Leishmaniasis treatment: update of possibilities for drug repurposing

Valter Viana Andrade-Neto¹, Edezio Ferreira Cunha-Junior¹, Viviane dos Santos Faioes¹, Thais Pereira Martins¹, Raphaela Lopes Silva¹, Leonor Laura Leon¹, Eduardo Caio Torres-Santos¹

¹Laboratorio de Bioquimica de Tripanosomatideos, Instituto Oswaldo Cruz, Fiocruz, Rio de Janeiro, Brasil

TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. Leishmaniases
 - 3.1. Clinical manifestations
 - 3.2. Treatment
- 4. Drug repurposing strategies
 - 4.1. New drugs repositioning candidates
 - 4.1.1. Antifungals
 - 4.1.2. Antiparasitic drugs
 - 4.1.3. Anticancer drugs
 - 4.1.4. Antidepressant drugs
 - 4.1.5. Antihypertensive drugs
 - 4.1.6. Antibiotics
 - 4.1.7. Other drugs
- 5. Clinical trials
- 6. Conclusion
- 7. Acknowledgement
- 8. References

1. ABSTRACT

The leishmaniases represent a public health problem in under-developed countries and are considered a neglected disease by the World Health Organization (WHO). They are cuased by Leishmania parasites with different clinical manifestations. Currently, there is no vaccine, and treatment is inefficient and is associated with both serious side effects often leading to resistance to the parasites. Thus, it is essential to search for new treatment strategies, such as drug repurposing, i.e., the use of drugs that are already used for other diseases. The discovery of new clinical applications for approved drugs is strategic for lowering the cost of drug discovery since human toxicity assays are already conducted. Here, we review a broad analysis of the different aspects of this approach for anti-leishmanial treatment.

2. INTRODUCTION

Drug repurposing (or repositioning) is a strategy within the drug development discovery process employed to find new uses for existing

drugs. This process has gained momentum in the past several years, mainly due to an increase in popularity and the influence of the pharmaceutical industry (1,2). Drug repositioning offers a possibility to bypass a dilemma faced by Big Pharma in the drug discovery process. This dilemma involves a productivity gap (discovery of a new chemical entity -NCE), despite the increasing research and investment in of new technologies, such as high-throughput screening (HTS), high-content screening (HCS), and combinatorial chemistry. Often, drug repurposing reduces the risk associated with development, which multiplies throughout the various stages of preclinical and clinical development, because pharmacokinetic and safety profiles have been already determined (3,4) (Figure 1). In recent years, drug repurposing was responsible for 30% of the new drugs and vaccines approved by the Food and Drug Administration (5). Interestingly, drug repurposing may extend the patent period of a drug from three to five years (6). The cost savings of repositioning an approved drug makes it an attractive strategy for neglected tropical diseases

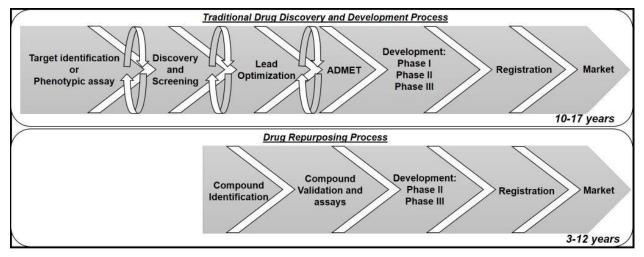


Figure 1. Traditional Drug Discovery and Development process x Drug Repurposing Process.

because of the limited research and development resources in this area. Thus, there have been a number of successful repurposing cases, mainly for the leishmaniases (e.g., pentamidine, amphotericin B, miltefosine and paromomycin were repositioned from other indications) (Figure 2) (7). In this review, different studies of the effectiveness of FDA-approved drugs on leishmaniases will be discussed.

3. LEISHMANIASES

The leishmaniases are a parasitic infectious disease caused by different species (20 species are considered pathogenic for humans) of the genus Leishmania and transmitted by Phlebotomus and Lutzomyia sandflies. The insect vector capable of transmitting the disease to different mammals (including humans) is Phlebotomus in the Old World (Europe, Asia and Africa) and Lutzomvia in the New World (Americas); in all, there are 30 species capable of transmitting the disease (8,9). The disease affects approximately 300 million people, causes 20 to 30 thousand deaths annually and occurs in 98 countries. Although the leishmaniases have very large geographical coverage. 7 countries account for 90% of the new cases: Bangladesh, Brazil, India, Ethiopia, Kenya, Nepal and Sudan. The number of new cases of the disease is uncertain: however, it is possible estimate an increase of 200,000-400,000 new cases of visceral leishmaniasis (VL) and 700,000-1,200,000 new cases of cutaneous leishmaniasis (10).

3.1. Clinical manifestations

The clinical forms of the disease, i.e., cutaneous, diffuse, disseminated, mucocutaneous and visceral, are categorized by parasite species and the immunological response of patients. *Localized* cutaneous *leishmaniasis* is the most common

form found in patients with tegumentary leishmaniasis and is characterized by one or more erythematous papules in the skin (11,12). Diffuse cutaneous leishmaniasis, also called anergic, presents multiple cutaneous lesions without a tendency to form ulcers and with typical lepromatous lesions. With chronic infections, the lesions have abundant parasites and persist for 20 years or more. Disseminated cutaneous leishmaniasis is characterized by multiple ulcerative or acneiform skin lesions and by a strongly positive leishmanin skin test (13-15). Mucocutaneous leishmaniasis is more common in the New World, with almost no cases found in the Old World. This clinical form usually occurs after the cutaneous manifestation with destruction of the mucosa and can progress until the destruction of cartilage, leading to the patient's respiratory compromise (16,17). Visceral leishmaniasis, also known as kala-azar, is present both in the New World and in the Old World. This is the most serious form of the disease and can lead to death. There are patients who are asymptomatic, but in general, the symptoms are fever, anorexia, diarrhea, hepatosplenomegaly, enlarged lymph nodes and vascular problems (11). The post-kala-azar form, which occurs only in the Old World, is a complication of the treatment of visceral leishmaniasis. It is characterized by the presence of small nodules on the face and body without pigmentation (18). Leishmania species that are pathogenic to humans and their clinical manifestations are listed in Table 1, according to Sereno and collaborators (8).

3.2. Treatment

Currently, the first line of drugs used in the treatment of the leishmaniases is the pentavalent antimonials. In 1912, the physician Gaspar Vianna, after observing the effect of trivalent antimony (tartar emetic) on certain trypanosomes, decided

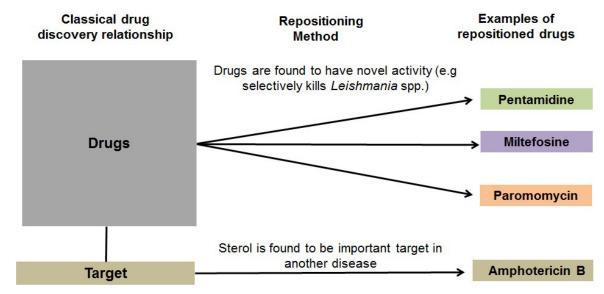


Figure 2. Examples of repositioned drugs for Leishmaniases.

Table 1. Species pathogenic to human, occurrence and clinical manifestation

Complex	Species	Ocurrence	Clinical manifestation
Viannia	L. shawi	NW ¹ / OW ²	CL ³
	L. peruviana	NW¹	CL³, MCL⁴
	L. panamensis	NW¹	CL³, MCL⁴
	L. lindenbergi	NW¹	CL ³
	L. naiffi	NW¹	CL ³
	L. braziliensis	NW¹	CL³, MCL⁴
	L. guyanensis	NW¹	CL³, MCL⁴
	L. lainsoni	NW¹	CL ³
Leishmania	L. venezuelensis	NW¹	CL ³
	L. tropica	OW ²	CL³, VL⁵
	L. mexicana	NW¹	CL ³ , DCL ⁶
	L. major	OW ²	CL ³
	L. infantum	NW ¹ / OW ²	VL⁵, CL³
	L. donovani	OW ²	VL⁵, PKDL ⁷
	L. amazonensis	NW¹	CL³, DCL ⁶ , MCL⁴, VL
	L. aethiopica	OW ²	CL ³ , DCL ⁶
Enrietti	L. martiniquensis	NW¹/ OW²	CL³, VL⁵
	L. siamensis	NW¹ / OW²	VL ⁵ , CL ³
Paraleishmania	L. colombiensis	NW¹	CL³, VL⁵

¹New World, ²Old World, ³Cutaneous leishmaniasis, ⁴Mucocutaneous leishmaniasis, ⁵Visceral leishmaniases, ⁶Difuse cutaneous leishmaniasis, ⁷De 14 de la contraction de la contracti

to reposition this compound for the treatment of cutaneous leishmaniasis cases diagnosed in São Paulo, Brazil (19). The development of pentavalent antimonials enabled a reduction in treatment toxicity (20). The formulations of the pentavalent antimonials commercially available, sodium stibogluconate (Pentostam®) and N-methyl glucamine antimoniate (Glucantime®) (Figure 3), have been used for the treatment of tegumentary leishmaniasis and visceral

leishmaniasis (21). The pentavalent antimonials are registered and licensed in Southeast Asia, Latin America, and some Mediterranean and African countries (22).

In addition, other drugs considered as a second choice have been used, such as amphotericin B, pentamidine, miltefosine and paromomycin (23) (Figure 3). Amphotericin B, an antifungal, has been

⁷Post-kala-azar dermal leishmaniasis

Figure 3. Approved drugs for Leishmaniases.

used as an alternative treatment for leishmaniases but has serious side effects. The liposomal formulation of amphotericin B made the drug less toxic and more effective, despite the high cost, and is the most used formulation for the treatment of visceral leishmaniasis (24). This formulation is licensed to use in India, USA, and Europe for treatment of the visceral form and is used as a second-line drug for the treatment of VL in East Africa and Brazil (22).

Pentamidine, a synthetic derivative of diamidine discovered while searching for hypoglycemic drugs, is used in cases of antimonial resistance as an alternative way of treating the leishmaniases; however, its side effects, which include cardiotoxic and metabolic disorders (diabetes) and cases of poor efficacy and resistance, have made this drug a poor option for the treatment of cutaneous leishmaniasis (25).

The only drug given orally today, miltefosine, was initially developed as an anticancer drug and was used in the treatment of visceral leishmaniasis in India, with patient cure rates of approximately 94%. However, after a decade of use in the country, several cases of recurrence and decreased efficacy of treatment have been described. Despite the advantage of being administered orally, the use of miltefosine is also linked to many side effects that are primarily related to the gastrointestinal tract and occasional hepatic toxicity and nephrotoxicity, in addition to its teratogenic potential (26).

Paromomycin is a broad-spectrum aminoglycoside antibiotic first used in the treatment of leishmaniases in 2002 as paromomycin sulfate. It is used in the treatment of the two forms of the disease and is administered parenterally for visceral leishmaniasis and parenterally and topically for cutaneous leishmaniasis. There are several advantages associated with the use of paromomycin. such as low cost, better efficacy, and fewer side effects. However, there is a likelihood of acquiring resistance with paromomycin monotherapy. Although not described in the clinic, there are reports of in vitro resistance in L. donovani and L. tropica (21). In addition to these drugs commonly used for the treatment of leishmaniases, the World Health Organization also recommends the use of some antifungal azoles as a second choice for the treatment of the disease, but only in specific cases of cutaneous form. For example. the use of fluconazole is recommended for systemic oral therapy of L. major infections in the Old World, while ketoconazole is recommended for the systemic treatment of L. mexicana infections in adults (27).

4. DRUG REPURPOSING STRATEGIES

Research into new uses for existing drugs is supported by two fundamental scientific concepts: finding new uses of a drug acting through the originally known target ("on-target") or finding new uses of a drug acting through a novel or unanticipated target ("off-target") (1, 5, 28). Target-based screening and

phenotypic screening are the two main approaches used in drug discovery, development and repositioning. In fact, the contribution of phenotypic screening to drug discovery has been higher than that of target-based screening (29). Recent partnerships between industry and academia through, for example, high-throughput studies with drug libraries, have brought about a range of advantages and a hope for neglected diseases because the support from organizations such as the Drug for Neglected Diseases initiative (DNDi), Big Pharma and open access chemical libraries (30-32). HTS campaigns, such as the one published by Peña and coworkers (1.8 million compounds), generate extensive information about compounds of different chemical classes with new biological activities against L. donovani, Trypanosoma cruzi and Trypanosoma brucei; specifically, approximately 200 compounds with activity against each parasite were selected based on the criteria of potency and selectivity (33). However. following this strategy with "premature repositioning" bias, Kaiser and coworkers evaluated 200 druglike and 200 probe-like compounds from the malaria box library against L. donovani, L. infantum, T. cruzi and T. brucei, identifying 55 hits for Human African trypanosomiasis, 21 hits for Chagas disease and 8 hits for VL (34), Recently, Kaiser and colleagues evaluated 100 FDA-approved drugs using the repositioning strategy; however, the author reports that the low return on promising drug repurposing options is related to the strategy of repositioning; i.e., it is low risk and high return (35). Despite the recent high-throughput studies on the discovery of drugs for the leishmaniases and other diseases caused by trypanosomatids, the majority of studies on drug repositioning for the leishmaniases have been conducted with classic in vitro (promastigote, axenic amastigote and intracellular amastigote) and in vivo assays. In this work, a review of studies on the repositioning possibilities of FDAapproved drugs is presented.

4.1. New drug repositioning candidates

4.1.1. Antifungals

Antifungal drugs have been widely used in antileishmanial activity assays, achieving substantial success in experimental animal models (Figure 4). The first class of antifungals described is the polyenic antibiotics. Amphotericin B, a drug belonging to this class, is licensed for the treatment of the leishmaniases. Another polyene drug is nystatin, whose antileishmanial activity was first reported in 1962 against L. donovani. Nystatin demonstrated antipromastigote activity against L. major with an IC $_{50}$ value of 9.76 µg/mL, inhibiting the entry of promastigotes into macrophages due to the sequestration of macrophage cholesterol and thus indicating the essential role of cholesterol in mediating the entry of the parasite (36–38).

The current treatments of the leishmaniases advocated by the WHO includes the use of some azole drugs as a second choice in the treatment of cutaneous leishmaniasis. However, there are many studies in which several azoles have been developed to treat experimental leishmaniases. Itraconazole and posaconazole were effective against promastigotes and amastigotes of L. amazonensis, with IC₅₀ values lower than 3 µM (39). Clotrimazole showed antileishmanial activity against promastigotes of L. infantum. An organometallic association between ruthenium and clotrimazole resulted in an increase in the antileishmanial activity in vitro compared to clotrimazole alone and demonstrated significant efficacy in in vivo treatment in the murine model of cutaneous leishmaniasis (40,41). Another azole drug, bifonazole, showed antileishmanial activity in L. infantum promastigotes, with an IC_{50} value of 8.97 uM. Econazole also showed antileishmanial activity in promastigotes and intracellular amastigotes of L. infantum, and voriconazole showed activity against different species: L. donovani, L. amazonensis and L. major (42,43). The antileishmanial activity of azoles can be influenced by the availability of an exogenous cholesterol source to the parasite. Parasites of L. amazonensis treated with ketoconazole and miconazole had decreased ergosterol synthesis accompanied by the accumulation of exogenous cholesterol, compensating for the low levels of endogenous sterols. When the parasites were incubated with delipidated fetal bovine serum, i.e., no availability of exogenous cholesterol, there was a 50% reduction in the ${\rm IC}_{\rm 50}$ values of ketoconazole and miconazole compared to the IC_{50} values of the parasites incubated with complete fetal bovine serum, indicating that this compensatory mechanism of exogenous cholesterol uptake may be a good pharmacological target (44).

Another class of antifungals, the allylamines, is composed of synthetic naphthalene derivatives. Two drugs of this class have been investigated for their antileishmanial activity: butenafine and terbinafine. A recent study showed that butenafine was able to eliminate promastigotes of L. amazonensis and L. braziliensis with the same efficacy as the reference drug, miltefosine (45). Many studies have also shown the antileishmanial activity of terbinafine. One of the early studies showed that the combination of terbinafine and another antifungal, ketoconazole, produced additive effects on the axenic growth of promastigotes and synergistic effects on the proliferation of intracellular amastigotes of L. amazonensis. When treated with terbinafine, a strain of L. braziliensis resistant to ketoconazole became more sensitive to azoles since terbinafine interfered in the synthesis of membrane sterols and treatment led to loss of cell viability. Treatment with terbinafine resulted in a decrease in the proliferation of *L. major* promastigotes, but terbinafine

Figure 4. Antifungal drugs

did not induce any inhibitory effect on the proliferation of *L. donovani* or *L. mexicana* promastigotes (46,47).

A controversial result was observed in experimental models of cutaneous leishmaniasis, the treatment with terbinafine of *L. major*-innfected BALB/c mice was effective in reducing lesions, while in the model *L. amazonensis*-infected C57BI/6 mice, this drug led to no significant reduction of both lesion size or parasite load when compared to the untreated control group (48,49). In the *in vivo* models of visceral infection, treatment of hamsters infected with *L. infantum chagasi* with terbinafine did not cause a difference in weight or spleen parasite load compared to the untreated control, with a significant decrease only observed when the drug was associated with glucantime (50,51).

Studies using other antifungals have been performed; for example, caspofungin, a cyclic lipo-hexapeptide, has been reported to be active against promastigotes of *L. tropica* (52). Amorolfine, which belongs to the morpholine class and is an inhibitor of delta-14-reductase and delta-8-delta-7-isomerase, was tested on intracellular amastigotes of *L. donovani*

and demonstrated an IC $_{50}$ lower than the standard antileishmanial drugs sodium stibogluconate and meglumine antimoniate (53). Ciclopirox olamine, a hydroxypyridone and an inhibitor of the enzyme deoxyhypusine hydroxylase, was tested on L. donovani, exhibiting an IC $_{50}$ value of 2.14 μ M in promastigotes and an IC $_{50}$ value of 1.09 μ M in intracellular amastigotes (35).

4.1.2. Antiparasitic drugs

Several antiparasitic drugs have been repositioned for leishmaniases treatment (Figure 5), including antimalarial drugs. Artemisinin is a secondary metabolite of *Artemisia annua* and is considered as one of the reference drug in the treatment of malaria, and some studies showed its efficacy in experimental visceral leishmaniasis caused by *L. donovani*. Macrophages infected with *Leishmania* spp. amastigotes and treated with artemisinin had a lower production of nitrite and nitric oxide synthase mRNA, promoting a greater protection of the host cell (54–57).

Another antimalarial, the aminoquinoline chloroquine, has been studied in a CBA mice model

Figure 5. Antiparasitic drugs

infected with *L. amazonensis*. Treatment with chloroquine reduced the size of the lesions and significantly decrease parasitism, showing that it is a promising drug for the treatment of leishmaniases (58). The paper published by Khan *et al.* in 2007 shows that chloroquine analogues are also effective on *L. donovani* promastigotes, including resistant strains (59). Mefloquine, another aminoquinoline, has activity against promastigotes with an IC $_{50}$ of 8.4 μ M. However, the use of mefloquine in mice and in the clinic studies does not provide good results, such as non-reduction of the lesions and no effects compared to that of the placebo groups (60–63).

Brazil and Gilbert, in 1976, showed the activity of oxamniquine, used for *Schistosoma mansoni* infections, in hamsters infected with *L. braziliensis* using only 5 doses, suggesting that further studies should be conducted to investigate the action of oxamniquine against *Leishmania* spp.(64).

Ivermectin is a broad-spectrum macrolide used in the treatment of onchocerciasis, strongy-

loidiasis, ascaridiasis and other diseases caused by worms. In addition, its effectiveness against the promastigote forms of *Leishmania* spp. has already been demonstrated. Studies show that its analogues have specific activity against promastigote and amastigote forms of the parasite and specifically do not affect enzymes in the host (65–67).

Metronidazole is derived from nitroimidazoles and has activity against bacterial infections, such as giardiasis, trichomoniasis and amebiasis. Some studies on the treatment of experimental cutaneous leishmaniasis have demonstrated a significant reduction in lesion size and in the number of parasites via histological examination. It is also a drug without *major* side effects and with easy administration and low cost, which reinforces the idea of using this drug in the treatment of cutaneous leishmaniasis (68–70).

Nitazoxanide is a nitrothiazole that has efficacy in the treatment of giardia and cryptosporidium, and some papers have shown its leishmanicidal activity in the promastigote and amastigote forms of *L. infantum*

Figure 6. Anticancer drugs

and *L. donovani*. In addition, BALB/c mice infected with *L. donovani* present a significant reduction in liver and spleen sizes in relation to the control group (71,72). In 2013, Mesquita *et al.* suggested that the use of nitazoxanide together with other drugs, such as amphotericin B, miltefosine and glucantime, should be considered as an alternative to the treatment of leishmaniases (73).

Imidocarb (used in the veterinary medicine) approved to treat anaplasmosis and babesiosis was effective alone or combined with levamisole (antihelmintic drug) decrease lesions and parasite loads of *L. amazonensis*-infected BALB/c mice (74).

4.1.3. Anticancer drugs

Antitumorals (Figure 6) also have previously demonstrated the ability to eliminate parasites of the genus *Leishmania*, and the leishmanicidal activity of these drugs will be discussed.

Hydroxyurea (a carbamyl hydroxamate) is used in treatment of myeloid leukemia and melanoma and displays leishmanicidal activity, leading to the death

of *L. mexicana in vitro* and inducing alterations in cell cycle by arresting the G₂/M phase in promastigotes. This drug is given orally and has advantages for a possible repositioning for the treatment of leishmaniasis (75).

Cisplatin is used in the treatment of ovarian and testicular cancer and has showed excellent leishmanicidal activity *in vitro* and *in vivo*. This antineoplastic drug induced alterations in the cell cycle in promastigotes and amastigotes with arrest of the S and G2 phases and mitochondrial membrane potential loss. In promastigotes, it also increases the levels of thiols and reactive oxygen species (76,77). Different groups have been using cisplatin in combination with antioxidants and immunomodulatory agents with promising results (78,79). In an attempt to reduce the adverse effects of cisplatin, carboplatin was developed and presented excellent activity against visceral leishmaniasis as well (80).

Carmustine and mitomycin-C, alkylating agents used in treatment of cancer, exhibited leishmanicidal activity *in vitro* against *L. donovani*, with the mechanism of action involving the inhibition of trypanothione reductase. The encapsulation of

mitomycin in nanospheres has been a novel approach used to decrease drug toxicity in the host cell and has shown promising results for repositioning strategies (81–83).

Doxorubicin, an anthraquinone used in the treatment of a wide variety of tumors with a mechanism of action on topoisomerase II and an ability to intercalate in DNA, has potent activity *in vitro* and in an experimental model of visceral leishmaniasis. The development of doxorubicin-loaded microparticles and nanocapsules showed promising results in the experimental model of visceral leishmaniasis (84–86).

Camptothecin is a quinoline and an inhibitor of topoisomerase I used in the treatment of a wide variety of tumors. The camptothecin derivatives topotecan and irinotecan were tested against promastigotes and intracellular amastigotes of *L. infantum*, and only the first drug had activity (87).

Paclitaxel is a tetracyclic diterpene used in the treatment of ovarian, breast and lung cancer. This drug leads to the death of promastigotes and intracellular amastigotes by action on tubulin and alterations in the cell cycle, arresting the G2/M phase of parasites (88,89). Sunitinib is used in the treatment of renal cell carcinoma, sorafenib in hepatocellular carcinoma and breast cancer and lapatinib in breast cancer. All these drugs were evaluated against Leishmania spp. and showed excellent activity in vitro and in experimental model of visceral leishmaniasis (90). Imatinib is used for the treatment of chronic myeloid leukemia, which inhibits the Abl/Arg kinase, a family of enzymes involved in endocytosis. This drug decreased in vitro infection of L. amazonensis, indicating that this enzyme is involved in phagocytosis and consequently in Leishmania infection. The treatment in vivo with imatinib resulted in mice with smaller lesions with fewer parasites (91). Dactolisib, an inhibitor of PI3K and mTOR kinase in clinical study as a possible anticancer drug, demonstrated activity in vitro against L. major and L. donovani. However, was ineffective to treat L.major-infected mice (92).

Other anticancer agents that could be cited include tamoxifen, which is used in the treatment of breast cancer and is a selective estrogen receptor modulator with activity against *L. amazonensis*, *L. braziliensis*, *L. infantum chagasi* and *L. major in vitro* and *in vivo*. In parasites, it induces cell death by apoptosis and, in infected macrophages, modifies the alkaline intravacuolar pH and increases drug action against amastigote forms (93–98). The combination of tamoxifen with reference drugs (amphotericin B or miltefosine) used in the treatment of leishmaniasis has also shown promising results (99,100). Raloxifene, an estrogen receptor modulating drug, was tested against the promastigotes of *L. amazonensis*, *L.*

infantum chagasi, L. donovani, L. braziliensis, L. major and L. mexicana, showing IC $_{50}$ values between 30.2 μ M and 38 μ M, and against amastigotes of L. amazonensis and L. infantum chagasi, showing IC $_{50}$ values between 8.8 μ M and 16.2 μ M, and the parasites exhibited autophagosomes and mitochondrial damage. In vivo efficacy trials of raloxifene in L. amazonensis-infected BALB/c mice showed a 41.7.% reduction in lesion size of the treated animals compared to the untreated animals and an 89.7.% reduction in parasitic load (101).

4.1.4. Antidepressant drugs

In addition to their characteristic effects, antidepressants of different classes have already demonstrated an excellent ability to eliminate parasites of the genus *Leishmania* (Figure 7).

Tricyclic antidepressants are indicated for the treatment of depression, obsessive compulsive disorder, panic attacks and chronic pain. Three drugs of this class have been studied in relation to their activity and mechanism of action: imipramine, clomipramine and amitriptyline. **Imipramine** demonstrates leishmanicidal activity, leading to the death of L. amazonensis in vitro and L. donovani in vitro and in vivo. The mechanism of action of imipramine in parasites involves alterations in the sterol profile and proton motive force, inhibition of trypanothione reductase and induction of apoptosis. It also has immunomodulatory capacity, inducing the production of TNF-α and IFN-y and decreasing that of IL-10 and TGB-β. Other work has demonstrated that imipramine inhibits IL-10 production by upregulating histone deacetylase, which inhibits the acetylation of the IL-10 promoter (102-105). Clomipramine also exhibited activity against L. donovani and L. major. The mechanism of action in parasites involves the inhibition of L-proline transport, disruption of the plasmatic membrane in promastigotes and inhibition of trypanothione reductase (106,107). Amitriptyline shows activity against *L. donovani* but is less potent than imipramine and clomipramine, causing a decrease in proline transport and depletion in ATP levels in these parasites (105). Interestingly, cyclobenzaprine (CBP), a tricyclic compound structurally related to the amitriptyline and used as muscle relaxer, recently demonstrated activity against L. infantum in vitro and in vivo by raising ROS levels in promastigotes (108).

Sertraline, a selective serotonin reuptake inhibitor used for the treatment of depression and obsessive compulsive, panic and anxiety disorder, shows promising activity against *L. donovani* with efficacy in oral therapy and induces a reduction in ATP levels and oxygen consumption in promastigotes (109). Mianserin is a tetracyclic antidepressant used in the treatment of depression, anxiety and schizophrenia.

Figure 7. Antidepressant drugs

This drug kills promastigotes and amastigotes of *L. donovani* by inhibiting the enzyme 3-hydroxy-3-methyl-glutaryl-CoA reductase (110).

Phenelzine and nialamide are monoamine oxidase inhibitors used in the treatment of depression and anxiety. Both demonstrated activity against visceral and cutaneous forms of Leishmania, but nialamide was more promising in a murine model of cutaneous leishmaniasis (111). Paroxetine is a selective serotonin reuptake inhibitor used in the treatment of depression and anxiety. Recently, it presented activity against promastigotes of L. infantum with an IC $_{\rm 50}$ equal to 2.2 $\mu \rm M.$ The mechanism of action of paroxetine in epimastigotes of T. cruzi involved the inhibition of the up take of putrescine, but in Leishmania spp., the mechanism, which culminates in cell death, has not yet been elucidated (112).

Diazepam is an antidepressant of benzodiazepines class, was active against promastigotes of $L.\ mexicana$ with IC₅₀ equal to 15mM. The treatment for 48h promoted ultrastructural changes indicating interference in cell division (113).

4.1.5. Antihypertensive drugs

Several antihypertensive drugs (Figure 8) have been evaluated against parasites of the genus

Leishmania in the search for the discovery of new therapeutic options.

Calcium channel blockers are a group of drugs often prescribed for the treatment of high blood pressure (hypertension) that acts by relaxing the arterial smooth muscle (114). Among the calcium channel blockers most used in clinical practice, azelnidipine, cilnidipine, lercanidipine, nicardipine, nitrendipine, nifedipine, nimodipine, and amlodipine were effective against *L. amazonensis*, *L. braziliensis*, *L. infantum chagasi*, and *L. major* (115,116). Different groups have studied using calcium channel blockers against *Leishmania* spp. and observed promising antiparasitic activities (114–118).

Nimodipine, a 1,4-dihydropyridine, was active against the promastigote and intracellular amastigote forms of *L. infantum chagasi*, although the promastigotes were less susceptible. Nimodipine also showed activity against other species of *Leishmania*, yet *L. major* was more susceptible than *L. amazonensis*. The treatment of nimodipine did not block amastigote penetration into the host cells but did cause ultrastructural damages, such as intracytoplasmic vacuoles, membrane blebbing, nuclear membrane bilayer detachment and augmented mitochondria (119).

Nifedipine and verapamil had inhibitory effects on macrophage infection by *L. donovani*,

Figure 8. Antihypertensive drugs

suggesting that the Ca²+ ion plays a role in the invasion process. In the treatment with verapamil, the number of parasites attached per infected macrophage and the number of infected macrophages was reduced, but no antiparasitic effect was observed. The pretreatment of macrophages with verapamil and nifedipine reduced their ability to bind the parasite (120). Other studies have demonstrated different results, with an increase in the percentage of infected macrophages and in the parasite load in the presence of nifedipine, suggesting that this drug impairs the microbicidal functions of macrophages (121). Another controversial result is related to the activity of verapamil, which inhibited Ca²+ uptake by *L. donovani* but did not show leishmanicidal activity against promastigotes and amastigotes (122).

Another interesting use of verapamil, a resistance reversing drug, is inhibiting drug efflux pump proteins such as P-glycoprotein in resistant parasites. Isolates of L. donovani resistant to sodium stibogluconate were treated with verapamil, which reversed the drug resistance and showed a significant reduction in IC_{50} . Verapamil alone did not have any effect on the parasites but could reverse the $in\ vitro$ drug resistance of L. donovani clinical isolates to sodium stibogluconate (123).

Amlodipine, lacidipine, verapamil diltiazem inhibited Ca2+ uptake by L. donovani, but only the first two had leishmanicidal activity against L. donovani promastigotes and amastigotes. This result demonstrated that there was no relationship between the leishmanicidal activity and the Ca2+ channel blocking action of these drugs because verapamil and diltiazem inhibited Ca2+ uptake but did not have activity against the parasites (122). Amlodipine and lacidipine were also effective against infections in BALB/c mice when administered orally, showing significant decreases in the spleen and liver weights and satisfactory reduction in the parasite burdens compared to those of the untreated controls (122). Another calcium-blocking agent, bepridil, demonstrated effectiveness against different visceral and cutaneous forms of Leishmania but failed to treat L. infantum chagasi-infected hamsters (117).

Goyal *et al.* reported the identification and biochemical characterization of an angiotensin-converting enzyme (ACE)-related dipeptidyl carboxypeptidase (DCP) in *L. donovani*. The ACE inhibitor, captopril, blocked both ACE activities, rat and leishmania, but the sensitivity of parasite's enzyme to this inhibitor is much less than that to the mammalian

one (124). Other studies tested the activity of enalapril and lisinopril against promastigotes and amastigotes of *L. amazonensis*, but no inhibition of parasite growth was observed (125).

In BALB/c mice infected with L. mexicana, the β-blocker propranolol demonstrated a 47% inhibition of footpad swelling, a significant reduction of the parasitic burden and a substantial increase in CD4+ and CD8+ splenic T lymphocytes producing IFN-y, which play an essential role in the elimination of these parasites. It was suggested that β-adrenergic receptor antagonists could be of therapeutic value. either as a treatment or as an adjuvant of vaccines for L. mexicana (126). The treatment of L. amazonensis promastigotes and amastigotes with other β-blockers, atenolol and metoprolol, did not show efficacy against the parasite (125), but in L. major infections, which are characterized by inflammation and hyperalgesia. peripheral hyperalgesia was induced during treatment with atenolol (127).

Sodium nitroprusside (SNP), a potent NO donor, promotes arterial and venous vasodilation and has been used in hypertensive emergencies. The treatment of *L. amazonensis* promastigotes and axenic amastigotes revealed a decrease in parasite number and showed morphological changes with loss of viability (128). SNP had increased leishmanicidal activity against *L. amazonensis*-infected macrophages, reducing the number of amastigotes and stimulating the levels of TNF-α, nitric oxide (NO) and 3-nitrotyrosine, thus suggesting that the drug is an exogenous source of NO (129).

Chlorthalidone and hydrochlorothiazide, two thiazide diuretics, and losartan, an angiotensin II receptor antagonist, were tested against L. amazonensis but did not demonstrate activity, showing no activity until the use of concentrations of 100 μ M in promastigotes and 25 μ M in amastigotes (125).

Ketanserin is an antihypertensive drug but also shows antidepressant action as a 5-HT2A/2C receptor antagonist. This drug leads to the death of promastigotes and intracellular amastigotes of *L. donovani* by inhibiting the enzyme 3-hydroxy-3-methylglutaryl-coenzyme A reductase, an important enzyme involved in ergosterol biosynthesis in this parasite (130).

4.1.6. Antibiotics

Antibiotics encompass several classes of molecules possessing leishmanicidal activity (Figure 9), within which is the paromomycin that is approved for the treatment of leishmaniasis in some countries

Penicillin is used in small concentrations in *Leishmania* spp. cultures in order to suppress bacterial contamination and thus does not have an effect on the parasites. The treatment of cutaneous leishmaniasis caused by *L. tropica* with large doses of penicillin, infiltration typical of leishmaniasis lesions remained, and no recovery was observed over a period of approximately 6 months. The largest dose of penicillin had no effect on lesions of *L. tropica* in humans, but experiments *in vitro* showed that penicillin inhibited the growth of the parasite (131). Oxacillin was also tested against *L. amazonensis* and did not have activity until used at concentrations of 100 μ M in promastigotes and 25 μ M in amastigotes (125).

Fluoroguinolones, a class of antibacterial agents that act through the inhibition of type II DNA topoisomerases (TOPII), showed leishmanicidalselective action against intracellular amastigotes of L. panamensis. Enoxacin and ciprofloxacin demonstrated to be the most selective agents for the parasites over human macrophages (132). Enoxacin, ciprofloxacin, ofloxacin, lomefloxacin, and norfloxacin displayed significant potencies and selectivity for *L. panamensis* TOPII (133). Marbofloxacin, a third generation fluoroquinolone for veterinary use, was tested in vitro against clinical strains of L. infantum alone or in combination with allopurinol. The combination between miltefosine and allopurinol caused a significant higher percentage of killed parasites than marbofloxacin plus allopurinol. Marbofloxacin was more efficacious when associated with allopurinol than alone after 72 h of incubation (134). Miltefosine was more efficacious than marbofloxacin, but when compared to meglumine antimoniate and sodium stibogluconate, marbofloxacin was more efficacious, with increases in NO and TNF-α observed after marbofloxacin treatment (135). Dogs naturally infected with visceral leishmaniasis and treated daily with a dose of 2 mg/kg marbofloxacin for different period of times, and the treatment for 28 days lead to clinical improvement and was considered, safe and efficient (136).

Among the aminoglycosides already tested, streptomycin and tobramycin have been shown to inhibit the growth of *L. donovani* parasites, whereas gentamycin did not inhibit the growth *in vitro*. Streptomycin and tobramycin demonstrated activity against intracellular amastigotes, with inhibitions of 63% and 52%, respectively (137).

Azithromycin, a semi-synthetic macrolide antibiotic chemically related to erythromycin and clarithromycin, has provided controversial results. Azithromycin demonstrated activity against promastigotes and amastigotes of *L. amazonensis*, *L. braziliensis*, *L. tropica*, *L. major* and *L. infantum chagasi* (138–140). The *in vivo* studies showed a decrease in the

Figure 9. Antibiotics drugs

parasite number following subcutaneous administration of azithromycin in BALB/c mice infected with L. major (139). The activity of azithromycin was also evaluated in golden hamsters infected with L. braziliensis and L. amazonensis using oral administration. NO activity against infections with L. amazonensis could be observed, but activity was demonstrated against L. braziliensis. However, azithromycin was unable to totally block the development of foot pad swelling, although the lesions were significantly smaller in the treated animals (141). The combination of azithromycin with glucantime or miltefosine was also evaluated. The combination of azithromycin and glucantime was not more efficient than treatment with glucantime alone in the experimental cutaneous leishmaniasis (142). Oral treatment with azithromycin plus miltefosine for a short period of 10 days in L. major-infected mice induced a dramatic clinical improvement, but relapse rapidly developed after the cessation of therapy (143). Clarithromycin exhibits antileishmanial properties that were more effective against promastigotes and intracellular amastigotes of L. tropica and L. donovani than azithromycin at lower concentration with minimal toxicity to mammalian cells (140,144). However, clarithromycin, when tested against promastigotes and amastigotes of L. amazonensis, did

not demonstrate activity (125). Clofazimine, originally developed for the treatment of tuberculosis and used in combination with rifampicin and dapsone as multidrug therapy (MDT) for the treatment of leprosy (145), demonstrated leishmanicidal activity (146). Clofazimine showed leishmanicidal activity against L. infantum and L. tropical, with IC $_{50}$ values of 4.48 μ M and 2.96 μ M, respectively, (147). Another antibiotic, tetracycline, was tested on the promastigote form of L. major and demonstrated leishmanicidal activity (148). Cephalexin, a cephalosporin, has been tested against promastigotes and amastigotes but did not show activity up to concentrations of 100 μ M in promastigotes and 25 μ M in amastigotes (125).

The combination of sulfadiazine, trimethoprim, and metronidazole or tinidazole showed good activity as an oral treatment for 12 to 25 weeks in nine patients with kala-azar, and no side effects were noticed (149). The combinations between trimethoprim and sulfamethoxazole and between isoniazid and rifampicin demonstrated no antileishmanial activity against intracellular amastigotes of *L. tropica* (less than or equal to 40% parasite elimination) but have been reported as being orally efficacious in other studies (146,150,151).

Figure 10. Miscellaneous drugs

Other studies showed that rifampicin, an antibiotic commonly used in the treatment of mycobacterial infections, significantly inhibited the growth of promastigotes and intracellular amastigotes of L. aethiopica, L. donovani, L. tropica and L. major (152,153). Rifampicin and amphotericin B were combined in vitro in intracellular amastigotes of L. tropica and showed a synergistic effect at several cell concentrations (154). Experiments in vivo were also conducted, and the treatment of mice infected with promastigotes of L. amazonensis with daily doses of 0.5 mg of rifampicin provided a significant reduction in the size of local lesions, but daily doses of 20 mg/kg in children or 1200 mg in adult patients infected with L. braziliensis did not bring any sign of improvement after 30 days of treatment (155).

Novel approved drug for for the treatment of MDR tuberculosis, Delamanid showed potent *in vitro* and *in vivo* activity against *L. donovani* and *L. infantum*. BALB/c mice infected with *L. donovani* treated with maximum dose of 30 mg/kg had suppressed infection in 99.5.%, given an estimated ED $_{50}$ and ED $_{90}$ of 7.3. and 21.5 mg/kg, respectively (156).

4.1.7. Other drugs

Other drugs have also been investigated for their antileishmanial activity (Figure 10). Disulfiram, used to support the treatment of chronic alcoholism, showed excellent activity against promastigotes and intracellular amastigotes with IC $_{\rm 50}$ values of 0.058 μM and 0.062 μM , respectively (157).

Cyclosporin, an immunosuppressant, is also a drug whose antileishmanial activity has been tested; in promastigotes of *L. donovani*, it caused a reduction in parasite growth in a dose-dependent manner, with a more than 5-fold decrease in parasite growth at the

highest concentration tested and with an IC $_{50}$ value between 15 and 20 μ M. Axenic amastigotes of *L. donovani* showed high susceptibility to the drug, which exhibited an IC $_{50}$ value between 5 and 10 μ M (158).

In another study, cyclosporin was able to reduce the infection rate of *L. major*-infected macrophages to less than 15% compared to that of untreated cultures, which had a 75% infection rate (159).

Hydroxyzine, an antagonist of histamine receptor H1 drug, was tested against L. infantum promastigotes and had an IC $_{50}$ value of 59.57 μ M (160). Omeprazole is a specific inhibitor of human gastric K⁺H⁺-ATPase. This drug had an inhibitory effect on K⁺H⁺-ATPase from L. donovani. The activity of this drug was tested on the promastigotes of L. donovani, and an IC $_{50}$ value of 50 μ M was obtained. Omeprazole also had a significant effect on the survival of intracellular amastigotes: at a concentration of 150 μ M, there were few or no parasites within the macrophages in comparison to untreated macrophages (161).

Ezetimibe, an antihypercholesterolemic drug, was active on promastigotes and amastigotes of L. amazonensis, with IC $_{50}$ values of 30 μ M and 20 μ M, respectively, were obtained. Analysis has shown that drug-treated parasites become rounded and that ezetimibe interfered with the ergosterol biosynthesis pathway. Assays of the combination of ezetimibe with the antifungals ketoconazole and miconazole showed a synergistic effect, with a significant decrease in the IC $_{90}$ values for intracellular amastigotes; for ketoconazole, there was a reduction from 11.3. μ M to 4.14 μ M, and for miconazole, there was a reduction from 11.5 μ M to 8.25 μ M, representing an improvement in the antileishmanial action of azoles (125).

Figure 11. Drugs in clinical trials involving dogs or humans.

5. CLINICAL TRIALS

Several groups have performed clinical trials using patients or dogs infected with different species of *Leishmania*, demonstrating a broad spectrum of clinical manifestations.

The efficacy of allopurinol (Figure 11) was evaluated in the treatment of 45 dogs with clinical signs indicating leishmaniases. In the treated group, a significant improvement in the general body condition was observed, such as reduction in peripheral lymphadenopathy, splenomegaly and cutaneous ulcerations. Comparing the allopurinol-treated group with the placebo group a significant decrease in the amastigotes in lymph node and bone marrow aspiration smears was observed (162). A controversial result was observed after treatment with allopurinol of adult patients with cutaneous leishmaniasis caused by L. panamensis and L. braziliensis, in which no effect was achieved (163). Another drug that was tested in dogs was domperidone, which is involved in the innate immune response and which could control early infections by Leishmania spp. Infected dogs were treated with an oral suspension of domperidone, which reduced the risk of developing clinical canine leishmaniasis (164).

The efficacy of rifampicin (Figure 9) and omeprazole (Figure 10) was determined through a randomized, double-blind, placebo-controlled study conducted on 50 patients with cutaneous leishmaniasis caused by *L. tropica*. The result demonstrated that 20% of patients had unhealed lesions in the group receiving rifampicin and omeprazole, whereas 82% of

patients had unhealed lesions at the end of 6 weeks in the placebo group (165).

Treatment with azithromycin (Figure 9) in patients with cutaneous leishmaniasis in two regions of Brazil, Araçuaí and Varzelândia in Minas Gerais state, and Manaus in Amazonas state, demonstrated different results. The first study in patients with mucosal leishmaniasis demonstrated that azithromycin is a good therapeutic option, whereas in the second one the drug showed a very low efficacy for cutaneous leishmaniasis (166,167). The other group that used azithromycin for the treatment of patients with Old World cutaneous leishmaniasis also observed no effect (168). Meanwhile, the effect of azithromycin on patients with cutaneous leishmaniasis caused by L. braziliensis was compared with that of meglumine antimoniate. Patients received oral azithromycin at 500 mg/day or intramuscular meglumine antimoniate at 10 mg Sb/kg/day for 28 days. The treatment with azithromycin showed moderate activity and provides evidence about the safety of this therapy (169).

Ergosterol biosynthesis inhibitors, such as itraconazole, ketoconazole, fluconazole and terbinafine, have been studied for many years (Figure 4), and some clinical trials have been carried out (170). A randomized, double-blind, placebo-controlled clinical trial using oral itraconazole evaluated 200 patients infected with *L. major*. After 8 weeks of treatment, no difference was observed compared to the placebo group (171). However, other controlled studies showed that oral itraconazole therapy was safe and effective for the treatment of cutaneous leishmaniasis caused by *L. major* (172).

Oral ketoconazole was tested for the treatment of cutaneous leishmaniasis and compared with intralesional administration of meglumine antimoniate. Ketoconazole treatment demonstrated a cure rate of 89% and a failure rate of 11% and seemed to be safe and effective for treatment, especially in children (173). The combination of intralesional sodium stibogluconate (SSG) and oral ketoconazole was demonstrated to be more effective than intralesional SSG alone, especially in cases with large plaque lesions and mucosal lesions. This combination is easy to use, effective, painless and free of *major* side effects (174). Other studies showed that the outcome of the ketoconazole treatment of cutaneous leishmaniasis was influenced by the parasite species. Patients infected with L. braziliensis and treated with ketoconazole showed a cure rate of 30%; however, patients infected with L. mexicana showed a cure rate of 89%. These differences highlight the importance of speciation in the treatment of leishmaniasis (175).

High doses of fluconazole were used in two studies: (i) a non-controlled open trial with patients that showed a cure rate of 75–100% for cutaneous leishmaniasis (176), and (ii) a randomized controlled trial to evaluate the efficacy and safety of oral fluconazole in patients infected with *L. braziliensis* that considered the drug ineffective (177). The treatment of patients infected with *L. major* with 400 mg of oral fluconazole daily was more efficient than treatment with 200 mg, but it showed side effects (178). Another antifungal, terbinafine, was tested as a topical formulation. Topical terbinafine used for the treatment of cutaneous leishmaniasis caused by *L. tropica* was more efficient than the placebo group, but the cure rate depended on the lesion type (179).

Imiquimod (Figure 11), which acts as an immunomodulatory agent, was tested in combination with meglumine antimoniate for the treatment of cutaneous leishmaniasis. Imiquimod indirectly stimulates the production of a T-cell type 1 response, and the application of 5% of drug in a cream for 4 weeks did not improve the response to the meglumine antimoniate treatment (180).

Pentoxifylline (Figure 11), a xanthine derivative used to treat peripheral vascular disease, inhibits tumor necrosis factor- α (TNF- α), attenuates the immune response and decreases tissue inflammation (181). This drug has been investigated alone or combined with meglumine antimoniate, mainly in cases of refractory mucocutaneous leishmaniasis or cutaneous leishmaniasis (17,182–184). The lesions of patients with mucocutaneous leishmaniasis caused by *L. braziliensis* receiving the combination treatment healed faster than those of the patients treated with the antimonial alone (185). The combination of

glucantime and orally administered pentoxifylline was used in the 10 patients with refractory mucosal leishmaniasis, and a re-epithelization of the mucosa was observed in 90% of patients (186). In another study, a randomized, double-blind trial for mucosal leishmaniasis caused by *L. braziliensis* was also conducted using oral pentoxifylline combined with glucantime. This combination accelerates the healing time of mucosal leishmaniasis with a 58% cure rate compared with control group and thus should be a therapeutic choice for the treatment of refractory mucosal leishmaniasis (187).

Fexinidazole (Figure 11) is a 5-nitroimidazole that has shown activity *in vitro* and *in vivo* against Chagas disease and human African trypanosomiasis (188–190). Currently, fexinidazole is in clinical development for the treatment of human sleeping sickness, with phase I completed (191).

Studies in vitro and in vivo have demonstrated that fexinidazole and its metabolites showed activity against visceral leishmaniasis caused by L. donovani and are activated by nitroreductases (192,193). Subsequently, fexinidazole was included in a clinical trial as an oral drug for the treatment of visceral leishmaniasis in Sudan. According to clinicaltrial. gov, fexinidazole is in a phase II trial to determine its efficacy as an oral treatment of visceral leishmaniasis in Sudanese adult patients. The treatment consists of 600 mg tablets given orally after the main daily meal at a daily dose of 1800 mg (3 tablets) once a day for 4 days, followed by 1200 mg (2 tablets) once a day for 6 days (194). This clinical trial appears as finished on clinicaltrial.gov; however, the results are not yet reported.

6. CONCLUSION

In this work, the main studies on the drug repositioning of FDA-approved drugs for cutaneous and visceral leishmaniasis were summarized. Drug repurposing is an attractive strategy for neglected tropical disease drug discovery and development, particularly for leishmaniases because a large part of the therapeutic arsenal consists of drugs that have been repurposed. This review presents drugs that have been identified by systematically searching the literature and, in figures, shows the distribution of pharmacological classes and available drugs for the purpose of repositioning. Recent studies using HTS and HCS approaches have contributed to the advancement of the classical drug discovery process and repurposing strategy for different diseases. Even with all the available new technologies, most of the studies identified in this review involve classical assays performed by groups worldwide. In Figure 12 are represented the main classes of drugs studied for repositioning, independent of having a promising

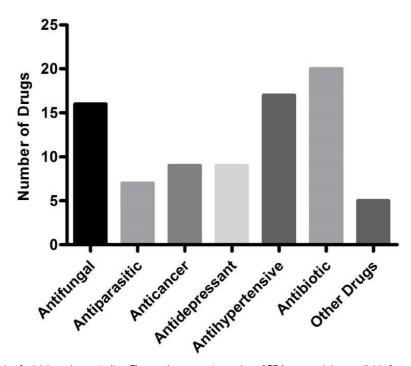


Figure 12. Drug repurposing for leishmaniases studies. The graph represents number of FDA-approval drug available for use against *Leishmania* spp. in vitro and/or in vivo.

result. The most advanced class in preclinical stages with positive results are the antitumorals. Tamoxifen as monotherapy or combinatory therapy (with reference drugs) has *in vivo* efficacy in different models of experimental leishmaniases, and *in vitro* resistance induction studies have shown that the parasites are not prone to develop resistance to tamoxifen. Fexinidazole is in phase II clinical studies for visceral leishmaniasis; however, the result of this trial was still not reported. The last drug repurposed for leishmaniases was 85097uifylline, used as an adjuvant therapy for cutaneous leishmaniasis in New World and Old World. Drug repurposing is a fast and cheap strategy for the discovery and development of new treatments for leishmaniases.

7. ACKNOWLEDGEMENTS

Valter Viana Andrade-Neto and Edezio Ferreira Cunha-Junior contributed equally to this paper. This work was supported by Fundação de Apoio à Pesquisa do Estado do Rio de Janeiro (Fellow and grant E-26/010.0.01828/2016 to EFCJ) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (Fellow to VVAN).

8. REFERENCES

 Deotarse, P. P., Jain, A. S., Baile, M. B., Kolhe, N. S. and Kulkarni, A. A. Drug Repositioning: A Review. *Int. J. Pharma Res. Rev.* 4, 51–58 (2015)

- Langedijk, J., Mantel-Teeuwisse, A. K., Slijkerman, D. S. and Schutjens, M.-H. D. B. Drug repositioning and repurposing: terminology and definitions in literature. *Drug Discov. Today* 20, 1027–1034 (2015) DOI: 10.1016/j.drudis.2015.05.001
- Ashburn, T. T. and Thor, K. B. Drug repositioning: identifying and developing new uses for existing drugs. *Nat. Rev. Drug Discov.* 3, 673–683 (2004)
 DOI: 10.1038/nrd1468
- Sharlow, E. R. Revisiting Repurposing. *Assay Drug Dev. Technol.* 14, 554–556 (2016) DOI: 10.1089/adt.2016.766
- 5. Jin, G. and Wong, S. T. C. Toward better drug repositioning: Prioritizing and integrating existing methods into efficient pipelines. *Drug Discov. Today* 19, 637–644 (2014) DOI: 10.1016/j.drudis.2013.11.005
- Smith, R. B. Repositioned drugs: Integrating intellectual property and regulatory strategies. *Drug Discov. Today Ther. Strateg.* 8, 131–137 (2012) DOI: 10.1016/j.ddstr.2011.06.008
- 7. Berenstein, A. J., Magariños, M. P., Chernomoretz, A. and Agüero, F. A Multilayer

- Network Approach for Guiding Drug Repositioning in Neglected Diseases. *PLoS Negl. Trop. Dis.* 10, e0004300 (2016) DOI: 10.1371/journal.pntd.0004300
- Akhoundi, M., Kuhls, K., Cannet, A., Votýpka, J., Marty, P., Delaunay, P. and Sereno, D. A Historical Overview of the Classification, Evolution, and Dispersion of *Leishmania* Parasites and Sandflies. *PLoS Negl. Trop. Dis.* 10, e0004349 (2016) DOI: 10.1371/journal.pntd.0004349
- Desjeux, P. Leishmaniasis: current situation and new perspectives. *Comp. Immunol. Microbiol. Infect. Dis.* 27, 305–318 (2004)
 DOI: 10.1016/j.cimid.2004.03.004
- Alvar, J., V?lez, I. D., Bern, C., Herrero, M., Desjeux, P., Cano, J., Jannin, J. and Boer, M. den. Leishmaniasis Worldwide and Global Estimates of Its Incidence. *PLoS One* 7, e35671 (2012) DOI: 10.1371/journal.pone.0035671
- Kevric, I., Cappel, M. A. and Keeling, J. H. New World and Old World Leishmania Infections. Dermatol. Clin. 33, 579–593 (2015)
 DOI: 10.1016/j.det.2015.03.018
- Oryan, A. and Akbari, M. Worldwide risk factors in leishmaniasis. *Asian Pac. J. Trop. Med.* 9, 925–932 (2016)
 DOI: 10.1016/j.apjtm.2016.06.021
- Hashiguchi, Y., Gomez, E. L., Kato, H., Martini, L. R., Velez, L. N. and Uezato, H. Diffuse and disseminated cutaneous leishmaniasis: clinical cases experienced in Ecuador and a brief review. *Trop. Med. Health* 44, 1–9 (2016) DOI: 10.1186/s41182-016-0002-0
- Vernal, S., De Paula, N.A., Gomes, C. M. and Roselino, A. M. Disseminated Leishmaniasis by *Leishmania* viannia Subgenus: A Series of 18 Cases in Southeastern Brazil. *Open Forum Infect. Dis.* 3, ofv184 (2016) DOI: 10.1093/ofid/ofv184
- Machado, P. R. L., Rosa, M. E. A., Guimarães, L. H., Prates, F. V. O., Queiroz, A., Schriefer, A. and Carvalho, E. M. Treatment of Disseminated Leishmaniasis With Liposomal Amphotericin B. Clin. Infect. Dis. 61, 945–949 (2015) DOI: 10.1093/cid/civ416

- Handler, M. Z., Patel, P. A., Kapila, R., Al-Qubati, Y. and Schwartz, R. A. Cutaneous and mucocutaneous leishmaniasis: Differential diagnosis, diagnosis, histopathology, and management. *J. Am. Acad. Dermatol.* 73, 911–26–8 (2015)
- 17. Amato, V. S., Tuon, F. F., Siqueira, A. M., Nicodemo, A. C. and Neto, V. A. Treatment of mucosal leishmaniasis in Latin America: Systematic review. *Am. J. Trop. Med. Hyg.* 77, 266–274 (2007)
- Mukhopadhyay, D., Dalton, J. E., Kaye, P. M. and Chatterjee, M. Post kala-azar dermal leishmaniasis: an unresolved mystery. *Trends Parasitol.* 30, 65–74 (2014) DOI: 10.1016/j.pt.2013.12.004
- Vianna, G. Comunicação à Sessão de 24 de abril de 1912 da Sociedade Brasileira de Dermatologia. Arch. Bras. Med. 1, 36–38 (1912)
- Frézard, F., Demicheli, C. and Ribeiro, R. R. Pentavalent antimonials: New perspectives for old drugs. *Molecules* 14, 2317–2336 (2009)
 DOI: 10.3390/molecules14072317
- 21. Sundar, S. and Chakravarty, J. An update on pharmacotherapy for leishmaniasis. *Expert Opin. Pharmacother.* 16, 237–52 (2015) DOI: 10.1517/14656566.2015.973850
- 22. DNDi. Current Treatments DNDi. at http://www.dndi.org/diseases-projects/leishmaniasis/leish-current-treatments/>
- Uliana, S. R. B., Trinconi, C. T. and Coelho, A. C. Chemotherapy of leishmaniasis: present challenges. *Parasitology* 1–17 (2017) DOI: 10.1017/S0031182016002523
- Chávez-Fumagalli, M. A., Ribeiro, T. G., Castilho, R. O., Fernandes, S. O. A., Cardoso, V. N., Coelho, C. S. P., Mendonça, D. V. C., Soto, M., Tavares, C. A. P., Faraco, A. A. G., Coelho, E. A. F., Chávez-Fumagalli, M. A., Ribeiro, T. G., Castilho, R. O., Fernandes, S. O. A., Cardoso, V. N., Coelho, C. S. P., Mendonça, D. V. C., Soto, M., Tavares, C. A. P., Faraco, A. A. G. and Coelho, E. A. F. New delivery systems for amphotericin B applied to the improvement of leishmaniasis treatment. Rev. Soc. Bras. Med. Trop. 48, 235–242 (2015)

DOI: 10.1590/0037-8682-0138-2015

- Hellier, I., Dereure, O., Tournillac, I., Pratlong, F., Guillot, B., Dedet, J. P. and Guilhou, J. J. Treatment of Old World cutaneous leishmaniasis by pentamidine isethionate. An open study of 11 patients. *Dermatology* 200, 120–3 (2000)
 DOI: 10.1159/000018343
- Sundar, S., Singh, A., Rai, M., Prajapati, V. K., Singh, A. K., Ostyn, B., Boelaert, M., Dujardin, J.-C. and Chakravarty, J. Efficacy of Miltefosine in the Treatment of Visceral Leishmaniasis in India After a Decade of Use. Clin. Infect. Dis. 55, 543–550 (2012) DOI: 10.1093/cid/cis474
- 27. Leishmaniases and Geneva. WHO Technical Report Series control of the leishmaniases. *II.World Heal. Organ. III.Series. ISBN* 978, 22–26 (2010)
- 28. Lim, H., Poleksic, A., Yao, Y., Tong, H., He, D., Zhuang, L., Meng, P. and Xie, L. Large-Scale Off-Target Identification Using Fast and Accurate Dual Regularized One-Class Collaborative Filtering and Its Application to Drug Repurposing. *PLoS Comput. Biol.* 12, e1005135 (2016)
 DOI: 10.1371/journal.pcbi.1005135
- 29. Swinney, D. C. and Anthony, J. How were new medicines discovered? *Nat. Rev. Drug Discov.* 10, 507–19 (2011) DOI: 10.1038/nrd3480
- Spangenberg, T., Burrows, J. N., Kowalczyk, P., McDonald, S., Wells, T. N. C. and Willis, P. The Open Access Malaria Box: A Drug Discovery Catalyst for Neglected Diseases. *PLoS One* 8, (2013) DOI: 10.1371/journal.pone.0062906
- 31. Edwards, A. M., Bountra, C., Kerr, D. J. and Willson, T. M. Open access chemical and clinical probes to support drug discovery. *Nat. Chem. Biol.* 5, 436–440 (2009) DOI: 10.1038/nchembio0709-436
- 32. Chatelain, E. and loset, J.-R. Drug discovery and development for neglected diseases: the DNDi model. *Drug Des. Devel. Ther.* 5, 175–81 (2011)
- 33. Peña, I., Pilar Manzano, M., Cantizani, J., Kessler, A., Alonso-Padilla, J., Bardera, A. I., Alvarez, E., Colmenarejo, G., Cotillo, I., Roquero, I., de Dios-Anton, F., Barroso, V., Rodriguez, A., Gray, D. W., Navarro, M., Kumar, V., Sherstnev, A., Drewry, D. H.,

- Brown, J. R., Fiandor, J. M. and Julio Martin, J. New compound sets identified from high throughput phenotypic screening against three kinetoplastid parasites: an open resource. *Sci. Rep.* 5, 8771 (2015) DOI: 10.1038/srep08771
- 34. Kaiser, M., Maes, L., Tadoori, L. P., Ioset, J.-R., Spangenberg, T. and Ioset, J.-R. Repurposing of the Open Access Malaria Box for Kinetoplastid Diseases Identifies Novel Active Scaffolds against Trypanosomatids. *J. Biomol. Screen.* 20, 634–645 (2015) DOI: 10.1177/1087057115569155
- 35. Kaiser, M., Mäser, P., Tadoori, L. P., Ioset, J. R., Brun, R. and Sullivan, D. J. Antiprotozoal activity profiling of approved drugs: A starting point toward drug repositioning. *PLoS One* 10, 1–16 (2015)
 DOI: 10.1371/journal.pone.0135556
- Ali, S. A., Iqbal, J., Nabeel, Khalil, Y., Manzoor, A., Bukhari, I., Ahmad, B. and Yasinzai, M. M. Leishmanicidal activity of Nystatin (mycostatin): a potent polyene compound. *J. Pak. Med. Assoc.* 47, 246–8 (1997)
- 37. Tewary, P., Veena, K., Pucadyil, T. J., Chattopadhyay, A. and Madhubala, R. The sterol-binding antibiotic nystatin inhibits entry of non-opsonized *Leishmania* donovani into macrophages. *Biochem. Biophys. Res. Commun.* 339, 661–6 (2006) DOI: 10.1016/j.bbrc.2005.11.062
- 38. Ghosh, B. K. and Chatterjee, A. N. Leishmanicidal activity of nystatin, a polyene antifungal antibiotic. I. The probable mechanism of action of nystatin on *Leishmania* donovani. *Antibiot. Chemother. (Northfield, III.)* 12, 204–6 (1962)
- 39. de Macedo-Silva, S. T., Urbina, J. A., de Souza, W. and Rodrigues, J. C. F. *In vitro* Activity of the Antifungal Azoles Itraconazole and Posaconazole against *Leishmania amazonensis*. *PLoS One* 8, e83247 (2013) DOI: 10.1371/journal.pone.0083247
- 40. Martínez, A., Carreon, T., Iniguez, E., Anzellotti, A., Sánchez, A., Tyan, M., Sattler, A., Herrera, L., Maldonado, R. A. and Sánchez-Delgado, R. A. Searching for new chemotherapies for tropical diseases: ruthenium-clotrimazole complexes display high in vitro activity against Leishmania major and Trypanosoma cruzi and low toxicity toward

- normal mammalian cells. *J. Med. Chem.* 55, 3867–77 (2012) DOI: 10.1021/jm300070h
- Iniguez, E., Varela-Ramirez, A., Martínez, A., Torres, C. L., Sánchez-Delgado, R. A. and Maldonado, R. A. Ruthenium-Clotrimazole complex has significant efficacy in the murine model of cutaneous leishmaniasis. *Acta Trop.* 164, 402–410 (2016)
 DOI: 10.1016/j.actatropica.2016.09.029
- 42. Kulkarni, M. M., Reddy, N., Gude, T. and McGwire, B. S. Voriconazole suppresses the growth of *Leishmania* species *in vitro*. *Parasitol. Res.* 112, 2095–9 (2013) DOI: 10.1007/s00436-013-3274-x
- Mesquita, J. T., da Costa-Silva, T. A., Borborema, S. E. T. and Tempone, A. G. Activity of imidazole compounds on Leishmania (L.) infantum chagasi: reactive oxygen species induced by econazole. Mol. Cell. Biochem. 389, 293–300 (2014) DOI: 10.1007/s11010-013-1954-6
- Andrade-Neto, V. V., Cicco, N. N. T., Cunha-Junior, E. F., Canto-Cavalheiro, M. M., Atella, G. C. and Torres-Santos, E. C. The pharmacological inhibition of sterol biosynthesis in *Leishmania* is counteracted by enhancement of LDL endocytosis. *Acta Trop.* 119, 194–8 (2011)
 DOI: 10.1016/j.actatropica.2011.05.001
- Bezerra-Souza, A., Yamamoto, E. S., Laurenti, M. D., Ribeiro, S. P. and Passero, L. F. D. The antifungal compound butenafine eliminates promastigote and amastigote forms of *Leishmania* (Leishmania) amazonensis and *Leishmania* (Viannia) braziliensis. Parasitol. Int. 65, 702–707 (2016) DOI: 10.1016/j.parint.2016.08.003
- Vannier-Santos, M. A., Urbina, J. A., Martiny, A., Neves, A. and de Souza, W. Alterations induced by the antifungal compounds ketoconazole and terbinafine in Leishmania. *J. Eukaryot. Microbiol.* 42, 337–46 (1995) DOI: 10.1111/j.1550-7408.1995.tb01591.x
- 47. Zakai, H. A., Zimmo, S. K. and Fouad, M. A. H. Effect of itraconazole and terbinafine on *Leishmania* promastigotes. *J. Egypt. Soc. Parasitol.* 33, 97–107 (2003)
- 48. Sampaio, R. N. R., Takano, G. H. S., Malacarne, A. C. B., Pereira, T. R. and

- de Magalhães, A. V. (*In vivo* Terbinafine inefficacy on cutaneous leishmaniasis caused by *Leishmania* (Leishmania) *amazonensis* in C57BL/6 mice). *Rev. Soc. Bras. Med. Trop.* 36, 531–3 (2003) DOI: 10.1590/S0037-86822003000400018
- Zakai, H. A. and Zimmo, S. K. Effects of itraconazole and terbinafine on *Leishmania major* lesions in BALB/c mice. *Ann. Trop. Med. Parasitol.* 94, 787–91 (2000)
 DOI: 10.1080/00034983.2000.11813603
- 50. Gangneux, J. P., Dullin, M., Sulahian, A., Garin, Y. J. F. and Derouin, F. Experimental evaluation of second-line oral treatments of visceral leishmaniasis caused by *Leishmania* infantum. *Antimicrob. Agents Chemother.* 43, 172–174 (1999)
- Simões-Mattos, L., Teixeira, M. J., Costa, D. C., Prata, J. R. C., Bevilaqua, C. M. L., Sidrim, J. J. C. and Rocha, M. F. G. Evaluation of terbinafine treatment in *Leishmania* chagasi-infected hamsters (Mesocricetus auratus). *Vet. Parasitol.* 103, 207–216 (2002) DOI: 10.1016/S0304-4017(01)00595-7
- 52. Limoncu, M. E., Eraç, B., Gürpınar, T., Özbilgin, A., Balcıoğlu, I. C. and Hoşgör-Limoncu, M. Investigation of *in vitro* antileishmanial activity of moxifloxacin, linezolid and caspofungin on *Leishmania* tropica promastigotes. *Turkiye parazitolojii Derg.* 37, 1–3 (2013) DOI: 10.5152/tpd.2013.01
- 53. Gebre-hiwot, A. and Frommel, D. The in-vitro anti-leishmanial activity of inhibitors of ergosterol biosynthesis. *J. Antimicrob. Chemother.* 32, 837–842 (1993) DOI: 10.1093/jac/32.6.837
- 54. Want, M. Y., Islamuddin, M., Chouhan, G., Ozbak, H. A., Hemeg, H. A., Dasgupta, A. K., Chattopadhyay, A. P. and Afrin, F. Therapeutic efficacy of artemisinin-loaded nanoparticles in experimental visceral leishmaniasis. *Colloids Surf. B. Biointerfaces* 130, 215–21 (2015) DOI: 10.1016/j.colsurfb.2015.04.013
- Sen, R., Bandyopadhyay, S., Dutta, A., Mandal, G., Ganguly, S., Saha, P. and Chatterjee, M. Artemisinin triggers induction of cell-cycle arrest and apoptosis in *Leishmania* donovani promastigotes. *J. Med. Microbiol.* 56, 1213–8 (2007) DOI: 10.1099/jmm.0.47364-0

- 56. Sen, R., Ganguly, S., Saha, P. and Chatterjee, M. Efficacy of artemisinin in experimental visceral leishmaniasis. *Int. J. Antimicrob. Agents* 36, 43–49 (2010) DOI: 10.1016/j.ijantimicag.2010.03.008
- 57. Ghaffarifar, F., Esavand Heydari, F., Dalimi, A., Hassan, Z. M., Delavari, M. and Mikaeiloo, H. Evaluation of Apoptotic and Antileishmanial Activities of Artemisinin on Promastigotes and BALB/C Mice Infected with *Leishmania* major. *Iran. J. Parasitol.* 10, 258–67
- Rocha, V. P. C., Nonato, F. R., Guimarães, E. T., Rodrigues de Freitas, L. A. and Soares, M. B. P. Activity of antimalarial drugs *in vitro* and in a murine model of cutaneous leishmaniasis. *J. Med. Microbiol.* 62, 1001–10 (2013)
 DOI: 10.1099/jmm.0.058115-0
- Khan, M. O. F., Levi, M. S., Tekwani, B. L., Wilson, N. H. and Borne, R. F. Synthesis of isoquinuclidine analogs of chloroquine: antimalarial and antileishmanial activity. *Bioorg. Med. Chem.* 15, 3919–25 (2007) DOI: 10.1016/j.bmc.2006.11.024
- Correia, D., Silva, C. A. and Matthes, A. G. Mefloquine in the treatment of cutaneous leishmaniasis. *Rev. Soc. Bras. Med. Trop.* 32, 585 (1999)
 DOI: 10.1590/S0037-86821999000500018
- 61. Galvão, L. O., Moreira, S., Medeiros, P., Lemos, G. J. P., Cunha, N. F., Antonino, R. M. P., Santos Filho, B. S. and Magalhães, A. V. Therapeutic trial in experimental tegumentary leishmaniasis caused by *Leishmania* (Leishmania) *amazonensis*. A comparative study between mefloquine and aminosidine. *Rev. Soc. Bras. Med. Trop.* 33, 377–382 (2000)
- 62. Hendrickx, E. P., Agudelo, S. P., Munoz, D. L., Puerta, J. A. and Velez Bernal, I. D. Lack of efficacy of mefloquine in the treatment of New World cutaneous leishmaniasis in Colombia. *Am. J. Trop. Med. Hyg.* 59, 889–92 (1998)
 DOI: 10.4269/ajtmh.1998.59.889
- 63. Laguna-Torres, V. A., Silva, C. A., Correia, D., Carvalho, E. M., Magalhaes, A. V and Macedo, V. de O. (Mefloquine in the treatment of cutaneous leishmaniasis in an endemic area of *Leishmania* (Viannia) *bra-*

- ziliensis). Rev. Soc. Bras. Med. Trop. 32, 529–532 (1999) DOI: 10.1590/S0037-86821999000500010
- 64. Brazil, R. P. and Gilbert, B. The action of oxamniquine on *Leishmania braziliensis braziliensis* in hamsters. *Rev. Inst. Med. Trop. Sao Paulo* 18, 87–8 (1976)
- 65. Noël, F., Pimenta, P. H. C., Dos Santos, A. R., Tomaz, E. C. L., Quintas, L. E. M., Kaiser, C. R., Silva, C. L. M. and Férézou, J. P. Δ2,3 -Ivermectin ethyl secoester, a conjugated ivermectin derivative with leish-manicidal activity but without inhibitory effect on mammalian P-type ATPases. *Naunyn. Schmiedebergs. Arch. Pharmacol.* 383, 101–107 (2011)
 DOI: 10.1007/s00210-010-0578-6
- 66. Santos, A. R. dos, Falcão, C. A. B., Muzitano, M. F., Kaiser, C. R., Rossi-Bergmann, B. and Férézou, J. P. Ivermectin-derived leishmanicidal compounds. *Bioorganic Med. Chem.*

DOI: 10.1016/j.bmc.2008.12.003

17, 496–502 (2009)

- Rasheid, K. A. and Morsy, T. A. Efficacy of ivermectin on the infectivity of *Leishmania* major promastigotes. *J Egypt Soc Parasitol* 28, 207–212 (1998)
- Griffiths, W. A. Use of metronidazole in cutaneous leishmaniasis. *Arch. Dermatol.* 112, 1791 (1976)
 DOI: 10.1001/archderm.1976.01630370071021
- Al-Waiz, M., Sharquie, K. E. and Al-Assir, M. Treatment of cutaneous leishmaniasis by intralesional metronidazole. *Saudi Med. J.* 25, 1512–3 (2004)
- 70. Belhadjali, H., Elhani, I., Youssef, M., Babba, H. and Zili, J. Traitement de la leishmaniose cutanée par le métronidazole : étude de 30 cas. *Presse Med.* 38, 325–326 (2009) DOI: 10.1016/j.lpm.2008.09.008
- Mesquita, J. T., Pinto, E. G., Taniwaki, N. N., Galisteo, A. J. and Tempone, A. G. Lethal action of the nitrothiazolyl-salicylamide derivative nitazoxanide via induction of oxidative stress in *Leishmania* (*L.*) *infantum*. *Acta Trop*. 128, 666–673 (2013)
 DOI: 10.1016/j.actatropica.2013.09.018
- 72. Zhang, R., Shang, L., Jin, H., Ma, C., Wu, Y., Liu, Q., Xia, Z., Wei, F., Zhu, X. Q. and Gao, H. *In vitro* and *in vivo* antileishmanial effica-

- cy of nitazoxanide against *Leishmania* donovani. *Parasitol. Res.* 107, 475–479 (2010) DOI: 10.1007/s00436-010-1906-y
- Mesquita, J. T., Tempone, A. G. and Reim??o, J. Q. Combination therapy with nitazoxanide and amphotericin B, Glucantime, miltefosine and sitamaquine against *Leishmania* (*Leishmania*) infantum intracellular amastigotes. *Acta Trop.* 130, 112–116 (2014) DOI: 10.1016/j.actatropica.2013.11.003
- Rodrigues, F. H., Afonso-Cardoso, S. R., Gomes, M. A. B., Beletti, M. E., Rocha, A., Guimarães, A. H. B., Candeloro, I. and de Souza, M. A. Effect of imidocarb and levamisole on the experimental infection of BALB/c mice by *Leishmania* (Leishmania) *amazonensis. Vet. Parasitol.* 139, 37–46 (2006) DOI: 10.1016/j.vetpar.2006.02.032
- Martinez-Rojano, H., Mancilla-Ramirez, J., Quiñonez-Diaz, L. and Galindo-Sevilla, N. Activity of hydroxyurea against *Leishmania* mexicana. *Antimicrob. Agents Chemother*. 52, 3642–7 (2008) DOI: 10.1128/AAC.00124-08
- Tavares, J., Ouaissi, M., Ouaissi, A. and Cordeiro-da-Silva, A. Characterization of the anti- *Leishmania* effect induced by cisplatin, an anticancer drug. *Acta Trop.* 103, 133–141 (2007)
 DOI: 10.1016/j.actatropica.2007.05.017
- Kaur, S., Sachdeva, H., Dhuria, S., Sharma, M. and Kaur, T. Antileishmanial effect of cisplatin against murine visceral leishmaniasis. *Parasitol. Int.* 59, 62–9 (2010)
 DOI: 10.1016/j.parint.2009.10.006
- 78. Sharma, M., Sehgal, R. and Kaur, S. Evaluation of nephroprotective and immunomodulatory activities of antioxidants in combination with cisplatin against murine visceral leishmaniasis. *PLoS Negl. Trop. Dis.* 6, e1629 (2012)
 DOI: 10.1371/journal.pntd.0001629
- 79. Sharma, M. and Kaur, S. Protective efficacy of antioxidants on cisplatin-induced tissue damage caused in *Leishmania* donovani infected BALB/c mice against murine visceral leishmaniasis. *J. Interdiscip. Histopathol.* 1, 121 (2013)
 DOI: 10.5455/jihp.20121219031901
- 80. Kaur, T., Makkar, P., Randhawa, K. and Kaur, S. Antineoplastic drug, carboplatin,

- protects mice against visceral leishmaniasis. *Parasitol. Res.* 112, 91–100 (2013) DOI: 10.1007/s00436-012-3108-2
- Shukla, A. K., Patra, S. and Dubey, V. K. Nanospheres Encapsulating Anti-Leishmanial Drugs for Their Specific Macrophage Targeting, Reduced Toxicity, and Deliberate Intracellular Release. Vector-Borne Zoonotic Dis. 12, 953–960 (2012) DOI: 10.1089/vbz.2011.0948
- 82. Shukla, A. K., Patra, S. and Dubey, V. K. Evaluation of selected antitumor agents as subversive substrate and potential inhibitor of trypanothione reductase: an alternative approach for chemotherapy of Leishmaniasis. *Mol. Cell. Biochem.* 352, 261–270 (2011)
 DOI: 10.1007/s11010-011-0762-0
- 83. van den Bogaart, E., Schoone, G. J., England, P., Faber, D., Orrling, K. M., Dujardin, J.-C., Sundar, S., Schallig, H. D. F. H. and Adams, E. R. Simple Colorimetric Trypanothione Reductase-Based Assay for High-Throughput Screening of Drugs against *Leishmania* Intracellular Amastigotes. *Antimicrob. Agents Chemother.* 58, 527–535 (2014) DOI: 10.1128/AAC.00751-13
- 84. Kansal, S., Tandon, R., Verma, P. R. P., Dube, A. and Mishra, P. R. Development of doxorubicin loaded novel core shell structured nanocapsules for the intervention of visceral leishmaniasis. *J. Microencapsul.* 30, 441–50 (2013) DOI: 10.3109/02652048.2012.752532
- 85. Kansal, S., Tandon, R., Verma, A., Misra, P., Choudhary, A. K., Verma, R., Verma, P. R. P., Dube, A. and Mishra, P. R. Coating doxorubicin-loaded nanocapsules with alginate enhances therapeutic efficacy against *Leishmania* in hamsters by inducing Th1-type immune responses. *Br. J. Pharmacol.* 171, 4038–50 (2014) DOI: 10.1111/bph.12754
- 86. Sett, R., Basu, N., Ghosh, A. K. and Das, P. K. Potential of Doxorubicin as an Antileishmanial Agent. *J. Parasitol.* 78, 350 (1992)
 DOI: 10.2307/3283487
- 87. Prada, C. F., Álvarez-Velilla, R., Balaña-Fouce, R., Prieto, C., Calvo-Álvarez, E., Escudero-Martínez, J. M., Requena, J. M.,

- Ordóñez, C., Desideri, A., Pérez-Pertejo, Y. and Reguera, R. M. Gimatecan and other camptothecin derivatives poison *Leishmania* DNA-topoisomerase IB leading to a strong leishmanicidal effect. *Biochem. Pharmacol.* 85, 1433–1440 (2013) DOI: 10.1016/j.bcp.2013.02.024
- 88. Doherty, T. M., Sher, A. and Vogel, S. N. Paclitaxel (Taxol)-induced killing of *Leishmania major* in murine macrophages. *Infect. Immun.* 66, 4553–6 (1998)
- Moulay, L., Robert-Gero, M., Brown, S., Gendron, M.-C. and Tournier, F. Sinefungin and Taxol Effects on Cell Cycle and Cytoskeleton of *Leishmania donovani* promastigotes. *Exp. Cell Res.* 226, 283–291 (1996) DOI: 10.1006/excr.1996.0229
- Sanderson, L., Yardley, V. and Croft, S. L. Activity of anti-cancer protein kinase inhibitors against *Leishmania* spp. *J. Antimicrob. Chemother.* 69, 1888–91 (2014)
 DOI: 10.1093/jac/dku069
- Wetzel, D. M., McMahon-Pratt, D. and Koleske, a. J. The Abl and Arg Kinases Mediate Distinct Modes of Phagocytosis and Are Required for Maximal *Leishmania* Infection. *Mol. Cell. Biol.* 32, 3176–3186 (2012) DOI: 10.1128/MCB.00086-12
- Diaz-Gonzalez, R., Kuhlmann, F. M., Galan-Rodriguez, C., da Silva, L. M., Saldivia, M., Karver, C. E., Rodriguez, A., Beverley, S. M., Navarro, M. and Pollastri, M. P. The Susceptibility of Trypanosomatid Pathogens to PI3/mTOR Kinase Inhibitors Affords a New Opportunity for Drug Repurposing. PLoS Negl. Trop. Dis. 5, e1297 (2011) DOI: 10.1371/journal.pntd.0001297
- 93. Miguel, D. C., Yokoyama-Yasunaka, J. K. U. and Uliana, S. R. B. Tamoxifen is effective in the treatment of *Leishmania amazonensis* infections in mice. *PLoS Negl. Trop. Dis.* 2, e249 (2008)

 DOI: 10.1371/journal.pntd.0000249
- 94. Eissa, M. M., Amer, E. I. and El Sawy, S. M. F. Leishmania major: activity of tamoxifen against experimental cutaneous leishmaniasis. Exp. Parasitol. 128, 382–90 (2011) DOI: 10.1016/j.exppara.2011.05.009
- 95. Miguel, D. C., Zauli-Nascimento, R. C., Yokoyama-Yasunaka, J. K. U., Katz, S.,

- Barbiéri, C. L. and Uliana, S. R. B. Tamoxifen as a potential antileishmanial agent: efficacy in the treatment of *Leishmania braziliensis* and *Leishmania* chagasi infections. *J. Antimicrob. Chemother.* 63, 365–8 (2009) DOI: 10.1093/jac/dkn509
- Miguel, D. C., Zauli-Nascimento, R. C., Yokoyama-Yasunaka, J. K. U., Pereira, L. I. A., Jerônimo, S. M. B., Ribeiro-Dias, F., Dorta, M. L. and Uliana, S. R. B. Clinical isolates of New World *Leishmania* from cutaneous and visceral leishmaniasis patients are uniformly sensitive to tamoxifen. *Int. J. Antimicrob. Agents* 38, 93–4 (2011) DOI: 10.1016/j.ijantimicag.2011.03.012
- Miguel, D. C., Yokoyama-Yasunaka, J. K. U., Andreoli, W. K., Mortara, R. A. and Uliana, S. R. B. Tamoxifen is effective against Leishmania and induces a rapid alkalinization of parasitophorous vacuoles harbouring Leishmania (Leishmania) amazonensis amastigotes. J. Antimicrob. Chemother. 60, 526–34 (2007) DOI: 10.1093/jac/dkm219
- Trinconi, C. T., Reimão, J. Q., Bonano, V. I., Espada, C. R., Miguel, D. C., Yokoyama-Yasunaka, J. K. U. and Uliana, S. R. B. Topical tamoxifen in the therapy of cutaneous leishmaniasis. *Parasitology* 1–7 (2017) DOI: 10.1017/S0031182017000130
- 99. Trinconi, C. T., Reimão, J. Q., Yokoyama-Yasunaka, J. K. U., Miguel, D. C. and Uliana, S. R. B. Combination therapy with tamoxifen and amphotericin B in experimental cutaneous leishmaniasis. *Antimicrob. Agents Chemother.* 58, 2608–13 (2014) DOI: 10.1128/AAC.01315-13
- 100. Trinconi, C. T., Reimão, J. Q., Coelho, A. C. and Uliana, S. R. B. Efficacy of tamoxifen and miltefosine combined therapy for cutaneous leishmaniasis in the murine model of infection with *Leishmania amazonensis*. *J. Antimicrob. Chemother.* 71, 1314–1322 (2016)
 DOI: 10.1093/jac/dkv495
- 101. Reimão, J. Q., Miguel, D. C., Taniwaki, N. N., Trinconi, C. T., Yokoyama-Yasunaka, J. K. U. and Uliana, S. R. B. Antileishmanial activity of the estrogen receptor modulator raloxifene. *PLoS Negl. Trop. Dis.* 8, e2842 (2014)
 DOI: 10.1371/journal.pntd.0002842

- 102. Andrade-Neto, V. V., Pereira, T. M., Canto-Cavalheiro, M. do and Torres-Santos, E. C. Imipramine alters the sterol profile in *Leishmania amazonensis* and increases its sensitivity to miconazole. *Parasit. Vectors* 9, 183 (2016)
 DOI: 10.1186/s13071-016-1467-8
- 103. Mukherjee, S., Mukherjee, B., Mukhopadhyay, R., Naskar, K., Sundar, S., Dujardin, J. C., Das, A. K. and Roy, S. Imipramine Is an Orally Active Drug against Both Antimony Sensitive and Resistant Leishmania donovani Clinical Isolates in Experimental Infection. PLoS Negl. Trop. Dis. 6, e1987 (2012)
 DOI: 10.1371/journal.pntd.0001987
- 104. Mukherjee, S., Mukherjee, B., Mukhopadhyay, R., Naskar, K., Sundar, S., Dujardin, J.-C. and Roy, S. Imipramine Exploits Histone Deacetylase 11 To Increase the IL-12/ IL-10 Ratio in Macrophages Infected with Antimony-Resistant *Leishmania* donovani and Clears Organ Parasites in Experimental Infection. *J. Immunol.* 193, 4083–4094 (2014) DOI: 10.4049/jimmunol.1400710
- 105. Zilberstein, D., Liveanu, V. and Gepstein, A. Tricyclic drugs reduce proton motive force in *Leishmania* donovani promastigotes. *Biochem. Pharmacol.* 39, 935–940 (1990) DOI: 10.1016/0006-2952(90)90210-C
- 106. Benson, T. J., McKie, J. H., Garforth, J., Borges, A., Fairlamb, A. H. and Douglas, K. T. Rationally designed selective inhibitors of trypanothione reductase. Phenothiazines and related tricyclics as lead structures. *Biochem. J.* 286 (Pt 1, 9–11 (1992)
- 107. Zilberstein, D. and Dwyer, D. M. Antidepressants cause lethal disruption of membrane function in the human protozoan parasite Leishmania. *Science* 226, 977–9 (1984)
 DOI: 10.1126/science.6505677
- 108. Cunha-Júnior, E. F., Andrade-Neto, V. V., Lima, M. L., da Costa-Silva, T. A., Galisteo Junior, A. J., Abengózar, M. A., Barbas, C., Rivas, L., Almeida-Amaral, E. E., Tempone, A. G. and Torres-Santos, E. C. Cyclobenzaprine Raises ROS Levels in *Leishmania infantum* and Reduces Parasite Burden in Infected Mice. *PLoS Negl. Trop. Dis.* 11, e0005281 (2017)

- 109. Palit, P. and Ali, N. Oral therapy with sertraline, a selective serotonin reuptake inhibitor, shows activity against *Leishmania* donovani. *J. Antimicrob. Chemother.* 61, 1120–1124 (2008)
 DOI: 10.1093/jac/dkn046
- 110. Dinesh, N., Kaur, P. K., Swamy, K. K. and Singh, S. Mianserin, an antidepressant kills *Leishmania* donovani by depleting ergosterol levels. *Exp. Parasitol.* 144, 84–90 (2014) DOI: 10.1016/j.exppara.2014.06.004
- 111. Evans, A. T., Croft, S. L., Peters, W. and Neal, R. A. Hydrazide antidepressants possess novel antileishmanial activity *in vitro* and *in vivo. Ann. Trop. Med. Parasitol.* 83, 19–24 (1989)
 DOI: 10.1080/00034983.1989.11812306
- 112. Alberca, L. N., Sbaraglini, M. L., Balcazar, D., Fraccaroli, L., Carrillo, C., Medeiros, A., Benitez, D., Comini, M. and Talevi, A. Discovery of novel polyamine analogs with anti-protozoal activity by computer guided drug repositioning. *J. Comput. Aided. Mol. Des.* 30, 305–321 (2016) DOI: 10.1007/s10822-016-9903-6
- 113. Dagger, F., Campos, Z., Rangel, H. and Roman, H. Antiproliferative effect of diazepam on *Leishmania* mexicana. *Mem. Inst. Oswaldo Cruz* 91, (1996)
- 114. Tamargo, J. and Ruilope, L. M. Investigational calcium channel blockers for the treatment of hypertension. *Expert Opin. Investig. Drugs* 25, 1295–1309 (2016) DOI: 10.1080/13543784.2016.1241764
- 115. Reimão, J. Q., Scotti, M. T. and Tempone, A. G. Anti-leishmanial and anti-trypanosomal activities of 1,4-dihydropyridines: *In vitro* evaluation and structure-activity relationship study. *Bioorganic Med. Chem.* 18, 8044–8053 (2010) DOI: 10.1016/j.bmc.2010.09.015
- 116. Reimão, J. Q. and Tempone, A. G. Investigation into in vitro anti-leishmanial combinations of calcium channel blockers and current anti-leishmanial drugs. Mem. Inst. Oswaldo Cruz 106, 1032–1038 (2011) DOI: 10.1590/S0074-02762011000800022
- 117. Reimão, J. Q., Colombo, F. A., Pereira-Chioccola, V. L. and Tempone, A. G. *In vitro* and experimental therapeutic studies of the calcium channel blocker bepridil: Detection

- of viable Leishmania (L.) chagasi by real-time PCR. Exp. Parasitol. 128, 111-115 (2011)
- DOI: 10.1016/j.exppara.2011.02.021
- 118. Shokri, A., Sharifi, I., Khamesipour, A., Nakhaee, N., Harandi, M. F., Nosratabadi, J., Parizi, M. H. and Barati, M. The effect of verapamil on in vitro susceptibility of promastigote and amastigote stages of Leishmania tropica to meglumine antimoniate. Parasitol. Res. 110, 1113-1117 (2012)

DOI: 10.1007/s00436-011-2599-6

- 119. Tempone, A. G., Taniwaki, N. N. and Reimão, J. Q. Antileishmanial activity and ultrastructural alterations of *Leishmania* (*L*.) *chagasi* treated with the calcium channel blocker nimodipine. Parasitol. Res. 105, 499-505 (2009) DOI: 10.1007/s00436-009-1427-8
- 120. Misra, S., Naskar, K., Sarkar, D. and Ghosh, D. K. Role of Ca2+ ion on Leishmaniaattachment. macrophage Mol. Biochem. 102, 13-18 (1991) DOI: 10.1007/BF00232154
- 121. Ganouly, N. K., Sodhi, S., Kaul, N., Kaur, S., Malla, N. and Mahaian, R. C. Effect of nifedipine on Leishmania donovani infection in-vivo and in-vitro: chemiluminescence responses of peritoneal macrophages and neutrophils. J. Pharm. Pharmacol. 43, 140-142 (1991)
 - DOI: 10.1111/j.2042-7158.1991.tb06652.x
- 122. Palit, P. and Ali, N. Oral therapy with amlodipine and lacidipine, 1,4-dihydropyridine derivatives showing activity against experimental visceral leishmaniasis. Antimicrob. Agents Chemother, 52, 374–377 (2008) DOI: 10.1128/AAC.00522-07
- 123. Valiathan, R., Dubey, M. L., Mahajan, R. C. and Malla, N. Leishmania donovani: Effect of verapamil on in vitro susceptibility of promastigote and amastigote stages of Indian clinical isolates to sodium stibogluconate. Exp. Parasitol. 114, 103-108 (2006) DOI: 10.1016/j.exppara.2006.02.015
- 124. Goyal, N., Duncan, R., Selvapandiyan, A., Debrabant, A., Baig, M. S. and Nakhasi, H. L. Cloning and characterization of angiotensin converting enzyme related dipeptidylcarboxypeptidase from Leishmania donovani. Mol. Biochem. Parasitol. 145, 147-157 (2006) DOI: 10.1016/j.molbiopara.2005.09.014

- 125. Andrade-Neto, V. V., Cunha-Júnior, E. F., Canto-Cavalheiro, M. M. do, Atella, G. C., Fernandes, T. de A., Costa, P. R. R. and Torres-Santos, E. C. Antileishmanial Activity of Ezetimibe: Inhibition of Sterol Biosynthesis, In vitro Synergy with Azoles, and Efficacy in Experimental Cutaneous Leishmaniasis. Antimicrob. Agents Chemother. 60, 6844–6852 (2016) DOI: 10.1128/AAC.01545-16
- 126. García-Miss, M. D. R., Mut-Martín, M. C. and Góngora-Alfaro, J. L. β-Adrenergic blockade protects BALB/c mice against infection with a small inoculum of Leishmania mexicana mexicana (LV4). Int. Immunopharmacol. 24, 59-67 (2015) DOI: 10.1016/j.intimp.2014.11.003
- 127. Karam, M. C., Merckbawi, R., Salman, S. and Mobasheri, A. Atenolol Reduces Leishmania major-Induced Hyperalgesia and TNF-α Without Affecting IL-1β or Keratinocyte Derived Chemokines (KC). Front. Pharmacol. 7, 1–10 (2016) DOI: 10.3389/fphar.2016.00022
- 128. Genestra, M., Soares-Bezerra, R. J., Gomes-Silva, L., Fabrino, D. L., Bellato-Santos, T., Castro-Pinto, D. B., Canto-Cavalheiro, M. M. and Leon, L. L. In vitro sodium nitroprusside-mediated toxicity towards Leishmania amazonensis promastigotes and axenic amastigotes. Cell Biochem. Funct. 26, 709-717 (2008) DOI: 10.1002/cbf.1496
- 129. Kawakami, N. Y., Tomiotto-Pellissier, F., Cataneo, A. H. D., Orsini, T. M., Thomazelli, A. P. F. D. S., Panis, C., Conchon-Costa, I. and Pavanelli. W. R. Sodium nitroprusside has leishmanicidal activity independent of iNOS. Rev. Soc. Bras. Med. Trop. 49, 68-73 (2016)DOI: 10.1590/0037-8682-0266-2015
- 130. Singh, S., Dinesh, N., Kaur, P. K. and
- Shamiulla, B. Ketanserin, an antidepressant, exerts its antileishmanial action via inhibition of 3-hydroxy-3-methylglutaryl coenzyme A reductase (HMGR) enzyme of Leishmania donovani. Parasitol. Res. 113, 2161-2168 (2014)
 - DOI: 10.1007/s00436-014-3868-y
- 131. Sagher, F., Zuckerman, A., Rein, C. R. and Kitchen, D. K. The effect of high concentrations of penicillin on Leishmania

- *tropica*, *in vivo* and *in vitro*. *Br. J. Dermatol*. 66, 246–251 (1954) DOI: 10.1111/j.1365-2133.1954.tb12628.x
- 132. Romero, I. C., Saravia, N. G. and Walker, J. Selective action of fluoroquinolones against intracellular amastigotes of *Leishmania* (*Viannia*) panamensis in vitro. J. Parasitol. 91, 1474–1479 (2005) DOI: 10.1645/GE-3489.1
- 133. Cortázar, T. M., Coombs, G. H. and Walker, J. Leishmania panamensis: Comparative inhibition of nuclear DNA topoisomerase II enzymes from promastigotes and human macrophages reveals anti-parasite selectivity of fluoroquinolones, flavonoids and pentamidine. Exp. Parasitol. 116, 475–482 (2007) DOI: 10.1016/j.exppara.2007.02.018
- 134. Farca, A. M., Miniscalco, B., Badino, P., Odore, R., Monticelli, P., Trisciuoglio, A. and Ferroglio, E. Canine leishmaniosis: In vitro efficacy of miltefosine and marbofloxacin alone or in combination with allopurinol against clinical strains of Leishmania infantum. Parasitol. Res. 110, 2509–2513 (2012)
 DOI: 10.1007/s00436-011-2792-7
- 135. Vouldoukis, I., Rougier, S., Dugas, B., Pino, P., Mazier, D. and Woehrlé, F. Canine visceral leishmaniasis: Comparison of *in vitro* leishmanicidal activity of marbofloxacin, meglumine antimoniate and sodium stibogluconate. *Vet. Parasitol.* 135, 137–146 (2006)
 DOI: 10.1016/j.vetpar.2005.09.003
- 136. Rougier, S., Vouldoukis, I., Fournel, S., Pérès, S. and Woehrlé, F. Efficacy of different treatment regimens of marbofloxacin in canine visceral leishmaniosis: A pilot study. *Vet. Parasitol.* 153, 244–254 (2008) DOI: 10.1016/j.vetpar.2008.01.041
- 137. Navin, T. R. and Pearson, R. D. Inhibition of *Leishmania donovani* growth by streptomycin and tobramycin. *Ann. Trop. Med. Parasitol.* 81, 731–3 (1987)
 DOI: 10.1080/00034983.1987.11812178
- 138. de Oliveira-Silva, F., de Morais-Teixeira, E. and Rabello, A. Antileishmanial activity of azithromycin against Leishmania (Leishmania) amazonensis, Leishmania (Viannia) braziliensis, and Leishmania (Leishmania) chagasi. Am. J. Trop. Med. Hyg. 78, 745–9 (2008)

- 139. Krolewiecki, A., Leon, S., Scott, P. and Abraham, D. Activity of azithromycin against *Leishmania major in vitro* and *in vivo. Am. J. Trop. Med. Hyg.* 67, 273–7 (2002) DOI: 10.4269/ajtmh.2002.67.273
- 140. Balcioglu, I. C., Ok, U. Z., Ozbel, Y., Girginkardesler, N. and Ozbilgin, A. The *in vitro* Effects of Azithromycin and Clarithromycin on Promastigotes and Amastigotes of *Leishmania* tropica. *Kafkas Univ. Vet. Fak. Derg.* 18, A115–A120 (2012)
- 141. Sinagra, Á., Luna, C., Abraham, D., Iannella, M. del C., Riarte, A. and Krolewiecki, A. J. The activity of azithromycin against *Leishmania* (*Viannia*) braziliensis and *Leishmania* (*Leishmania*) amazonensis in the golden hamster model. Rev. Soc. Bras. Med. Trop. 40, 627–630 (2007) DOI: 10.1590/S0037-86822007000600005
- 142. Sampaio, R. N. R., Lucas, I. C. and Costa Filho, A. V. da. The use of azythromycin and N-methyl glucamine for the treatment of cutaneous Leishmaniasis caused by *Leishmania* (*Leishmania*) amazonensis in C57BL6 mice. *An. Bras. Dermatol.* 84, 125–8 (2009) DOI: 10.1590/S0365-05962009000200004
- 143. Amer, E. I., Eissa, M. M. and Mossallam, S. F. Oral azithromycin versus its combination with miltefosine for the treatment of experimental Old World cutaneous leishmaniasis. *J. Parasit. Dis.* 40, 475–484 (2016) DOI: 10.1007/s12639-014-0529-0
- 144. Roy, K., Das, S., Mondal, S., Roy, A. K. and Bera, T. The *in vitro* effect of clarithromycin on amastigote of *Leishmania* donovani. *Int. J. Drug Dev. Res.* 5, (2013)
- 145. Barteselli, A., Casagrande, M., Basilico, N., Parapini, S., Rusconi, C. M., Tonelli, M., Boido, V., Taramelli, D., Sparatore, F. and Sparatore, A. Clofazimine analogs with antileishmanial and antiplasmodial activity. *Bioorganic Med. Chem.* 23, 55–65 (2015) DOI: 10.1016/j.bmc.2014.11.028
- 146. Neal, R. A. and Croft, S. L. An in-vitro system for determining the activity of compounds against the intracellular amastigote form of *Leishmania donovani*. *J. Antimicrob. Chemother*. 14, 463–475 (1984)
 DOI: 10.1093/jac/14.5.463
- 147. Arbiser, J. L. and Moschella, S. L. Clofazimine: A review of its medical uses

- and mechanisms of action. *J. Am. Acad. Dermatol.* 32, 241–247 (1995) DOI: 10.1016/0190-9622(95)90134-5
- 148. Katiyar, S. K. and Edlind, T. D. Enhanced antiparasitic activity of lipophilic tetracyclines: role of uptake. *Antimicrob. Agents Chemother.* 35, 2198–2202 (1991) DOI: 10.1128/AAC.35.11.2198
- 149. Bano, P. and Shahab, S. M. A combination of sulphadiazine, trimethoprim and metronidazole or tinidazole in kala-azar. *J. Assoc. Physicians India* 42, 535–6 (1994)
- 150. Berman, J. D. and Lee, L. S. Activity of oral drugs against *Leishmania tropica* in human macrophages *in vitro*. *Am J Trop Med Hyg* 32, 947–951 (1983) DOI: 10.4269/ajtmh.1983.32.947
- 151. Livshin, R., Weinrauch, L., Even-Paz, Z. and El-On, J. Efficacy of Rifampicin and Isoniazid in Cutaneous Leishmaniasis. *Int. J. Dermatol.* 26, 55–59 (1987) DOI: 10.1111/j.1365-4362.1987.tb04578.x
- 152. Arora, S. K., Sinha, R. and Sehgal, S. Use of *in vitro* method to assess different brands of anti-leishmanial drugs. *Med. Microbiol. Immunol.* 180, 21–27 (1991) DOI: 10.1007/BF00191697
- 153. El-On, J., Pearlman, E., Schnur, L. F. and Greenblatt, C. L. Chemotherapeutic activity of rifampicin on leishmanial amastigotes and promastigotes *in vitro. Isr. J. Med. Sci.* 19, 240–5 (1983)
- 154. El-On, J., Messer, G. and Greenblatt, C. L. Growth inhibition of *Leishmania* tropica amastigotes *in vitro* by rifampicin combined with amphotericin B. *Ann Trop Med Parasitol* 78, 93–98 (1984) DOI: 10.1080/00034983.1984.11811782
- 155. Peters, W., Shaw, J. J., Lainson, R., Robinson, B. L. and Leão, A. F. Potentiating action of rifampicin and isoniazid against *Leishmania mexicana amazonensis. Lancet* 317, 1122–1124 (1981)
 DOI: 10.1016/S0140-6736(81)92296-0
- 156. Patterson, S., Wyllie, S., Norval, S., Stojanovski, L., Simeons, F. R., Auer, J. L., Osuna-Cabello, M., Read, K. D. and Fairlamb, A. H. The anti-tubercular drug delamanid as a potential oral treatment for visceral leishmaniasis. *Elife* 5, 1–21 (2016) DOI: 10.7554/eLife.09744

- 157. Peniche, A. G., Renslo, A. R., Melby, P. C. and Travi, B. L. Antileishmanial Activity of Disulfiram and Thiuram Disulfide Analogs in an *Ex vivo* Model System Is Selectively Enhanced by the Addition of Divalent Metal Ions. *Antimicrob. Agents Chemother.* 59, 6463–70 (2015)
 DOI: 10.1128/AAC.05131-14
- 158. Yau, W.-L., Blisnick, T., Taly, J.-F., Helmer-Citterich, M., Schiene-Fischer, C., Leclercq, O., Li, J., Schmidt-Arras, D., Morales, M. A., Notredame, C., Romo, D., Bastin, P. and Späth, G. F. Cyclosporin A Treatment of *Leishmania* donovani Reveals Stage-Specific Functions of Cyclophilins in Parasite Proliferation and Viability. *PLoS Negl. Trop. Dis.* 4, e729 (2010) DOI: 10.1371/journal.pntd.0000729
- 159. Meissner, U., Jüttner, S., Röllinghoff, M. and Gessner, A. Cyclosporin A-mediated killing of *Leishmania major* by macrophages is independent of reactive nitrogen and endogenous TNF-alpha and is not inhibited by IL-10 and 13. *Parasitol. Res.* 89, 221–7 (2003)
- 160. Pinto, E. G., da Costa-Silva, T. A. and Tempone, A. G. Histamine H1-receptor antagonists against *Leishmania* (L.) infantum: an *in vitro* and *in vivo* evaluation using phosphatidylserine-liposomes. *Acta Trop.* 137, 206–10 (2014) DOI: 10.1016/j.actatropica.2014.05.017
- 161. Jiang, S., Meadows, J., Anderson, S. A. and Mukkada, A. J. Antileishmanial Activity of the Antiulcer Agent Omeprazole. *Antimicrob. Agents Chemother.* 46, 2569–2574 (2002) DOI: 10.1128/AAC.46.8.2569-2574.2002
- 162. Koutinas, A. F., Saridomichelakis, M. N., Mylonakis, M. E., Leontides, L., Polizopoulou, Z., Billinis, C., Argyriadis, D., Diakou, N. and Papadopoulos, O. A randomised, blinded, placebo-controlled clinical trial with allopurinol in canine leishmaniosis. *Vet. Parasitol.* 98, 247–261 (2001) DOI: 10.1016/S0304-4017(01)00399-5
- 163. Velez, I., Agudelo, S., Hendrickx, E., Puerta, J., Grogl, M., Modabber, F. and Berman, J. Inefficacy of allopurinol as monotherapy for Colombian cutaneous leishmaniasis. A randomized, controlled trial. *Ann. Intern. Med.* 126, 232–6 (1997) DOI: 10.7326/0003-4819-126-3-199702010-00010
- 164. Sabaté, D., Llinás, J., Homedes, J., Sust, M. and Ferrer, L. A single-centre, open-label,

- controlled, randomized clinical trial to assess the preventive efficacy of a domperidone-based treatment programme against clinical canine leishmaniasis in a high prevalence area. *Prev. Vet. Med.* 115, 56–63 (2014)
- DOI: 10.1016/j.prevetmed.2014.03.010
- 165. Kochar, D. K., Saini, G., Kochar, S. K., Sirohi, P., Bumb, R. A., Mehta, R. D. and Purohit, S. K. A double blind, randomised placebo controlled trial of rifampicin with omeprazole in the treatment of human cutaneous leishmaniasis. J. Vector Borne Dis. 43, 161–7 (2006)
- 166. Silva-Vergara, M. L., Silva, L. D. A., Maneira, F. R. Z., Da Silva, A. G. and Prata, A. Azithromycin in the treatment of mucosal Leishmaniasis. *Rev. Inst. Med. Trop. Sao Paulo* 46, 175–177 (2004) DOI: 10.1590/S0036-46652004000300011
- 167. Teixeira, A. C., Paes, M. G., Guerra, J. D. O., Prata, A. and Silva-Vergara, M. L. Low efficacy of azithromycin to treat cutaneous leishmaniasis in Manaus, AM, Brazil. Rev. Inst. Med. Trop. Sao Paulo 49, 235–8 (2007) DOI: 10.1590/S0036-46652007000400008
- 168. Daoud, S. and Boushi, L. Azithromycin, ineffective in the treatment of old-world cutaneous leishmaniasis. *Int. J. Dermatol.* 45, 1126–1128 (2006)
 DOI: 10.1111/j.1365-4632.2006.02885.x
- 169. Krolewiecki, A. J., Romero, H. D., Cajal, S. P., Abraham, D., Mimori, T., Matsumoto, T., Juarez, M. and Taranto, N. J. A randomized clinical trial comparing oral azithromycin and meglumine antimoniate for the treatment of American cutaneous leishmaniasis caused by *Leishmania* (Viannia) *braziliensis*. *Am. J. Trop. Med. Hyg.* 77, 640–6 (2007)
- 170. Khatami, A., Firooz, A., Gorouhi, F. and Dowlati, Y. Treatment of acute Old World cutaneous leishmaniasis: A systematic review of the randomized controlled trials. *J. Am. Acad. Dermatol.* 57, 335.e1–335.e29 (2007) DOI: 10.1016/j.jaad.2007.01.016
- 171. Nassiri-Kashani, M., Firooz, A., Khamesipour, A., Mojtahed, F., Nilforoushzadeh, M., Hejazi, H., Bouzari, N. and Dowlati, Y. A randomized, double-blind, placebo-controlled clinical trial of itraconazole in the treatment of cutaneous leishmaniasis. *J. Eur. Acad. Dermatol. Venereol.* 19, 80–3 (2005) DOI: 10.1111/j.1468-3083.2004.01133.x

- 172. Dogra, J., Aneja, N., Lal, B. B. and Mishra, S. N. Cutaneous Leishmaniasis In India. *Int. J. Dermatol.* 29, 661–662 (1990) DOI: 10.1111/j.1365-4362.1990.tb02593.x
- 173. Salmanpour, R., Handjani, F. and Nouhpisheh, M. K. Comparative study of the efficacy of oral ketoconazole with intra-lesional meglumine antimoniate (Glucantime) for the treatment of cutaneous leishmaniasis. *J. Dermatolog. Treat.* 12, 159–162 (2001) DOI: 10.1080/09546630152607899
- 174. El-Sayed, M. and Anwar, A. Intralesional sodium stibogluconate alone or its combination with either intramuscular sodium stibogluconate or oral ketoconazole in the treatment of localized cutaneous leishmaniasis: A comparative study. *J. Eur. Acad. Dermatology Venereol.* 24, 335–340 (2010) DOI: 10.1111/j.1468-3083.2009.03417.x
- 175. Navin, T. R., Arana, B. A., Arana, F. E., Berman, J. D. and Chajon, J. F. Placebo-Controlled Clinical Trial of Sodium Stibogluconate (Pentostam) versus Ketoconazole for Treating Cutaneous Leishmaniasis in Guatemala. *J. Infect. Dis.* 165, 528–534 (1992) DOI: 10.1093/infdis/165.3.528
- 176. Newlove, T., Guimaraes, L. H., Morgan, D. J., Alcantara, L., Glesby, M. J., Carvalho, E. M. and Machado, P. R. Antihelminthic Therapy and Antimony in Cutaneous Leishmaniasis: A Randomized, Double-Blind, Placebo-Controlled Trial in Patients Co-Infected with Helminths and *Leishmania braziliensis*. *Am. J. Trop. Med. Hyg.* 84, 551–555 (2011) DOI: 10.4269/ajtmh.2011.10-0423
- 177. Prates, F. V. D. O., Dourado, M. E. F., Silva, S. C., Schriefer, A., Guimarães, L. H., Brito, M. D. G. O., Almeida, J., Carvalho, E. M. and Machado, P. R. L. Fluconazole in the Treatment of Cutaneous Leishmaniasis Caused by *Leishmania braziliensis*: A Randomized Controlled Trial. *Clin. Infect. Dis.* 64, 67–71 (2017) DOI: 10.1093/cid/ciw662
- 178. Emad, M., Hayati, F., Fallahzadeh, M. K. and Namazi, M. R. Superior efficacy of oral fluconazole 400 mg daily versus oral fluconazole 200 mg daily in the treatment of cutaneous *Leishmania major* infection: A randomized clinical trial. *J. Am. Acad. Dermatol.* 64, 606–608 (2011) DOI: 10.1016/j.jaad.2010.04.014

- 179. Farajzadeh, S., Heshmatkhah, A., Vares, B., Mohebbi, E., Mohebbi, A., Aflatoonian, M., Eybpoosh, S., Sharifi, I., Aflatoonian, M. R., Shamsi Meymandi, S., Fekri, A. R. and Mostafavi, M. Topical terbinafine in the treatment of cutaneous leishmaniasis: triple blind randomized clinical trial. *J. Parasit. Dis.* 40, 1159–1164 (2016)
 DOI: 10.1007/s12639-014-0641-1
- 180. Firooz, A., Khamesipour, A., Ghoorchi, M. H., Nassiri-Kashani, M., Eskandari, S. E., Khatami, A., Hooshmand, B., Gorouhi, F., Rashighi-Firoozabadi, M. and Dowlati, Y. Imiquimod in combination with meglumine antimoniate for cutaneous leishmaniasis. *Arch. Dermatol.* 142, 1575–1579 (2006) DOI: 10.1001/archderm.142.12.1575
- 181. Meng, Y., Squires, H., Stevens, J. W., Simpson, E., Harnan, S., Thomas, S., Michaels, J., Stansby, G. and O'Donnell, M. E. Cost-Effectiveness of Cilostazol, Naftidrofuryl Oxalate, and Pentoxifylline for the Treatment of Intermittent Claudication in People With Peripheral Arterial Disease. *Angiology* 65, 190–197 (2014) DOI: 10.1177/0003319712474335
- 182. Brito, G., Dourado, M., Polari, L., Celestino, D., Carvalho, L. P., Queiroz, A., Carvalho, E. M., Machado, P. R. L. and Passos, S. Clinical and immunological outcome in cutaneous leishmaniasis patients treated with pentoxifylline. *Am. J. Trop. Med. Hyg.* 90, 617–620 (2014) DOI: 10.4269/ajtmh.12-0729
- 183. Sadeghian, G. and MA, N. The effect of combination therapy with systemic meglumine antimoniate (Glucantime) and pentoxifylline the treatment of cutaneous leishmaniasis. *Iran. J. dermatology* 9, 2 (2006)
- 184. Báfica, A., Oliveira, F., Freitas, L. A. R., Nascimento, E. G. and Barral, A. American Cutaneous Leishmaniasis unresponsive to antimonial drugs: Successful treatment using combination of N-methilglucamine antimoniate plus pentoxifylline. *Int. J. Dermatol.* 42, 203–207 (2003) DOI: 10.1046/j.1365-4362.2003.01868.x
- 185. Almeida, O. L. S. and Santos, J. B. Advances in the treatment of cutaneous leishmaniasis in the new world in the last ten years: a systematic literature review. *An. Bras. Dermatol.* 86, 497–506 (2011) DOI: 10.1590/S0365-05962011000300012

- 186. Lessa, H. A., Machado, P., Lima, F., Cruz, Á. A., Bacellar, O., Guerreiro, J. and Carvalho, E. M. Successful treatment of refractory mucosal leishmaniasis with pentoxifylline plus antimony. *Am. J. Trop. Med. Hyg.* 65, 87–89 (2001)

 DOI: 10.4269/ajtmh.2001.65.87
- 187. Machado, P. R. L., Lessa, H., Lessa, M., Guimaraes, L. H., Bang, H., Ho, J. L. and Carvalho, E. M. Oral Pentoxifylline Combined with Pentavalent Antimony: A Randomized Trial for Mucosal Leishmaniasis. *Clin. Infect. Dis.* 44, 788–793 (2007) DOI: 10.1086/511643
- 188. Bahia, M. T., de Andrade, I. M., Martins, T. A. F., Nascimento, Á. F. da S. do, Diniz, L. de F., Caldas, I. S., Talvani, A., Trunz, B. B., Torreele, E. and Ribeiro, I. Fexinidazole: A Potential New Drug Candidate for Chagas Disease. *PLoS Negl. Trop. Dis.* 6, (2012) DOI: 10.1371/journal.pntd.0001870
- 189. Burri, C. Chemotherapy against human African trypanosomiasis: Is there a road to success? *Parasitology* 137, 1987–1994 (2010)
 DOI: 10.1017/S0031182010001137
- 190. Kaiser, M., Bray, M. A., Cal, M., Trunz, B. B., Torreele, E. and Brun, R. Antitrypanosomal activity of fexinidazole, a new oral nitro-imidazole drug candidate for treatment of sleeping sickness. *Antimicrob. Agents Chemother.* 55, 5602–5608 (2011) DOI: 10.1128/AAC.00246-11
- 191. Singh Grewal, A., Pandita, D., Bhardwaj, S. and Lather, V. Recent Updates on Development of Drug Molecules for Human African Trypanosomiasis. *Curr. Top. Med. Chem.* 16, 2245–2265 (2016) DOI: 10.2174/15680266166661604131253
- 192. Wyllie, S., Patterson, S., Stojanovski, L., Simeons, F. R. C., Norval, S., Kime, R., Read, K. D. and Fairlamb, a. H. The Anti-Trypanosome Drug Fexinidazole Shows Potential for Treating Visceral Leishmaniasis. *Sci. Transl. Med.* 4, 119re1–119re1 (2012)
- 193. Wyllie, S., Patterson, S. and Fairlamb, A. H. Assessing the essentiality of *Leishmania* donovani nitroreductase and its role in nitro drug activation. *Antimicrob. Agents Chemother.* 57, 901–906 (2013) DOI: 10.1128/AAC.01788-12

194. ClinicalTrials.gov. Trial to Determine Efficacy of Fexinidazole in Visceral Leihmaniasis Patients in Sudan. at https://clinicaltrials.gov/ct2/show/NCT01980199?term=%22fex inidazole%22+AND+%22leishmaniasis%22 &rank=1

Key Words: Leishmaniasis, Treatment; Drug Repurposing, Review

Send correspondence to: Eduardo Caio Torres-Santos, Laboratorio de Bioquimica de Tripanosomatideos, Instituto Oswaldo Cruz, FIOCRUZ, Rio de Janeiro, Brasil, Tel: 55–21–3865–8247, Fax: 55–21–38362141, E-mail: ects@ioc.fiocruz.br