Nanoparticles in wound healing; from hope to promise, from promise to routine

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ABSTRACT

Chronic non-healing wounds represent a growing problem due to their high morbidity and cost. Despite recent advances in wound healing, several systemic and local factors can disrupt the weighed physiologic healing process. This paper critically reviews and discusses the role of nanotechnology in promoting the wound healing process. Nanotechnologybased materials have physicochemical, optical and biological properties unique from their bulk equivalent. These nanoparticles can be incorporated into scaffolds to create nanocomposite smart materials, which promote wound healing through their antimicrobial. as well as selective anti- and pro-inflammatory, and pro-angiogenic properties. Owed to their high surface area, nanoparticles have also been used for drug delivery as well as gene delivery vectors. In addition, nanoparticles affect wound healing by influencing

collagen deposition and realignment and provide approaches for skin regeneration and wound healing.

1. INTRODUCTION

Wounds result from disruption of the normal anatomical epithelial lined tissue barriers and may be caused by trauma, tissue resection, or burns (6). Some wounds fail to heal in a timely fashion and become chronic as a result of co-existing conditions such as diabetes or peripheral vascular disease. Failure to heal might also result from post-operative wound infections which are estimated to affect up to 4% of patients who undergo surgery. Chronic non-healing wounds represent a growing health and economic burden and are associated with a high morbidity that adds significantly to the cost of medical care (7). The gold

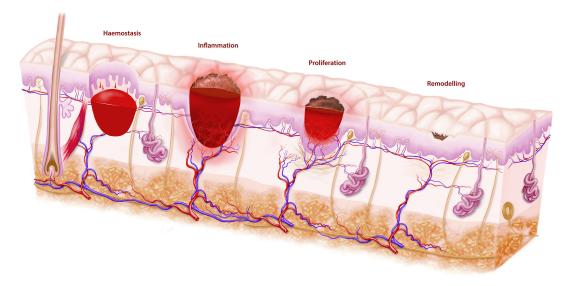


Figure 1. Schematic depiction of nanoparticles.

standard for the treatment of non-healing skin wounds is the transplantation of autologous skin. This strategy, however, might not be suitable in certain cases due to the lack of a donor site. In such cases, engineered skin substitutes are an alternative to autologous skin transplantation. Clearly, there is a need to develop strategies to promote wound healing and prevent scarring (8).

The use of cell therapy with or without of growth factors in experimental models has prevailed some positive results, but these therapies have not moved to clinical setting due to complications in scalable fabrication and storage, high costs, regulatory issues, and lack of standardisation. Moreover, their effectiveness and safety have not been demonstrated fully.

Nanotechnology is a rapidly expanding multidisciplinary scientific field, which combines the disciplines of material science and engineering. Nanoparticles (NPs), usually ranging in dimension from 1-100 nanometers (nm), have properties unique from their bulk equivalent. They possess unique physicochemical, optical and biological properties, which can be manipulated suitable for desired applications. Since ancient times, elements such as silver, gold, copper and titanium were used to treat a number of human conditions. More recently, researchers have developed insight in and awareness of nanoparticles and how these could be used for drug delivery, diagnostic and imaging, biosensor, and cosmetic purposes (9). Several nanomaterials for biological applications have been intensively investigated during the last several decades. These have included liposomes, dendrimers, quantum dots, fullerenes, carbon nanotubes, graphene, iron and titanium oxide, and gold and silver nanoparticles

(Figure 1). Recently NP-based delivery of ions, such as calcium and oxygen has been used to promote angiogenesis (10). The application of nanomaterial-based scaffold with controlled delivery of calcium ions or oxygen would promote differentiation of ADSC to endothelial cells and angiogenesis (10).

Nanoparticles can be incorporated into biomaterials and scaffolds to create nanocomposite smart materials (Figure 1), which can aid wound healing through their antimicrobial (1), selective anti- and proinflammatory (2), and pro-angiogenic properties (3). They can be used as gene delivery vectors altering intracellular gene expression and protein synthesis related to the wound healing process (4). In addition, they can affect the wound healing process by influencing collagen deposition and realignment (5).

Wound healing either occurs by primary intention, where the wound edges are approximated and sutured, or by secondary intention, where the wound is left open to heal by a combination of granulation tissue formation, contraction, and reepithelialisation. The wound healing process includes the subsequent and overlapping phases of haemostasis, inflammation, proliferation, and remodelling (Figure 2) (11). Haemostasis involves vasoconstriction, the formation of a platelet plug, and platelet degranulation. Inflammation occurs in the first two to three days after injury and involves the release of pro-inflammatory factors by platelets, which enhance inflammatory cell proliferation and migration. The proliferation phase overlaps the inflammatory phase and lasts up to 4 weeks. Here, inflammatory cells release chemoattractants to fibroblasts, which migrate into the wound to deposit ground substance, type III collagen, and elastin. Angiogenesis occurs simultaneously. Finally, remodelling can last up to a year or longer and involves

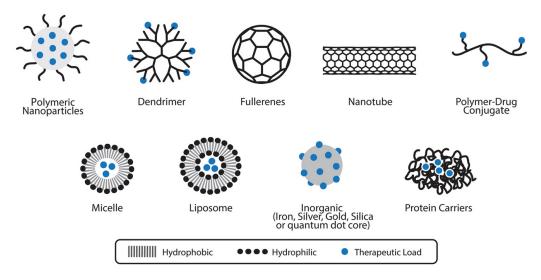


Figure 2. The four stages of wound healing; haemostasis, inflammation, proliferation, and remodelling.

re-arrangement and organisation of collagen fibres, as well as replacement of type III by type I collagen. It is a fine equilibrium between the inflammatory, proliferative, and remodelling phases that results in satisfactory wound healing.

Systemic and local factors that disrupt the weighed physiologic healing process can impede wound healing. Systemic factors are either congenital or acquired. Congenital factors include a range of genetic disorders associated with defective collagen synthesis, increased collagen degradation, defective elastin synthesis, prelamin an accumulation, and increased telomere decay. Acquired systemic factors include conditions such as diabetes mellitus, smoking, old age, vitamin deficiencies, and use of anti-inflammatory drugs. Examples of local factors are an infection, radiation, trauma, and poor tissue blood and neural supply.

Contrarily, excessive scarring following injury can result from a disruption in the equilibrium between the different wound healing phases. Keloids and hypertrophic scars differ from the healthy skin by a rich vasculature, high mesenchymal cell density, and thickened epidermal cell layer (12). In addition, they contain an abnormally high density of fibroblasts and unidirectional collagen fibrils (12). A prolonged or excessive inflammatory phase is believed to cause the onset of excessive scarring. Keloid scars can cause pain, pruritis, contractions, and are generally unaesthetic. Current treatment options include massage therapy. pressure garments, silicone gel sheeting, intralesional corticosteroid/5-fluorouracil injection, laser therapy, cryotherapy, radiotherapy, and surgery.

Both lack of appropriate healing and excessive scarring remain a common concern and

an on-going challenge for clinicians. The incidence of refractory wounds is rising as a consequence of the ageing population, making the improvement of wound treatment a major healthcare issue (13). Current advances in wound healing aim to enhance regeneration and decrease scarring.

Nanotechnology has the potential to revolutionise the treatment of wounds through therapeutically active wound dressings using nanoparticles for the delivery of drugs, growth factors and pro-angiogenesis compounds such as calcium ions. Table 1 summarises recent studies investigating *in vivo* application of NPs in wound healing.

2. NANOPARTICLES IN WOUND HEALING

2.1. Nanoparticles with antimicrobial properties

Challenges facing the management of refractory wounds are often associated with microbial contamination and infection. The eradication of these is crucial for timely wound healing (33). The rise of multi-drug resistant pathogens has led to increasing use of nanoparticle-based anti-microbial remedies.

Currently, the metallic nanoparticles are thoroughly explored and extensively investigated as potential antimicrobials. The antimicrobial activity of the nanoparticles is known to be a function of the surface area in contact with the microorganisms.

2.1.1. Anti-microbial activity of silver nanoparticles (AgNP)

The most vastly investigated nanoparticle with antimicrobial properties is silver. Several studies have confirmed the efficacy of AgNP and biomaterial

Table 1. Nanoparticles and wound healing preclinical studies.

Nanoparticle/ Nanoscaffold	Preclinical model	Wound	Procedure	Outcome	
PLA nanosheets with AgSD	Mouse	Partial-thickness burns	Antimicrobial properties and cell viability assays	AgSD significantly reduced MRSA contamination both <i>in vivo</i> and <i>in vitro</i>	
Pectin/copper exchanged faujasite	Rat and NIH3T3 fibroblast cell line	Burns	Assessment of membrane morphology, thermal stability, swelling and degradation	Cell viability of 89% was achieved, + improvement in wound healing & re-epithelialisation	
NAC-SNO-NPs	Mouse	Burns	Histological examination of burn wounds for collagen deposition	Acceleration of the transition from inflammatory to proliferative wound healing	
Quantum dots	Mouse	Laceration	In vivo optical system for assessment of wound healing.	Effective system for visualisation of wound healing.	
Gold NPs (Au NPs)	Rat	Burns	Wound healing with Au NPs with microcurrent	Improved tissue repair due to enhanced mitochondrial function	
Gelatin NPs	Rat	Full thickness laceration	Collagen and hyaluronic acid nanofibrous skin equivalent, with controllable release of angiogenetic factors	Acceleration of wound closure rate and elevated collagen deposition	
AgNPs	Rat	Excision wound	Microwave irradiation of Naringi crenulata leaf extracts to synthesise bioactive AgNPs	Very effective wound repair and potential for tropical wounds	
AgNPs coated with BC nanofibers	Rat	Partial thickness wound	Investigation of AgNP-BC for antibacterial properties and cytocompatibility	Reduction in inflammation and promotion of scald wound healing	
Hypericin nanoparticles (HYNPs)	Rat	Infected excision wound	Antibacterial activity of hypericin	Improved epithelialisation, keratinisation, collagen deposition	
Gelatin nanofibres	Rat	Excision wound	Development of gelatin nanofibrous mat loaded with epigallocatechin gallate / polyvinyl alcohol hydrogel	Significant increased in angiogenesis, re-epithilialisation and collagen synthesis	
Pirfenidone NPs	Rat	Alkali burn in cornea	Assessment of corneal re-epithelialisation, haze and collagen deposition	Reduced collagen synthesis, prevented scarring/fibrosis, +improved corneal healing	
Copper and zinc NPs	Rat	Soft tissue full layer excision wound	Wounds were either aseptic or infected	Regeneration attributed to antibacterial properties	
Fibrin NPs coated with chitosan	Rat	Excision wound	Swelling, biodegradation, porosity, platelet activation and blood clotting	Faster wound healing and re- epithelialisation	
AgNPs in alginate fibres	Mouse	Excision wound (2cm)	Investigation of AgNPs alone and AgNPs in alginate fibres with regards to wound healing	AgNPs in alginate fibres promoted fibroblast migration to the wound and increased epidermal	
Elastin-like peptides & KGF (self-assembly)	Mouse	Excision wound	Assessment of efficacy of NPs in wounds of diabetic mice	These NPs offer a beneficial effect on chronic wounds	
Fullerenes (carbon nanospheres)	Mouse & human skin, ex vivo	Skin irritation	Anti-inflammatory and anti-oxidant properties of fullerenes	Cell migration mediated human wound closure. Accelerated wound healing	
AgNPs as a dressing	Dog	Severe burns (50% of TBSA)	AgNPs along with VAC dressing were assessed in wound healing	VAC and AgNPs successfully treated the dog	
Mesoporous Silica NPs (MSN)	Rat	Achilles tendon injury	The effect of PDGF administration via MSN	Significant faster healing with PDGF incorporation	
Lecithin NPs	Rat	Burns	Dihydroquecetin immobilised with lecithin NPs for healing of burns	Limitation of secondary necrotic zones in wounds and improvement of skin regeneration	

Abbreviations: AgSD, silver sulfadiazine; PLA, poly (lactic acid); NPs, nanoparticles; NAC-SNO-NPs, N-acetylcysteine S-nitrosothiol nanoparticles; KGF, keratinocyte growth factor; VAC, vacuum assisted closure; TBSA, total body surface area; HA, hyaluronic acid.; BC, bacterial cellulose.

composites against bacterial contamination and infection (34-39). In terms of assessing the antibacterial efficacy of AgNPs of different sizes and surface conditions against Escherichia coli, Ag-resistant E. coli, Staphylococcus aureus, methicillin-resistant S.

aureus (MRSA), and Salmonella sp, AgNP synthesized by base reduction with unmodified surfaces (sizes: 20, 50 and 80 nm) are toxic to all bacterial strains. AgNPs synthesised by base reduction followed by phosphate buffer washes (sizes: 20, 50 and 80 nm) and carboncoated AgNPs (sizes: 25 and 35 nm) are toxic to all bacterial strains except Aq-resistant E. coli (40). Stable silver nanoparticles (AgNPs) generated via the active involvement of Bryonia laciniosa have shown antibacterial activity against both Gram negative and positive bacteria with no cytotoxicity observed in vitro (41). In addition, they lead to effective cytokine modulation. Preclinical wound healing showed AgNPs induced improved wound contracting ability in rats (41). Furthermore, AgNPs and carboxymethylcellulose gel formulation prepared by the reduction of silver nitrate in situ examined in simulated wound experiments showed that the gel was effective against the growth of both Gram-negative and positive strains including methicillin-resistant Staphylococcus aureus (MRSA) (42). Proliferation studies of human skin cells confirmed cytocompatibility of the composite (42).

In addition to anti-bacterial properties, AgNPs have also been shown to be effective against viral and fungal pathogens, including hepatitis B and HIV viruses (43-47). It is generally believed that various forms of Ag inactivate viruses by denaturing enzymes via reactions with carboxyl, amino, sulfhydryl, phosphate, and imidazole groups (48-52).

2.1.2. AgNPs combined with matrices as topical wound dressings

Silver-based dressings are currently in clinical use and have been evaluated thoroughly, including Ag-alginates, Ag-collagen preparations, Ag-hydrogels, Ag-hydrocolloids, Ag-fabrics, Ag-foams, and Ag creams and powders. However, the type of silver in many of these dressings is not specified. Several use ionic Ag whereas Acticoat uses nanocrystalline Ag. A retrospective analysis of burn wounds managed with Acticoat in humans showed a significant reduction in wound healing times for deep partial thickness burns compared to conventional paraffin gauze dressings (53). Generally, AqNP based dressings have shown superior results to more traditional Ag dressings such as Ag sulfadiazine cream (36, 54). Due to their beneficial antimicrobial activity and cytocompatibility. AgNPs often combined with hydrogels as silver nanocomposites have been widely investigated as antimicrobial wound dressings.

The use of nanocrystalline silver dressing (Acticoat) for the management of microbial contamination in cultured skin substitutes grafted to full-thickness wounds in athymic mice suggested that Acticoat may be suitable as a protective dressing to reduce contamination of cultured skin substitutes (55). AgNP polyvinyl alcohol (PVA) nanocomposite fibres for wound healing purposes showed significant inhibition of Gram-positive and negative bacteria (56). Preclinical studies combining AgNPs with a wide range of electrospun biomaterials as wound

dressing materials have confirmed the antimicrobial properties of these constructs, as well as their positive effects on expediting the wound healing process. Electrospun poly (dopamine methacrylamide-comethyl methacrylate) nanofibres functionalised with AgNPs through catechol redox chemistry showed effective AqNPs size and amount control with the minimum degree of aggregation. These dressings showed desirable antimicrobial activity against Grampositive and negative bacteria. Following a rapid AgNP release in the first 24 hours, a sustained release was observed in the next 5 days. Preclinical study of the nanocomposite in full thickness skin wounds in rats showed expedited healing compared to controls (57). AgNP incorporated in the electrospun scaffold of a copolymer blend showed that a nanofibre membrane with good hydrophilicity and high porosity considerably facilitates in vivo wound healing especially at the early healing stage (58). Fibrous mats of electrospun poly (vinyl alcohol), chitosan oligosaccharides, and AgNP were also shown to accelerate in vivo wound healing over that of control gauze (37).

In addition to elecrospinning, scaffolds fabricated using other techniques combined with AgNPs have been evaluated preclinically. Silver and chitosan nanocomposite dressings, fabricated using a nanometre and self-assembly technology, tested on rats with deep partial thickness wounds showed significantly increased the rate of wound healing compared to Ag sulfadiazine with lower Ag levels in blood and tissues (59). Cellulose-chitosan-AqNP composite wound dressing showed faster wound healing in experimental wounds of rats compared to untreated control (60). Comparison between ionic and nanocrystalline silver and distilled water dressings showed increased wound contracture in rats treated with silver-based dressings (61). Similarly, guar gum alkylamine impregnated with AgNPs evaluated in rodents showed faster healing compared to a commercially available silver alginate cream (62). The nano biomaterial was observed to promote wound closure by inducing proliferation and migration of the keratinocytes at the wound site (62). Topical application of silver nanoparticles prepared from Naringi crenulata leaf extracts and evaluated preclinically in rats showed accelerated healing (20). AgNP-containing activated carbon fibres exhibited good biocompatibility in vitro and improved healing and collagen and granulation tissue deposition of infected wounds in vivo (38), AqNP incorporated into alginate fibres were shown to promote fibroblast migration, reduce inflammation, and improve wound healing both preclinical and in vitro (27). In vivo evaluation of AgNPs for wound healing showed these to exhibit antimicrobial properties in addition to the reduction in wound inflammation and modulation of fibrogenic cytokines (39). Biosynthesis of AgNPs using the Phytophthora infestans microorganisms showed stability and enhanced wound contraction

ability in an *in vivo* excision wound model compared to Ag sulfadiazine (54). Synthesis of AgNPs using two glycosaminoglycans (chondroitin sulfate and acharan sulfate) as reducing agents supported the stability of these composites without any noticeable aggregation. A murine model of wound healing demonstrated that topical application of these nanocomposites stimulated wound closure and accelerated the deposition of granulation tissue and collagen at much lower Ag concentrations than commercial Ag sulfadiazine (36). Incorporation of silver-clay nanohybrid in poly (sulfobetaine) resulted in high, sustained, and diffusion-controlled antimicrobial activity of the silver-eluting polymer with antifouling properties resisting protein adsorption (63).

A matter that should be considered in the field of antimicrobial nanocomposite dressings is the selectivity towards microbes versus cytotoxicity to host cells and tissue. Silver preparations (55, 64), antimicrobial peptides (65), and antimicrobial photodynamic therapy (66) have been investigated for possible cytotoxicity towards human cells. AqNPs like other biocides are non-specific in action and cytotoxic to both microbial and human cells. They have been shown to be cytotoxic at high concentrations in vitro. The mechanisms underlying this cytotoxicity are believed to be agglomeration in cell nuclei and cytoplasm with induced intracellular oxidative stress (67), induced apoptosis via a mitochondrial pathway through ROS and JNK (68, 69), and DNA-repair geneup regulation suggesting associated DNA damage (70, 71). Hence, dose regulation by entrapment in a matrix that utilises special drug carrier systems, as well as slow-release drug delivery systems may be useful in preventing their agglomeration. An in vitro study combining Ag sulfadiazine loaded lipid nanoparticles with chitosan found no cytotoxicity toward dermal fibroblasts and keratinocytes, suggesting that lipid encapsulation of Ag sulfadiazine prevents cytotoxicity (72). Similarly, polymeric micelles obtained by selfassembling of chitosan developed as carriers for Ag sulfadiazine showed a marked increase of Ag sulfadiazine concentration in micelle dispersion and reduced cytotoxicity (73). Nevertheless, it should not be neglected that several in vitro studies have attempted to approximate lethal and sublethal doses of silver solutions for keratinocytes, and values ranging from 7x10-4% to 55x10-4% in solution have been reported as toxic (74). A widely accepted mechanism of toxicity includes the release of Ag+ which readily interact with functional groups in proteins and can lead to enzymatic dysfunction and membrane damage, which manifest as nascent toxicity, symptomatically (75). Therefore, ensuring the dose is safe is equally important with administering a therapeutic dose, and the demonstrated cytotoxic potential of silver nanoparticles at higher doses should not be neglected.

Additionally, several preclinical studies have evaluated the toxicity of AgNPs. Toxicity studies of Acticoat in athymic mice with full-thickness wounds showed Acticoat to be toxic within 1 day, but *in vivo* exposure for a week did not injure the skin substitute or inhibit wound healing (55). A study looking at the safety of silver dressings including nanocrystalline silver in the treatment of MRSA-infected full thickness wounds in Sprague-Dawley and streptozotocin-induced diabetic rats showed that silver dressings induced slight liver damage in the diabetic rats (76). Although changes in serum chemistry caused by silver were observed, this did not indicate silver deposition in the organs and the hazards of silver-containing dressings were thought to be insignificant (76).

2.1.3. Recent Nanoparticles with antimicrobial properties

In addition to AgNPs, several other nanoparticles have been demonstrated to possess antimicrobial properties. Copper (Cu), graphene oxide, graphene, titanium oxide (TiO2), fibrin, polycationic NPs, and zinc oxide (ZnO) are amongst these nanoparticles (1, 77-83) often combined with biocompatible scaffolds as wound dressings (Figure 3). Photothermal treatment with the aid of NPs has also been described in the literature (22, 84), highlighting the efficacy of these constructs as antibiotic-free antimicrobial remedies promoting wound healing.

Graphene-based nanoparticles, in particular, have shown to promote wound healing through their antimicrobial properties (84-86). In addition to these properties, they can be combined with other materials to form nanocomposites, used for stem cell and/or growth factor delivery (87, 88), to enhance the bioactivity of materials (89-91), and to promote angiogenesis (92, 93). The intracellular formation of reactive oxygen and nitrogen species as well as activation of phospho-eNOS and phospho-Akt are believed to be the underlying mechanisms for graphene induced angiogenesis and antimicrobial properties (93). The results of these studies confirm the important role graphene NPs can play in wound healing.

2.2. Nanoparticles and angiogenesis

Angiogenesis, the formation of new blood vessels, plays a vital role in several physiological and pathological processes in the body. Angiogenesis is imperative for wound repair because new vessels provide nutrients and oxygen to support the actively proliferating cells.

Although several studies have suggested that gold nanoparticles (AuNPs) have anti-angiogenic properties (94-96), more recently, AuNPs have been

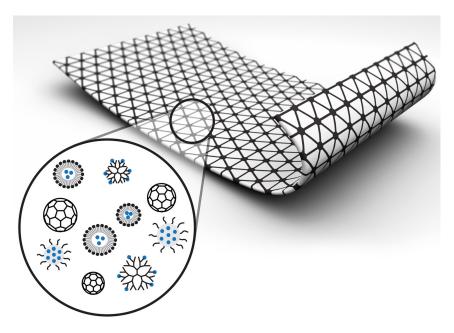


Figure 3. Nanomaterials incorporated into scaffolds for use in biomedical applications.

synthesised following a green chemistry approach, demonstrating their high stability and biocompatibility, as well as their excellent pro-angiogenic activity, through a series of *in vitro* and *in vivo* assays. Formation of reactive oxygen species and activation of p-Akt were believed to be the probable mechanism of angiogenesis (3). Similarly, the combination of AuNP, epigallocatechin gallate, and α -lipoic acid significantly accelerated diabetic cutaneous wound healing through angiogenesis regulation and anti-inflammatory effects (97). The combination led to an initial increase in vascular endothelial growth factor (VEGF) and angiopoietin-2 but not angiopoietin-1 expression (97).

It is highly likely that AuNPs modulate angiogenesis in healing wounds. Further preclinical studies are needed to draw conclusions about their angiogenic effects. Beside inherent angiogenic properties of NPs, these materials can also be used as drug delivery vectors for various factors, including some stimulating angiogenesis (98).

2.3. Nanoparticles and drug delivery

Nanoparticles have become potent drug delivery systems that have attracted much attention and interest as efficient carriers for various active compounds. The specific characteristics of nanoparticles make their use as drug delivery systems an interesting and suitable strategy. It is well established that growth factors (GFs) and other bioactive compounds play an important role in wound healing, inducing cell proliferation and migration, angiogenesis, and collagen deposition (99). There is

a substantial potential for combining these factors and appropriate cells to treat wounds.

The majority of NP-based drug delivery systems aim to deliver growth factors to the wound site. Several studies have designed and described the effects of NP vectors that deliver VEGF, recombinant human epidermal growth factor (rhEGF), platelet-derived growth factor (PDGF), basic fibroblast growth factor (bFGF), granulocyte colony stimulating factor (GCSF), keratinocyte growth factor (KGF), and platelet-rich plasma (PRP), which contains an array of GFs.

Constructs incorporating VEGF, EGF, bFGF, and PDGF either directly embedded in collagenhyaluronic acid (HA) nanofibrous matrices or encapsulated in gelatin NPs have also been made. Collagen-HA-gelatin NP constructs had similar mechanical properties to the human skin. In addition. the design of a particle-in-fibre structure allowed a slow controlled release of the GFs for up to 1 month. *In vitro*, these constructs stimulated the growth and maturation of endothelial cells. Application of these composite GF delivery systems on wounds of diabetic rats was associated with accelerated closure rate, together with elevated collagen deposition and enhanced maturation of vessels (19). Similarly, poly (ether) urethanepolydimethylsiloxane/fibrin scaffolds containing poly (lactic-co-glycolic acid) (PLGA) NPs loaded with VEGF and bFGF accelerated wound closure in genetically diabetic mice compared to scaffolds without growth factors or containing unloaded PLGA NPs. However. the closure rate was similar to that observed in mice treated with scaffolds containing free VEGF and bFGF. Both scaffolds containing growth factors induced

complete re-epithelialization, with enhanced granulation tissue formation and maturity and collagen deposition compared to the other groups (100). Others designed a dual GF delivery system based on electrospun chitosan and poly (ethylene oxide) matrices loaded with VEGF and embedded with PDGF encapsulated poly (lacticcoglycolic acid) (PLGA) NPs. In vitro studies revealed that the nanofibrous composites delivered VEGF quickly and PDGF in a delayed manner. They supported fibroblast growth, exhibited anti-bacterial activities, and preclinical significantly accelerated wound healing by promoting angiogenesis, increasing re-epithelialization and controlling granulation tissue formation. For later stages of healing, evidence also supported quicker collagen deposition and earlier remodelling (101). Mohandas et al. developed VEGF-loaded fibrin NPs, incorporated into chitosan-HA sponges as dressings for diabetic wounds. The nanocomposites combined with human umbilical vein endothelial cells induced capillary-like tube formation in vitro, which was absent in control sponges, suggesting that the VEGF-fibrin NP-chitosan-HA constructs have the potential to induce angiogenesis (102).

PLGA-rhEGF NPs showed a controlled release of rhEGF encapsulated in the NPs and enhanced rhEGF effects on cell proliferation whilst shortening preclinical wound healing time in diabetic rats when compared to controls (free EGF, NPs, and phosphate-buffered saline (PBS). The PLGA-EGF NPs were uniform and dispersible and EGF release lasted for 24 hours (103). A similar topical delivery system composed of lipid NPs and rhEGF showed excellent bioactivity in vitro even higher than that of free rhEGF. In vivo examination in both diabetic mice and a porcine model showed improved healing evidenced by the number of arranged microvasculature, fibroblast migration and proliferation, collagen deposition and evolution of the inflammatory response, whilst rhEGF plasma levels were almost undetectable (104-106).

KGF, combined with elastin-like peptides fabricated using self-assembly to form a fusion protein and applied to wounds of diabetic mice, was associated with enhanced re-epithelialisation and granulation compared to controls (28). Another study utilised GCSF-loaded dextran NPs coated with PLGA and spray-painted on haemostatic gauze as a scaffold for application in post tumour resection wounds and demonstrated enhanced haemostasis and blood neutrophil counts in vivo (107). PRP contains many GFs and can also be used to enhance wound healing and GF delivery. PRP combined with heparin-PLGA NPs and fibrin gel was associated with a prolonged PDGF release compared to PRP and fibrin gel alone (108). Examination of the construct in a murine model resulted in much faster wound closure, as well as dermal and epidermal regeneration compared with PRP-fibrin gel and heparin-PLGA fibrin gel. Heparin-PLGA-fibrin gel PRP also accelerated angiogenesis (108).

In the same way, other bioactive molecules and compounds can be incorporated into NPs for sustained release at the wound site. These compounds may include antibiotics, analgesics, and peptides. A system based on HA and lipid NPs for the delivery of Astragaloside IV, the active compound of Astragali Radix (the root of Astragalus membranaceous plant) was used to accelerate wound healing and reduce scars (109). This construct enhanced the migration and proliferation of keratinocytes and increased drug uptake on fibroblasts *in vitro* (p < 0.01). It strengthened wound healing and inhibited scar formation *in vivo* by increasing wound closure rate (p < 0.05) and contributing to angiogenesis and collagen remodelling (110).

Delivery of topically applied opioids using NP can lead to efficient pain reduction. Opioids encapsulated in lipid and dendritic NPs yielded enhanced load delivery compared to unloaded NPs and free morphine (111). Interestingly, transforming growth factor beta1 (TGF- β 1) was taken up by dendritic NPs. Opioid-lipid NPs enhanced keratinocyte migration, whereas opioid-dendritic NPs did not inhibit this. Another morphine-lipid NP delivery system tested in a human-based 3D-wound healing model showed accelerated re-epithelialisation and wound healing, suggesting that in addition to analgesic effects, opioids may improve wound healing (112).

LL37, an endogenous human host defence peptide that modulates wound healing and angiogenesis and has anti-microbial properties, encapsulated in PLGA NPs displayed antimicrobial activity against Escherichia coli (113). *In vivo* examination of the nanocomposite showed that treatment with PLGA-LL37 NPs significantly accelerated wound healing compared to PLGA or LL37 alone. PLGA-LL37 NP-treated wounds were characterised by advanced granulation tissue formation with higher collagen deposition, reepithelialisation and neovascularisation (113). PLGA-LL37 NP improved angiogenesis, significantly upregulated IL-6 and VEGF expression and modulated the inflammatory wound response (113).

Nanocarriers comprised of clarithromycin encapsulated in chitosan NPs are biocompatible *in vitro*, as well as able to increase the concentration of clarithromycin compared to a saturated water solution (114).

In conclusion, NPs can be effectively used for the targeted and sustainable delivery of several drugs, GFs, and other bioactive compounds, which can play an important role in the wound healing process.

2.4. Nanoparticles and immunomodulation

Cell populations and the complex signalling pathways that regulate the stages of wound healing depend on an intact and functional immune system.

Macrophages are pivotal cells in orchestrating the healing of wounds (115, 116). They debride unhealthy tissue and phagocytose invading bacteria. Macrophages are further activated within the wound to secrete a variety of cytokines that recruit other cells and enhance their proliferation, thereby promoting regeneration. Injection of activated macrophages into wounds of patients was reported to promote wound healing (117). Essential to wound healing, depletion of wound T lymphocytes decreases wound strength and collagen content, while selective depletion of the CD8+ suppressor subset of T lymphocytes enhances wound healing (118). Lymphocytes also exert a downregulating effect on fibroblast collagen synthesis via several secreted lymphokines (interferon gamma (IFN-y), tumour necrosis factor alpha (TNF- α), and interleukin-1 (IL-1) and by cell-to-cell contact. Finetuning of the immune system response can aid the process of wound healing. Nanoparticles that modulate the immune response and inflammation phase of wound healing have been vastly studied in the literature.

AuNPs are among the NPs studied for their immunomodulatory properties. Silica-gold core-shell NPs (SiO2-Au) applied to wounds in vivo promoted wound healing, which was potentially related to the anti-inflammatory and anti-oxidation properties of AuNPs (119), although an exact mechanism was not detailed. In vitro evaluation did not show any cytotoxicity of the composites (119). A similar study examining the effect of AuNPs combined with epigallocatechin gallate, and α-lipoic acid on wound healing in diabetic mice showed that the nanocomposites significantly accelerated diabetic wound healing through angiogenesis regulation and anti-inflammatory effects (97). Immunoblotting showed a significant decrease of CD68 expression whilst VEGF significantly increased following treatment (97).

In addition to AuNPs, AgNPs also play an anti-inflammatory role in wound healing. Treatment of porcine wounds with nanocrystalline Ag significantly increased induction of apoptosis in the inflammatory cells present in the dermis (120). Additionally, decreased levels of pro-inflammatory cytokines TNF-a and IL-8, and increased levels of anti-inflammatory cytokine IL-4, EGF, KGF, and KGF-2 were observed (120). A similar study investigating dendrimers, as well as AqNPs. showed that both these NPs had antiinflammatory properties. When combined in the form of dendrimer-AgNP composites the anti-inflammatory properties were further augmented, resulting in faster wound healing in vivo (2). Others (121) describe that nanocrystalline Ag dressing decreases adversely high levels of matrix metalloproteinases (MMP) -9, a proteolytic enzyme involved in wound healing. High MMP-9 levels promote TNF-α, IL-8, and TGF-β, all associated with exaggerated ongoing inflammation.

Low levels impede keratinocyte migration (121). When used in a situation of minimal inflammation, these dressings may undesirably decrease the low levels of MMP-9 and adversely affect epithelialization.

Several other NPs have been investigated for their immunomodulatory effects. These include dendrimers (122), TiO2 NPs (80, 123), fullerenes (29), curcumin (124), tea tree oil NPs (125), and Cu NPs (83). As described previously, NPs can also be used as drug delivery tools loaded with immunomodulatory compounds such as α -gal (126) and indomethacin (127).

2.5. Nanoparticles and collagen deposition

AgNPs impregnated polyelectrolyte multilayer (PEM) have been shown to be non-cytotoxic yet bactericidal in vitro. A full-thickness, excisional murine wound healing model in both normal and spontaneously diabetic mice showed mildly increased collagen deposition in the silver dressing treated animals (128). Other studies have shed light to the excellent structural alignment of the collagen, as well as its increased deposition in improving tensile properties of tissues such as skin, after administration of silver nanoparticles post-wound healing (5). Silver is not the only nanoparticle able to induce collagen synthesis. Calcium-based nanoparticles also cause contracture of collagen lattices and stimulate fibroblast activity (129). This may show great potential and it will not be unlikely to see calcium nanoparticles used for wound healing in the future. Returning to the realm of the metals, with a more noble approach, one should not overwrite gold nanoparticles (AuNPs) from the scene. AuNPs have been used particularly for imaging of scars, as they are adept at accumulating at those sites, particularly large myocardial scars (130). In combination with other nanoparticles from this immense arsenal, these localisers of scars may prove very promising in the near future. Some managed to develop a platform of sustained release of nitric oxide, using nanoparticles, to achieve a reduction in inflammation, along with a marked increase in collagen deposition, accelerating wound healing (131). Other interesting designs have produced similar effects; carbon nanotube on polystyrene and polyaniline copolymer increased collagen gelation in a dose-dependent manner, whilst leaving D-periodicity and average fibril diameter of this immense protein, largely unchanged (132). An appreciation of the size of collagen molecules, along with the cross-linking required, results in more justice to the complexity of the process and the role of nanoparticles.

2.6. Gene delivery with nanoparticles for acceleration of wound healing

Nanoparticles have also been used in studies of genes for acceleration of wound healing. Several

groups have tried different methods, with vascular endothelial growth factor (VEGF) transfection being the most common one. Viral vectors have been used to administer VEGF to diabetic patients and an effect in wound healing was observed as soon as 6 days post-transfection (133). Viral vectors, despite the amount of attenuation or other measures which reduce the likelihood of generating an immune response, should always be treated with caution. As a result, most groups focused on non-viral transfections, using nanoparticles; Peng et al. introduced VEGF into bone mesenchymal stem cells, using β-cyclodextrin linked polyethyleneimine. The results were more than promising, as wound closure and increased collagen formation were observed 72hours post transfection (134). They also looked at the effects of VEGF in a gelatin scaffold in terms of skin re-epithelialisation and collagen synthesis, both of which were increased (135). Another way to facilitate wound closure apart from inducing angiogenesis is to minimise local motility in the cellular level. Fidgeting-like 2 is a severin enzyme involved in this process, and inhibition with target delivery of siRNA using nanoparticles accelerates healing (4, 136). Intradermal delivery of key genes such as the sonic hedgehog has been performed, as well. The mode was biodegradable cationic poly (\(\beta\)-amino ester) nanoparticles, which resulted in greater transfection compared to the commonly used Lipofectamine 2000 (137). Other carriers, such as β-cyclodextrin and poly (amidoamine) dendron atoms have also been tested and proven successful in promoting wound healing in streptozocin-induced diabetic mice (138). Finally. hypoxia inducible factor (HIF-1α) has been entrapped in fibrin to improve the healing process in full thickness wounds, by being a potential inducer of VEGF (139). Although the evidence in this field is recent and limited. some fundamental questions regarding the safety and efficacy of this technique shall be elucidated in the vears to come.

2.7. The role of nitric oxide nanoparticles in wound healing

The presence of Nitric Oxide (NO) plays a significant role in the wound healing process through modulation of angiogenesis, collagen deposition, and keratinocyte proliferation (131, 138-140). NO, and its reactive nitrogen species derivatives are effective in killing pathogens (141, 142) and potentially form a useful preventive and therapeutic strategy against skin infections (140).

NO synthetase (NOS) knockout mice have shown impairments in the wound healing processes that were only ameliorated after the addition of excess L-arginine substrate or re-introduction of the NOS gene through transfection (143). NO-releasing hydrogels and nanocomposites have shown significant antimicrobial activity with an acceleration of infected

wound healing both *ex* and *in vivo* (144, 145). NO release decreases suppurative inflammation (131) and collagen degradation, minimises the bacterial burden (145), and inhibits fibroblasts to a lesser extent than clinically administered concentrations of antiseptics like povidone iodine (144). NO nanoparticles significantly accelerate wound healing (146) through modification of leukocyte migration and increasing tumour growth factor-β production with a subsequent promotion of angiogenesis (122, 139), leading to increased fibroblast migration and collagen deposition (131, 146). In infected wounds, stained NO nanoparticle-treated tissue depicts decreased neutrophil infiltrate and bacterial load, as well as rapid healing (145, 147-148).

Consequently, the fact that NO nanoparticles greatly expedite wound healing is not surprising. Together, these data suggest that NO nanoparticles have the potential to serve as a novel category of applied antimicrobials for the treatment of infected wounds and may also function as a novel wound healing strategy in the setting of immunocompromised states associated with defective wound healing.

2.8. Stem cell delivery with the aid of nanoparticles

This field is certainly one occupying high ambitions and hopes in the horizon. The domain is two-sided, as seen from the studies discussed below since nanoparticles can be used to guide stem cells. but so can the latter be utilised as a medium to deliver nanoparticle properties. Researchers have tried different combinations of various stem cell types with nanoparticles and managed to achieve localised drug delivery, inhibit neovascularisation where appropriate (e.g. diabetic retinopathy) and reconstruct traumadistorted surfaces (e.g. ocular reconstruction) (149). Another group injected human MSCs with quantum dot NPs and seeded them onto a fibrin suture. When applied to the myocardium of rats, fibrosis was greater in non-hMSC seeded sutures, and the quantum dots provided a satisfactory media for imaging in both cases (150). Apart from soft tissue, success with stem cells and nanoparticles has also been recorded in bone. Ferucarbotran labelled MSCs guided with an extracorporeal device were successfully guided to the site of a rat bone fracture (151). Furthermore, PLGA has been used to deliver a plasmid for expression of bone morphogenetic protein into rabbit adipocytes (152). The technique offered significant healing advantage to the treated rats over the control group, in terms of chondrogenesis. Others modified adipocyte stem cells to express VEGF in a murine hindlimb model of ischaemia. The transfection was through nanoparticles, making them here the "therapeutic substance" delivered to the tissue of interest (153). This is possible due to the well described migratory properties of stem cells. However, it is noteworthy

Table 2. Clinical trials of nanoparticles in wound healing

Nanoparticle/ Nanoscaffold	Type of Wound	Procedure	Outcome	Year/ref
AgNPs and AgSD	Partial thickness burns	RCT of AgNPs compared to AgSD in 54 pts	AgNP was superior to topical AgSD for wound healing	2014 (158)
Aquacel Ag dressing	Venous leg ulcers	Multi-centre RCT comparing Aquacel Ag to Urgotul in 281 pts	Better wound healing progression with Aquacel Ag	2012 (159)
Ag nylon (Silverlon)	Colorectal surgical wound	Prospective RCT comparing Ag with gauze dressings in pts undergoing colorectal op	Silverlon was safe and three times more effective	2011 (160)
AgNPs (nanocrystaline silver)	Leg ulcers	RCT of AgNPs compared with cadexomer iodine in 281 pts	Similar antibacterial properties. Healing within 2 weeks AgNPs	2010 (161)
Ag hydrofibre	Pilonidal sinus	RCT of Ag hydrofibres compared to sponge dressings in 43 pts	Ag hydrofibre is more cost- effective & wound healing	2010 (162)
AgNP (Acticoat)	Freshly grafted burn	RCT of Acticoat compared to the standard management in 20 pts	Acticoat was cheaper & the effect on wound healing was similar	2007 (163)
AgNPs Partial thickness burn		RCT of AgNPs compared to 1% AgSD in 191 pts	Similar bacterial colonisation. AgNPs significantly reduced healing time	2006 (164)
AgNPs	Superficial burn wounds	120 patients were randomised to receive: AgNPs/carbon fibre/hydrogel/ Vaseline gauze dressings	Water retention capacity was significantly higher in carbon fibre dressing	2007 (165)

Abbreviations: Pt, patient; AgNP, silver nanoparticles; AgSD, silver sulfadiazine; op, operations

to consider the limitations in this field; although application of nanoparticles offers great potential, the closer interaction with stem cells requires further investigation. Some nanoparticles, indeed, interfere with gene expression amongst other intracellular process in stem cells and thus, further research is required in this field to optimise the delivery. For instance, Au- and Ag- based nanoparticles affect the growth of embryonic neuronal stem cells (154). More specifically, whilst Au- based nanoparticles could be considered for delivering stem cells, they have been widely utilised to induce effects of interest in stem cells; an example being altered osteogenesis when culturing human mesenchymal stem cells with gold nanoparticles in vitro (155). In summary, different nanoparticles can have different effects on the proliferation and differentiation of stem cells and this has been exploited by scientists in previous experiments (156). However, it needs to be taken into consideration when attempting to deliver stem cells via nanoparticles and intend to avoid such interactions.

Perhaps an even better way to demonstrate the two-way relationship between stem cells and nanoparticles is through two eminent studies in the field. More specifically, one group have controlled the migration of mesenchymal stem cells (MSCs) magnetically, via filling them with Si-, Au- and Fenanoparticles. This technique allowed for very effective magnetic guiding to the site of atherosclerosis, and using photothermal therapy, the results were superior to conventional stenting (157). Finally, Peng et al. reveal the other side of the same spectrum, in a study of wound healing in the skin. By transfecting

epidermal stem cells with β -cyclodextrin linked to polyethyleneimines, they managed demonstrated acceleration of hair follicle regeneration, skin reepithelialisation, dermal collagen synthesis and VEGF synthesis, therefore concluding how nanoparticles can be integrated into stem cells for use as a gene reservoir in wound healing applications (137).

3. CLINICAL TRIALS OF NANOPARTICLES IN WOUND HEALING

A brief list of clinical trials in patients using AgNPs is summarised in Table 2, focusing on one major study for the different types of wounds.

Whilst the amount as well as the variety of the studies conducted in randomised patients (i.e. RCTs) is still not significant compared to preclinical studies, the results so far are promising. In particular, silver nanoparticles are undoubtedly at the core of human research due to their therapeutic properties. However, their potential for cytotoxicity shall be taken into consideration when tailoring treatment to a particular patient. Studies to come will shed more light on other candidates used in preclinical trials. The latter may serve a major role alongside AgNPs in the years to come, enriching the armamentarium for wound healing.

4. CONCLUSION AND FUTURE PERSPECTIVES

The field of nanotechnology as applied to wound healing is moving at a rapid pace. With further advances, it is likely that breakthroughs in nano-inspired treatments will significantly improve wound healing

in the foreseeable future. Perhaps the most exciting aspects of nanotechnology, as applied to wounds, would be advances in NP-based growth factors delivery systems for angiogenesis, as well as NPs' inherent anti-microbial properties resulting in efficient skin regeneration. Therefore, it can be reasonably concluded that nanotechnology-based remedies will be the next frontier poised for breakthroughs in unmet clinical needs of skin regeneration and wound healing. The ideal wound dressing should have good flexibility, good mechanical strength, large porosity, and be non-adherent to the wound surface. They should also provide a cooling sensation and a moist environment. whilst acting as a barrier to microbes. This is a multibillion pound industry with a large number of companies as well as academics working towards accelerating wound healing products based on nanotechnology.

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6. REFERENCES

- Lu B, Li T, Zhao H, Li X, Gao C, Zhang S, Xie E. Graphene-based composite materials beneficial to wound healing. *Nanoscale* 4, 2978-82 (2012) DOI: 10.1039/c2nr11958g
- 2. Liu X, Hao W, Lok CN, Wang YC, Zhang R, Wong KK. Dendrimer encapsulation enhances anti-inflammatory efficacy of silver nanoparticles. *J Pediatr Surg* 49, 1846-51 (2014)

DOI: 10.1016/j.jpedsurg

- Nethi SK, Mukherjee S, Veeriah V, Barui AK, Chatterjee S, Patra CR. Bioconjugated gold nanoparticles accelerate the growth of new blood vessels through redox signaling. *Chem Commun (Camb)* 50, 14367-70 (2014)
 DOI: 10.1039/c4cc06996j
- Charafeddine RA, Makdisi J, Schairer D, O'Rourke BP, Diaz-Valencia JD, Chouake J, Kutner A, Krausz A, Adler B, Nacharaju P, Liang H, Mukherjee S, Friedman JM, Friedman A, Nosanchuk JD, Sharp DJ. Fidgetin-Like 2: A Microtubule-Based Regulator of Wound Healing. *J Invest Dermatol* 135, 2309-18 (2015) DOI: 10.1038/jid.2015.94

- Kwan KH, Liu X, To MK, Yeung KW, Ho CM, Wong KK. Modulation of collagen alignment by silver nanoparticles results in better mechanical properties in wound healing. *Nanomedicine* 7, 497-504 (2011) DOI: 10.1016/j.nano.2011.01.003
- Diegelmann RF. Cellular and biochemical aspects of normal and abnormal wound healing: an overview. J Urol 157, 298-302 (1997)
 DOI: 10.1016/S0022-5347(01)65364-3
- Metcalfe AD, Ferguson MWJ. Tissue engineering of replacement skin: the crossroads of biomaterials, wound healing, embryonic development, stem cells and regeneration. J R Soc Interface 4, 413-37 (2007)

DOI: 10.1098/rsif.2006.0179

- Atala A, Irvine DJ, Moses M, Shaunak S. Wound Healing Versus Regeneration: Role of the Tissue Environment in Regenerative Medicine. MRS Bull 35, (2010) DOI: 10.1557/mrs2010.528
- Syed A, Ahmad A. Extracellular biosynthesis of platinum nanoparticles using the fungus Fusarium oxysporum. Colloids Surf B Biointerfaces 97, 27-31 (2011) DOI: 10.1016/j.colsurfb.2012.03.026
- Vila OF, Bago JR, Navarro M, Alieva M, Aguilar E, Engel E, Planell J, Rubio N, Blanco J. Calcium phosphate glass improves angiogenesis capacity of poly (lactic acid) scaffolds and stimulates differentiation of adipose tissue-derived mesenchymal stromal cells to the endothelial lineage. *J* Biomed Mater Res A 101 932-41 (2013) DOI: 10.1002/jbm.a.34391
- 11. Niessen FB, Spauwen PH, Schalkwijk J, Kon M. On the nature of hypertrophic scars and keloids: a review. *Plast Reconstr Surg* 104, 1435-58 (1999)
- 12. Kischer CW, Brody GS. Structure of the collagen nodule from hypertrophic scars and keloids. *Scan Electron Microsc* 3, 371-6 (1981)
- Fonder MA, Lazarus GS, Cowan DA, Aronson-Cook B, Kohli AR, Mamelak AJ. Treating the chronic wound: A practical approach to the care of nonhealing wounds and wound care dressings. *J Am Acad Dermatol* 58, 185-206 (2008) DOI: 10.1016/j.jaad.2007.08.048

14. Ito K, Saito A, Fujie T, Nishiwaki K, Miyazaki H, Kinoshita M, Saitoh D, Ohtsubo S, Takeoka S. Sustainable antimicrobial effect of silver sulfadiazine-loaded nanosheets on infection in a mouse model of partialthickness burn injury. Acta Biomater 24, 87-95 (2015)

DOI: 10.1016/j.actbio.2015.05.035

- 15. Ninan N, Muthiah M, Park IK, Elain A, Wong TW, Thomas S, Grohens Y. In vitro and In vivo Evaluation of Pectin/Copper Exchanged Faujasite Composite Membranes. J Biomed Nanotechnol 11, 1550-67 (2015)
- 16. Landriscina A, Musaev T, Rosen J, Ray A. Nacharaiu P. Nosanchuk JD. Friedman AJ. N-acetylcysteine S-nitrosothiol Nanoparticles Prevent Wound Expansion and Accelerate Wound Closure in a Murine Burn Model. J Drugs Dermatol 14, 726-32 (2015)
- 17. Hsiao WT, Lin LH, Chiang HJ, Ou KL, Cheng HY. Biomedical electrosurgery devices containing nanostructure for minimally invasive surgery: reduction of thermal injury and acceleration of wound healing for liver cancer. J Mater Sci Mater Med 26, 77 (2015) DOI: 10.1007/s10856-015-5416-4
- 18. Silveira PC, Venancio M, Souza PS, Victor EG, de Souza Notoya F, Paganini CS, Streck EL, da Silva L, Pinho RA, Paula MM. Iontophoresis with gold nanoparticles improves mitochondrial activity and oxidative stress markers of burn wounds. Mater Sci Eng C Mater Biol Appl 44, 380-5 (2014) DOI: 10.1016/j.msec.2014.08.045
- 19. Lai HJ. Kuan CH. Wu HC. Tsai JC. Chen TM, Hsieh DJ, Want WT. Tailored design of electrospun composite nanofibers with staged release of multiple angiogenic growth factors for chronic wound healing. Acta Biomater 10, 4156-66 (2014) DOI: 10.1016/j.actbio.2014.05.001
- 20. Bhuvaneswari T, Thiyagarajan M, Geetha N, Venkatachalam P. Bioactive compound loaded stable silver nanoparticle synthesis microwave irradiated from aqueous extracellular leaf extracts of Naringi crenulata and its wound healing activity in experimental rat model. Acta Trop 134, 55-61 (2014) DOI: 10.1016/j.actatropica.2014.03.009
- 21. Wu J, Zheng Y, Wen X, Lin Q, Chen X, Wu Z. Silver nanoparticle/bacterial cellulose

- gel membranes for antibacterial wound dressing: investigation in vitro and in vivo. Biomed Mater 9, 035005 (2014) DOI: 10.1088/1748-6041/9/3/035005
- 22. Nafee N. Youssef A. El-Gowelli H. Asem H. Kandil S. Antibiotic-free nanotherapeutics: hypericin nanoparticles thereof improved in vitro and in vivo antimicrobial photodynamic therapy and wound healing. Int J Pharm 454, 249-58 (2013) DOI: 10.1016/j.ijpharm.2013.06.067
- 23. Jaiswal M, Gupta A, Agrawal AK, Jassal M, Dinda AK, Koul V. Bi-layer composite dressing of gelatin nanofibrous mat and poly vinyl alcohol hydrogel for drug delivery and wound healing application: in-vitro and in-vivo studies. J Biomed Nanotechnol 9, 1495-508 (2013)
- 24. Chowdhury S, Guha R, Trivedi R, Kompella UB, Konar A, Hazra S. Pirfenidone nanoparticles improve corneal wound healing and prevent scarring following alkali burn. PLoS One 8, e70528 (2013) DOI: 10.1371/journal.pone.0070528
- 25. Kariakina EV. Gladkova EV. Babushkina IV. Belova SV, Matveeva OV, Mamonova IA. (Reparative regeneration of rat soft tissues is affected by a wound-healing composite) Ross Fiziol Zh Im I M Sechenova 99, 737-44 (2013)
- 26. Sudheesh Kumar PT, Raj NM, Praveen G, Chennazhi KP, Nair SV, Jayakumar R. In vitro and in vivo evaluation of microporous chitosan hydrogel/nanofibrin composite bandage for skin tissue regeneration. Tissue Eng Part A 19, 380-92 (2013) DOI: 10.1089/ten.TEA.2012.0376
- 27. Neibert K, Gopishetty V, Grigoryev A, Tokarev I, Al-Hajaj N, Vorstenbosch J, Philip A, Minko S, Mayasinger D. Woundhealing with mechanically robust and biodegradable hydrogel fibers loaded with silver nanoparticles. Adv Healthc Mater 1, 621-30 (2012) DOI: 10.1002/adhm.201200075
- 28. Koria P, Yagi H, Kitagawa Y, Megeed Z, Nahmias Y, Sheridan R, Yarmush ML. Self-assembling elastin-like peptides growth factor chimeric nanoparticles for the treatment of chronic wounds. Proc Natl Acad Sci U S A 108, 1034-9 (2011) DOI: 10.1073/pnas.1009881108

- 29. Zhou Z, Joslin S, Dellinger A, Ehrich M, Brooks B, Ren Q, Rodeck U, Lenk R, Kepley CL. A novel class of compounds with cutaneous wound healing properties. *J Biomed Nanotechnol* 6, 605-11 (2010)
- Mullally C, Carey K, Seshadri R. Use of a nanocrystalline silver dressing and vacuum-assisted closure in a severely burned dog. *J Vet Emerg Crit Care (San Antonio)* 20, 456-63 (2010)
 DOI: 10.1111/j.1476-4431.2010.00564.x.
- 31. Suwalski A, Dabboue H, Delalande A, Bensamoun SF, Canon F, Midoux P, Saillant G, Klatzmann D, Salvetat JP, Pichon C. Accelerated Achilles tendon healing by PDGF gene delivery with mesoporous silica nanoparticles. *Biomaterials* 31, 5237-45 (2010) DOI: 10.1016/j.biomaterials.2010.02.077
- 32. Naumov AA, Shatalin YV, Potselueva MM. Effects of a nanocomplex containing antioxidant, lipid, and amino acid on thermal burn wound surface. *Bull Exp Biol Med* 149, 62-6 (2010)
- Robson MC. Wound infection. A failure of wound healing caused by an imbalance of bacteria. Surg Clin North Am 77, 637-50 (1997)
 DOI: 10.1016/S0039-6109(05)70572-7
- Paladini F, Meikle ST, Cooper IR, Lacey J, Perguini V, Santin M. Silver-doped selfassembling di-phenylalanine hydrogels as wound dressing biomaterials. *J Mater Sci Mater Med* 24, 2461-72 (2013) DOI: 10.1007/s10856-013-4986-2
- 35. Wren AW, Coughlan A, Hassanzadeh P, Towler MR. Silver coated bioactive glass particles for wound healing applications. *J Mater Sci Mater Med* 23, 1331-41 (2012) DOI: 10.1007/s10856-012-4604-8.
- Im AR, Kim JY, Kim HS, Cho S, Park Y, Kim YS. Wound healing and antibacterial activities of chondroitin sulfate- and acharan sulfate-reduced silver nanoparticles. *Nanotechnology* 24, 395102 (2013) DOI: 10.1088/0957-4484/24/39/395102
- 37. Li C, Fu R, Yu C, Li Z, Guan H, Hu D, Zhao D, Lu L. Silver nanoparticle/chitosan oligosaccharide/poly (vinyl alcohol) nanofibers as wound dressings: a preclinical study. *Int J Nanomedicine* 8, 4131-45 (2013) DOI: 10.2147/IJN.S51679

- 38. Lin YH, Hsu WS, Chung WY, Ko TH, Lin JH. Evaluation of various silver-containing dressing on infected excision wound healing study. *J Mater Sci Mater Med* 25, 1375-86 (2014)
 DOI: 10.1007/s10856-014-5152-1
- Tian J, Wong KK, Ho CM, Lok CN, Yu WY, Che CM, Chiu JF, Tam PK. Topical delivery of silver nanoparticles promotes wound healing. *ChemMedChem* 2, 129-36 (2007)
 DOI: 10.1002/cmdc.200600171
- Samberg ME, Orndorff PE, Monteiro-Riviere NA. Antibacterial efficacy of silver nanoparticles of different sizes, surface conditions and synthesis methods. *Nanotoxicology* 5, 244-53 (2011) DOI: 10.3109/17435390.2010.525669
- Dhapte V, Kadam S, Moghe A, Pokharkar V. Probing the wound healing potential of biogenic silver nanoparticles. *J Wound Care* 23, 431-436 (2014)
 DOI: 10.12968/jowc.2014.23.9.431
- 42. Das A, Kumar A, Patil NB, Viswanathan C, Ghosh D. Preparation and characterization of silver nanoparticle loaded amorphous hydrogel of carboxymethylcellulose for infected wounds. *Carbohydr Polym* 130, 254-61 (2015)
 DOI: 10.1016/j.carbpol.2015.03.082
- Lara HH, Ayala-Nunez NV, Ixtepan-Turrent L, Rodriguez-Padilla C. Mode of antiviral action of silver nanoparticles against HIV-1. J Nanobiotechnology 8, 1 (2010) DOI: 10.1186/1477-3155-8-1
- 44. Elechiguerra JL, Burt JL, Morones JR, Camacho-Bragado A, Gao X, Lara HH, Yacaman MJ. Interaction of silver nanoparticles with HIV-1. *J Nanobiotechnology* 3, 6 (2005) DOI: 0.1186/1477-3155-3-6
- 45. Lu L, Sun RW, Chen R, Hui CK, Ho CM, Luk JM, Lau GK, Che CM. Silver nanoparticles inhibit hepatitis B virus replication. *Antivir Ther* 13, 253-62 (2008)
- Sun RW, Chen R, Chung NP, Ho CM, Lin CL, Che CM. Silver nanoparticles fabricated in Hepes buffer exhibit cytoprotective activities toward HIV-1 infected cells. *Chem Commun* (*Camb*) 40, 5059-61 (2005) DOI: 10.1039/B510984A

- Mallmann EJ, Cunha FA, Castro NMF, Maciel AM, Menezes EA, Fechine PBA. Antifungal activity of silver nanoparticles obtained by green synthesis. Rev Inst Med Trop Sao Paulo 57, 165-7 (2015) DOI: 10.1590/S0036-46652015000200011
- 48. Baker C, Pradhan A, Pakstis L, Pochan DJ, Shash SI. Synthesis and antibacterial properties of silver nanoparticles. *J Nanosci Nanotechnol* 5, 244-9 (2005)
- 49. Rai M, Yadav A, Gade A. Silver nanoparticles as a new generation of antimicrobials. *Biotechnol Adv* 27, 76-83 (2009) DOI: 10.1016/j.biotechadv.2008.09.002
- 50. Aymonier C, Schlotterbeck U, Antonietti L, Zacharias P, Thomann R, Tiller JC, Mecking S. Hybrids of silver nanoparticles with amphiphilic hyperbranched macromolecules exhibiting antimicrobial properties. *Chem Commun (Camb)* 24, 3018-9 (2002)
- Ruparelia JP, Chatterjee AK, Duttagupta SP, Mukherji S. Strain specificity in antimicrobial activity of silver and copper nanoparticles. *Acta Biomater* 4, 707-16 (2008) DOI: 10.1016/j.actbio.2007.11.006
- 52. Borkow G, Gabbay J. Putting copper into action: copper-impregnated products with potent biocidal activities. *FASEB J* 18, 1728-30 (2004)
 DOI: 10.1096/fj.04-2029fje
- 53. Gravante G, Montone A, A retrospective analysis of ambulatory burn patients: focus on wound dressings and healing times. *Ann R Coll Surg Engl* 92, 118-23 (2010) DOI: 10.1308/003588410X12518836439001
- 54. Thirumurugan G, Veni VS, Ramachandran S, Rao JV, Dhanaraju MD. Superior wound healing effect of topically delivered silver nanoparticle formulation using eco-friendly potato plant pathogenic fungus: synthesis and characterization. *J Biomed Nanotechnol* 7, 659-66 (2011)
- 55. Supp AP, Neely AN, Supp DM, Warden GD, Boyce ST. Evaluation of cytotoxicity and antimicrobial activity of Acticoat Burn Dressing for management of microbial contamination in cultured skin substitutes grafted to athymic mice. *J Burn Care Rehabil* 26, 238-46 (2005) DOI: 10.1097/01.BCR.0000162152.28330.6C

- 56. Nguyen TH, Kim YH, Song HY, Lee BT. Nano Ag loaded PVA nano-fibrous mats for skin applications. *J Biomed Mater Res B Appl Biomater* 96, 225-33 (2011) DOI: 10.1002/jbm.b.31756
- 57. GhavamiNejad A, Unnithan AR, Sasikala ARK, Samarikhalaj M, Thomas RG, Jeong YY, Nasseri S, Murugesan P, Wu D, Park CH, Kim CS. Mussel-Inspired Electrospun Nanofibers Functionalized with Size-Controlled Silver Nanoparticles for Wound Dressing Application. ACS Appl Mater Interfaces 7, 12176-83 (2015) DOI: 10.1021/acsami.5b02542
- Liu X, Lin T, Fang J, Yao G, Zhao H, Dodson M, Wang X. *In vivo* wound healing and antibacterial performances of electrospun nanofibre membranes. *J Biomed Mater Res A* 94, 499-508 (2010)
 DOI: 10.1002/jbm.a.32718
- 59. Lu S, Gao W, Gu HY. Construction, application and biosafety of silver nanocrystalline chitosan wound dressing. *Burns* 34, 623-8 (2008) DOI: 10.1016/j.burns.2007.08.020
- Ahamed MI, Sankar S, Kashif PM, Basha SK, Sastry TP. Evaluation of biomaterial containing regenerated cellulose and chitosan incorporated with silver nanoparticles. *Int J Biol Macromol* 72, 680-6 (2015)
 DOI: 10.1016/i.ijbiomac.2014.08.055
- 61. Prestes MA, Ribas CA, Ribas Filho JM, Moreira LB, Boldt AB, Brunstolin EV, Castanho LS, Bernardi JA, Dias FC. Wound healing using ionic silver dressing and nanocrystalline silver dressing in rats. *Acta Cir Bras* 27, 761-7 (2012)
 DOI: 10.1590/S0102-86502012001100004
- 62. Ghosh Auddy R, Abdullah MF, Das S, Roy P, Datta S, Mukherjee A. New guar biopolymer silver nanocomposites for wound healing applications. *Biomed Res Int*, 912458 (2013) DOI: 10.1155/2013/912458
- 63. Depan D, Misra RD. Hybrid Nanoscale Architecture of Wound Dressing with Super Hydrophilic, Antimicrobial, and Ultralow Fouling Attributes. *J Biomed Nanotechnol* 11, 306-18 (2015)
- 64. Samberg ME, Oldenburg SJ, Monteiro-Riviere NA. Evaluation of silver nanoparticle

toxicity in skin *in vivo* and keratinocytes *in vitro*. *Environ Health Perspect* 118, 407-13 (2010)

DOI: 10.1289/ehp.0901398

- 65. Jacobsen F, Mohammadi-Tabrisi A, Hirsch T, Mittler D, Mygind PH, Sonksen CP, Ravenatos D, Kristensen HH, Gatermann S, Lehnhardt M, Daigeler A, Steinau HU, Steinstraesser L. Antimicrobial activity of the recombinant designer host defence peptide P-novispirin G10 in infected full-thickness wounds of porcine skin. *J Antimicrob Chemother* 59, 493-8 (2007) DOI: 10.1093/jac/dkl513
- Soukos NS, Ximenez-Fyvie LA, Hamblin MR, Socransky SS, Hasan T. Targeted antimicrobial photochemotherapy. *Antimicrob Agents Chemother* 42, 2595-601 (1998)
- 67. Kim JS, Kuk E, Yu KN, Kim JH, Park SJ, Lee HJ, Kim SH, Park YK. Antimicrobial effects of silver nanoparticles. *Nanomedicine* 3, 95-101 (2007)
 DOI: 10.1016/j.nano.2006.12.001
- 68. Hsin YH, Chen CF, Huang S, Shih TS, Lai PS, Chueh PJ. The apoptotic effect of nanosilver is mediated by a ROS- and JNK-dependent mechanism involving the mitochondrial pathway in NIH3T3 cells. *Toxicol Lett* 173, 130-9 (2008) DOI: 10.1016/j.toxlet.2008.04.015
- Park EJ, Yi J, Kim Y, Choi K, Park K. Silver nanoparticles induce cytotoxicity by a Trojan-horse type mechanism. *Toxicol In* vitro 24, 872-8 (2010) DOI: 10.1016/j.tiv.2009.12.001
- Kawata K, Osawa M, Okabe S. In vitro toxicity of silver nanoparticles at noncytotoxic doses to HepG2 human hepatoma cells. Environ Sci Technol 43, 6046-51 (2009)
- Miura N, Shinohara Y. Cytotoxic effect and apoptosis induction by silver nanoparticles in HeLa cells. *Biochem Biophys Res Commun* 390, 733-7 (2009)
 DOI: 10.1016/j.bbrc.2009.10.039
- 72. Sandri G, Bonferoni MC, D'Autilia F, Rossi S, Ferrari F, Grisoli P, Sorrenti M, Catenacci L, Del Fante C, Perotti C, Caramella C. Wound dressings based on silver sulfadiazine solid lipid nanoparticles for tissue repairing. Eur J Pharm Biopharm 84, 84-90 (2013) DOI: 10.1016/j.ejpb.2012.11.022

- 73. Dellera E, Bonferoni MC, Sandri G, Rossi S, Ferrari F, Del Fante C, Perotti C, Grisoli P, Caramela C. Development of chitosan oleate ionic micelles loaded with silver sulfadiazine to be associated with platelet lysate for application in wound healing. *Eur J Pharm Biopharm* 88, 643-50 (2014) DOI: 10.1016/j.ejpb.2014.07.015
- 74. Maghsoudi H, Monshizadeh S, Mesgari M. A comparative study of the burn wound healing properties of saline-soaked dressing and silver sulfadiazine in rats. *Indian J Surg* 73, 24-27 (2011)
- 75. Hollinger MA. Toxicological aspects of topical silver pharmaceuticals. *Crit Rev Toxicol*. 26, 255-60 (1996)
- Yeo ED, Yoon SA, Oh SR, Choi YS, Lee YK.
 Degree of the hazards of silver-containing
 dressings on MRSA-infected wounds
 in Sprague-Dawley and streptozotocin induced diabetic rats. Wounds 27, 95-102
 (2015)
- Gopal A, Kant V, Gopalakrishnan A, Tandan SK, Kumar D. Chitosan-based copper nanocomposite accelerates healing in excision wound model in rats. *Eur J Pharmacol* 731, 8-19 (2014) DOI: 10.1016/j.ejphar.2014.02.033
- Atar-Froyman L, Sharon A, Weiss EI, Houri-Haddad Y, Kesler-Shvero D, Domb AJ, Pilo R, Beyth N. Anti-biofilm properties of wound dressing incorporating nonrelease polycationic antimicrobials. *Biomaterials* 46, 141-8 (2015)
 DOI: 10.1016/j.biomaterials.2014.12.047
- P TSK, Lakshmanan VK, Raj M, Biswas R, Hiroshi T, Nair SV, Jayakumar R. Evaluation of wound healing potential of beta-chitin hydrogel/nano zinc oxide composite bandage. *Pharm Res* 30, 523-37 (2013) DOI: 10.1007/s11095-012-0898-y
- 80. Peng CC, Yang MH, Chiu WT, Chiu CH, Yang CS, Chen YW, Chen KC, Peng RY. Composite nano-titanium oxide-chitosan artificial skin exhibits strong wound-healing effect-an approach with anti-inflammatory and bactericidal kinetics. *Macromol Biosci* 8, 316-27 (2008)
 DOI: 10.1002/mabi.200700188
- 81. Kumar PT, Lakshamanan VK, Anilkumar TV, Ramya C, Reshmi P, Unnikrishnan AG, Nair

- SV, Jayakumar R. Flexible and microporous chitosan hydrogel/nano ZnO composite bandages for wound dressing: *in vitro* and *in vivo* evaluation. *ACS Appl Mater Interfaces* 4, 2618-29 (2012) DOI: 10.1021/am300292v
- 82. Vedakumari WS, Prabu P, Sastry TP. Chitosan-Fibrin Nanocomposites as Drug Delivering and Wound Healing Materials. *J Biomed Nanotechnol* 11, 657-67 (2015)
- 83. Tiwari M, Narayankan K, Thakar MB, Jagani HV, Venkata Rao J. Biosynthesis and wound healing activity of copper nanoparticles. *IET Nanobiotechnol* 8, 230-7 (2014) DOI: 10.1049/iet-nbt.2013.0052
- 84. Shahnawaz Khan M, Abdelhamid HN, Wu HF. Near infrared (NIR) laser mediated surface activation of graphene oxide nanoflakes for efficient antibacterial, antifungal and wound healing treatment. *Colloids Surf B Biointerfaces* 127, 281-91 (2015) DOI: 10.1016/j.colsurfb.2014.12.049
- 85. Shams E, Yeganeh H, Naderi-Manesh H, Gharibi R, Mohammad Hassan Z. Polyurethane/siloxane membranes containing graphene oxide nanoplatelets as antimicrobial wound dressings: in vitro and in vivo evaluations. *J Mater Sci Mater Med* 28, 75 (2017)
- 86. Palmieri V, Papi M, Conti C, Ciasca G, Maulucci G, De Spirito M. The future development of bacteria fighting medical devices: the role of graphene oxide. *Expert Rev Med Devices* 13, 1013-1019 (2016)
- 87. Zhou Y, Chen R, He T, Xu K, Du D, Zhao N, Cheng X, Yang J, Shi H, Lin Y. Biomedical Potential of Ultrafine Ag/AgCl Nanoparticles Coated on Graphene with Special Reference to Antimicrobial Performances and Burn Wound Healing. ACS Appl Mater Interfaces 8,15067-75 (2016)
- 88. Li Z, Wang H, Yang B, Sun Y, Huo R. Threedimensional graphene foams loaded with bone marrow derived mesenchymal stem cells promote skin wound healing with reduced scarring. *Mater Sci Eng C Mater Biol Appl* 57, 181-8 (2015)
- 89. Liu T, Dan W, Dan N, Liu X, Liu X, Peng X. A novel grapheme oxide-modified collagen-chitosan bio-film for controlled growth factor

- release in wound healing applications. *Mater Sci Eng C Mater Biol Appl* 77, 202-211 (2017)
- 90. Wang CH, Guo ZS, Pang F, Zhang LY, Yan M, Yan JH, Li KW, Li XJ, Li Y, Bi L, Han YS. Effects of graphene modification on the bioactivation of polyethylene-terephthalate-based artificial ligaments. *ACS Appl Mater Interfaces* 7, 15263-76 (2015)
- Nishida E, Miyaji H, Kato A, Takita H, Iwanaga T, Momose T, Ogawa K, Murakami S, Sugaya T, Kawanami M. Graphene oxide scaffold accelerates cellular proliferative response and alveolar bone healing of tooth extraction socket. *Int J Nanomedicine* 11, 2265-77 (2016)
- 92. Fan L, Yi J, Tong J, Zhou X, Ge H, Zou S, Wen H, Nie M. Preparation and characterization of oxidized konjac glucomannan/carboxymethyl chitosan/graphene oxide hydrogel. *Int J Biol Macromol* 91, 358-67 (2016)
- 93. Mukherjee S, Sriram P, Barui AK, Nethi SK, Veeriah V, Chatterjee S, Suresh KI, Patra CR. Graphene Oxides Show Angiogenic Properties. *Adv Healthc Mater* 4,1722-32 (2015)
- Mukherjee P, Bhattacharya R, Wang P, Wang L, Basu S, Nagy JA, Atala A, Mukhopadhyay D, Soker S. Antiangiogenic properties of gold nanoparticles. *Clin Cancer Res* 11, 3530-4 (2005)
 DOI: 10.1158/1078-0432
- 95. Kim JH, Kim MH, Jo DH, Yu SY, Lee TG, Kim JH. The inhibition of retinal neovascularization by gold nanoparticles via suppression of VEGFR-2 activation. *Biomaterials* 32, 1865-71 (2011) DOI: 10.1016/j.biomaterials.2010.11.030
- Kalishwaralal K, Barathmanikanth S, Pandian SR, Deepak V, Gurunathan S. Silver nano - a trove for retinal therapies. *J Control Release* 145, 76-90 (2010) DOI: 10.1016/j.jconrel.2010.03.022
- 97. Chen SA, Chen HM, Yao YD, Hung CF, Tu CS, Liang YJ. Topical treatment with antioxidants and Au nanoparticles promote healing of diabetic wound through receptor for advance glycation end-products. *Eur J Pharm Sci* 47, 875-83 (2012) DOI: 10.1016/j.ejps.2012.08.018

- 98. Karponis D, Azzawi M, Seifalian A. An arsenal of magnetic nanoparticles; perspectives in the treatment of cancer. *Nanomed (Lond)* 11, 2215-32 (2016)
- 99. Secco M, Bueno C Jr, Vieira NM, Almeida C, Pelatti M, Zucconi E, Bartolini P, Vainzof M, Miyabara EH, Okamoto OK, Zatz M. Systemic delivery of human mesenchymal stromal cells combined with IGF-1 enhances muscle functional recovery in LAMA2 dy/2j dystrophic mice. Stem Cell Rev 9, 93-109 (2013)

DOI: 10.1007/s12015-012-9380-9

- 100. Losi P, Briganti E, Errico C, Lisella A, Sanguinetti E, Chiellini F, Soldani G. Fibrinbased scaffold incorporating VEGF- and bFGF-loaded nanoparticles stimulates wound healing in diabetic mice. Acta Biomater 9, 7814-21 (2013) DOI: 10.1016/j.actbio.2013.04.019
- 101. Xie Z, Paras CB, Weng H, Punnakitikashem P, Su LC, Vu K, Tang L, Yang J, Nguyen KT. Dual growth factor releasing multi-functional nanofibers for wound healing. *Acta Biomater* 9, 9351-9 (2013) DOI: 10.1016/j.actbio.2013.07.030
- 102. Mohandas A, Anisha BS, Chennazhi KP, Jayakumar R. Chitosan-hyaluronic acid/ VEGF loaded fibrin nanoparticles composite sponges for enhancing angiogenesis in wounds. *Colloids Surf B Biointerfaces* 127, 105-13 (2015) DOI: 10.1016/j.colsurfb.2015.01.024
- 103. Chu Y, Yu D, Wang P, Xu J, Li D, Ding M. Nanotechnology promotes the full-thickness diabetic wound healing effect of recombinant human epidermal growth factor in diabetic rats. *Wound Repair Regen* 18, 499-505 (2010)

DOI: 10.1111/j.1524-475X.2010.00612.x

- 104. Gainza G, Pastor M, Aguirre JJ, Villullas S, Pedraz JL, Hernandez RM, Igartua M. A novel strategy for the treatment of chronic wounds based on the topical administration of rhEGF-loaded lipid nanoparticles: *In vitro* bioactivity and *in vivo* effectiveness in healing-impaired db/db mice. *J Control Release* 185, 51-61 (2014) DOI: 10.1016/j.jconrel.2014.04.032
- 105. Gainza G, Bonafonte DC, Moreno B, Aguirre JJ, Gutierrez FB, Villullas S,

Pedraz JL, Igartua M, Hernandez RM. The topical administration of rhEGF-loaded nanostructured lipid carriers (rhEGF-NLC) improves healing in a porcine full-thickness excisional wound model. *J Control Release* 197, 41-7 (2015)

DOI: 10.1016/j.jconrel.2014.10.033

- 106. Gainza G, Chu WS, Guy RH, Pedraz JL, Hernandez RM, Delgado-Charro B, Igartua M. Development and in vitro evaluation of lipid nanoparticle-based dressings for topical treatment of chronic wounds. *Int J Pharm* 490, 404-11 (2015) DOI: 10.1016/j.ijpharm.2015.05.075
- 107. Yuan W, Liu Z. Surgical wound healing using hemostatic gauze scaffold loaded with nanoparticles containing sustained-release granulocyte colony-stimulating factor. *Int J Nanomedicine* 6, 3139-49 (2011) DOI: 10.2147/IJN.S26006
- 108. La WG, Yang HS. Heparin-conjugated poly (lactic-co-glycolic acid) nanospheres enhance large-wound healing by delivering growth factors in platelet-rich plasma. *Artif Organs* 39, 388-94 (2015) DOI: 10.1111/aor.12389
- 109. Chen X, Peng LH, Li N, Li QM, Li P, Fung KP, Leung PC, Gao JQ. The healing and antiscar effects of astragaloside IV on the wound repair *in vitro* and *in vivo*. *J Ethnopharmacol* 139, 721-7 (2012)
 DOI: 10.1016/j.jep.2011.11.035
- 110. Chen X, Peng LH, Shan YH, Li N, Li QM, Wei W, Liang WQ, Gao JQ. Astragaloside IV-loaded nanoparticle-enriched hydrogel induces wound healing and anti-scar activity through topical delivery. *Int J Pharm* 447, 171-81 (2013)
- 111. Wolf NB, Kuchler S, Radowski MR, Blaschke T, Kramer KD, Weindl G, Kleuser B, Haag R, Schafer-Korting M. Influences of opioids and nanoparticles on in vitro wound healing models. *Eur J Pharm Biopharm* 73, 34-42 (2009)

DOI: 10.1016/j.ejpb.2009.03.009

112. Kuchler S, Wolf NB, Heilmann S, Weindl G, Helfmann J, Yahya MM, Stein C, Schafer-Korting M. 3D-wound healing model: influence of morphine and solid lipid nanoparticles. *J Biotechnol* 148, 24-30 (2010) DOI: 10.1016/j.jbiotec.2010.01.001

- 113. Chereddy KK, Her CH, Comune M, Moia C, Lopes A, Porporato PE, Vanacker J, Lam MC, Steinstraesser L, Sonveaux P, Zhu H, Ferreira LS, Vandermeulen G, Preat V. PLGA nanoparticles loaded with host defense peptide LL37 promote wound healing. *J Control Release* 194, 138-47 (2014) DOI: 10.1016/j.jconrel.2014.08.016
- 114. Bonferoni MC, Sandri G, Dellera E, Rossi S, Ferrari F, Mori M, Caramela C. Ionic polymeric micelles based on chitosan and fatty acids and intended for wound healing. Comparison of linoleic and oleic acid. *Eur J Pharm Biopharm* 87, 101-6 (2014) DOI: 10.1016/j.ejpb.2013.12.018
- 115. Singer AJ, Clark RA. Cutaneous wound healing. *N Engl J Med* 341, 738-46 (1999) DOI: 10.1056/NEJM199909023411006
- 116. DiPietro LA. Wound healing: the role of the macrophage and other immune cells. *Shock* 4, 233-40 (1995)
- 117. Danon D, Madjar J, Edinov E, Knyszynski A, Brill S, Diamantshtein L, Shinar E. Treatment of human ulcers by application of macrophages prepared from a blood unit. *Exp Gerontol* 32, 633-41 (1997) DOI: 10.1016/S0531-5565(97)00094-6
- 118. Barbul A, Regan MC. The regulatory role of T lymphocytes in wound healing. *J Trauma* 30, S97-100 (1990)
- 119. Li X, Wang H, Rong H, Li W, Luo Y, Tian K, Quan D, Wang Y, Jiang L. Effect of composite SiO (2) @AuNPs on wound healing: in vitro and vivo studies. J Colloid Interface Sci 445, 312-9 (2015) DOI: 10.1016/j.jcis.2014.12.084
- 120. Nadworny PL, Landry BK, Wang J, Tredget EE, Burrell. Does nanocrystalline silver have a transferable effect? *Wound Repair Regen* 18, 254-65 (2010)
 DOI: 10.1111/j.1524-475X.2010.00579.x
- 121. Widgerow AD. Nanocrystalline silver, gelatinases and the clinical implications. *Burns* 36, 965-74 (2010)
 DOI: 10.1016/j.burns.2010.01.010
- 122. Barata T, Teo I, Lalwani S, Simanek EE, Zloh M, Shaunak S. Computational design principles for bioactive dendrimer based constructs as antagonists of the TLR4-MD-

- 2-LPS complex. *Biomaterials* 32, 8702-11 (2011)
- DOI: 10.1016/j.biomaterials.2011.07.085
- 123. Archana D, Dutta J, Dutta PK. Evaluation of chitosan nano dressing for wound healing: characterization, *in vitro* and *in vivo* studies. *Int J Biol Macromol* 57, 193-203 (2013) DOI: 10.1016/j.ijbiomac.2013.03.002
- 124. Chereddy KK, Coco R, Memvanga PB, Ucakar B, des Rieux A, Vandermeulen G, Preat V. Combined effect of PLGA and curcumin on wound healing activity. *J Control Release* 171, 208-15 (2013) DOI: 10.1016/j.jconrel.2013.07.015
- 125. Flores FC, De Lima JA, Da Silva CR, Benvegnu D, Ferreira J, Burger ME, Beck RC, Rolim CM, Rocha MI, Da Veiga ML, Da Silva Cde B. Hydrogels Containing Nanocapsules and Nanoemulsions of Tea Tree Oil Provide Antiedematogenic Effect and Improved Skin Wound Healing. *J Nanosci Nanotechnol* 15, 800-9 (2015)
- 126. Wigglesworth KM, Racki WJ, Mishra R, Szomolanyi-Tsuda E, Greiner DL, Galili U. Rapid recruitment and activation of macrophages by anti-Gal/alpha-Gal liposome interaction accelerates wound healing. *J Immunol* 186, 4422-32 (2011) DOI: 10.4049/jimmunol.1002324
- 127. Ferreira AM, Mattu C, Ranzato E, Ciardelli G. Bioinspired porous membranes containing polymer nanoparticles for wound healing. *J Biomed Mater Res A* 102, 4394-405 (2014) DOI: 10.1002/jbm.a.35121
- 128. Guthrie KM, Agarwal A, Teixeira LB, Dubielzig RR, Abbott NL, Murphy CJ, Singh H, McAnulty JF, Schurr MJ. Integration of silver nanoparticle-impregnated polyelectrolyte multilayers into murine-splinted cutaneous wound beds. *J Burn Care Res* 34, e359-67 (2013)
 - DOI: 10.1097/BCR.0b013e31827e7ef9
- 129. Kawai K, Larson BJ, Ishise H, Carre AL, Nishimoto S, Longaker M, Lorenz HP. Calcium-based nanoparticles accelerate skin wound healing. *PLoS One* 6, e27106 (2011)

 DOI: 10.1371/journal.pone.0027106
- 130. Danila D, Johnson E, Kee P. CT imaging of myocardial scars with collagen-targeting

gold nanoparticles. Nanomedicine 9, 1067-76 (2013)

DOI: 10.1016/j.nano.2013.03.009

131. Blecher K, Martinez LR, Tuckman-Vernon C, Nacharaju P, Schairer D, Chouake J, Friedman JM, Alfieri A, Guha C, Nosanchuk JD, Friedman AJ. Nitric oxide-releasing nanoparticles accelerate wound healing in NOD-SCID mice. Nanomedicine 8, 1364-71

DOI: 10.1016/j.nano.2012.02.014

132. Dombi GW, Purohit K, Matin LM, Yang SC. Collagen gel formation in the presence of a carbon nanobrush. J Mater Sci Mater Med 26. 5356 (2015)

DOI: 10.1007/s10856-014-5356-4

- 133. Kwon MJ, An S, Choi S, Nam K, Jung HS, Yoon CS, Ko JH, Jun HJ, Kim TK, Jung SJ, Park JH, Lee Y, Park JS. Effective healing of diabetic skin wounds by using nonviral gene therapy based on minicircle vascular endothelial growth factor DNA and a cationic dendrimer. J Gene Med 14 272-8, (2012) DOI: 10.1002/jgm.2618
- 134. Peng LH, Wei W, Shan YH, Zhang TY, Zhang CZ, Wu JH, Yu L, Lin J, Liang WQ, Khang G, Gao JQ. beta-Cyclodextrin-Linked Polyethylenimine Nanoparticles Facilitate Gene Transfer and Enhance the Angiogenic Capacity of Mesenchymal Stem Cells for Wound Repair and Regeneration. J Biomed Nanotechnol 11, 680-90 (2015)
- 135. Peng LH, Wei W, Qi XT, Shan YH, Zhang FJ, Chen X, Zhu QY, Yu L, Liang WQ, Gao JQ. Epidermal stem cells manipulated by pDNA-VEGF165/CYD-PEI nanoparticles loaded gelatin/beta-TCP matrix as a therapeutic agent and gene delivery vehicle for wound healing. Mol Pharm 10, 3090-102 (2013) DOI: 10.1021/mp400162k
- 136. Jozic I, Daunert S, Tomic-Canic M, Pastar I. Nanoparticles for Fidgety Cell Movement and Enhanced Wound Healing. J Invest Dermatol 135, 2151-3 (2015) DOI: 10.1038/jid.2015.237
- 137. Park HJ, Lee J, Kim MJ, Kang TJ, Jeong Y, Um SH, Cho SW. Sonic hedgehog intradermal gene therapy using a biodegradable poly (beta-amino esters) nanoparticle to enhance wound healing. Biomaterials 33, 9148-56

DOI: 10.1016/j.biomaterials.2012.09.005

138. Li N, Luo HC, Yang C, Deng JJ, Ren M, Xie XY, Lin DZ, Yan L, Zhang LM. Cationic star-shaped polymer as an siRNA carrier for reducing MMP-9 expression in skin fibroblast cells and promoting wound healing in diabetic rats. Int J Nanomedicine 9, 3377-87 (2014)

DOI: 10.2147/IJN.S66368

139. Trentin D. Hall H. Weschler S. Hubbell JA. Peptide-matrix-mediated gene transfer of an oxygen-insensitive hypoxia-inducible factor-1alpha variant for local induction of angiogenesis. Proc Natl Acad Sci U S A 103, 2506-11 (2006)

DOI: 10.1073/pnas.0505964102

- 140. Bruch-Gerharz D, Ruzicka T, Kolb-Bachofen V. Nitric oxide and its implications in skin homeostasis and disease - a review. Arch Dermatol Res 290, 643-51 (1998)
- 141. Ammons MC. Anti-biofilm strategies and the need for innovations in wound care. Recent Pat Antiinfect Drug Discov 5, 10-7 (2010) DOI: 10.2174/157489110790112581#stha sh.nK0YIH8C.dpuf
- 142. Martinez-Ruiz A, Lamas S. S-nitrosylation: a potential new paradigm in signal transduction. Cardiovasc Res 62, 43-52
 - DOI: 10.1016/j.cardiores.2004.01.013
- 143. Weller RB. Nitric oxide-containing nanoparticles as an antimicrobial agent and enhancer of wound healing. J Invest Dermatol 129, 2335-7 (2009) DOI: 10.1038/jid.2009.149
- 144. Dave RN, Joshi HM, Venugopalan VP. Biomedical evaluation of a novel nitrogen oxides releasing wound dressing. J Mater Sci Mater Med 23, 3097-106 (2012) DOI: 10.1007/s10856-012-4766-4
- 145. Martinez LR, Han G, Chacko M, Mihu MR, Jacobson M, Gialanella P, Friedman AJ, Nosanchuk JD, Friedman JM. Antimicrobial and healing efficacy of sustained release nitric oxide nanoparticles against Staphylococcus aureus skin infection. J Invest Dermatol 129, 2463-9 (2009) DOI: 10.1038/jid.2009.95
- 146. Han G, Nguyen LN, Macherla C, Chi Y, Friedman JM, Nosanchuk JD, Martinez LR. Nitric oxide-releasing nanoparticles accelerate wound healing by promoting

- fibroblast migration and collagen deposition. Am J Pathol 180, 1465-73 (2012) DOI: 10.1016/j.ajpath.2011.12.013
- 147. Han G, Martinez LR, Mihu MR, Friedman AJ, Friedman JM, Nosanchuk JD. Nitric oxide releasing nanoparticles are therapeutic for Staphylococcus aureus abscesses in a murine model of infection. *PLoS One* 4, e7804 (2009)

 DOI: 10.1371/journal.pone.0007804
- 148. Mihu MR, Sandkovsky U, Han G, Friedman JM, Nosanchuk JD, Martinez LR. The use of nitric oxide releasing nanoparticles as a treatment against Acinetobacter baumannii in wound infections. *Virulence* 1, 62-7 (2010) DOI: 10.4161/viru.1.2.10038
- 149. Hsu CC, Peng CH, Hung KH, Lee YY, Lin TC, Jang SF, Liu JH, Chen YT, Woung LC, Qang CY, Tsa CY, Chiou SH, Chen SJ, Chang YL. Stem Cell Therapy for Corneal Regeneration Medicine and Contemporary Nanomedicine for Corneal Disorders. *Cell Transplant* 24, 1915-30 (2015) DOI: 10.3727/096368914X685744
- 150. Tao ZW, Favreau JT, Guyette JP, Hansen KJ, Lessard J, Burford E, Pins GD, Gaudette GR. Delivering stem cells to the healthy heart on biological sutures: effects on regional mechanical function. *J Tissue Eng Regen Med*, (2014) DOI: 10.1002/term.1904
- 151. Kodama A, Kamei N, Kamei G, Kongcharoensombat W, Ohkawa S, Nakabayashi A, Ochi M. In vivo bioluminescence imaging of transplanted bone marrow mesenchymal stromal cells using a magnetic delivery system in a rat fracture model. *J Bone Joint Surg Br* 94, 998-1006 (2012)

 DOI: 10.1302/0301-620X.94B7.28521
- 152. Shi J, Zhang X, Zhu J, Pi Y, Hu X, Zhou C, Ao Y. Nanoparticle delivery of the bone morphogenetic protein 4 gene to adipose-derived stem cells promotes articular cartilage repair *in vitro* and *in vivo*. *Arthroscopy* 29, 2001-2011 e2 (2013) DOI: 10.1016/j.arthro.2013.09.076
- 153. Keeney M, Deveza L, Yang F. Programming stem cells for therapeutic angiogenesis using biodegradable polymeric nanoparticles. *J Vis Exp* 79, e50736 (2013) DOI: 10.3791/50736.

- 154. Soderstjerna E, Johansson F, Klefbohm B, Englund Johansson U. Gold- and Silver- nanoparticles affect the growth characteristics of human embryonic neural precursor cells. *PLoS One* 8, e58211 (2013)
- 155. Liu X, He W, Fang Z, Kienzle A, Feng Q. Influence of silver nanoparticles on osteogenic differentiation of human mesenchymal stem cells. *J Biomed Nanotechnol* 10, 1277-85 (2014)
- 156. Ilie I, Ilie R, Mocan T, Bartos D, Mocan D. Influence of nanomaterials on stemmm cell differentiation: designing an appropriate nanobiointerface. *Int J Nanomedicine* 7, 2211-2225 (2012)
- 157. Kharlamov AN, Tyurnina AE, Veselova VS, Kovtun OP, Shur VY, Gabinsky JL. Silicagold nanoparticles for atheroprotective management of plaques: results of the NANOM-FIM trial. *Nanoscale* 7, 8003-15 (2015)
 DOI: 10.1039/c5nr01050k
- 158. Adhya A, Bain J, Ray O, Harza A, Adhikari S, Dutta S, Ray S, Majumdar BK. Healing of burn wounds by topical treatment: A randomized controlled comparison between silver sulfadiazine and nano-crystalline silver. *J Basic Clin Pharm* 6, 29-34 (2014) DOI: 10.4103/0976-0105.145776
- 159. Harding K, Gottrup F, Jawien A, Mikosinski J, Twardowska-Saucha K, Kaczmarek S, Sopata M, Shearman C, Pieronne A, Kommala D. A prospective, multicentre, randomised, open label, parallel, comparative study to evaluate effects of AQUACEL (R) Ag and Urgotul (R) Silver dressing on healing of chronic venous leg ulcers. *Int Wound J* 9, 285-94 (2012) DOI: 10.1111/j.1742-481X.2011.00881.x
- 160. Krieger BR, Davis DM, Sanchez JE, Mateka JJ, Nfonsam VN, Frattini JC, Marcet JE. The use of silver nylon in preventing surgical site infections following colon and rectal surgery. *Dis Colon Rectum* 54, 1014-9 (2011) DOI: 10.1097/DCR.0b013e31821c495d
- 161. Miller CN, Newall N, Kapp SE, Lewin G, Karimi L, Carville K, Gliddon T, Santamaria NM. A randomized-controlled trial comparing cadexomer iodine and nanocrystalline silver on the healing of leg ulcers. Wound Repair Regen 18, 359-67 (2010)
 DOI: 10.1111/j.1524-475X.2010.00603.x

- 162. Oram Y, Kahraman F, Karincaoglu Y, Koyuncu E. Evaluation of 60 patients with pilonidal sinus treated with laser epilation after surgery. *Dermatol Surg* 36, 88-91 (2010)
 DOI: 10.1111/j.1524-4725.2009.01387.x
- 163. Silver GM, Robertson SW, Halerz MM, Conrad P, Supple KG, Gamelli RL. A silver-coated antimicrobial barrier dressing used postoperatively on meshed autografts: a dressing comparison study. *J Burn Care Res* 28, 715-9 (2007)
 DOI: 10.1097/BCR.0B013E318148C9E4
- 164. Chen J, Han CM, Lin XW, Tang ZJ, Su SJ. (Effect of silver nanoparticle dressing on second degree burn wound) *Zhonghua Wai Ke Za Zhi* 44, 50-2 (2006)
- 165. Chen J, Han CM, Su GL, Tang ZJ, Su SJ, Lin XW. Randomized controlled trial of the absorbency of four dressings and their effects on the evaporation of burn wounds. *Chin Med J (Engl)* 120, 1788-91 (2007)

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