

Review

The Significance of the Ocular Adverse Effect Induced by Systemic Taxane Application

Ya-Ting Ye^{1,2,3,†}, Zi-Yi Zhou^{1,†}, Li-Shi Wen¹, Yu Sun², Zhao-Jie Chu^{2,3,*}, Guo-Rui Dou^{1,*}

¹Department of Ophthalmology, Eye Institute of Chinese PLA, Xijing Hospital, Fourth Military Medical University, 710032 Xi'an, Shaanxi, China

²Department of Chinese Materia Medica, College of Life Sciences, Northwestern University, 710069 Xi'an, Shaanxi, China

³Department of Ophthalmology, The First Affiliated Hospital of Northwest University (Xi'an First Hospital), 710002 Xi'an, Shaanxi, China

*Correspondence: fierywang@126.com (Guo-Rui Dou); chuzhaojie2009@163.com (Zhao-Jie Chu)

†These authors contributed equally.

Academic Editor: Shikun He

Submitted: 7 March 2022 Revised: 20 April 2022 Accepted: 22 April 2022 Published: 31 May 2022

Abstract

In recent years, in-depth research on anti-tumor therapy has brought the emergence of new active chemotherapeutic agents and combination regimens. However, as one of them, taxane drugs are widely used in clinical practice, but it should be noted that many side reactions caused by their application bring some difficulties to routine management. Among the side reactions related to taxane anti-tumor therapy, ocular adverse reactions are occasionally reported and are not life-threatening but may seriously affect patients' life quality. Thus, the continuation, reduction and cessation of taxane chemotherapy still need to be further evaluated by ophthalmologists and oncologists once the side effects show up. To prevent ocular side reactions, close attention should be paid to complications during medication. To facilitate the oncology department and ophthalmologists to comprehensively understand the ophthalmic adverse reactions of taxane drugs and their possible mechanisms and improve drug use efficiency, we collected relevant literature and reviewed and provided some suggestions for the monitoring and managing of ophthalmic toxicity.

Keywords: paclitaxel; taxane; eye diseases; drug-related side effects and adverse reactions

1. Introduction

As the first natural phytochemical drug approved by Food and Drug Administration (FDA), paclitaxel (Taxol) is an anti-tumor drug with high efficacy and low toxicity. Paclitaxel is used to treat breast cancer, ovarian, lung cancer, sarcoma, lymphoma and other diseases [1,2]. Breast cancer is the most common cancer among women in most countries. The number of women diagnosed with breast cancer will increase to almost 3.2 million per year by 2030 [3]. Taxane is a first- and second-line anticancer drug in treating breast cancer, ovarian cancer and non-small cell lung cancer. However, served as an anti-tumor drug, the number of reports on Paclitaxel's adverse side effects is increasing rapidly, mainly including bone marrow suppression, allergic reaction, digestive system reaction, numbness of the hand and foot, liver injury, and etc. Moreover, in recent years, there have been increasing reports of adverse ocular reactions caused by taxane drugs, such as vision loss [4], dry eye [5,6], lacrimal duct obstruction [7], conjunctivitis [8], glaucoma [9], optic neuropathy [10–12], epiphora [13], macular edema [14,15], blindness caused by combination with cisplatin [16], and so on. Therefore, clinicians must be aware of potential vision-threatening complications [17]. Prompt consultation with an ophthalmologist can lead to early detection, proper diagnosis, and appropriate therapeutic measures. Dose reduction or discontinuation of incriminated drugs may help reduce the severity and the dura-

tion of side effects. Generally, the ocular side effects caused by taxane chemotherapy can be relieved after stopping the administration of chemotherapy [18]. All those side effects mentioned above may not threaten the life span of the patients while may cause various troubles in their life after anti-cancer treatment. Herein, we review the relevant literatures and reports to conclude a comprehensive understanding of the ocular adverse side effects caused by taxane drugs, and also discuss underlining toxic mechanisms in order to provide a few instructions to its clinical administration.

2. Taxane

Paclitaxel is the first microtubule-stabilizing drug of the taxane family isolated from the yew root. In 1967, Mansukh Wani and Monroe Wall isolated and identified the active ingredient from the bark of *T. brevifolia*. They named it taxol, based on its species of origin and the presence of hydroxyl groups [19]. Paclitaxel entered phase I clinical trials in 1894 and phase II clinical trials in 1895. In the United States, the Phase III clinical trial was completed in-ears from 1983 and was approved by the FDA in 1992 [1]. Paclitaxel has been obtained by chemical semi-synthesis, total chemical synthesis, tissue and cell culture, microbial fermentation, and biosynthesis. In 1995, docetaxel, the second family member, was waived for medical use. Due to the poor water-solubility of paclitaxel be-



fore, solvent-increasing polyoxyethylene castor oil and anhydrous ethanol will be added when it is used [20]. As the solvent cause severe allergic reactions, there were various limitations to clinical use.

Currently, the paclitaxel family includes traditional paclitaxel, paclitaxel liposome, paclitaxel nanoparticle albumin-bound (NAB) (Abraxane), and docetaxel (Taxotere). Unlike other microtubule-stabilizing anticancer drugs which prevent the assembly of tubulin into microtubules, it is a microtubule stabilizer (anticontractile agent) and a mitotic inhibitor (antiproliferative agent) [19]. Paclitaxel promotes the assembly of tubulin to microtubules and prevents the dissociation of microtubules and blocks cell cycle progression, prevents mitosis, and inhibits the growth of cancer cells [1]. Thus, paclitaxel has become a widely accepted chemotherapeutic drug in treating ovarian cancer, breast cancer, non-small cell lung cancer, and other malignant solid tumors [2].

3. The Symptoms of Ocular Side Effects Induced by Taxane

According to the Drug Instructions, the injectable paclitaxel (albumin-bound) specification states that eye/visual adverse events occurred in 13% population (48/366) of clinical studies in the United States and Europe, while severe cases occurred in 1% population. The major diseases were keratitis and blurred vision, which were usually reversible. In a phase I clinical study of Chinese patients, 1 out of 104 patients developed transient blurred vision and diplopia. In a randomized controlled clinical study of Chinese patients with metastatic breast cancer, 4 out of 100 patients developed mild blurred vision, which was transient and self-healing. Noguchi has conducted a retrospective study of risk factors concerning ocular disorders caused by paclitaxel and NAB-paclitaxel [21]. This retrospective study targeted patients who were newly treated with paclitaxel or NAB-paclitaxel at Kyoto Okamoto Memorial Hospital between April 1 2012, and March 31 2017 of 128 subjects, 13 (10.2%) had ocular disorders with symptoms ranging from grades 1 and 2 (Eye disorders and peripheral were graded by using the Common Terminology Criteria for Adverse Events version are grade 1, which are asymptomatic, asymptomatic only detected during examination, or asymptomatic but not affecting function without treatment. Grade 2 is characterized by symptoms that affect function but do not interfere with daily activities, requiring correction or simple drug treatment). The symptoms mainly of these 13 patients included conjunctivitis or subconjunctival hemorrhage (3.1%), visual acuity reduction (2.3%), blurred vision and eye pain (1.6% each), eye mucus, blepharitis, sty, watering eyes, photopsia, and muscae volitantes (0.8% each). Meanwhile, Noguchi conducted a retrospective study on the risk factors of eye diseases caused by docetaxel administration every 3 weeks [22]. This case-control study targeted newly administered docetaxel patients at the Ky-

oto Okamoto Memorial Hospital between July 1 2015 and June 30 2018. Of the 89 subjects, 7 (7.9%) had eye disorders. Another retrospective analysis of clinical characteristics, treatment regimens, and concurrent systemic adverse reactions was performed in patients treated with taxanes at a single center from January 1 2010 to February 29 2020. In this study, 22 (1.1%) of 1918 patients experienced ophthalmic side effects [23]. Cancer chemotherapy has the potential to produce acute and chronic damage in any organ system. However, some organs are more sensitive than others. In this context, the eye is usually considered a sanctuary site, but has a potentially high degree of sensitivity to toxic substances. Ocular toxicity induced by cancer chemotherapy is not uncommon. Still, the broad spectrum of reactions to injury displayed by the eye reflects the unique anatomical, physiological, and biochemical features of this essential organ.

4. The Ocular Disorders Induced by Taxane (Table 1)

4.1 Ocular Surface Diseases

The most common symptoms of ocular adverse effects are discomfort of the eyelids, conjunctiva, and lacrimal gland organs, including dry eye [5,6], epiphora [13], nasolacrimal duct obstruction [29], lacrimal duct obstruction [29–31], erosive conjunctivitis and punctate stenosis [8], corneal epithelial lesions [27,32], chalazion [26], corneal disorder [27] and limbal stem cell deficiency [7]. According to previous reports, the nasolacrimal duct obstruction may be related to the interstitial fibrosis of lacrimal duct mucosa [29]. It was presumable that conjunctiva, keratopathy, and dry eye might be associated with the cytotoxicity of the drug. The drug inhibits cell proliferation in the cornea and eye surface, leading to stem cell dysfunction, and then triggers inflammation of the conjunctiva and epithelial defects [32]. Docetaxel-induced meibomian duct inflammation and blockage are the likely causes of chalazion [26]. However, the mechanisms of the ocular surface and accessory organ injury still need to be investigated.

4.2 Cataract

Intraocular tissue (lens and vitreous) damage caused by taxane is rare. Kuwata *et al.* [28] evaluated the ocular toxicity of different doses of paclitaxel to Sprague-Dawley (SD) rats of different ages. The results showed that the amount of 4 or 8 mg/kg of paclitaxel injected into 0-day-old rats resulted in apoptotic epithelial cells of the lens, and the lens fibers were degenerative on the 7th day. These phenomena suggested the possible occurrence of cataracts. However, no ocular lesions were observed being injured at the dose of 2 mg/kg of paclitaxel injection. Also, no ocular lesions were observed injured in rats injected intraperitoneally with 4 mg/kg paclitaxel at 14 days of age and 8 mg/kg paclitaxel at 12 to 18 weeks of age. These results suggested that the ocular toxicity of paclitaxel may be

Table 1. Ocular disorders and patient complaints caused by taxane chemotherapy.

Ocular disorders	Main symptoms	Incidence rate
Dry eyes/ocular surface discomfort	Foreign body sensation Grit or sand in eyes	51.7% [6]
Limbal stem cell deficiency	Eye pain; Blurred vision in both eyes [7]	NA
Conjunctivitis	Eye redness and lacrimation [8]	NA
Glaucoma	Vision loss [9,24]	NA
Optic Neuropathy	Transient obscurations of vision lasting seconds [10]	NA
	Vision reduce, narrowed field of vision, Color vision is impaired [11]	
	Loss of visual acuity with blind spots [25]	
Epiphora	Not described	12.5% [13]
		7.9% [22]
Macular edema	Visually impaired/Sudden, catastrophic vision loss	0.5% [14]
Chalazion	Eye irritation and dryness [26]	NA
Corneal disorder	Blurred vision [27]	NA
Cataract	Not described [28]	NA

NA, not applicable.

dose-related and age-related, and attention should be paid to the ocular toxicity of paclitaxel in the early developmental stage. But the mechanisms are still underwater.

4.3 Macular Edema

The first symptom of macular edema caused by taxane is impaired visual acuity. Kaya *et al.* [14] analyzed 202 patients who received taxane-based therapy to treat various cancer. Taxane-related cystoid macular edema (CME) was detected only in one patient on paclitaxel. Generally, the proportion of taxane-related maculopathy was 0.5% (1/202) of all patients in the taxane group. In a case report and literature review shared by Alvarez-Fernandez *et al.* [33], a 73-year-old patient with metastatic breast cancer developed macular edema after being treated with paclitaxel, which resolved after drug withdrawal. In addition, 57 cases of paclitaxel-related macular edema reported in 52 literatures were analyzed. The median time of occurrence of macular edema after taking the drug among these patients was 4.25 months. Among these patients, 92.86% were diagnosed with macular edema and bilateral vision loss in the initial examination, and most of their symptoms reversed after paclitaxel withdrawal. So far, we have retrieved 18 cases of taxane-induced macular edema that spontaneously resolved after stopping chemotherapy [33–48].

However, this particular macular edema, unlike another common macular edema such as diabetic macular edema (DME), macular edema secondary to retinal vein occlusion or uveitis, presents unique clinical characteristics. In taxane-induced macular edema, optical coherence tomography (OCT) showed intraretinal cystoid spaces and dome-shaped configuration of the fovea in both eyes. However, there was no evidence of any fluorescein leakage shown on fluorescein angiography (FA) in both eyes. In addition, most taxane-induced macular edema will resolve

spontaneously after drug withdrawal. The pathogenesis of this type of macular edema is still unclear. So far, its specific mechanisms are not well studied, except for several speculations in several case reports [4,15,34,36,38,41,49–56]. Among them, the primary underlining mechanism is mainly focused on the toxicity of Müller cells and etinal pigment epithelial (RPE) cells in the retina. Paclitaxel may lead to cellular dysfunction and accumulation of intracellular fluid [4], resulting in the disturbing of the nerve sensory osmotic gradient maintained by Müller cells in the retina. Also, paclitaxel would cause minimal impairment to the blood-retina barrier (BRB), where RPE cells play an essential role in maintaining and participating in phagocytosis and visual circulation metabolism [15].

4.4 Retinal Dysplasia

Meanwhile, through experiments on animals, Kuwata *et al.* [28] found that ocular toxicity of different doses of paclitaxel to newborn SD rats of different ages also showed that retinal dysplasia occurred after intraperitoneal injection of 4 or 8 mg/kg paclitaxel in 0-day-old SD rats. This suggested that the ocular side effects of paclitaxel were related to the duration and concentration of the drug.

4.5 Optic Neuropathy

Hofstra *et al.* [25] mentioned a case where an ovarian cancer patient experienced a sudden vision loss with scotomas after receiving six chemotherapy cycles of intraperitoneal paclitaxel infusion. Physical examination by an ophthalmologist revealed bilateral left-sided hemianopsia because of a prolonged vascular spasm resulting in a neurological defect of the optical cortex. When paclitaxel was infused, the patient experienced scotomata, small luminous dots or “flies” in the visual fields of both eyes, lasting a few minutes to several hours. The speculation is that pa-

clitaxel may damage the optic nerve [57]. Sediman *et al.* [10] estimated that among 25 breast cancer patients who received paclitaxel at 250–275 mg/m² for the first time, 6 of them experienced the flashing of stars or fireworks in the whole field of vision 3 h after the chemotherapy which usually lasts 15 min to 3 h [10]. After the attack, there were no significant chronic sequelae. And this phenomenon occurred again when the same or slightly lower dose of chemotherapy (less than 275 mg/m² but not less than 250 mg/m²) was received but wouldn't appear when the amount was less than 250 mg/m². It was considered a transient optic nerve vasogenic reaction induced by paclitaxel, dose-dependent and reversible. Visual electrophysiology was also evaluated in 14 breast cancer patients undergoing paclitaxel chemotherapy. ERG b-wave latency increased significantly. 7 patients showed abnormal ERG, oscillating potentials, 30 Hz flashing light response, and visual evoked potentials (VEP) monitoring in different combinations. Twelve patients presented transient dark spots and blurred vision with abnormal oscillating potentials. The mechanisms of visual symptoms and electrophysiological changes during paclitaxel administration are vascular dysregulation in the retina or ischemic mechanisms when the optic nerve is involved [58]. In a study of 47 patients who received paclitaxel for non-small lung cancer, three showed abnormal VEP, showing a significant decrease in P100 amplitude and a slight increase in latency. The abnormal growth in P100 latency of VEP is considered to be typical demyelinating optic neuropathy [59].

It should be noted that optic neuropathy may also be induced by taxane-induced glaucoma, although further study was needed. A young female patient with metastatic breast cancer who underwent chemotherapy of docetaxel in combination with both low-dose and high-pulse corticosteroid treatment. She developed symptomatic vision loss during the course of treatment and had high intraocular pressure, optic nerve cupping, and bilateral visual-field loss and was then diagnosed as open-angle glaucoma [9]. As docetaxel was discontinued with local treatment by a β -blocker drop (betaxolol), her intraocular pressure decreased. Steroid-induced glaucoma should be considered [24]. However, as the patient was treated again with docetaxel and without corticosteroids, her fluid retention occurred after two cycles and open-angle glaucoma recurred with intraocular pressures of 35 mmHg and 40 mmHg after the third cycle. Therefore, the author assumed that taxane-induced glaucoma was the most probable diagnosis.

5. The Mechanisms of Ocular Side Effects Induced by Taxane

5.1 The Effects of Taxane on Cell Proliferation and Apoptosis

Paclitaxel inhibited cell viability and proliferation in a dose-dependent manner. Paclitaxel may inhibit the proliferation, differentiation, and apoptosis of human adi-

pose stem cells (hASC) regulated by the tumor necrosis factor- α (TNF- α) pathway [60]. At the same time, paclitaxel can significantly inhibit endothelial cell differentiation and the formation of new blood vessels, resulting in poor wound healing in cancer patients receiving chemotherapy [60]. Upon paclitaxel treatment, caspase-2, caspase-3, and caspase-8 were activated in BCBL-1 cells. The upregulation of Bax also accompanied Paclitaxel-induced apoptosis. Also, paclitaxel downregulated the expression of Bcl-2 protein slightly. Paclitaxel-induced apoptosis of BCBL-1 cells is primarily mediated by the caspase activation pathway [61]. Paclitaxel could inhibit breast cancer cells proliferation and invasion and promote MCF-7 cell apoptosis by downregulating the expression of p-AKT (Thr308) and p-AKT (Ser473) in the PI3K/AKT signaling pathway [62]. A few studies investigated the effect on non-tumor cells like endothelial cells [63–65]. Further studies are needed on the pathway through which paclitaxel affects the proliferation and apoptosis of RPE cells and Müller cells.

5.2 The Effects of Taxane on Microtubule

Microtubules are involved in many vital functions like signal transduction and intracellular transport. Paclitaxel can bind to the interior of microtubules, thus making them more stable, interfering with the regular cycling of microtubule depolymerization and repolymerization. In dividing cells such as cancer cells, this disrupts normal spindle dynamics, interferes with mitosis, and ultimately leads to cell death [66]. Meanwhile, paclitaxel can interfere with normal cellular functions, including cell division and microtubule-dependent intra-cellular and axonal transport. Axons are enriched in microtubules where increased microtubule stability may cause neurotoxicity. Microtubule-dependent transport is critical for axonal function and health, both in the central and peripheral nervous systems; as such, disruption of axonal transport is implicated in various central nervous system disorders and peripheral neuropathies [67,68]. Thus, neurotoxicity could be induced primarily by affecting local effects in the distal axon.

5.3 The Effects of Taxane on Oxidative and Mitochondrial Stress

It has been reported that paclitaxel treatment increased the production of reactive oxygen species (ROS) in BCBL-1 cells, resulting in cytotoxicity. As is known to all, caspase-2 was activated in paclitaxel-treated BCBL-1 cells, and caspase-2 can induce mitochondrial dysfunction directly or indirectly [61]. In addition, paclitaxel can also induce mitochondrial swelling and transport dysfunction in peripheral nerves [69,70]. It impairs not only the axonal transport of mitochondria but also their morphology and function. The paclitaxel-induced alterations in the permeability transition pore opening in mitochondria then cause changes in calcium flow. Calcium flow changes induce deficiencies in the mitochondrial respiratory chain, which

leads to ATP deficits. Furthermore, the response to oxidative stress is impaired by paclitaxel treatment, further increasing the ATP deficits [71]. Mitochondrial damage and ROS are closely related since mitochondrial damage causes the production of ROS, i.e., H_2O_2 formation. ROS, in turn, causes damage to the mitochondria, inducing DNA fragmentation and mitochondrial membrane potential loss [72]. Hence, ROS plays a crucial role in the oxidative stress reaction and is upregulated after paclitaxel therapy [73]. In the end, paclitaxel decreases retrograde endosomal/lysosomal flux at late cell stages [69].

5.4 The Effects of Taxane on Cell Degeneration

Paclitaxel causes degeneration of both central and peripheral axon branches of dorsal root ganglia in mice, but this is dose-dependent. At lower doses (lower than the maximum tolerated dose), dark inclusions, as well as clear vacuolations in the cytoplasm of neurons and satellite cells, can be observed. At relatively high doses (at the maximum tolerated dose), a proportion of the cytoplasm of dorsal root ganglion (DRG) neurons appeared much darker than that of normal neurons, and a few degenerating neuronal cells were reported [66,70]. The enhanced stabilization of the microtubule is likely to be the effect of paclitaxel that initiates the downstream neurotoxic, while the degeneration of DRG neurons caused by taxol is mainly due to paclitaxel changes in tubulin modifications [69].

5.5 The Effects of Taxane on Stem Cell Regeneration

Paclitaxel induces the classical development of stem cell-like cells. It promotes the regeneration of neurons, which may lead to rapid and sustainable functional improvement in Spinal cord injury (SCI) rats. When it comes to the differentiation of neural stem cells, researchers found that paclitaxel reduces scar formation after SCI and enhances intrinsic axonal regeneration. Paclitaxel-triggered neuronal differentiation occurs through the Wnt/ β -catenin signaling pathway [74,75]. However, paclitaxel diminishes adipose-derived stem cells (ASCs) function. The cytotoxic effects of paclitaxel on ASCs should not be overlooked, for it may promote apoptosis and inhibit cell proliferation. At the same time, some functions can be restored in a short time after yew is eluted [76]. Meanwhile, bone marrow-derived mesenchymal stem cells (MSCs) were also susceptible to the taxane compound taxol. While MSCs remained metabolically viable, they were severely impaired in both their proliferation and their functional capabilities after exposure to paclitaxel. Paclitaxel reduced cell migration, delays in cellular adhesion, and significant dose-dependent inhibition of the stem cells' multi-lineage differentiation potential [77]. All these mentioned above were the researches accomplished in another disease model. The specific mechanisms of stem cell regeneration in ocular side effects induced by paclitaxel are still unclear.

5.6 The Effects of Taxane on Ischemia

Paclitaxel chemotherapy may cause chest pain, and bradycardia, resulting in myocardial ischemia. Symptoms improved when chemotherapy was stopped [78]. In addition, it may reduce ischemic ventricular arrhythmias (Vas) by stabilizing microtubules to protect connexin 43 (Cx43), and the dynamic movement of Cx43 is regulated by microtubules and their related proteins. Dysfunction of Cx43 in ischemic myocardium is associated with Vas [79]. Also, cerebral ischemia is another side effect that should be paid enough attention to [80]. On the other hand, paclitaxel enhances the myocardial protective effect of myocardial ischemia preconditioning through stabilizing microtubules of cardiomyocytes and promoting HIF-1 α localization in the nucleus [81]. As is known to all, many ocular diseases are caused by retinal ischemia, and paclitaxel therapy is reported to be related to ischemia in other tissues. That's why we consider ischemia as potential damage that paclitaxel caused to the retina.

5.7 The Effects of Taxane on Cellular Junctions

Intercellular communication occurs through specific membrane pores called gap junctions. Taxol significantly affects gap-junction-mediated intercellular communication (GJIC) by impairing the transfer of Cx43 to the plasma membrane by stabilizing microtubules. Reversibility of taxol-induced reduction of GJIC as well as microtubular and Cx43 arrangements were demonstrated after taxol depletion from the nutrient medium. However, taxol does not immediately reduce GJIC, depleting connexin reserves and thus affecting GJIC [82]. Müller cells and RPE are critical to sustaining the BRB. As we mentioned before, taxane may disturb the functions of these cells. Thus the cellular junctions may be influenced as well.

5.8 The Effects of Taxane on Tissue Fibrosis

Paclitaxel has an anti-fibrosis effect. In the thioacetamide (TAA)-induced rat liver fibrosis model, paclitaxel prevented TAA-induced liver fibrosis in rats, possibly by inhibiting the expression of transforming growth factor- β 1 (TGF- β 1) and platelet-derived growth factor-BB (PDGF-BB) and subsequently suppressing the apoptosis and the manifestation of tissue inhibitor of metalloproteinase-1 (TIMP-1) [83]. However, as a result of the systemic effects of the docetaxel drug, puncta and canaliculi undergo fibrosis, which in turn causes tears [84]. At the same time, docetaxel chemotherapy can cause extensive fibrosis of the lacrimal sac and the nasal mucosa resulting in nasolacrimal duct obstruction [29]. Whether similar phenomena can be observed in ocular tissues is still required to be explored.

6. Prevention and Treatment

For patients under taxane administration, it is recommended to have a baseline examination of ophthalmology before treatment, including visual acuity, intraocular pres-

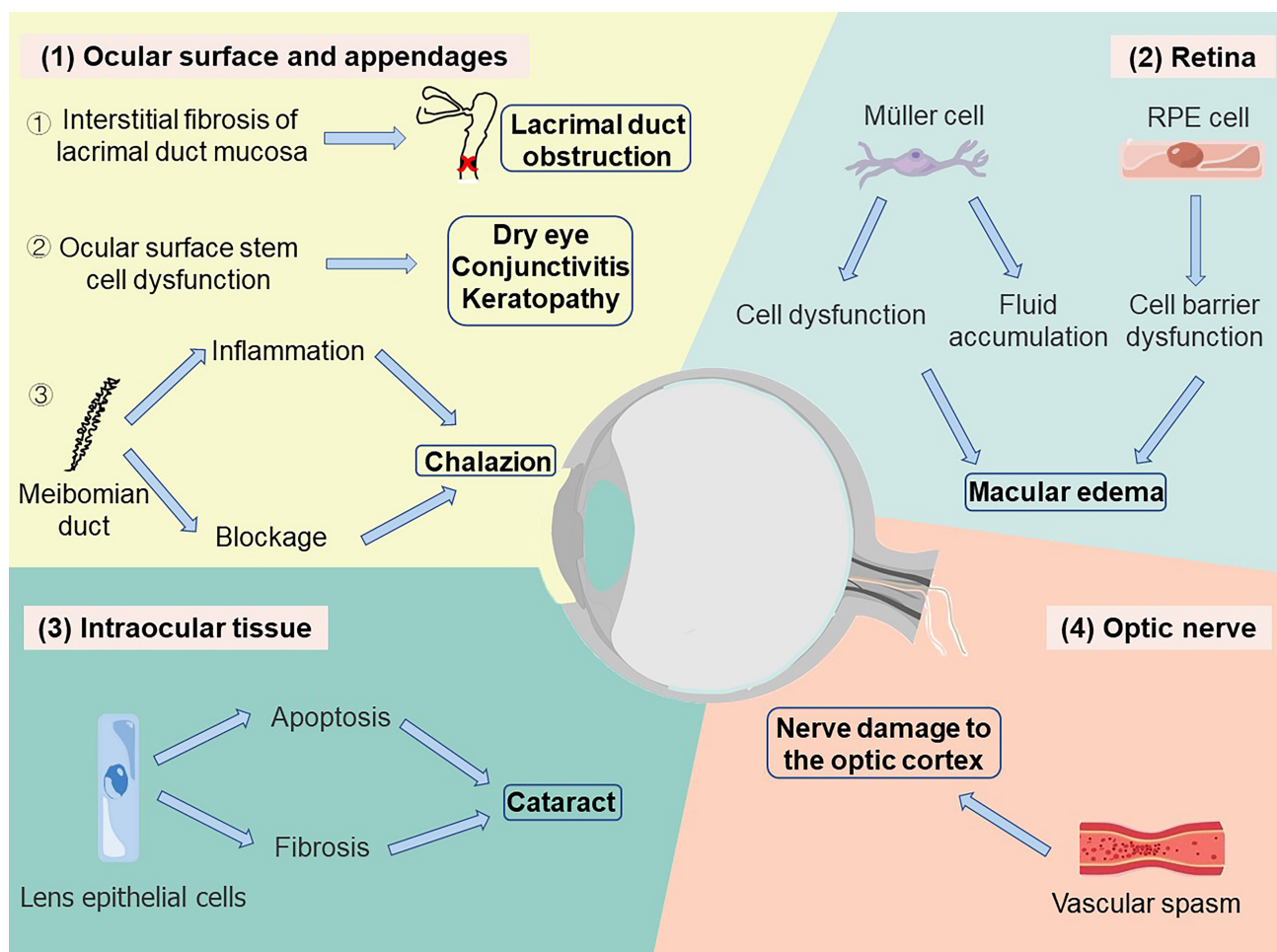


Fig. 1. Taxane-induced ocular diseases and their mechanisms. (1) Diseases in ocular surface: ① Taxane promotes interstitial fibrosis of lacrimal duct mucosa and leads to lacrimal duct obstruction. ② Taxane inhibits cell proliferation in the cornea and eye surface, causing the dysfunction of stem cells, which may result in dry eye, conjunctivitis, and keratopathy. ③ Taxane leads to chalazion by inducing the inflammation and blockage of the meibomian duct. (2) Diseases in retina: Taxane results in the dysfunction of Müller cells and the accumulation of intracellular fluid as well as damage of RPE barrier function which might cause further macular edema. (3) Diseases in intraocular tissue: Taxane induces the apoptosis of lens epithelial cells and the formation of lens fibers, leading to cataracts. (4) Diseases in optic nerve: Taxane injection may cause transient vascular spasm, resulting in neurological defects of the optical cortex.

sure, fundus, color vision, visual field examination, and etc. and to closely observe whether there are new ocular symptoms during the treatment process. Patients with ocular symptoms in the course of medication should be referred to have ocular consultancy and have a complete ocular checkup in time. For the ocular surface diseases caused by paclitaxel drugs, such as dry eye, conjunctivitis, keratitis, and etc. Usually, it is recommended to use artificial tears, non-steroidal anti-inflammatory drugs, antibiotic eye ointment and other symptomatic treatments. Lacrimal duct obstruction or narrow, tear overflow can be used for lacrimal duct irrigation or lacrimal duct exploration [32]. The adverse side effects on the ocular surface caused by paclitaxel are reversible and can be recovered after withdrawal and symptomatic treatment, within several days to 5 months [32].

7. Conclusion and Perspective

In conclusion, the adverse ocular side effects of taxane drugs mainly appear on the ocular surface, ocular appendage and intraocular tissues, with various clinical manifestations such as dry eye, conjunctivitis, keratitis, macular edema, retinal injury, optic nerve injury, and etc (see in Fig. 1). The underlining mechanisms remain to be further investigated. The macular edema induced by taxane drugs may be related to the dysfunction of RPE cells and Müller cells. Other adverse reactions may be associated with the cytotoxicity of paclitaxel. If baseline eye exams were available, it would be helpful for clinicians to identify and diagnose whether eye disorders are caused by taxane chemotherapy. The concern on baseline exams has been raised in several cases [23,85]. We also highlight those ocular symptoms should be closely observed during taxane

chemotherapy. Chemotherapy regimens should be adjusted according to comprehensive evaluation, and symptomatic treatments should also be given to avoid irreversible damage.

Author Contributions

YTY and ZYZ wrote the manuscript. LSW, and YS participate to assist this work. GRD and ZJC reviewed the manuscript. GRD provided thoughtful comments and suggestions during this review preparation. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Thanks to all the peer reviewers for their opinions and suggestions.

Funding

This article is supported by the National Natural Science Foundation of China (81970814, 81670863), Professional Promotion Program of Xijing Hospital (XJZT19ML19), Key Research and Development Program of Shaanxi Province (2021SF-335), Science and Technology Project of Xi'an Health Committee (2020yb07).

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Weaver BA. How Taxol/paclitaxel kills cancer cells. *Molecular Biology of the Cell*. 2014; 25: 2677–2681.
- [2] Marupudi NI, Han JE, Li KW, Renard VM, Tyler BM, Brem H. Paclitaxel: a review of adverse toxicities and novel delivery strategies. *Expert Opinion on Drug Safety*. 2007; 6: 609–621.
- [3] Ginsburg O, Bray F, Coleman MP, Vanderpuye V, Eniu A, Kotha SR, *et al*. The global burden of women's cancers: a grand challenge in global health. *The Lancet*. 2017; 389: 847–860.
- [4] Joshi MM. Paclitaxel Maculopathy. *Archives of Ophthalmology*. 2007; 125: 709.
- [5] Gore M, Rustin G, Slevin M, Gallagher C, Penson R, Osborne R, *et al*. Single-agent paclitaxel in patients with previously untreated stage IV epithelial ovarian cancer. *British Journal of Cancer*. 1997; 75: 710–714.
- [6] Chiang JCB, Goldstein D, Trinh T, Au K, Park SB, Krishnan AV, *et al*. A cross-sectional study of ocular surface discomfort and corneal nerve dysfunction after paclitaxel treatment for cancer. *Scientific Reports*. 2021; 11: 1786.
- [7] Sekhon A, Wang JYF, Tan JCH, Holland SP, Yeung SN. Limbal stem cell deficiency secondary to systemic paclitaxel (Taxol) for breast cancer: a case report. *BMC Ophthalmology*. 2020; 20: 400.
- [8] Skolnick CA, Doughman DJ. Erosive Conjunctivitis and Punctal Stenosis Secondary to Docetaxel (Taxotere) Eye & Contact Lens: Science & Clinical Practice. 2003; 29: 134–135.
- [9] del Pilar Bernal M, Loftfield K, Nussdorf JD. Taxane-induced glaucoma. *The Lancet*. 2000; 355: 577.
- [10] Seidman AD, Barrett S, Canezo S. Photopsia during 3-hour paclitaxel administration at doses \geq 250 mg/m². *Journal of Clinical Oncology*. 1994; 12: 1741–1742.
- [11] Noguchi Y, Kawashima Y, Kawara H. An Undeniable Case of Optic Neuropathy Due to Cabazitaxel. *Cancer and Chemotherapy*. 2016; 43: 777–779.
- [12] Yuan X, Feng Y, Li D, Li M. Unilateral visual impairment in a patient undergoing chemotherapy: a case report and clinical findings. *BMC Ophthalmology*. 2019; 19: 236.
- [13] Noguchi Y, Kawashima Y, Kawara H. A Retrospective Analysis of Epiphora Due to Docetaxel. *Cancer and Chemotherapy*. 2016; 43: 737–741.
- [14] Kaya M, Atas F, Gulsum Guc Z, Oztup I, Durak I, Saatci AO. A cross-sectional optical coherence tomography study in patients on taxane-based therapy and a case report with the literature review. *Cutaneous and Ocular Toxicology*. 2020; 39: 287–293.
- [15] Nomi N, Ota M, Fukumura M, Nuno Y, Hatano M, Wakuta M, *et al*. Indocyanine green angiography findings of cystoid macular edema secondary to paclitaxel therapy. *Japanese Journal of Ophthalmology*. 2018; 62: 163–167.
- [16] Shen M. A case of vision loss after the chemotherapy of cisplatin combined with paclitaxel. *Chinese Journal of New Drugs*. 2009; 18: 1815–1816.
- [17] Al-Tweigeri T, Nabholz J, Mackey JR. Ocular toxicity and cancer chemotherapy: a review. *Cancer*. 1996; 78: 1359–1373.
- [18] Yokoe T, Fukada I, Kobayashi K, Shibayama T, Miyagi Y, Yoshida A, *et al*. Cystoid Macular Edema during Treatment with Paclitaxel and Bevacizumab in a Patient with Metastatic Breast Cancer: A Case Report and Literature Review. *Case Reports Oncology*. 2017; 10: 605–612.
- [19] Wall ME, Wani MC. Camptothecin and taxol: discovery to clinic-thirteenth Bruce F. Cain Memorial Award Lecture. *Cancer Research*. 1995; 55: 753–760.
- [20] Koudelka S, Turánek J. Liposomal paclitaxel formulations. *Journal of Controlled Release*. 2012; 163: 322–334.
- [21] Noguchi Y, Kawashima Y, Maruyama M, Kawara H, Tokuyama Y, Uchiyama K, *et al*. Risk Factors for Eye Disorders Caused by Paclitaxel: a Retrospective Study. *Biological and Pharmaceutical Bulletin*. 2018; 41: 1694–1700.
- [22] Noguchi Y, Kawashima Y, Maruyama M, Kawara H, Tokuyama Y, Uchiyama K, *et al*. Current status of eye disorders caused by docetaxel administration every 3 weeks: a case-control study in Japanese patients. *Journal of Oncology Pharmacy Practice*. 2020; 26: 655–665.
- [23] Fortes BH, Liou H, Dalvin LA. Ophthalmic adverse effects of taxanes: the Mayo Clinic experience. *European Journal of Ophthalmology*. 2022; 32: 602–611.
- [24] Fabre-Guillevin E, Tchen N, Anibaldi-Charpiat M, Calluau L, Ravaud A. Taxane-induced glaucoma. *The Lancet*. 1999; 354: 1181–1182.
- [25] Hofstra LS, de Vries EG, Willemse PH. Ophthalmic toxicity following paclitaxel infusion. *Annals of Oncology*. 1997; 8: 1053.
- [26] Gupta S, Silliman CG, Trump DL. Docetaxel-induced Meibomian duct inflammation and blockage leading to chalazion formation. *Prostate Cancer and Prostatic Diseases*. 2007; 10: 396–397.
- [27] Hosotani Y, Morimatsu T, Takata M. A Case of a Corneal Disorder after Breast Cancer Treatment with Nab-paclitaxel. *Nippon Ganka Gakkai Zasshi*. 2016; 120: 449–453.
- [28] Kuwata M, Yoshizawa K, Matsumura M, Takahashi K, Tsubura A. Ocular toxicity caused by Paclitaxel in neonatal sprague-dawley rats. *In Vivo*. 2009; 23: 555–560.
- [29] Esmaeli B, Burnstine MA, Ahmadi MA, Prieto VG. Docetaxel-Induced Histologic Changes in the Lacrimal Sac and the Nasal Mucosa. *Ophthalmic Plastic & Reconstructive Surgery*. 2003; 19: 305–308.

- [30] Esmaeli B, Hidaji L, Adinin RB, Faustina M, Coats C, Arbuckle R, *et al.* Blockage of the lacrimal drainage apparatus as a side effect of docetaxel therapy. *Cancer*. 2003; 98: 504–507.
- [31] Yamagishi T, Ochi N, Yamane H, Hasebe S, Takigawa N. Epiphora in lung cancer patients receiving docetaxel: a case series. *BMC Research Notes*. 2014; 7: 322.
- [32] Lee HS, Ha JY, Choi W, Yoon KC. Bilateral Corneal Epithelial Lesions Associated with Paclitaxel. *Optometry and Vision Science*. 2016; 93: 1333–1336.
- [33] Álvarez-Fernández D, Cubillas-Martín M, Álvarez-Suárez ML, Viescas-Fernández MJ, Medina-Mejías MR, Rodríguez-Balsera C. Macular oedema associated with taxanes: a case report and literature review. *Archivos De La Sociedad Española De Oftalmología (English Edition)*. 2020; 95: 485–495.
- [34] Alves Pereira S, Vale C, Moreira J, Sampaio F. Macular Cystoid Edema Induced by Nab-Paclitaxel. *Acta Médica Portuguesa*. 2022; 35: 294–297.
- [35] Chang S, Tsai SH, Chen L, Chan W. Paclitaxel-induced cystoid macular oedema. *Acta Ophthalmologica*. 2018; 96: e649–e650.
- [36] Haider A, Bababeygy SR, Lu SY. Cystoid Macular Edema And Macular Pigmentation Associated With Nab-Paclitaxel Therapy. *Retinal Cases & Brief Reports*. 2015; 9: 220–222.
- [37] Smith SV. Cystoid Macular Edema Secondary to Albumin-Bound Paclitaxel Therapy. *Archives of Ophthalmology*. 2008; 126: 1605.
- [38] Kanakis M, Georgalas I, Makatsoris T, Pharmakakis N. Taxane Induced Cystoid Macular Edema: Case Report and Integrated Pathogenic Theory. *Current Drug Safety*. 2019; 14: 43–47.
- [39] Kato K, Nagashima R, Sugimoto M, Ikesugi K, Matsubara H, Kondo M. Case of cystoid macular edema induced by systemic administration of paclitaxel: evaluations with electroretinograms. *Documenta Ophthalmologica*. 2021; 143: 229–235.
- [40] Baek J, Lee J, Ra H. Ultra-widefield angiographic imaging of albumin-bound paclitaxel-induced cystoid macular edema. *Indian Journal of Ophthalmology*. 2019; 67: 2058.
- [41] Matsuoka N, Hasebe H, Mayama T, Fukuchi T. Sub-Tenon Injections of Triamcinolone Acetonide had Limited Effect on Cystoid Macular Edema Secondary to Nanoparticle Albumin-Bound-Paclitaxel (Abraxane) Case Reports in Ophthalmological Medicine. 2015; 2015: 1–4.
- [42] Murphy CG, Walsh JB, Hudis CA, Lake D, Theodoulou M. Cystoid macular edema secondary to nab-paclitaxel therapy. *Journal of Clinical Oncology*. 2010; 28: e684–687.
- [43] Park E, Goldberg NR, Adams S. Nab-paclitaxel-induced cystoid macular edema in a patient with pre-existing optic neuropathy. *Anti-Cancer Drugs*. 2016; 27: 580–584.
- [44] Rahimy E, Sarraf D. Cystoid Macular Edema Secondary to Nanoparticle Albumin-Bound Paclitaxel Therapy. *Ophthalmic Surgery, Lasers and Imaging Retina*. 2013; 44: 187–189.
- [45] Rao RC, Choudhry N. Cystoid macular edema associated with chemotherapy. *Canadian Medical Association Journal*. 2016; 188: 216–216.
- [46] Risard SM, Pieramici DJ, Rabena MD. CYSTOID MACULAR EDEMA SECONDARY To PACLITAXEL (ABRAXANE) RETINAL Cases & Brief Reports. 2009; 3: 383–385.
- [47] Sridhar J, Shahlaee A, Ehmann D, Samara WA, Rahimy E, Ho AC, *et al.* En Face Optical Coherence Tomography and Optical Coherence Tomography Angiography Imaging of Taxane-Associated Cystoid Macular Edema. *Ophthalmic Surgery, Lasers and Imaging Retina*. 2016; 47: 176–179.
- [48] Tanaka Y, Bando H, Hara H, Ito Y, Okamoto Y. Cystoid macular edema induced by nab-paclitaxel. *Breast Cancer*. 2015; 22: 324–326.
- [49] Enzsoly A, Kammerer K, Nemeth J, Schneider M. Bilateral cystoid macular edema following docetaxel chemotherapy in a patient with retinitis pigmentosa: a case report. *BMC Ophthalmology*. 2015; 15: 32.
- [50] Koo NK, Kim YC. A Case of Paclitaxel-induced Maculopathy Treated with Methazolamide. *Korean Journal of Ophthalmology*. 2012; 26: 394.
- [51] Kuznetcova TI, Cech P, Herbolt CP. The mystery of angiographically silent macular oedema due to taxanes. *International Ophthalmology*. 2012; 32: 299–304.
- [52] Rahman HT, Yeh S, Bergstrom CS. Cystoid Macular Edema without Leakage Secondary to Nab-Paclitaxel (Abraxane): Clinical Experience with Intravitreal Bevacizumab. *Journal of Ocular Pharmacology and Therapeutics*. 2013; 29: 360–362.
- [53] Shih C, Lee Y. Impaired retinal pigment epithelium in paclitaxel-induced macular edema. *Medicine*. 2018; 97: e11229.
- [54] Tapia Quijada HE, Quijada Fumero E, Mesa Lugo FI, Serrano García M, Betancor Caro N. Nepafenac for cystoid macular oedema secondary to paclitaxel. *Archivos De La Sociedad Española De Oftalmología (English Edition)*. 2021; 96: 434–437.
- [55] Teitelbaum BA, Tresley ADJ. Cystic Maculopathy with Normal Capillary Permeability Secondary to Docetaxel. *Optometry and Vision Science*. 2003; 80: 277–279.
- [56] Telander DG, Sarraf D. Cystoid Macular Edema with Docetaxel Chemotherapy and the Fluid Retention Syndrome. *Seminars in Ophthalmology*. 2007; 22: 151–153.
- [57] Capri G, Munzone E, Tarenzi E, Fulfaro F, Gianni L, Caraceni A, *et al.* Optic Nerve Disturbances: a New Form of Paclitaxel Neurotoxicity. *JNCI Journal of the National Cancer Institute*. 1994; 86: 1099–1101.
- [58] Scaioli V, Caraceni A, Martini C, Curzi S, Capri G, Luca G. Electrophysiological evaluation of visual pathways in paclitaxel-treated patients. *Journal of Neuro-Oncology*. 2006; 77: 79–87.
- [59] Agustoni F, Platania M, Vitali M, Zilembo N, Haspinger E, Sinno V, *et al.* Emerging toxicities in the treatment of non-small cell lung cancer: Ocular disorders. *Cancer Treatment Reviews*. 2014; 40: 197–203.
- [60] Choron RL, Chang S, Khan S, Villalobos MA, Zhang P, Carpenter JP, *et al.* Paclitaxel impairs adipose stem cell proliferation and differentiation. *Journal of Surgical Research*. 2015; 196: 404–415.
- [61] Wang Y, Chen C, Chung S, Chiou Y, Lo H. Involvement of oxidative stress and caspase activation in paclitaxel-induced apoptosis of primary effusion lymphoma cells. *Cancer Chemotherapy and Pharmacology*. 2004; 54
- [62] Li G, Xu D, Sun J, Zhao S, Zheng D. Paclitaxel inhibits proliferation and invasion and promotes apoptosis of breast cancer cells by blocking activation of the PI3K/AKT signaling pathway. *Advances in Clinical and Experimental Medicine*. 2020; 29: 1337–1345.
- [63] Michailidou M, Brown HK, Lefley DV, Evans A, Cross SS, Coleman RE, *et al.* Microvascular Endothelial Cell Responses in vitro and in vivo: Modulation by Zoledronic Acid and Paclitaxel? *Journal of Vascular Research*. 2010; 47: 481–493.
- [64] Banerjee S, Xu H, Fuh E, Nguyen KT, Garcia JA, Brilakis ES, *et al.* Endothelial progenitor cell response to antiproliferative drug exposure. *Atherosclerosis*. 2012; 225: 91–98.
- [65] Mishra GP, Nguyen D, Alani AWG. Inhibitory Effect of Paclitaxel and Rapamycin Individual and Dual Drug-Loaded Polymeric Micelles in the Angiogenic Cascade. *Molecular Pharmaceutics*. 2013; 10: 2071–2078.
- [66] Tasnim A, Rammelkamp Z, Slusher AB, Wozniak K, Slusher BS, Farah MH. Paclitaxel causes degeneration of both central and peripheral axon branches of dorsal root ganglia in mice. *BMC Neuroscience*. 2016; 17: 47.
- [67] Quasthoff S, Hartung HP. Chemotherapy-induced peripheral neuropathy. *Journal of Neurology*. 2002; 249: 9–17.
- [68] Argyriou AA, Bruna J, Marmiroli P, Cavaletti G.

Chemotherapy-induced peripheral neurotoxicity (CIPN): an update. *Critical Reviews in Oncology Hematology*. 2012; 82: 51–77.

- [69] Gornstein EL, Schwarz TL. Neurotoxic mechanisms of paclitaxel are local to the distal axon and independent of transport defects. *Experimental Neurology*. 2017; 288: 153–166.
- [70] Wozniak KM, Nomoto K, Lapidus RG, Wu Y, Carozzi V, Cavalletti G, *et al.* Comparison of Neuropathy-Inducing Effects of Eribulin Mesylate, Paclitaxel, and Ixabepilone in Mice. *Cancer Research*. 2011; 71: 3952–3962.
- [71] Shim HS, Bae C, Wang J, Lee K, Hankerd KM, Kim HK, *et al.* Peripheral and central oxidative stress in chemotherapy-induced neuropathic pain. *Molecular Pain*. 2019; 15: 174480691984009.
- [72] Yang Y, Karakhanova S, Hartwig W, D’Haese JG, Philippov PP, Werner J, *et al.* Mitochondria and Mitochondrial ROS in Cancer: Novel Targets for Anticancer Therapy. *Journal of Cellular Physiology*. 2016; 231: 2570–2581.
- [73] Duggett NA, Griffiths LA, McKenna OE, de Santis V, Yongsanguanchai N, Mokori EB, *et al.* Oxidative stress in the development, maintenance and resolution of paclitaxel-induced painful neuropathy. *Neuroscience*. 2016; 333: 13–26.
- [74] Li X, Fan C, Xiao Z. A collagen microchannel scaffold carrying paclitaxel-liposomes induces neuronal differentiation of neural stem cells through Wnt/beta-catenin signaling for spinal cord injury repair. *Biomaterials*. 2018; 183: 114–127.
- [75] Nazemi Z, Nourbakhsh MS, Kiani S, Heydari Y, Ashtiani MK, Daemi H, *et al.* Co-delivery of minocycline and paclitaxel from injectable hydrogel for treatment of spinal cord injury. *Journal of Controlled Release*. 2020; 321: 145–158.
- [76] Harris WM, Zhang P, Plastini M, Ortiz T, Kappy N, Benites J, *et al.* Evaluation of function and recovery of adipose-derived stem cells after exposure to paclitaxel. *Cytotherapy*. 2017; 19: 211–221.
- [77] Münz F, Lopez Perez R, Trinh T, Sisombath S, Weber K, Wuchter P, *et al.* Human mesenchymal stem cells lose their functional properties after paclitaxel treatment. *Scientific Reports*. 2018; 8: 312.
- [78] Schrader C, Keussen C, Bewig B, Lins M. Symptoms and signs of an acute myocardial ischemia caused by chemotherapy with Paclitaxel (Taxol) in a patient with metastatic ovarian carcinoma. *European Journal of Medical Research*. 2005; 10: 498–501.
- [79] Wang D, Xing W, Wang X, Zhu H. Taxol stabilizes gap junctions and reduces ischemic ventricular arrhythmias in rats in vivo. *Molecular Medicine Reports*. 2015; 11: 3243–3248.
- [80] Miller K, Wang M, Gralow J, Dickler M, Cobleigh M, Perez EA, *et al.* Paclitaxel plus Bevacizumab versus Paclitaxel alone for Metastatic Breast Cancer. *New England Journal of Medicine*. 2007; 357: 2666–2676.
- [81] Guo H, Zheng M, Jiao YB, *et al.* Paclitaxel enhances the protective effect of myocardial ischemia preconditioning on ischemia/reperfusion injury in aged rat. *Zhonghua Xin Xue Guan Bing Za Zhi*. 2018; 46: 719–724.
- [82] Giessmann D, Theiss C, Breipohl W, Meller K. Decreased gap junctional communication in neurobiotin microinjected lens epithelial cells after taxol treatment. *Anatomy and Embryology*. 2005; 209: 391–400.
- [83] Sharawy MH, El-Awady MS, Makled MN. Protective effects of paclitaxel on thioacetamide-induced liver fibrosis in a rat model. *Journal of Biochemical and Molecular Toxicology*. 2021; 35: e22745.
- [84] Esmali B, Valero V, Ahmadi MA, Booser D. Canalicular stenosis secondary to docetaxel (taxotere). *Ophthalmology*. 2001; 108: 994–995.
- [85] Makri OE, Georgalas I, Georgakopoulos CD. Drug-induced macular edema. *Drugs*. 2013; 73: 789–802.