ligands to membrane channel receptors by producing sequence alignments across many species at the functional-element level (78).

Its ability to respond to ROS has made TRPM2 a potential therapeutic target for chronic inflammation, neurodegenerative diseases, and OS-related pathologies. The physiological and pathophysiological context of ROS-mediated events makes TRPM2 a promising target for the development of therapeutic tools of inflammatory and degenerative diseases. In fact, because OS contributes to numbers of pathophysiological conditions, it is meaningful to investigate TRPM2, because it may be an important potential target to discover new drugs to cure diseases with aging especially neurologic disease. Though the mechanisms of TRPM2 activation and regulation has gained significant interest, the relationship between channel activation, intracellular Ca2+ rise and cell death is still unclear (68), a problem compounded by uncertainty of the second messengers involved and the limited specificity of the pharmacological blockers, then it's very imperative to find or design a new drug to regulate the channel and to do advance research. Peptide omimetics is the approach of reproducing the biological activity or binding properties in a smaller molecule, like peptides or modified peptides which were designed to mimic the desired region. The structural basis of peptide-protein binding strategies also analysis and found most peptides do not induce conformational changes on their partner upon binding. thus minimizing the entropic cost of binding. Peptides display interfaces that are better packed than proteinprotein interfaces and contain significantly more hydrogen bonds, mainly those involving the peptide backbone. Additionally, "hot spot" residues contribute most of the binding energy (79), and these residues can be accurately predict (80). Finally, peptides tend to bind in the largest pockets available on the protein surface (79). Peptides designed to mimic the VEGF binding site to its receptor VEGFR-2 has shown effective function to inhibit endothelial cell proliferation, migration and network formation (81). As far as concerned TRPM2, peptides could be design similar to or peculiar binding to the NUDT9-H domain, which could inhibit or activate TRPM2 channel gating. As peptides cannot last for a long time, the designed peptide drugs has efficacy neither gradual nor irreversible, but reversible and can be controlled through dose and administration time.

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12. REFERENCES

- JM Phang, W Liu, O Zabirnyk: Proline metabolism and microenvironmental stress. Annu Rev Nutr 30, 441-463 (2010) DOI: 10.1146/annurev.nutr.012809.104638
- JHJ Hoeijmaker: DNA damage, aging, and cancer. N Engl J Med 361 (15), 1475-1485 (2009)

DOI: 10.1056/NEJMra0804615

 KE Wellen, CB Thompson: Cellular metabolic stress: considering how cells respond to nutrient excess. *Mol Cell* 40 (2),323-332 (2010)

DOI: 10.1016/j.molcel.2010.10.004

- 4. M Yanik, O Erel, M Kati: The relationship between potency of oxidative stress and severity of depression. *Acta Neuropsychiatrica* 16 (4), 200-203 (2004) DOI: 10.1111/j.0924-2708.2004.00090.x
- M Fransen, M Nordgren, B Wang, O Apanasets: Role of peroxisomes in ROS/ RNS-metabolism: implications for human disease. *Biochim Biophys Acta* 1822 (9), 1363-1373 (2012) DOI: 10.1016/j.bbadis.2011.12.001
- EA Kosenko, IN Solomadin, LA Tikhonova, VP Reddy, G Aliev, YG Kaminsky: Pathogenesis of Alzheimer disease: Role of oxidative stress, amyloid-β peptides, systemic ammonia and erythrocyte energy

- metabolism. CNS Neurol Disord Drug Targets 13 (1), 112-119 (2014) DOI: 10.2174/18715273113126660130
- AL Perraud, A Fleig, CA Dunn, LA Bagley, P Launay, C Schmitz, AJ Stokes, Q Zhu, MJ Bessman, R Penner, JP Kinet, AM Scharenberg: ADPR gating of the calciumpermeable LTRPC2 channel revealed by Nudix motif homology. *Nature* 411,595-599 (2001) DOI: 10.1038/35079100
- AL Perraud, B Shen, CA Dunn, K Rippe, MK Smith, MJ Bessman, BL Stoddard, AM Scharenberg: NUDT9, a member of the Nudix hydrolase family, is an evolutionarily conserved mitochondrial ADPR pyrophosphatase. J Biol Chem 278,1794-1801 (2003) DOI: 10.1074/jbc.M205601200
- B Tóth, L Iordanov, L Csanády: Putative chanzyme activity of TRPM2 cation channel is unrelated to pore gating. *Proc Natl Acad Sci U S A* 111 (47),16949-16954 (2014) DOI: 10.1073/pnas.1412449111
- CH Tan, PA McNaughton: The TRPM2 ion channel is required for sensitivity to warmth. *Nature* 536 (7617), 460-463 (2016) DOI: 10.1038/nature19074
- K Song, H Wang, GB Kamm, J Pohle, F de Castro, Reis, P Heppenstall, H Wende, J Siemens: The TRPM2 channel is a hypothalamic heat sensor that limits fever and can drive hypothermia. Science 353 (6306),1393-1398 (2016)
 DOI: 10.1126/science.aaf7537
- X Zeng, SC Sikka, L Huang, C Sun, C Xu, D Jia, AB Abdel-Mageed, JE Pottle, JT Taylor, M Li: Novel role for the transient receptor potential channel TRPM2 in prostate cancer cell proliferation. *Prostate Cancer Prostatic Dis* 13,195-201 (2010)
 DOI: 10.1038/pcan.2009.55
- K Uchida, M Tominaga: The role of TRPM2 in pancreatic β-cells and the development of diabetes. *Cell Calcium*56 (5),332-339 (2014) DOI: 10.1016/j.ceca.2014.07.001
- Y Jang, SH Lee, B Lee, S Jung, A Khalid, K Uchida: TRPM2, a Susceptibility Gene for Bipolar Disorder, Regulates Glycogen Synthase Kinase-3 Activity in the Brain. J Neurosci 35 (34),11811–11823 (2015) DOI: 10.1523/JNEUROSCI.5251-14.2015

- 15. VG Ostapchenko, M Chen, MS Guzman, YF Xie, N Lavine, J Fan, FH Beraldo, AC Martyn, JC Belrose, Y Mori, JF MacDonald, VF Prado, MA Prado, MF Jackson: The Transient Receptor Potential Melastatin 2 (TRPM2) Channel Contributes to β-Amyloid Oligomer-Related Neurotoxicity and Memory Impairment. *J Neurosci* 35 (45),15157-15169 (2015) DOI: 10.1523/JNEUROSCI.4081-14.2015
- CS Tautermann, DE Gloriam: Editorial overview: New technologies: GPCR drug design and function-exploiting the current (of) structures. *Curr Opin Pharmacol* 30,vii-x (2016) DOI: 10.1016/j.coph.2016.07.012
- K Nagamine, J Kudoh, S Minoshima, K Kawasaki, S Asakawa, F Ito, N Shimizu: Molecular cloning of a novel putative Ca2+channel protein (TRPC7) highly expressed in brain. *Genomics* 54,124-131 (1998) DOI: 10.1006/geno.1998.5551
- RE Straub, T Lehner, Y Luo, JE Loth, W Shao, L Sharpe, JR Alexander, K Das, R Simon, RR Fieve: A possible vulnerability locus for bipolar affective disorder on chromosome 21q22.3. *Nat Genet* 8,291-296 (1994) DOI: 10.1038/ng1194-291
- T Uemura, J Kudoh, S Noda, S Kanba, N Shimizu: Characterization of human and mouse TRPM2 genes: identification of a novel N-terminal truncated protein specifically expressed in human striatum. Biochem. *Biophys Res Commun* 328,1232-1243 (2005) DOI: 10.1016/j.bbrc.2005.01.086
- DR Rosen, T Siddique, D Patterson, DA Figlewicz, P Sapp, A Hentati, D Donaldson, J Goto, JP O'Regan, HX Deng: Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature* 362,59-62 (1993) DOI: 10.1038/362059a0
- 21. JZ Bai, J Lipski: Differential expression of TRPM2 and TRPV4 channels and their potential role in oxidative stress-induced cell death in organotypic hippocampal culture. *Neurotoxicology* 31 (2),204-214 (2010) DOI: 10.1016/j.neuro.2010.01.001
- E Fonfria, PR Murdock,FS Cusdin, CD Benham, RE Kelsell, S McNulty: Tissue distribution profiles of the human TRPM cation channel family. *J Recept Signal Transduct Res* 26, 159-178 (2006)
 DOI: 10.1080/10799890600637506

- 23. XR Yang, MJ Lin, LS McIntosh, JS Sham: Functional expression of transient receptor potential melastatin- and vanilloid-related channels in pulmonary arterial and aortic smooth muscle. Am J Physiol Lung Cell Mol Physiol 290,L1267-1276 (2006) DOI: 10.1152/ajplung.00515.2005
- 24. FJ Kühn, G Knop, A Lückhoff: The transmembrane segment S6 determines cation versus anion selectivity of TRPM2 and TRPM8. *J Biol Chem* 282,27598-27609 (2007) DOI: 10.1074/jbc.M702247200
- 25. FJ Kühn, C Kühn, M Naziroglu, A Lückhoff: Role of an N-terminal splice segment in the activation of the cation channel TRPM2 by ADPR and hydrogen peroxide. *Neurochem Res* 34,227-233 (2009) DOI: 10.1007/s11064-008-9755-0
- R Xia, ZZ Mei, HJ Mao, W Yang, L Dong, H Bradley, DJ Beech, LH Jiang: Identification of pore residues engaged in determining divalent cationic permeation in transient receptor potential melastatin subtype channel 2. *J Biol Chem* 283,27426-27432 (2008) DOI: 10.1074/jbc.M801049200
- ZZ Mei, LH Jiang: Requirement for the N-terminal coiled-coil domain for expression and function, but not subunit interaction of, the ADPR-activated TRPM2 channel. *J Membr Biol* 230,93-99 (2009)
 DOI: 10.1007/s00232-009-9190-4
- 28. C Schmitz, AL Perraud: The TRPM cation channels in the immune context. *Curr Pharm Des* 11,2765-2778 (2005)
 DOI: 10.2174/1381612054546851
- ZZ Mei, LH Jiang: Conserved cysteine residues in the pore region are obligatory for human TRPM2 channel function. Am. *J Physiol Cell Physiol* 291,C1022-1028 (2006)
 DOI: 10.1152/ajpcell.00606.2005
- TK Kim, JH Nam, WG Ahn, NH Kim, HY Ham, CW Hong, JS Nam, J Lee, SO Huh, L So, SJ Kim, DK Song: Lys¹¹¹⁰ of TRPM2 is critical for channel activation. *Biochem J* 455 (3),319-327 (2013)
 DOI: 10.1042/BJ20130303
- 31. Y Maruyama, T Ogura, K Mio, S Kiyonaka, K Kato, Y Mori, S Sato: Three-dimensional reconstruction using transmission electron microscopy reveals a swollen, bell-shaped structure of transient receptor potential me-

- lastatin type 2 cation channel. *J Biol Chem* 282,36961-36970 (2007) DOI: 10.1074/jbc.M705694200
- W Zhang, X Chu, Q Tong, JY Cheung, K Conrad, K Masker, BA Miller: A novel TRPM2 isoform inhibits calcium influx and susceptibility to cell death. *J Biol Chem* 278,16222-16229 (2003)
 DOI: 10.1074/jbc.M300298200
- 33. E Wehage, J Eisfeld, I Heiner, E Jüngling, C Zitt, A Lückhoff, A: Activation of the cation channel long transient receptor potential channel 2 (LTRPC2) by hydrogen peroxide. A splice variant reveals a mode of activation independent of ADP-ribose. *J Biol Chem* 277, 23150–23156 (2002) DOI: 10.1074/jbc.M112096200
- 34. MC Hermosura, AM Cui, RC Go, B Davenport, CM Shetler, JW Heizer, C Schmitz, G Mocz, RM Garruto, AL Perraud: Altered functional properties of a TRPM2 variant in Guamanian ALS and PD. *Proc. Natl. Acad. Sci.* U S A. 105,18029-18034 (2008) DOI: 10.1073/pnas.0808218105
- 35. C Xu, PP Li, RG Cooke, SV Parikh,K Wang, JL Kennedy, JJ Warsh: TRPM2 variants and bipolar disorder risk: confirmation in a family-based association study. *Bipolar Disord* 11,1-10 (2009) DOI: 10.1111/j.1399-5618.2008.00655.x
- H Knowles,Y Li, AL Perraud: The TRPM2 ion channel, an oxidative stress and metabolic sensor regulating innate immunity and inflammation. *Immunol Res* 55,241–248 (2013) DOI: 10.1007/s12026-012-8373-8
- 37. Q Tong, W Zhang, K Conrad, K Mostoller, JY Cheung, BZ Peterson, BA Miller: Regulation of the transient receptor potential channel TRPM2 by the Ca²⁺ sensor calmodulin. *J Biol Chem* 281,9076-9085 (2006) DOI: 10.1074/jbc.M510422200
- 38. J Starkus, A Beck, A Fleig, R Penner: Regulation of TRPM2 by extra- and intracellular calcium. *J. Gen. Physiol* 130,427-440 (2007) DOI: 10.1085/jgp.200709836
- 39. FJ Kühn, C Kühn, M Winking, DC Hoffmann, A Lückhoff: ADP-Ribose Activates the TRPM2 Channel from the Sea Anemone Nematostella vectensis Independently of theNUDT9H Domain. *PLoS One* 11 (6),e0158060 (2016) DOI: 10.1371/journal.pone.0158060

- ME Olah, MF Jackson, H Li, Y Perez, HS Sun, S Kiyonaka, Y Mori, M Tymianski, JF MacDonald: Ca²⁺-dependent induction of TRPM2 currents in hippocampal neurons. *J Physiol* 587,965–979 (2009)
 DOI: 10.1113/jphysiol.2008.162289
- 41. YF Xie, JC Belrose, G Lei, M Tymianski, Y Mori, JF Macdonald, MF Jackson: Dependence of NMDA/GSK-3beta mediated metaplasticity on TRPM2 channels at hippocampal CA3-CA1 synapses. *Mol Brain* 4,44 (2011)

DOI: 10.1186/1756-6606-4-44

- 42. A Beck, M Kolisek, LA Bagley, A Fleig, R Penner: Nicotinic acid adenine dinucleotide phosphate and cyclic ADP-ribose regulate TRPM2 channels in T lymphocytes. *FASEB J* 20, 962–964 (2006) DOI: 10.1096/fj.05-5538fje
- 43. M Kolisek, A Beck, A Fleig, R Penner: Cyclic ADP-ribose and hydrogen peroxide synergize with ADP-ribose in the activation of TRPM2 channels. *Mol Cell* 18, 61–69 (2005) DOI: 10.1016/j.molcel.2005.02.033
- 44. O Grubisha, LA Rafty CL Takanishi, X Xu, L Tong, AL Perraud, AM Scharenberg, JM Denu: Metabolite of SIR2 reaction modulates TRPM2 ion channel. *J Biol Chem* 281, 14057-14065 (2006) DOI: 10.1074/jbc.M513741200
- 45. B Tóth, I lordanov, L Csanády: Ruling out pyridine dinucleotides as true TRPM2 channel activators reveals novel direct agonist ADP-ribose-2'-phosphate. *J Gen Physiol* 45 (5),419-430 (2015) DOI: 10.1085/jgp.201511377
- 46. Y Hara, M Wakamori, M Ishii, E Maeno, M Nishida, T Yoshida, H Yamada, S Shimizu, E Mori, J Kudoh, N Shimizu, H Kurose, Y Okada, K Imoto, Y Mori: LTRPC2 Ca²⁺-permeable channel activated by changes in redox status confers susceptibility to cell death. *Mol Cell* 9, 163-173 (2002) DOI: 10.1016/S1097-2765(01)00438-5
- 47. FJ Kühn, A Lückhoff: Sites of the NUDT9-H domain critical for ADPR activation of the cation channel TRPM2. *J Biol Chem* 279,46431-46437 (2004)
 DOI: 10.1074/jbc.M407263200
- 48. AL Perraud, CL Takanishi, B Shen, S Kang, MK Smith, C Schmitz, HM Knowles, D Fer-

- raris, W Li, J Zhang, BL Stoddard, AM Scharenberg: Accumulation of free ADPR from mitochondria mediates oxidative stress-induced gating of TRPM2 cation channels. *J Biol Chem* 280,6138-6148 (2005) DOI: 10.1074/jbc.M411446200
- 49. CW Hong, TK Kim, HY Ham, JS NamYH Kim, H Zheng, B Pang, TK Min, JS Jung, SN Lee,: Lysophosphatidylcholine increases neutrophil bactericidal activity by enhancement of azurophil granule-phagosome fusion via glycine/GlyRα2/TRPM2/p38 MAPK signaling. *J Immunol* 184, 4401–4413 (2010) DOI: 10.4049/jimmunol.0902814
- K Hill, CD Benham, S McNulty, AD Randall: Flufenamic acid is a pH-dependent antagonist of TRPM2 channels. *Neuropharmacology* 47,450-460 (2004) DOI: 10.1016/j.neuropharm.2004.04.014
- 51. K Hill, S McNulty, AD Randa: Inhibition of TRPM2 channels by the antifungal agents clotrimazole and econazole. Naunyn. Schmiedebergs. *Arch Pharmacol* 370, 227-237 (2004) DOI: 10.1007/s00210-004-0981-y
- R Kraft, C Grimm, H Frenzel, C Harteneck: Inhibition of TRPM2 cation channels by N-(p-amylcinnamoyl)anthranilic acid. *Br J Pharmacol* 148,264-273 (2006) DOI: 10.1038/sj.bjp.0706739
- 53. MR Bari, S Akbar, M Eweida, FJP Kühn, AJ Gustafsson, A Lückhoff, MS Islam: H₂O₂-induced Ca²⁺ influx and its inhibition by N-(p-amylcinnamoyl) anthranilic acid in the beta-cells: involvement of TRPM2 channels. *J Cell Mol Med* 13 (9B), 3260-3267 (2009) DOI: 10.1111/j.1582-4934.2009.00737.x
- 54. E Kheradpezhouh, GJ Barritt, GY Rychkov: Curcumin inhibits activation of TRPM2 channels in rat hepatocytes. *Redox Biol* 7,1-7 (2016)
 DOI: 10.1016/j.redox.2015.11.001
- 55. E Sözbir, M Nazıroğlu: Diabetes enhances oxidative stress-induced TRPM2 channel activity and its control by N-acetylcysteine in rat dorsal root ganglion and brain. *Metab Brain Dis* 31 (2),385-393 (2016) DOI: 10.1007/s11011-015-9769-7
- 56. C Moreau, T Kirchberger, JM Swarbrick, SJ Bartlett, R Fliegert, T Yorgan, A Bauche, A

Harneit, AH Guse, BV Potter: Structure-activity relationship of adenosine 5'-diphosphoribose at the transient receptor potential melastatin 2 (TRPM2) channel: rational design of antagonists. *J Med Chem* 56 (24),10079-10102 (2013)

DOI: 10.1021/jm401497a

K Togashi, H Inada, M Tominaga: Inhibition of the transient receptor potential cation channel TRPM2 by 2-aminoethoxydiphenyl borate (2-APB) Br J Pharmacol 153,1324-1330 (2008)

DOI: 10.1038/sj.bjp.0707675

- W Yang, J Zou, R Xia, ML Vaal, VA Seymour, J Luo, DJ Beech, LH Jiang: State-dependent inhibition of TRPM2 channel by acidic pH. *J Biol Chem* 285, 30411–30418 (2010) DOI: 10.1074/jbc.M110.139774
- M Naziroğlu: New molecular mechanisms on the activation of TRPM2 channels by oxidative stress and ADPR. *Neurochem Res* 32,1990-2001 (2007)
 DOI: 10.1007/s11064-007-9386-x
- F Schuber, FE Lund: Structure and enzymology of ADP-ribosyl cyclases: conserved enzymes that produce multiple calcium mobilizing metabolites. *Curr Mol Med* 4,249-261 (2004)

DOI: 10.2174/1566524043360708

61. CM Hecquet, GU Ahmmed, SM Vogel, AB Malik: Role of TRPM2 channel in mediating H₂O₂-induced Ca²⁺ entry and endothelial hyperpermeability. *Circ Res* 102,347-355 (2008)

DOI: 10.1161/CIRCRESAHA.107.160176

- 62. W Zhang, Q Tong, K Conrad, J Wozney, JY Cheung, BA Miller: Regulation of TRP channel TRPM2 by the tyrosine phosphatase PTPL1. Am J Physiol Cell Physiol 292,C1746-1758 (2007) DOI: 10.1152/ajpcell.00569.2006
- 63. CM Hecquet, GU Ahmmed, AB Malik: TRPM2 channel regulates endothelial barrier function. *Adv Exp Med Biol* 661,155-167 (2010) DOI: 10.1007/978-1-60761-500-2 10
- 64. F Qian, P Huang, L Ma, A Kuznetsov, N Tamarina, LH Philipson: TRP genes: candidates for nonselective cation channels and store-operated channels in insulin-secreting cells. *Diabetes* 51 Suppl 1,S183-189 (2002) DOI: 10.2337/diabetes.51.2007.S183

- 65. Y Mizukami, WS Jo, EM Duerr, M Gala, J Li, X Zhang, MA Zimmer, O Iliopoulos, LR Zukerberg, Y Kohgo, MP Lynch, BR Rueda, DC Chung: Induction of interleukin-8 preserves the angiogenic response in HIF-1alpha-deficient colon cancer cells. *Nat Med* 11,992-997 (2005) DOI: 10.1038/nm1294
- 66. S Yamamoto, N Takahashi, Y Mori: Chemical physiology of oxidative stress-activated TRPM2 and TRPC5 channels. *Prog Biophys Mol Biol* 103,18-27 (2010) DOI: 10.1016/j.pbiomolbio.2010.05.005
- 67. J Zhong, S Amina, M Liang, S Akther, T Yuhi, T Nishimura, C Tsuji, T Tsuji, HX Liu, M Hashii, F Furuhara, S Yokoyama, Y Yamamoto, H Okamoto, YJ Zhao, HC Lee, M Tominaga, O Lopatina, H Higashida: Cyclic ADP-Ribose and Heat Regulate Oxytocin Release via CD38 and TRPM2 in the Hypothalamus during Social or Psychological Stress in Mice. *Front Neurosci* 10,304 (2016) DOI: 10.3389/fnins.2016.00304
- 68. JA Wilkinson, JL Scragg, JP Boyle, B Nilius, C Peers: H₂O₂-stimulated Ca²⁺ influx via TRPM2 is not the sole determinant of subsequent cell death. *Pflugers Arch* 455,1141-1151 (2008)
 DOI: 10.1007/s00424-007-0384-2
- 69. CR Dass, PF Choong: Biophysical delivery of peptides: applicability for cancer therapy. *Peptides* 27,3479-3488 (2006) DOI: 10.1016/j.peptides.2006.08.016
- RC Ladner, AK Sato, J Gorzelany, M de Souza: Phage display-derived peptides as therapeutic alternatives to antibodies. *Drug Discov oday* 9,525-529 (2004) DOI: 10.1016/S1359-6446(04)03104-6
- ML Tan, PF Choong, CR Dass: Recent developments in liposomes, microparticles and nanoparticles for protein and peptide drug delivery. *Peptides* 31,184-193 (2010)
 DOI: 10.1016/j.peptides.2009.10.002
- 72. J Audie, C Boyd: The synergistic use of computation, chemistry and biology to discover novel peptide-based drugs: the time is right. *Cur Pharm Des* 16, 567-582 (2010) DOI: 10.2174/138161210790361425
- 73. P Järver, I Mäger, U Langel: In vivo bio-distribution and efficacy of peptide mediated

1437

delivery. *Trends Pharmacol* Sci 31,528-35 (2010)

DOI: 10.1016/j.tips.2010.07.006

- 74. P Pechon, A Tartar, MK Dunn: Peptide Report Development and Trends for Peptide Therapeutics (2010) [doi not found]
- T Pawson, P Nash: Assembly of cell regulatory systems through protein interaction domains. Science 300,445-452 (2003)
 DOI: 10.1126/science.1083653
- L Parthasarathi, F Casey, A Stein, P Aloy, DC Shields: Approved drug mimics of short peptide ligands from protein interaction motifs. *J Chem Inf Model* 48,1943-1948 (2008) DOI: 10.1021/ci800174c
- 77. P Molek, B Strukelj, T Bratkovic: Peptide phage display as a tool for drug discovery: targeting membrane receptors. *Molecules* 16,857-87 (2011) DOI: 10.3390/molecules16010857
- K Sonmez, NT Zaveri, IA Kerman, S Burke, CR Neal, X Xie, SJ Watson, L Toll: Evolutionary sequence modeling for discovery of peptide hormones. *PLoS Comput Biol* 5,e1000258 (2009) DOI: 10.1371/journal.pcbi.1000258
- 79. N London, D Movshovitz-Attias, O Schueler-Furman: The structural basis of peptide-protein binding strategies. *Structure* 18,188-199 (2010)
 DOI: 10.1016/j.str.2009.11.012
- E Petsalaki, A Stark, E García-Urdiales, RB Russell: Accurate prediction of peptide binding sites on protein surfaces. *PLoS Comput Biol* 5,e1000335 (2009) DOI: 10.1371/journal.pcbi.1000335
- 81. D Vicari, KC Foy, EM Liotta, PT Kaumaya: Engineered conformation-dependent VEGF peptide mimics are effective in inhibiting VEGF signaling pathways. *J Biol Chem* 286,13612-13625 (2011) DOI: 10.1074/jbc.M110.216812

Abbreviations: 2-APB: 2-aminoethoxydiphenyl borate; 6TM: Six trans-membrane; ACA: N-(p-amylcinnamoyl) anthranilic acid; ADPR: Adenosine diphospho-ribose; AMP: adenosine monophosphate; BD: Bipolar disorder; FFA: flufenamic acid; H2O2: hydrogen peroxide; NAD: nicotinamide adenine dinucleotide; NADases: NAD+ glycohydrolases; NUDT9-H: Nucleoside

diphosphate linked moiety X-type motif 9 homology; O2: Oxygen; PARP: poly-adenine dinucleotide phosphate-ribose polymerase; ROS: Reactive oxygen species; SNP: single nucleotide polymorphisms; SOD: superoxide dismutase; SSF-TRPM2: striatum short form transient receptor potential (melastatin) 2; TNF α : tumor necrosis factor α ; TRP: The transient receptor potential channel; TRPC7: Transient receptor potential-related channel 7; TRPM2: The transient receptor potential (melastatin) 2; TRPM2-WT: The transient receptor potential (melastatin) 2 wild type.

Key Words: Transient receptor potential melastatin 2, Oxidative stress, Calciumpermeable channel, Drug design, Review

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1438 © 1996-2017