The role of T cells in the development of Henoch-Schonlein purpura

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1. ABSTRACT

Henoch-Schonlein purpura (HSP) is an IgA-mediated disorder that most commonly occurs in children. Its etiology and pathogenesis remain unknown. In recent years, numerous studies have pointed to a dysfunction of T cells in the pathogenesis of HSP. Here, we will review the epidemiology, clinical and molecular characteristics of HSP, as well as abnormalities of Th cell subsets in this disorder. Finally, we will discuss the key factors that are involved in Th cell differentiation as potential novel targets for the prevention and treatment of HSP.

2. INTRODUCTION

Henoch-Schonlein purpura (HSP), also known as immunoglobulin A (IgA) vasculitis (1), is an IgA-mediated disorder that causes inflammation of small blood vessels, leading to hemorrhage in the skin, joints, intestines and kidneys. HSP can affect anyone, including elderly people (2), but it is most common in children between the ages of two and six years (3-5). It occurs about twice as often in boys as girls (6). The incidence of HSP in children is about 20 per 100,000 children per year, making it the most common form of vasculitis in children (4). The most striking feature of HSP is a purplish rash, typically on the lower legs and buttocks. HSP can also cause abdominal pain and aching joints. The condition usually improves on its own, but medical care is generally needed if the disorder affects the kidneys (3, 7). Severe HSP nephritis (HSPN) remains the major cause of morbidity and mortality among children with HSP (8). In a systematic review, 34.2% of HSP patients were found to have had renal involvement (9). Cases of HSP may occur anytime throughout the year, but some studies have found that fewer cases occur during the summer months, with most cases occurring in autumn and winter (10).

Multiple standards exist for defining HSP, including the 1990 American College of Rheumatology classification (11-13) and the 1994 Chapel Hill Consensus Conference nomenclature of vasculitides (14, 15). More recent classifications include the 2006 European League Against Rheumatism and Pediatric Rheumatology Society classification criteria (16, 17). Recently, a modified semiguantitative classification was found to be more sensitive than the classical International Study of Kidney Disease in Children system for predicting the outcome in cases of HSPN (18). Like most human illnesses, HSP is an etiologicallycomplex disease, with environmental factors triggering the disease in genetically predisposed individuals. The current diagnostic criteria have high sensitivity and specificity, and most HSP patients are accurately diagnosed. However, although the prognosis is generally good, the failure of HSP treatment remains a key problem. To reduce associated morbidity and mortality and improve patient quality of life, further research is needed to improve our understanding of the pathogenetic mechanisms underlying HSP.

3. CHARACTERISTICS OF HSP

The pathophysiological features of HSP are not well understood but appear to involve the deposition of immune complexes containing IgA and complement component 3 (C3) in arterioles, capillaries and venules (19). The activation of the complement system, in part through the alternative pathway, may result in increased plasma levels of C3a and C5a; in particular, the increased level of C5a is thought to play a role in disease pathogenesis by activating the endothelium of cutaneous small vessels (19). Increased synthesis of IgA in response to antigens processed by the mucosa-associated immune system also appears to contribute to disease development. HSPN and IgA nephropathy are related diseases, with both resulting from glomerular deposition of aberrantly glycosylated IgA. Although both nephritides present with similar histological findings and IgA abnormalities, they display pathophysiological differences that have important therapeutic implications (20, 21). HSP nephritis is mainly characterized by acute episodes of glomerular inflammation with endocapillary and mesangial proliferation, fibrin deposits and epithelial crescents that can heal spontaneously or lead to chronic lesions. In contrast, IgA nephropathy normally presents with slowly progressing mesangial lesions resulting from continuous low-grade deposition of macromolecular IgA1 (22). Further, IgA nephropathy has a predilection for young adults while HSP is more predominant among children, and IgA nephropathy typically only affects the kidneys, while HSP is a systemic disease (23-25).

The dominant clinical features of HSP are cutaneous purpura (100%), arthritis (82%), abdominal pain (63%), gastrointestinal bleeding (33%) and nephritis (40%) (26). These symptoms can be summarized as four main characteristics (27, 28). The first is rash (purpura). Reddish-purple spots which look like bruises are the most distinctive and universal sign of HSP. The rash develops mainly on the buttocks, legs and feet, but can also appear on the arms, face and trunk and may be worse in areas of pressure, such as the sock line and waistline (29, 30). The second characteristic is swollen sore joints (arthritis). Patients with HSP often have pain and swelling around the joints, mainly in the knees and ankles. Joint pain sometimes precedes the classical rash by one or two weeks. These symptoms subside when the disease clears and leave no lasting damage (31). The third characteristic is gastrointestinal symptoms which many children with HSP develop, including abdominal pain. nausea, vomiting or bloody stools. These symptoms sometimes occur before the rash appears, so they can be helpful in the early diagnosis of HSP (32, 33). The final characteristic is kidney involvement. In most cases, this is revealed by protein or blood in the urine, which may not be evident unless a urine test is performed.

This normally dissipates once the illness passes, but in a few cases kidney disease may develop and persist. Before these symptoms begin, patients may have two to three weeks of fever, headache and muscular aches and pains. Other organs, such as the brain, heart and lungs, may also be affected (34, 35). There is a significant correlation between the severity of renal involvement and pathological grading and scoring of HSPN; in particular, the severity of proteinuria is a significant determinant of renal pathologies (36). The exact cause of this phenomenon is unknown. It usually resolves within several weeks and requires no treatment apart from symptom control, but may relapse in a third of cases and cause irreversible kidney damage in about one in a hundred cases.

4. INVOLVEMENT OF CD4⁺ T CELLS IN HSP DEVELOPMENT

The immune system is believed to play a role in targeting the blood vessels involved in HSP. An abnormal immune response to an infection may be a factor in many cases. In recent years, the role of T cells in the pathogenesis of HSP has become a focus of research. It has been suggested that leukocyturia is associated with post-infectious glomerulonephritis (GN), interstitial nephritis and renal allograft rejection. In addition, prominent infiltration of T cells and macrophages is commonly observed in the renal tissues of patients with GN, accompanied by cellular crescent formation and/or interstitial cell infiltration (37). In urine from patients with different forms of GN, including IgA nephropathy, HSPN and anti-neutrophil cytoplasmic antibody-associated GN, T cells appeared together with macrophages. T cells from urine were mainly CD45RA⁻, CD45RO⁺ and CD62L (L-selectin), which are the phenotypic features of effector T cells (37). The findings suggest that the appearance of effector T cells in urine may reflect the cellular immune reaction that occurs in the kidneys of patients with GN, which is accompanied by active cell infiltration.

CD4⁺ T cells play critical roles in mediating adaptive immunity to a variety of pathogens. They are also involved in autoimmunity, asthma and allergic responses as well as in tumor immunity. Recently, these cells have been found to contribute to the pathogenesis of HSP. Results from 32 HSP patients and 25 healthy donors revealed that freshly isolated CD4⁺ T cells from patients with HSP expressed higher levels of OX40 than cells from healthy individuals (38). The levels of soluble OX40L (Sox40l) in the sera of patients with HSP were also much higher than in controls. Importantly, significantly elevated levels of OX40 on CD4⁺ T cells and Sox40L in sera were detected in patients with HSPN compared to patients without nephritis, indicating that both OX40 upregulation and an increase in Sox40I were closely associated with disease activity in these patients (38).

As OX40/OX40L is a costimulatory pathway that can promote T-cell activation and prolong survival, their upregulation indicates the involvement of CD4⁺ T cells in the pathology of HSP.

During T cell receptor (TCR) activation in a particular cytokine milieu, naive CD4⁺ T cells may differentiate into different subsets of T helper (Th) cells. There are at least four types of Th cells: Th1, Th2, Th17 and regulatory T (Treg) cells, as defined by their pattern of cytokine production and function. Th1, Th2 and Th17 cells are important for eradicating intracellular pathogens, helminth and extracellular bacteria/fungi, respectively. Th1 and Th17 cells are also involved in many types of autoimmune diseases, whereas Th2 cells contribute to allergic responses (39). Treg cells are critical in maintaining self-tolerance and in modulating immune responses to infections (40). CD4⁺ T cells are also important in the induction and control of immunoglobulin class switching and somatic hypermutation. These events occur mainly within germinal centers (GCs), and CD4⁺ T cells that enter GCs to mediate their helper function for antibody production are often designated T follicular helper (Tfh) cells (41). Children with HSP show T-cell disorders and abnormalities in Th cell differentiation.

4.1. Th1 cells

Th1 cells are the host immunity effectors against intracellular bacteria and protozoa. They are triggered by interleukin 12 (IL-12) and IL-2, and their effector cytokine is interferon gamma (IFN- γ). The key Th1 transcription factors are signal transducer and activator of transcription 4 (STAT4) and T-bet. Th1 overactivation against autoantigens will cause type 4 delayed-type hypersensitivity; tuberculin reactions and type 1 diabetes also belong to this category of autoimmunity (39).

There are findings showing that in urine from patients with IgA nephropathy, HSPN or anti-neutrophil cvtoplasmic antibody-associated GN. T cells were present along with macrophages. These urine cells expressed mRNA for Th lymphocyte 1 cytokines, IL-2 and/or IFN-y, indicating the involvement of Th1 cells in the pathology of HSP (37). Furthermore, renal biopsy specimens from 22 pediatric patients diagnosed with HSP were compared to normal renal tissue taken during nephrectomy in 20 pediatric patients diagnosed with Wilms tumor (42). Immunohistochemical analysis of IFN-y expression showed that glomeruli and tubules in HSP patients had significantly higher IFN-y expression than those in control patients. This suggests that IFN-v may contribute to HSP in children and provides indirect evidence that Th1 cells may be involved in the pathogenesis of HSP. Another study suggesting Th1 predominance in HSP studied the transcriptional factor T-bet, which regulates the differentiation of Th lymphocytes into the Th1 subset (43). The relative expression of T-bet was significantly higher in the urinary sediment from patients with HSP than in healthy controls. Moreover, a significant increase in T-bet expression was observed in glomeruli biopsy specimens from all patients studied. When the patients received immunosuppressive therapy, expression of T-bet was reduced (43), suggesting a predominant role for Th1 cells in the development of HSP. However, other studies have found no evidence that Th1 is involved in the pathogenesis of HSP (44, 45).

4.2. Th2 cells

Th2 cells are the host immunity effectors against extracellular parasites, including helminths. They are triggered by IL-4 and their effector cytokines are IL-4, IL-5, IL-9, IL-10 and IL-13. The key Th2 transcription factors are STAT6 and GATA3 (46). IL-4 is the positive feedback cytokine for Th2 cell differentiation, and overactivation of Th2 in response to autoantigen causes Type1 IgE-mediated allergy and hypersensitivity. Allergic rhinitis, atopic dermatitis and asthma belong to this category of autoimmunity (39).

The transcriptional factor GATA-3, which regulates the differentiation of Th lymphocytes into the Th2 subset, was examined in HSP patients (43). The relative expression of GATA-3 was significantly lower in the urinary sediment of patients than in control subjects. When the patients received immunosuppressive therapy, the expression of GATA-3 remained static (43), which suggested that Th2 cells were not be involved in the pathogenesis of HSP. In another study, histone modification patterns in peripheral blood mononuclear cells (PBMCs) from HSP patients were analyzed and the expression of inflammatory cytokines (IFN-y, IL-2, IL-4, IL-6 and IL-13), transcription factors (T-bet, GATA-3 and TIM-1) and chemokines (CXCL4 and CXCL10) were investigated (47). Results showed that histone H3 acetylation and methylation were significantly enhanced in PBMCs from HSP patients. In particular, there were marked increases in histone H3 acetylation and H3 lysine 4 trimethylation at the IL-4 locus in these patients. In addition, the expression levels of IL-4, IL-6, IL-13, GATA-3, TIM-1 and CXCL4 were increased, indicating that abnormal histone modifications may have led to the Th1/Th2 cvtokine imbalance in HSP. and hence that Th2 cells may contribute to the pathogenesis of HSP. The same conclusion can be drawn from another study which showed that toll-like receptor 2 (TLR2) and TLR4 overactivation induced HSP-related renal impairment, and further indicated that activated TLR2 and TLR4 may mediate the pathogenesis of HSP by upregulating a Th2-type immune response (45). Finally, a study of 42 children with acute HSP and 30 healthy children showed that the plasma levels of IL-4, IFN-y and IL-17 were significantly higher in the

HSP group compared to the controls. And the TLR6 protein expression levels in the monocytes of the HSP group significantly positively correlated with the serum IL-4 and IL-17 levels, but not with the serum levels of IFN- γ . Thus, the activation of TLR6 may be involved in the immunopathogenesis of HSP by upregulating the immune responses of Th2 and Th17 cells (48). However, further studies will be required to confirm Th2 cell involvement in the pathology of HSP.

4.3. Th17 cells

Th17 cells are a subset of Th cells, and are developmentally distinct from Th1 and Th2 lineages in that they produce IL-17. They are related to Treg cells because the signals that cause Th17 to differentiate inhibit Treg differentiation (49). Transforming growth factor beta (TGF- β), IL-6, IL-21 and IL-23 contribute to Th17 formation in mice and humans. Key factors involved in the differentiation of Th17 cells are STAT3 and retinoic acid receptor-related orphan receptor gamma (ROR γ) and alpha (ROR α) (50). Th17 cells can alter their differentiation program, ultimately giving rise to either protective or pro-inflammatory pathogenic cells.

IL-17 and Th17 cells are known to be involved in many autoimmune diseases, and studies have shown that IL-17 and Th17 cells may be involved in the pathogenesis of childhood HSP. For example, it has been shown that children with acute HSP have significantly higher serum levels of IL-17, IL-6 and TGF- β than healthy controls (44, 51). These patients also had more Th17 cells but not Th1 cells in peripheral blood, indicating that Th17 cells and serum IL-17 may each contribute in part to HSP and that upregulation of Th17 cells may perpetuate the inflammatory response in HSP. Immunohistochemical analysis of renal biopsy specimens from children with HSP demonstrated significantly higher IL-17 expression (42). Another study of 42 children with acute HSP and 30 healthy children also showed that plasma levels of IL-17 were significantly higher in patients than in healthy controls (48). Thus, the activation of TLR6 may be involved in the immunopathogenesis of HSP by upregulating the immune responses of Th2 and Th17 cells. Taken together, these studies suggest a role for Th17 cells in HSP.

4.4. Treg cells

Tregs are a subpopulation of T cells that modulate the immune system, maintain tolerance to self-antigens and prevent autoimmune disease. Tregs come in many forms, with the well-understood being those that express CD4, CD25 and forkhead box P3 (FOXP3). Tregs are immunosuppressive and generally suppress or downregulate induction and proliferation of effector T cells (52). The cytokine TGF- β has been found to be essential for Treg differentiation from naive CD4⁺ T cells and for the maintenance of Treg homeostasis (53, 54). The immunosuppressive cytokines TGF- β and IL-10 have also been implicated in Treg function.

Treg cells have been implicated in a wide range of autoimmune disorders, such as rheumatoid arthritis, autoimmune liver disease, systemic lupus and immune-mediated diabetes (53, 55, 56). A study of 63 children with HSP showed that both FoxP3 and TGF- β 1 mRNA expression was significantly lower than in healthy controls, indicating lowered Treg activity (57). Another study reported a lower frequency of Treg cells and reduced IL-10 concentration in HSP patients compared to healthy controls (51). However, further studies will be required to fully elucidate the role of Treg cells in HSP.

4.5. Tfh cells

Tfh cells are found in the periphery of B cell follicles and are identified by their constitutive expression of the B cell follicle-homing receptor, CXC chemokine receptor 5 (CXCR5) (58). They are critical for the activation of B cells, antibody class switching and GC formation (59). The inducible T-cell co-stimulator ICOS or CD278 was shown to provide a particularly critical signal for Tfh cells (60). It has been reported that ICOS induces the secretion of IL-21 by activated CD4⁺ T cells and that IL-21 plays a crucial role in the development of Tfh cells and GCs (61-63). B-cell lymphoma-6 (Bcl-6) and programmed death 1 (PD-1) have also been identified as transcription factors in Tfh cells. Thus, Tfh cells are characterized by the expression of CXCR5, ICOS, PD-1, Bcl-6 and IL-21 (59). However, different studies define blood Tfh cells in different ways. Although some studies have defined circulating human Tfh cells as the total population of CXCR5⁺ CD4⁺ T cells, other studies have investigated subsets of these cells, such as CXCR5+ ICOS⁺, CXCR5⁺ ICOShi, CXCR5⁺ PD1⁺, CXCR5⁺ PD1hi, CXCR5⁺ ICOS⁺ PD1⁺, CXCR5⁺ CD57⁺ and CXCR5⁺ IL-21⁺ cells (64-73).

Tfh cells play crucial roles in regulating immune responses. Studies have shown that the frequency of circulating CXCR5⁺ CD4⁺ Tfh cells coexpressing ICOS was significantly higher in children with acute HSP than in healthy controls, whereas the frequency of CXCR5⁺ CD4⁺ Tfh cells expressing PD-1 was not increased in these patients (74, 75). Moreover, serum levels of IL-21, IL-6, IgA and C3 were also significantly higher in HSP children than in healthy controls. A positive correlation was observed between the frequency of circulating ICOS⁺ CXCR5⁺ CD4⁺ Tfh cells and the serum levels of IL-21 or IgA (74). Additionally, mRNA expression levels of IL-21, IL-6 and Bcl-6 were also significantly increased in peripheral blood from children with acute HSP compared to healthy controls (74). Following treatment, the numbers of CD4⁺ CXCR5⁺, CD4⁺ CXCR5⁺ PD-1⁺ and CD4⁺ CXCR5⁺ ICOS⁺ Tfh cells, as well as serum levels of IL-21 were significantly reduced (75). Taken together, these findings suggest that Tfh cells and their associated molecules may play critical roles in the pathogenesis of HSP, and thus can be considered as possible therapeutic targets.

Taken together, increasing numbers of Th1, Th2 and Tfh cells, together with decreasing Treg cell numbers may contribute in part to the pathogenesis of HSP. Further research into the differentiation of Th cells can be expected to provide more information regarding the pathogenesis of HSP and to identify novel targets for disease control and prevention.

5. KEY FACTORS AFFECTING CD4⁺ T CELL DIFFERENTIATION AS THERAPEUTIC TARGETS

The various functions of mature CD4⁺ T cells are achieved through the differentiation of naive CD4⁺ T cells following stimulation by their cognate antigens presented by competent antigen-presenting cells. This results in the formation of subsets of effector and/ or memory cells with specialized phenotypes. Our knowledge of Th cells has expanded greatly. T cell heterogeneity and plasticity open new opportunities for targeting or redirecting specific subsets of cells in autoimmune and allergic diseases. These targets may become clinically feasible when we more fully understand the regulation of Th cell subsets and their relationship to one another.

Cytokines play critical roles in determining Th cell differentiation. The distinctive differentiated states of the various CD4 effector/regulatory subpopulations are determined largely by the set of transcription factors they express and the genes they transcribe. The induction of distinctive patterns of gene expression may be achieved in several ways, but in vitro, the major determinants of the differentiated state of the cell are the set of cytokines present during the TCR-mediated activation process. A combination of cytokines is required for the differentiation of each lineage: IL-12 and IFN-y for Th1; IL-4 and IL-2/IL-7/ thymic stromal lymphopoietin for Th2; TGF-B and IL-6/IL-21/IL-23 for Th17; TGF-β and IL-2 for Treg; and IL-6 and IL-21 for Tfh cells (Figure 1). One of the effector cytokines produced by each subset of Th cells further promotes the differentiation process, providing a powerful positive amplification loop. The IL-1 family of cytokines may also participate in inducing TCRindependent effector-cytokine production by Th cells, including IL-18 for Th1, IL-33 for Th2 and IL-1 for Th17 cells.

Master transcription factors and STAT proteins are indispensable for Th cell fate determination and cytokine production. GATA3, the Th2 master regulator, was the first master regulator to be identified (76, 77). GATA3 expression is upregulated or downregulated during Th2 or Th1 differentiation, respectively (77-79). Deleting Gata3 from fully differentiated Th2 cells by the introduction of retrovirally-encoded Cre has only a modest effect on IL-4 production but completely blocks the production of IL-5 and IL-13, consistent with GATA3 binding directly to the IL-5 (80) and IL-13 (81, 82) promoters, but only binding IL-4 enhancers (83). T-bet is a major factor for inducing IFN-v production and Th1 cell differentiation (84). T-bet induces IFN-y partly through remodeling the *lfng* gene and by upregulating IL-12R_β2 expression, thus promoting both IFN-y expression and selective Th1 cell expansion in response to IL-12 (85, 86). The differential requirement for T-bet in IFN-y production by CD4 and CD8 T cells may be explained by the heightened expression of another T-box family member, Eomesodemin (Eomes), in CD8 T cells (87). IL-21 inhibition of Th1 cell IFN-y production may be mediated by suppression of Eomes but not by suppression of T-bet (88), suggesting that Eomes is also upregulated during Th1 differentiation and is involved in optimal IFN-y production by CD4 T cells. Foxp3 has been reported to be the master transcriptional regulator for Tregs (89, 90). Continuous expression of Foxp3 in Tregs is required to maintain the suppressive activity of these cells (91). Limiting Foxp3 expression appears to divert cells that would have differentiated into Tregs into becoming Th2-like cells, implying a close relationship between the Th2 and Treg lineages (92). Th17 cells do not express GATA3 or T-bet. Instead, they express high levels of RORy (50, 93, 94). RORyt is the master regulator of Th17 cells, and RORvt-deficient mice are partially resistant to experimental autoimmune encephalomvelitis (95). Bcl-6 is a transcriptional repressor. It is frequently translocated and hypermutated in diffuse large B cell lymphoma and is critical for GC B cell differentiation and thus GC formation (96). Bcl-6 is also expressed in Tfh cells and is critical for Tfh cell differentiation. It is necessary and sufficient to induce Tfh-related molecules, including CXCR5, PD-1, IL-6R and IL-21R (97-99). Bcl-6 also suppresses the expression of Th1, Th2 and Th17 cytokines.

As mentioned above, the major signaling pathway triggered by cytokines is the activation of the STAT family of proteins. STATs play critical roles in the differentiation and expansion of Th cells. Activation of STAT1 by IFN- γ is important for the induction of T-bet during *in vitro* Th1 differentiation (100, 101). The existence of a positive feedback loop—in which IFN- γ , acting through T-bet, induces more IFN- γ —indicates that STAT1 serves as a critical mediator for the amplification of *in vitro* Th1 responses. STAT2 forms

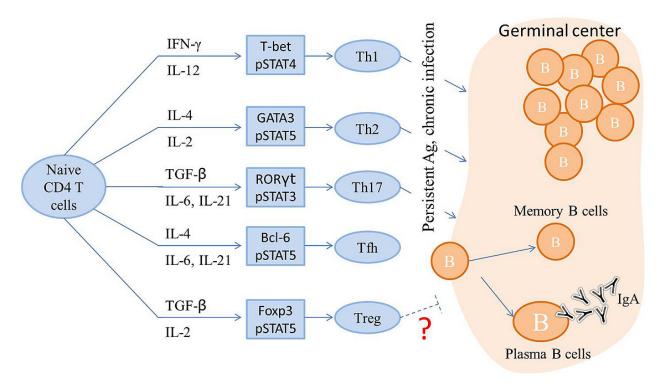


Figure 1. Differentiation of Th cells and their role in B cell function. Upon TCR activation triggered by antigen-presenting cells, naive CD4 T cells differentiate into distinct Th lineages in the context of combinations of cytokines. The differentiation processes involve upregulation of master transcriptional regulators and activation of STAT proteins. With the help of different Th cells, B cell is activated and migrates into the germinal center and proliferates, followed by differentiation further to memory B cells or antibody secreting plasma cells. Plasma B cells then secret IgA which mediates HSP.

a heterodimeric complex with STAT1 in response to type I IFNs. IL-6, IL-21 and IL-23 cytokines that are involved in Th17 cell differentiation, amplification and maintenance also induce TCR-independent, cyclosporine A-independent IL-17A production through the activation of STAT3 (102-105). STAT3 binds to IL17 (106) and IL21 (107) and is responsible for the induction of RORyt and IL-23R (103, 108, 109). STAT4 expression is higher in Th1 compared to Th2 cells (117). Activated STAT4 can directly induce IFN-y production and the expression of IL-12R β 2 and T-bet during Th1 differentiation (54, 117). Low levels of STAT5 activation are sufficient for cell proliferation and survival; however, strong STAT5 signaling is required for Th2 differentiation (110, 111). STAT5 activation by IL-2 is also critical for Treg development (112-114). STAT5 may contribute to Foxp3 induction by binding to its promoter (114, 115). STAT5 activation also regulates the activity of the Bcl6 promoter in B cells (116). In view of the expression of Bcl-6 in Tfh cells, this regulation raises the possibility that such an effect may be important for Tfh cell differentiation. STAT6 is the major signal transducer in IL-4-mediated Th2 differentiation and expansion (117-119). In vitro, STAT6 activation is necessary and sufficient for inducing high levels of expression of the Th2 master regulator gene, GATA3 (120, 121).

In summary, a collaboration exists between the master regulators and STAT family members in T cell differentiation and expansion: T-bet and STAT4 for Th1; GATA3 and STAT5 for Th2; RORyt and STAT3 for Th17; Foxp3 and STAT5 for Treg; and Bcl-6 and STAT5 for Tfh (Figure 1). Other transcription factors are either secondary to master regulators and STAT proteins or are responsible for the induction of master regulators; these include Runx family members (Runx1, Runx2, and Runx3) (122-125), IFN regulatory factor family members (IRF4 and IRF1) (126, 127), Gfi-1 (128, 129), Ikaros family members (Ikaros, Helios, Aiolos, Eos, and Pegasus) (130), c-Maf (131), and others like HIx (a transcription factor induced by T-bet) (85), Ets-1 (a cofactor for T-bet during Th1 differentiation) (132, 133) and Blimp-1 (an important transcription factor induced in Th2 cells) (134, 135).

6. CONCLUSIONS

Like most human illnesses, HSP is an etiologically-complex disease. Although the prognosis is generally good, the failure to find effective treatment strategies for HSP remains a key problem. The immune system is believed to play a role in targeting the blood vessels involved. In recent years, the role of T cells in the pathogenesis of HSP has become a

focus of research, but information regarding Th cell involvement in HSP is still incomplete. Many signaling molecules and transcription factors shown to be critical for Th cell differentiation in mouse models are also defective in human diseases related to abnormal Th cell differentiation. Linking animal models with clinical studies should provide greater insight into the details of Th cell differentiation. As abnormalities in Th cell differentiation are present in HSP, it is important to elucidate pathologies in Th cell differentiation to fully understand this disease.

7. REFERENCES

- 1. Jennette, J.C., R.J. Falk, P.A. Bacon, N. Basu, M.C. Cid, F. Ferrario, L.F. Flores-Suarez, W.L. Gross, L. Guillevin, E.C. Hagen, G.S. Hoffman, D.R. Jayne, C.G. Kallenberg, P. Lamprecht, C.A. Langford, R.A. Luqmani, A.D. Mahr, E.L. Matteson, P.A. Merkel, S. Ozen, C.D. Pusey, N. Rasmussen, A.J. Rees, D.G. Scott, U. Specks, J.H. Stone, K. Takahashi, and R.A. Watts: 2012 revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides. Arthritis Rheum 65:1-11 (2013) DOI: 10.1002/art.37715
- 2. Min, Z., R.R. Garcia, M. Murillo, J.M. Uchin, and N. Bhanot: Vancomycin-associated Henoch-Schonlein purpura. J Infect Chemother 23:180–184 (2017) DOI: 10.1016/j.jiac.2016.08.012
- 3. Delbet, J.D., J. Hogan, B. Aoun, I. Stoica, R. Salomon, S. Decramer, I. Brocheriou, G. Deschenes, and T. Ulinski: Clinical outcomes in children with Henoch-Schonlein purpura nephritis without crescents. Pediatr Nephrol (2017)

DOI: 10.1007/s00467-017-3604-9

- Gardner-Medwin, J.M., P. Dolezalova, 4. C. Cummins, and T.R. Southwood: Incidence of Henoch-Schonlein purpura, Kawasaki disease, and rare vasculitides in children of different ethnic origins. Lancet 360:1197-202 (2002) DOI: 10.1016/S0140-6736(02)11279-7
- 5. Blanco. R., V.M. Martinez-Taboada, V. Rodriguez-Valverde, M. Garcia-Fuentes. and M.A. Gonzalez-Gay: Henoch-Schonlein purpura in adulthood and childhood: two different expressions of the same syndrome. Arthritis Rheum 40:859–64 (1997) DOI: 10.1002/art.1780400513

- 6. Piram, M. and A. Mahr: Epidemiology of immunoglobulin A vasculitis (Henoch-Schonlein): current state of knowledge. Curr *Opin Rheumatol* 25:171–8 (2013) DOI: 10.1097/BOR.0b013e32835d8e2a
- 7. Feng, D., W.Y. Huang, S. Hao, X.L. Niu, P. Wang, Y. Wu, and G.H. Zhu: A singlecenter analysis of Henoch-Schonlein purpura nephritis with nephrotic proteinuria in children. Pediatr Rheumatol Online J 15:15 (2017) DOI: 10.1186/s12969-017-0146-4
- Saulsbury, F.T.: Clinical update: Henoch-8. Schonlein purpura. *Lancet* 369:976–8 (2007) DOI: 10.1016/S0140-6736(07)60474-7
- Narchi, H.: Risk of long term renal impairment 9. and duration of follow up recommended for Henoch-Schonlein purpura with normal or minimal urinary findings: a systematic review. Arch Dis Child 90:916-20 (2005) DOI: 10.1136/adc.2005.074641
- 10. Saulsbury, F.T.: Epidemiology of Henoch-Schonlein purpura. Cleve Clin J Med 69 Suppl 2:SII87–9 (2002) DOI: 10.3949/ccjm.69.Suppl 2.SII87
- 11. Mills, J.A., B.A. Michel, D.A. Bloch, L.H. Calabrese, G.G. Hunder, W.P. Arend, S.M. Edworthy, A.S. Fauci, R.Y. Leavitt, J.T. Lie, and et al.: The American College of Rheumatology 1990 criteria for the classification of Henoch-Schonlein purpura. Arthritis Rheum 33:1114–21 (1990) DOI: 10.1002/art.1780330809
- 12. Ortiz-Sanjuan, F., R. Blanco, J. Loricera, J.L. Hernandez, T. Pina, V. Calvo-Rio, L. Alvarez, M.C. Gonzalez-Vela, J. Rueda-Gotor, M.A. Gonzalez-Lopez, and M.A. Gonzalez-Gay: Reappraisal of the 1990 American College of Rheumatology criteria for the classification of cutaneous vasculitis: an analysis based on 766 patients. Clin Exp Rheumatol 32:S51-4 (2014)
- 13. Bloch, D.A., B.A. Michel, G.G. Hunder, D.J. McShane, W.P. Arend, L.H. Calabrese, S.M. Edworthy, A.S. Fauci, J.F. Fries, R.Y. Leavitt, and et al.: The American College of Rheumatology 1990 criteria for the classification of vasculitis. Patients and methods. Arthritis Rheum 33:1068-73 (1990)

DOI: 10.1002/art.1780330803

- Jennette, J.C., R.J. Falk, K. Andrassy, P.A. Bacon, J. Churg, W.L. Gross, E.C. Hagen, G.S. Hoffman, G.G. Hunder, C.G. Kallenberg, and *et al.*: Nomenclature of systemic vasculitides. Proposal of an international consensus conference. *Arthritis Rheum* 37:187–92 (1994) DOI: 10.1002/art.1780370206
- Murali, N.S., R. George, G.T. John, S.M. Chandi, M. Jacob, L. Jeyaseelan, P.P. Thomas, and C.K. Jacob: Problems of classification of Henoch Schonlein purpura: an Indian perspective. *Clin Exp Dermatol* 27:260–3 (2002) DOI: 10.1046/j.1365-2230.2002.01063.x
- Aalberse, J., K. Dolman, G. Ramnath, R.R. Pereira, and J.C. Davin: Henoch Schonlein purpura in children: an epidemiological study among Dutch paediatricians on incidence and diagnostic criteria. *Ann Rheum Dis* 66:1648–50 (2007) DOI: 10.1136/ard.2006.069187
- Ozen, S., N. Ruperto, M.J. Dillon, A. Bagga, K. Barron, J.C. Davin, T. Kawasaki, C. Lindsley, R.E. Petty, A.M. Prieur, A. Ravelli, and P. Woo: EULAR/PReS endorsed consensus criteria for the classification of childhood vasculitides. *Ann Rheum Dis* 65:936–41 (2006) DOI: 10.1136/ard.2005.046300
- Koskela, M., E. Ylinen, E.M. Ukonmaanaho, H. Autio-Harmainen, P. Heikkila, J. Lohi, O. Jauhola, J. Ronkainen, T. Jahnukainen, and M. Nuutinen: The ISKDC classification and a new semiquantitative classification for predicting outcomes of Henoch-Schonlein purpura nephritis. *Pediatr Nephrol* (2017) DOI: 10.1007/s00467-017-3608-5
- 19. Yang, Y.H., I.J. Tsai, C.J. Chang, Y.H. Chuang, H.Y. Hsu, and B.L. Chiang: The interaction between circulating complement proteins and cutaneous microvascular endothelial cells in the development of childhood Henoch-Schonlein Purpura. *PLoS One* 10:e0120411 (2015) DOI: 10.1371/journal.pone.0120411
- 20. Komatsu, H., S. Fujimoto, N. Yoshikawa, H. Kitamura, H. Sugiyama, and H. Yokoyama: Clinical manifestations of Henoch-Schonlein purpura nephritis and IgA nephropathy: comparative analysis of data from the Japan

Renal Biopsy Registry (J-RBR) *Clin Exp Nephrol* 20:552–60 (2016) DOI: 10.1007/s10157-015-1177-0

- Mao, S., X. Xuan, Y. Sha, S. Zhao, C. Zhu, A. Zhang, and S. Huang: Clinico-pathological association of Henoch-Schoenlein purpura nephritis and IgA nephropathy in children. *Int J Clin Exp Pathol* 8:2334–42 (2015)
- 22. Davin, J.C. and R. Coppo: Henoch-Schonlein purpura nephritis in children. *Nat Rev Nephrol* 10:563–73 (2014) DOI: 10.1038/nrneph.2014.126
- 23. Rai, A., C. Nast, and S. Adler: Henoch-Schonlein purpura nephritis. *J Am Soc Nephrol* 10:2637–44 (1999)
- Pillebout, E., E. Thervet, G. Hill, C. Alberti, P. Vanhille, and D. Nochy: Henoch-Schonlein Purpura in adults: outcome and prognostic factors. *J Am Soc Nephrol* 13:1271–8 (2002) DOI: 10.1097/01. ASN.0000013883.99976.22
- Yoshikawa, N., H. Ito, K. Yoshiya, C. Nakahara, S. Yoshiara, O. Hasegawa, S. Matsuyama, and T. Matsuo: Henoch-Schoenlein nephritis and IgA nephropathy in children: a comparison of clinical course. *Clin Nephrol* 27:233–7 (1987)
- Saulsbury, F.T.: Henoch-Schonlein purpura in children. Report of 100 patients and review of the literature. *Medicine (Baltimore)* 78:395–409 (1999) DOI: 10.1097/00005792-199911000-00005
- Trnka, P.: Henoch-Schonlein purpura in children. J Paediatr Child Health 49:995– 1003 (2013) DOI: 10.1111/jpc.12403
- Cameron, J.S.: Henoch-Schonlein purpura: clinical presentation. *Contrib Nephrol* 40:246–9 (1984) DOI: 10.1159/000409757
- Paydary, K., S. Emamzadeh Fard, A.H. Mahboubi, V. Ziaee, M.H. Moradinejad, and A.M. Kajbafzadeh: Penile Skin Involvement as the First Presentation of Henoch-Schonlein Purpura Report of Nine Cases and Review of Literature. *Iran J Pediatr* 25:e2177 (2015) DOI: 10.5812/ijp.2177

- 30. Nussinovitch, M., D. Prais, Y. Finkelstein, and I. Varsano: Cutaneous manifestations of Henoch-Schonlein purpura in young children. *Pediatr Dermatol* 15:426–8 (1998) DOI:10.1046/j.1525-1470.1998.1998015426.x
- Duman, M.A., N.S. Duru, B. Caliskan, H. Sandikci, and F. Cengel: Lumbar Swelling as the Unusual Presentation of Henoch-Schonlein Purpura in a Child. *Balkan Med J* 33:360–2 (2016) DOI: 10.5152/balkanmedj.2016.150208
- 32. Zhang, Y. and X. Huang: Gastrointestinal involvement in Henoch-Schonlein purpura. *Scand J Gastroenterol* 43:1038–43 (2008) DOI: 10.1080/00365520802101861
- Nay, J., C.O. Menias, V.M. Mellnick, and D.M. Balfe: Gastrointestinal manifestations of systemic disease: a multimodality review. *Abdom Imaging* 40:1926–43 (2015) DOI: 10.1007/s00261-014-0334-3
- Dos Santos, D., F.W. Langer, T. Dos Santos, G. Rafael Tronco Alves, M. Feiten, and W. Teixeira de Paula Neto: Posterior reversible encephalopathy syndrome as a complication of Henoch-Schonlein purpura in a sevenyear-old girl. *Scott Med J* 62:34–37 (2017) DOI: 10.1177/0036933017690467
- 35. Bellantoni, A., P. Lo Presti, A. Giordano, M. Chiatto, M. Matta, and G. Perri: (A pediatric case of Schoenlein-Henoch purpura with clinical, serologic and electrocardiographic signs of myocardial damage) *G Ital Cardiol (Rome)* 14:622–5 (2013)
- 36. Nickavar, A., M. Mehrazma, and A. Lahouti: Clinicopathologic correlations in Henoch-Schonlein nephritis. *Iran J Kidney Dis* 6:437–40 (2012)
- Sakatsume, M., Y. Xie, M. Ueno, H. Obayashi, S. Goto, I. Narita, N. Homma, K. Tasaki, Y. Suzuki, and F. Gejyo: Human glomerulonephritis accompanied by active cellular infiltrates shows effector T cells in urine. *J Am Soc Nephrol* 12:2636–44 (2001)
- Qin, W., W. Hongya, C. Yongjing, X. Fang, M. Yue, Z. Xuekun, L. Xiaozhong, and Z. Xueguang: Increased OX40 and soluble OX40 ligands in children with Henoch-Schonlein purpura: association with renal involvement. *Pediatr Allergy Immunol* 22:54–9 (2011) DOI: 10.1111/j.1399-3038.2010.01111.x

- 39. Zhu, J. and W.E. Paul: CD4 T cells: fates, functions, and faults. *Blood* 112:1557–69 (2008) DOI: 10.1182/blood-2008-05-078154
- Belkaid, Y. and K. Tarbell: Regulatory T cells in the control of host-microorganism interactions (*) *Annu Rev Immunol* 27:551–89 (2009)
 DOI: 10.1146/annurev.immunol.021908. 132723
- King, C., S.G. Tangye, and C.R. Mackay: T follicular helper (TFH) cells in normal and dysregulated immune responses. *Annu Rev Immunol* 26:741–66 (2008) DOI: 10.1146/annurev.immunol.26.021607. 090344
- Gulhan, B., D. Orhan, G. Kale, N. Besbas, and S. Ozen: Studying cytokines of T helper cells in the kidney disease of IgA vasculitis (Henoch-Schonlein purpura) *Pediatr Nephrol* 30:1269–77 (2015) DOI: 10.1007/s00467-015-3051-4
- 43. Tsuruga, K., S. Watanabe, E. Oki, T. Aizawa-Yashiro, H. Yoshida, T. Imaizumi, E. Ito, and H. Tanaka: Imbalance towards Th1 pathway predominance in purpura nephritis with proteinuria. *Pediatr Nephrol* 26:2253–8 (2011) DOI: 10.1007/s00467-011-1996-5
- 44. Jen, H.Y., Y.H. Chuang, S.C. Lin, B.L. Chiang, and Y.H. Yang: Increased serum interleukin-17 and peripheral Th17 cells in children with acute Henoch-Schonlein purpura. *Pediatr Allergy Immunol* 22:862–8 (2011) DOI: 10.1111/j.1399-3038.2011.01198.x
- 45. Chang, H., Q.Y. Zhang, Y. Lin, N. Cheng, and S.Q. Zhang: Correlation of TLR2 and TLR4 expressions in peripheral blood mononuclear cells to Th1- and Th2-type immune responses in children with henochschonlein purpura. *Int J Clin Exp Med* 8:13532–9 (2015)
- 46. Wan, Y.Y.: GATA3: a master of many trades in immune regulation. *Trends Immunol* 35:233–42 (2014) DOI: 10.1016/j.it.2014.04.002
- Luo, S., G. Liang, P. Zhang, M. Zhao, and Q. Lu: Aberrant histone modifications in peripheral blood mononuclear cells from patients with Henoch-Schonlein purpura. *Clin Immunol* 146:165–75 (2013) DOI: 10.1016/j.clim.2012.12.009

- Chang, H., Y. Cao, Y.I. Lin, H. Zhu, Y. Fu, X. Chen, and Q. Zhang: Association between toll-like receptor 6 expression and auxiliary T cells in the peripheral blood of pediatric patients with allergic purpura. *Exp Ther Med* 10:1536–1540 (2015) DOI: 10.3892/etm.2015.2710
- 49. Hartigan-O'Connor, D.J., L.A. Hirao, J.M. McCune, and S. Dandekar: Th17 cells and regulatory T cells in elite control over HIV and SIV. *Curr Opin HIV AIDS* 6:221–7 (2011) DOI: 10.1097/COH.0b013e32834577b3
- Ivanov, II, B.S. McKenzie, L. Zhou, C.E. Tadokoro, A. Lepelley, J.J. Lafaille, D.J. Cua, and D.R. Littman: The orphan nuclear receptor RORgammat directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell* 126:1121–33 (2006) DOI: 10.1016/j.cell.2006.07.035
- Chen, O., X.B. Zhu, H. Ren, Y.B. Wang, and R. Sun: The imbalance of Th17/Treg in Chinese children with Henoch-Schonlein purpura. *Int Immunopharmacol* 16:67–71 (2013) DOI: 10.1016/j.intimp.2013.03.027
- Bettelli, E., Y. Carrier, W. Gao, T. Korn, T.B. Strom, M. Oukka, H.L. Weiner, and V.K. Kuchroo: Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* 441:235–8 (2006) DOI: 10.1038/nature04753
- Miyara, M., Y. Ito, and S. Sakaguchi: TREGcell therapies for autoimmune rheumatic diseases. *Nat Rev Rheumatol* 10:543–51 (2014) DOI: 10.1038/nrrheum.2014.105
- 54. Chen, W.: Tregs in immunotherapy: opportunities and challenges. *Immunotherapy* 3:911–4 (2011) DOI: 10.2217/imt.11.79
- 55. Dejaco, C., C. Duftner, B. Grubeck-Loebenstein, and M. Schirmer: Imbalance of regulatory T cells in human autoimmune diseases. *Immunology* 117:289–300 (2006) DOI: 10.1111/j.1365-2567.2005.02317.x
- 56. Long, S.A. and J.H. Buckner: CD4+FOXP3+ T regulatory cells in human autoimmunity: more than a numbers game. *J Immunol* 187:2061–6 (2011) DOI: 10.4049/jimmunol.1003224

- Donadio, M.E., E. Loiacono, L. Peruzzi, A. Amore, R. Camilla, F. Chiale, L. Vergano, A. Boido, M. Conrieri, M. Bianciotto, F.M. Bosetti, and R. Coppo: Toll-like receptors, immunoproteasome and regulatory T cells in children with Henoch-Schonlein purpura and primary IgA nephropathy. *Pediatr Nephrol* 29:1545–51 (2014) DOI: 10.1007/s00467-014-2807-6
- Fazilleau, N., L. Mark, L.J. McHeyzer-Williams, and M.G. McHeyzer-Williams: Follicular helper T cells: lineage and location. *Immunity* 30:324–35 (2009) DOI: 10.1016/j.immuni.2009.03.003
- 59. Jogdand, G.M., S. Mohanty, and S. Devadas: Regulators of Tfh Cell Differentiation. *Front Immunol* 7:520 (2016) DOI: 10.3389/fimmu.2016.00520
- Akiba, H., K. Takeda, Y. Kojima, Y. Usui, N. Harada, T. Yamazaki, J. Ma, K. Tezuka, H. Yagita, and K. Okumura: The role of ICOS in the CXCR5+ follicular B helper T cell maintenance *in vivo. J Immunol* 175:2340–8 (2005) DOI: 10.4049/jimmunol.175.4.2340
- Terrier, B., N. Costedoat-Chalumeau, M. Garrido, G. Geri, M. Rosenzwajg, L. Musset, D. Klatzmann, D. Saadoun, and P. Cacoub: Interleukin 21 correlates with T cell and B cell subset alterations in systemic lupus erythematosus. J Rheumatol 39:1819–28 (2012) DOI: 10.3899/irheum.120468
- Bauquet, A.T., H. Jin, A.M. Paterson, M. Mitsdoerffer, I.C. Ho, A.H. Sharpe, and V.K. Kuchroo: The costimulatory molecule ICOS regulates the expression of c-Maf and IL-21 in the development of follicular T helper cells and TH-17 cells. *Nat Immunol* 10:167–75 (2009) DOI: 10.1038/ni.1690
- Vogelzang, A., H.M. McGuire, D. Yu, J. Sprent, C.R. Mackay, and C. King: A fundamental role for interleukin-21 in the generation of T follicular helper cells. *Immunity* 29:127–37 (2008) DOI: 10.1016/j.immuni.2008.06.001
- 64. Chevalier, N., D. Jarrossay, E. Ho, D.T. Avery, C.S. Ma, D. Yu, F. Sallusto, S.G. Tangye, and C.R. Mackay: CXCR5 expressing human central memory CD4 T cells and their relevance for humoral immune responses. *J Immunol* 186:5556–68 (2011) DOI: 10.4049/jimmunol.1002828

- Ma, C.S., D.T. Avery, A. Chan, M. Batten, J. Bustamante, S. Boisson-Dupuis, P.D. Arkwright, A.Y. Kreins, D. Averbuch, D. Engelhard, K. Magdorf, S.S. Kilic, Y. Minegishi, S. Nonoyama, M.A. French, S. Choo, J.M. Smart, J. Peake, M. Wong, P. Gray, M.C. Cook, D.A. Fulcher, J.L. Casanova, E.K. Deenick, and S.G. Tangye: Functional STAT3 deficiency compromises the generation of human T follicular helper cells. *Blood* 119:3997–4008 (2012) DOI: 10.1182/blood-2011-11-392985
- Deenick, E.K., A. Chan, C.S. Ma, D. Gatto, P.L. Schwartzberg, R. Brink, and S.G. Tangye: Follicular helper T cell differentiation requires continuous antigen presentation that is independent of unique B cell signaling. *Immunity* 33:241–53 (2010) DOI: 10.1016/j.immuni.2010.07.015
- 67. Bossaller, L., J. Burger, R. Draeger, B. Grimbacher, R. Knoth, A. Plebani, A. Durandy, U. Baumann, M. Schlesier, A.A. Welcher, H.H. Peter, and K. Warnatz: ICOS deficiency is associated with a severe reduction of CXCR5+CD4 germinal center Th cells. *J Immunol* 177:4927–32 (2006) DOI: 10.4049/jimmunol.177.7.4927
- Ma, J., C. Zhu, B. Ma, J. Tian, S.E. Baidoo, C. Mao, W. Wu, J. Chen, J. Tong, M. Yang, Z. Jiao, H. Xu, L. Lu, and S. Wang: Increased frequency of circulating follicular helper T cells in patients with rheumatoid arthritis. *Clin Dev Immunol* 2012:827480 (2012) DOI: 10.1155/2012/827480
- 69. Liu, R., Q. Wu, D. Su, N. Che, H. Chen, L. Geng, J. Chen, W. Chen, X. Li, and L. Sun: A regulatory effect of IL-21 on T follicular helper-like cell and B cell in rheumatoid arthritis. *Arthritis Res Ther* 14:R255 (2012) DOI: 10.1186/ar4100
- Pallikkuth, S., A. Parmigiani, S.Y. Silva, V.K. George, M. Fischl, R. Pahwa, and S. Pahwa: Impaired peripheral blood T-follicular helper cell function in HIV-infected nonresponders to the 2009 H1N1/09 vaccine. *Blood* 120:985–93 (2012) DOI: 10.1182/blood-2011-12-396648
- Saito, R., H. Onodera, H. Tago, Y. Suzuki, M. Shimizu, Y. Matsumura, T. Kondo, and Y. Itoyama: Altered expression of chemokine receptor CXCR5 on T cells of myasthenia gravis patients. *J Neuroimmunol* 170:172–8 (2005) DOI: 10.1016/j.jneuroim.2005.09.001

- Tackenberg, B., J. Kruth, J.E. Bartholomaeus, K. Schlegel, W.H. Oertel, N. Willcox, B. Hemmer, and N. Sommer: Clonal expansions of CD4+ B helper T cells in autoimmune myasthenia gravis. *Eur J Immunol* 37:849–63 (2007) DOI: 10.1002/eji.200636449
- 73. Wang, J., Y. Shan, Z. Jiang, J. Feng, C. Li, L. Ma, and Y. Jiang: High frequencies of activated B cells and T follicular helper cells are correlated with disease activity in patients with new-onset rheumatoid arthritis. *Clin Exp Immunol* 174:212–20 (2013) DOI: 10.1111/cei.12162
- 74. Xie, J., Y. Liu, L. Wang, G. Ruan, H. Yuan, H. Fang, J. Wu, and D. Cui: Expansion of Circulating T Follicular Helper Cells in Children with Acute Henoch-Schonlein Purpura. *J Immunol Res* 2015:742535 (2015) DOI: 10.1155/2015/742535
- Zhang, Z., S. Zhao, L. Zhang, R. Crew, N. Zhang, X. Sun, and Y. Jiang: A higher frequency of CD4 (+)CXCR5 (+) T follicular helper cells in patients with newly diagnosed Henoch-Schonlein purpura nephritis. *Int Immunopharmacol* 32:8–15 (2016) DOI: 10.1016/j.intimp.2015.12.037
- Zheng, W. and R.A. Flavell: The transcription factor GATA-3 is necessary and sufficient for Th2 cytokine gene expression in CD4 T cells. *Cell* 89:587–96 (1997) DOI: 10.1016/S0092-8674(00)80240-8
- 77. Zhang, D.H., L. Cohn, P. Ray, K. Bottomly, and A. Ray: Transcription factor GATA-3 is differentially expressed in murine Th1 and Th2 cells and controls Th2-specific expression of the interleukin-5 gene. *J Biol Chem* 272:21597–603 (1997) DOI: 10.1074/jbc.272.34.21597
- Ouyang, W., S.H. Ranganath, K. Weindel, D. Bhattacharya, T.L. Murphy, W.C. Sha, and K.M. Murphy: Inhibition of Th1 development mediated by GATA-3 through an IL-4-independent mechanism. *Immunity* 9:745–55 (1998) DOI: 10.1016/S1074-7613(00)80671-8
- 79. Usui, T., J.C. Preiss, Y. Kanno, Z.J. Yao, J.H. Bream, J.J. O'Shea, and W. Strober: T-bet regulates Th1 responses through essential effects on GATA-3 function rather than on

IFNG gene acetylation and transcription. *J Exp Med* 203:755–66 (2006) DOI: 10.1084/jem.20052165

- Siegel, M.D., D.H. Zhang, P. Ray, and A. Ray: Activation of the interleukin-5 promoter by cAMP in murine EL-4 cells requires the GATA-3 and CLE0 elements. *J Biol Chem* 270:24548–55 (1995) DOI: 10.1074/jbc.270.41.24548
- Kishikawa, H., J. Sun, A. Choi, S.C. Miaw, and I.C. Ho: The cell type-specific expression of the murine IL-13 gene is regulated by GATA-3. *J Immunol* 167:4414–20 (2001) DOI: 10.4049/jimmunol.167.8.4414
- Yamashita, M., M. Ukai-Tadenuma, M. Kimura, M. Omori, M. Inami, M. Taniguchi, and T. Nakayama: Identification of a conserved GATA3 response element upstream proximal from the interleukin-13 gene locus. *J Biol Chem* 277:42399–408 (2002) DOI: 10.1074/jbc.M205876200
- Agarwal, S., O. Avni, and A. Rao: Cell-typerestricted binding of the transcription factor NFAT to a distal IL-4 enhancer *in vivo*. *Immunity* 12:643–52 (2000) DOI: 10.1016/S1074-7613(00)80215-0
- Szabo, S.J., S.T. Kim, G.L. Costa, X. Zhang, C.G. Fathman, and L.H. Glimcher: A novel transcription factor, T-bet, directs Th1 lineage commitment. *Cell* 100:655–69 (2000) DOI: 10.1016/S0092-8674(00)80702-3
- Mullen, A.C., A.S. Hutchins, F.A. High, H.W. Lee, K.J. Sykes, L.A. Chodosh, and S.L. Reiner: HIx is induced by and genetically interacts with T-bet to promote heritable T (H)1 gene induction. *Nat Immunol* 3:652–8 (2002) DOI: 10.1038/ni807
- Mullen, A.C., F.A. High, A.S. Hutchins, H.W. Lee, A.V. Villarino, D.M. Livingston, A.L. Kung, N. Cereb, T.P. Yao, S.Y. Yang, and S.L. Reiner: Role of T-bet in commitment of TH1 cells before IL-12-dependent selection. *Science* 292:1907–10 (2001)
- Pearce, E.L., A.C. Mullen, G.A. Martins, C.M. Krawczyk, A.S. Hutchins, V.P. Zediak, M. Banica, C.B. DiCioccio, D.A. Gross, C.A. Mao, H. Shen, N. Cereb, S.Y. Yang, T. Lindsten, J. Rossant, C.A. Hunter, and

S.L. Reiner: Control of effector CD8+ T cell function by the transcription factor Eomesodermin. *Science* 302:1041–3 (2003) DOI: 10.1126/science.1090148

- Suto, A., A.L. Wurster, S.L. Reiner, and M.J. Grusby: IL-21 inhibits IFN-gamma production in developing Th1 cells through the repression of Eomesodermin expression. *J Immunol* 177:3721–7 (2006) DOI: 10.4049/jimmunol.177.6.3721
- Fontenot, J.D., M.A. Gavin, and A.Y. Rudensky: Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol* 4:330–6 (2003) DOI: 10.1038/ni904
- Hori, S., T. Nomura, and S. Sakaguchi: Control of regulatory T cell development by the transcription factor Foxp3. *Science* 299:1057–61 (2003) DOI: 10.1126/science.1079490
- 91. Williams, L.M. and A.Y. Rudensky: Maintenance of the Foxp3-dependent developmental program in mature regulatory T cells requires continued expression of Foxp3. *Nat Immunol* 8:277–84 (2007) DOI: 10.1038/ni1437
- 92. Wan, Y.Y. and R.A. Flavell: Regulatory T-cell functions are subverted and converted owing to attenuated Foxp3 expression. *Nature* 445:766–70 (2007) DOI: 10.1038/nature05479
- Park, H., Z. Li, X.O. Yang, S.H. Chang, R. Nurieva, Y.H. Wang, Y. Wang, L. Hood, Z. Zhu, Q. Tian, and C. Dong: A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol* 6:1133–41 (2005) DOI: 10.1038/ni1261
- 94. Harrington, L.E., R.D. Hatton, P.R. Mangan, H. Turner, T.L. Murphy, K.M. Murphy, and C.T. Weaver: Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 6:1123–32 (2005) DOI: 10.1038/ni1254
- Yang, X.O., B.P. Pappu, R. Nurieva, A. Akimzhanov, H.S. Kang, Y. Chung, L. Ma, B. Shah, A.D. Panopoulos, K.S. Schluns, S.S. Watowich, Q. Tian, A.M. Jetten, and C. Dong: T helper 17 lineage differentiation is programmed by orphan nuclear receptors

ROR alpha and ROR gamma. Immunity 28:29-39 (2008) DOI: 10.1016/j.immuni.2007.11.016

- 96. Dent. A.L., A.L. Shaffer, X. Yu. D. Allman. and L.M. Staudt: Control of inflammation, cytokine expression, and germinal center formation by BCL-6. Science 276:589-92 (1997)DOI: 10.1126/science.276.5312.589
- 97. Chtanova, T., S.G. Tangye, R. Newton, N. Frank, M.R. Hodge, M.S. Rolph, and C.R. Mackay: T follicular helper cells express a distinctive transcriptional profile, reflecting their role as non-Th1/Th2 effector cells that provide help for B cells. J Immunol 173:68-78 (2004) DOI: 10.4049/jimmunol.173.1.68
- 98. Nurieva, R.I., Y. Chung, G.J. Martinez, X.O. Yang, S. Tanaka, T.D. Matskevitch, Y.H. Wang, and C. Dong: Bcl6 mediates the development of T follicular helper cells. Science 325:1001-5 (2009) DOI: 10.1126/science.1176676
- 99. Yu, D., S. Rao, L.M. Tsai, S.K. Lee, Y. He, E.L. Sutcliffe, M. Srivastava, M. Linterman, L. Zheng, N. Simpson, J.I. Ellyard, I.A. Parish, C.S. Ma, Q.J. Li, C.R. Parish, C.R. Mackay, and C.G. Vinuesa: The transcriptional repressor Bcl-6 directs T follicular helper cell lineage commitment. Immunity 31:457-68 (2009)

DOI: 10.1016/j.immuni.2009.07.002

- 100. Afkarian, M., J.R. Sedy, J. Yang, N.G. Jacobson, N. Cereb, S.Y. Yang, T.L. Murphy, and K.M. Murphy: T-bet is a STAT1-induced regulator of IL-12R expression in naive CD4+ T cells. Nat Immunol 3:549–57 (2002) DOI: 10.1038/ni794
- 101. Lighvani, A.A., D.M. Frucht, D. Jankovic, H. Yamane, J. Aliberti, B.D. Hissong, B.V. Nguyen, M. Gadina, A. Sher, W.E. Paul, and J.J. O'Shea: T-bet is rapidly induced by interferon-gamma in lymphoid and myeloid cells. Proc Natl Acad Sci USA 98:15137-42 (2001) DOI: 10.1073/pnas.261570598
- 102. Veldhoen, M., R.J. Hocking, C.J. Atkins, R.M. Locksley, and B. Stockinger: TGFbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. Immunity 24:179-89 (2006)DOI: 10.1016/j.immuni.2006.01.001

- 103. Nurieva, R., X.O. Yang, G. Martinez, Y. Zhang, A.D. Panopoulos, L. Ma, K. Schluns, Q. Tian, S.S. Watowich, A.M. Jetten, and C. Dong: Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature 448:480-3 (2007) DOI: 10.1038/nature05969
- 104. Guo, L., G. Wei, J. Zhu, W. Liao, W.J. Leonard, K. Zhao, and W. Paul: IL-1 family members and STAT activators induce cytokine production by Th2, Th17, and Th1 cells. Proc Natl Acad Sci U S A 106:13463–8 (2009) DOI: 10.1073/pnas.0906988106
- 105. Mangan, P.R., L.E. Harrington, D.B. O'Quinn, W.S. Helms, D.C. Bullard, C.O. Elson, R.D. Hatton, S.M. Wahl, T.R. Schoeb, and C.T. Weaver: Transforming growth factor-beta induces development of the T (H)17 lineage. Nature 441:231-4 (2006) DOI: 10.1038/nature04754
- 106. Chen, Z., A. Laurence, Y. Kanno, M. Pacher-Zavisin, B.M. Zhu, C. Tato, A. Yoshimura, L. Hennighausen, and J.J. O'Shea: Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells. Proc Natl Acad Sci USA 103:8137–42 (2006) DOI: 10.1073/pnas.0600666103
- 107. Wei, L., A. Laurence, K.M. Elias, and J.J. O'Shea: IL-21 is produced by Th17 cells and drives IL-17 production in a STAT3dependent manner. J Biol Chem 282:34605-10 (2007) DOI: 10.1074/jbc.M705100200
- 108. Zhou, L., Ivanov, II, R. Spolski, R. Min, K. Shenderov, T. Egawa, D.E. Levy, W.J. Leonard, and D.R. Littman: IL-6 programs T (H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat Immunol 8:967–74 (2007) DOI: 10.1038/ni1488
- 109. Yang, X.O., A.D. Panopoulos, R. Nurieva, S.H. Chang, D. Wang, S.S. Watowich, and C. Dong: STAT3 regulates cytokinemediated generation of inflammatory helper T cells. J Biol Chem 282:9358-63 (2007) DOI: 10.1074/jbc.C600321200
- 110. Cote-Sierra, J., G. Foucras, L. Guo, L. Chiodetti, H.A. Young, J. Hu-Li, J. Zhu, and W.E. Paul: Interleukin 2 plays a central role in Th2 differentiation. Proc Natl Acad Sci U SA 101:3880-5 (2004) DOI: 10.1073/pnas.0400339101

- 111. Zhu, J., J. Cote-Sierra, L. Guo, and W.E. Paul: Stat5 activation plays a critical role in Th2 differentiation. Immunity 19:739-48 (2003)DOI: 10.1016/S1074-7613(03)00292-9
- 112. Davidson, T.S., R.J. DiPaolo, J. Andersson, and E.M. Shevach: Cutting Edge: IL-2 is essential for TGF-beta-mediated induction of Foxp3+ T regulatory cells. J Immunol 178:4022-6 (2007) DOI: 10.4049/jimmunol.178.7.4022
- 113. Burchill, M.A., J. Yang, K.B. Vang, J.J. Moon, H.H. Chu, C.W. Lio, A.L. Vegoe, C.S. Hsieh, M.K. Jenkins, and M.A. Farrar: Linked T cell receptor and cytokine signaling govern the development of the regulatory T cell repertoire. Immunity 28:112-21 (2008) DOI: 10.1016/j.immuni.2007.11.022
- 114. Burchill, M.A., J. Yang, C. Vogtenhuber, B.R. Blazar, and M.A. Farrar: IL-2 receptor betadependent STAT5 activation is required for the development of Foxp3+ regulatory T cells. J Immunol 178:280–90 (2007) DOI: 10.4049/jimmunol.178.1.280
- 115. Yao, Z., Y. Kanno, M. Kerenvi, G. Stephens, L. Durant, W.T. Watford, A. Laurence, G.W. Robinson, E.M. Shevach, R. Moriggl, L. Hennighausen, C. Wu, and J.J. O'Shea: Nonredundant roles for Stat5a/b in directly regulating Foxp3. Blood 109:4368-75 (2007) DOI: 10.1182/blood-2006-11-055756
- 116. Scheeren, F.A., M. Naspetti, S. Diehl, R. Schotte, M. Nagasawa, E. Wijnands, R. Gimeno, F.A. Vyth-Dreese, B. Blom, and H. Spits: STAT5 regulates the selfrenewal capacity and differentiation of human memory B cells and controls Bcl-6 expression. Nat Immunol 6:303-13 (2005) DOI: 10.1038/ni1172
- 117. Kaplan, M.H., U. Schindler, S.T. Smiley, and M.J. Grusby: Stat6 is required for mediating responses to IL-4 and for development of Th2 cells. Immunity 4:313–9 (1996) DOI: 10.1016/S1074-7613(00)80439-2
- 118. Shimoda, K., J. van Deursen, M.Y. Sangster, S.R. Sarawar, R.T. Carson, R.A. Tripp, C. Chu, F.W. Quelle, T. Nosaka, D.A. Vignali, P.C. Doherty, G. Grosveld, W.E. Paul, and J.N. Ihle: Lack of IL-4-induced Th2 response and IgE class switching in mice with disrupted Stat6 gene. Nature 380:630–3 (1996) DOI: 10.1038/380630a0

- 119. Takeda, K., T. Tanaka, W. Shi, M. Matsumoto, M. Minami, S. Kashiwamura, K. Nakanishi, N. Yoshida, T. Kishimoto, and S. Akira: Essential role of Stat6 in IL-4 signalling. Nature 380:627–30 (1996) DOI: 10.1038/380627a0
- 120. Kurata, H., H.J. Lee, A. O'Garra, and N. Arai: Ectopic expression of activated Stat6 induces the expression of Th2-specific cytokines and transcription factors in developing Th1 cells. Immunity 11:677-88 (1999)DOI: 10.1016/S1074-7613(00)80142-9
- 121. Zhu, J., L. Guo, C.J. Watson, J. Hu-Li, and W.E. Paul: Stat6 is necessary and sufficient for IL-4's role in Th2 differentiation and cell expansion. J Immunol 166:7276-81 (2001) DOI: 10.4049/jimmunol.166.12.7276
- 122. Egawa, T., R.E. Tillman, Y. Naoe, I. Taniuchi, and D.R. Littman: The role of the Runx transcription factors in thymocyte differentiation and in homeostasis of naive T cells. J Exp Med 204:1945-57 (2007) DOI: 10.1084/jem.20070133
- 123. Zhang, F., G. Meng, and W. Strober: Interactions among the transcription factors Runx1, RORgammat and Foxp3 regulate the differentiation of interleukin 17-producing T cells. Nat Immunol 9:1297-306 (2008) DOI: 10.1038/ni.1663
- 124. Klunker, S., M.M. Chong, P.Y. Mantel, O. Palomares, C. Bassin, M. Ziegler, B. Ruckert, F. Meiler, M. Akdis, D.R. Littman, and C.A. Akdis: Transcription factors RUNX1 and RUNX3 in the induction and suppressive function of Foxp3+ inducible regulatory T cells. J Exp Med 206:2701-15 (2009) DOI: 10.1084/jem.20090596
- 125. Rudra, D., T. Egawa, M.M. Chong, P. Treuting, D.R. Littman, and A.Y. Rudensky: Runx-CBFbeta complexes control expression of the transcription factor Foxp3 in regulatory T cells. Nat Immunol 10:1170-7 (2009) DOI: 10.1038/ni.1795
- 126. Lohoff, M., H.W. Mittrucker, S. Prechtl, S. Bischof, F. Sommer, S. Kock, D.A. Ferrick, G.S. Duncan, A. Gessner, and T.W. Mak: Dysregulated T helper cell differentiation in the absence of interferon regulatory factor Proc Natl Acad Sci U S A 99:11808-12 (2002)

DOI: 10.1073/pnas.182425099

- 127. Kano, S., K. Sato, Y. Morishita, S. Vollstedt, S. Kim, K. Bishop, K. Honda, M. Kubo, and T. Taniguchi: The contribution of transcription factor IRF1 to the interferon-gammainterleukin 12 signaling axis and TH1 versus TH-17 differentiation of CD4+ T cells. *Nat Immunol* 9:34–41 (2008) DOI: 10.1038/ni1538
- 128. Zhu, J., D. Jankovic, A. Grinberg, L. Guo, and W.E. Paul: Gfi-1 plays an important role in IL-2-mediated Th2 cell expansion. *Proc Natl Acad Sci U S A* 103:18214–9 (2006) DOI: 10.1073/pnas.0608981103
- 129. Zhu, J., T.S. Davidson, G. Wei, D. Jankovic, K. Cui, D.E. Schones, L. Guo, K. Zhao, E.M. Shevach, and W.E. Paul: Downregulation of Gfi-1 expression by TGF-beta is important for differentiation of Th17 and CD103+ inducible regulatory T cells. *J Exp Med* 206:329–41 (2009) DOI: 10.1084/jem.20081666
- 130. Quirion, M.R., G.D. Gregory, S.E. Umetsu, S. Winandy, and M.A. Brown: Cutting edge: Ikaros is a regulator of Th2 cell differentiation. *J Immunol* 182:741–5 (2009) DOI: 10.4049/jimmunol.182.2.741
- 131. Kim, J.I., I.C. Ho, M.J. Grusby, and L.H. Glimcher: The transcription factor c-Maf controls the production of interleukin-4 but not other Th2 cytokines. *Immunity* 10:745–51 (1999)
 DOI: 10.1016/S1074-7613(00)80073-4
- 132. Grenningloh, R., B.Y. Kang, and I.C. Ho: Ets-1, a functional cofactor of T-bet, is essential for Th1 inflammatory responses. *J Exp Med* 201:615–26 (2005) DOI: 10.1084/jem.20041330
- 133. Moisan, J., R. Grenningloh, E. Bettelli, M. Oukka, and I.C. Ho: Ets-1 is a negative regulator of Th17 differentiation. *J Exp Med* 204:2825–35 (2007) DOI: 10.1084/jem.20070994
- 134. Cimmino, L., G.A. Martins, J. Liao, E. Magnusdottir, G. Grunig, R.K. Perez, and K.L. Calame: Blimp-1 attenuates Th1 differentiation by repression of ifng, tbx21, and bcl6 gene expression. *J Immunol* 181:2338–47 (2008) DOI: 10.4049/jimmunol.181.4.2338
- 135. Wang, L., N. van Panhuys, J. Hu-Li, S. Kim, G. Le Gros, and B. Min: Blimp-1 induced

by IL-4 plays a critical role in suppressing IL-2 production in activated CD4 T cells. *J Immunol* 181:5249–56 (2008) DOI: 10.4049/jimmunol.181.8.5249

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Abbreviations: HSP, Henoch-Schonlein purpura; HSPN, Henoch-Schonlein purpura nephritis; Th, T helper; Treg cells, regulatory T cells; Tfh, t follicular helper; C3, complement component 3; GN, glomerulonephritis; IFN- γ , interferongamma; TGF- β , transforming growth factor beta; PBMCs, peripheral blood mononuclear cells; ROR γ , retinoic acid receptor-related orphan receptors gamma; GC, germinal center; PD-1, programmed death-1; STAT, signaling transducer and activator of transcription.

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