

THE ESSENTIAL ROLE OF INFLAMMATION AND INDUCED GENE EXPRESSION IN THE PATHOGENIC PATHWAY OF ALZHEIMER S DISEASE

Lars Nilsson¹, Jack Rogers² and Huntington Potter¹

Department of Neurobiology¹, Harvard Medical School and Division of Hematology, Department of Medicine², Brigham and Women s Hospital, Harvard Medical School, Boston, MA 02115, USA

Received 3/30/98 Accepted 4/3/98

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Amyloid-associated proteins
4. Cytokines
5. Epidemiology
6. Pathological chaperones the accelerator hypothesis
7. Acknowledgements
8. References

1. ABSTRACT

Alzheimer's disease is among the most common diseases of advanced age affecting almost one out of ten individuals who survive beyond the age of 65 years, and another 10% for each additional decade of the life-span. The prognosis of the disease is an inexorable decline of mental functions leading to complete dependence on caretakers in the late stages of the disease. Alzheimer's disease will become a steadily increasing financial health-care problem in the industrialized world with the increasing longevity and ageing of the population. To-date there are no effective therapeutics. However, during the last years promising findings suggest that anti-inflammatory treatment strategies might be efficient. Here, we will review the experimental and epidemiological findings which support the idea that inflammatory mechanisms play an important role in Alzheimer's disease pathogenesis. The review of the experimental findings will be focussed on the amyloid-associated proteins, alpha₁-antichymotrypsin and apolipoprotein E, as well as the major cytokines. In addition, the epidemiological studies on non-steroidal anti-inflammatory drugs and traumatic head injury will be summarized. We hypothesize a pathogenic model for Alzheimer's disease in which the expression of amyloid-associated proteins/pathological chaperones, induced by inflammatory cytokines, plays an essential role in accelerating the disease progress, and suggest potential targets for drug discovery based on such a model.

2. INTRODUCTION

Alzheimer's disease is a devastating neurodegenerative disorder clinically characterized by an insidious onset, a progressive decline of multiple cognitive functions, and ultimately fatal loss of mental functions. The disease is defined by characteristic neuropathological lesions – proteinaceous deposits – which are primarily found in the hippocampus and the parietotemporal parts of the cerebral cortex. These lesions are the neuritic plaques, which are largely composed of extracellular deposits of

beta-amyloid peptides, and the intraneuronal neurofibrillary tangles consisting of twisted filaments of the cytoskeletal tau protein.

During the last five years, Alzheimer's disease research has been greatly benefited by molecular genetic discoveries of pathogenic genes, which have allowed us to begin to decipher the molecular mechanisms of the disorder. Initially, missense mutations in the amyloid precursor protein (APP) gene were found, although these cases were extremely rare (1-2). Two more recently identified pathogenic genes, presenilin-1 and presenilin-2, account for a much larger portion of the early-onset familial cases (3-7). These findings have brought the metabolism of the amyloid precursor protein, a ubiquitously expressed transmembrane protein, into focus for two reasons. Cleaved fragments of the amyloid precursor protein, beta-amyloid peptides, are the major constituents of the amyloid deposits (8-9), and all Alzheimer pathogenic mutations have been shown to increase the production of beta-amyloid, in most cases by generating more of the longer beta-amyloid peptides 1-42(43) (10-12). Thus therapeutic strategies aimed at intervening in the formation of beta-amyloid peptides or the aggregation of beta-amyloid filaments have been suggested (13-16). The major obstacle to these strategies is the nonspecific mechanism by which beta-amyloid peptides probably are generated (17) and the rapid turnover rate of the amyloid precursor protein (18).

It has become increasingly appreciated that inflammatory mechanisms in the brain are as important as beta-amyloid production in the Alzheimer pathogenic pathway, and that these mechanisms perhaps are more promising drug targets. The evidence that anti-inflammatory treatment strategies would be beneficial comes from epidemiological and experimental findings. Here we will review this literature in the context of a model for the Alzheimer pathogenic pathway in which expression

Pathogenic pathway of Alzheimer's disease

of amyloid-associated proteins/pathological chaperones, orchestrated by inflammatory cytokines, plays an essential role in accelerating the disease progress.

3. AMYLOID-ASSOCIATED PROTEINS

The first hint that Alzheimer's disease pathology might involve an inflammatory reaction came from the observation of reactive astrocytes and microglia in affected brain regions in the initial neuropathological description (19). However the absence of standard features of inflammation such as swelling and lymphocyte infiltration argued against this description. The breakthrough came in the 1980's when the Alzheimer amyloid deposits were found to contain, in addition to beta-amyloid peptides, other proteins that are normally observed during inflammation and its associated acute phase response. Activated complement components were the first such inflammation-associated proteins identified in the Alzheimer amyloid deposits (20). However, it was quite possible that such proteins had leaked into the brain from the circulation and were not indicative of an endogenous inflammation in the brain. Soon thereafter, our lab showed that the acute phase protein alpha₁-antichymotrypsin (ACT) was an integral component of the amyloid filaments (21). ACT formed stable complexes with beta-amyloid peptide *in vitro*, that were resistant to boiling in SDS and beta-mercaptoethanol (22-23). Furthermore ACT was found to be specifically associated with beta-amyloid containing deposits but not with other types of amyloidosis (24), and to be generated through overexpression from astrocytes surrounding the plaque structures (25-26). ACT levels in serum and cerebrospinal fluid (CSF) have been suggested as diagnostic markers for Alzheimer's disease, since elevated levels have been observed in AD but not in other neurodegenerative disorders (27-29), while at the same time various other acute phase proteins have been found unchanged (30). The serum and CSF levels have not been found to correlate, indicating that serum ACT is most probably derived from the periphery. The increase in serum levels of ACT is likely not a very early marker of Alzheimer's disease, since normal levels have been found in young Down's syndrome patients (31-32).

Another primarily astrocyte-derived protein of clear relevance for Alzheimer disease pathology, and possibly reflective of inflammatory mechanisms, is apolipoprotein E (apoE). This lipoprotein was early-on detected in amyloid plaques of the brain parenchyma with immunohistochemistry (33-34). However apoE did not receive major attention until Allen Roses and Judes Poirier and their coworkers highlighted it as a beta-amyloid binding protein in the cerebrospinal fluid, and the ApoE4 allele as a susceptibility gene for the development of Alzheimer's disease (35-37). In addition, Poirier and coworkers discovered that apoE serves important functions with respect to cholesterol and phospholipid metabolism following injury of the central nervous system (38). Later studies have confirmed that apoE forms stable complexes with beta-amyloid *in vitro* as well as in purified amyloid filaments from Alzheimer disease brain (39), and that the

apoE4 allele is a major risk factor for late-onset Alzheimer's disease (40).

4. CYTOKINES

Because glial cell activation is such a prominent feature of Alzheimer disease pathology, substances with the ability to promote or modulate these changes were early investigated. Cytokines belong to a growing family of polypeptides associated with injury and inflammatory responses in the periphery as well as in the central nervous system. They are present at very low concentrations under normal physiological conditions and are rapidly induced during various pathological states. High-affinity cytokine receptors allow low concentrations of released cytokine to exert potent effects, which can be either beneficial or detrimental to neuronal survival. For instance, interleukin-1 (IL-1) has been shown to protect neurons from excitotoxic damage by release of nerve growth factor *in vitro* (41). However, a substantial literature implicates the IL-1-cleaving enzyme (ICE) and other homologous caspases in apoptotic cell death (42).

The cytokine which has been most studied and implicated with respect to Alzheimer disease pathology is interleukin-1 (IL-1). IL-1 was early-on shown to stimulate astroglial proliferation and angiogenesis in response to traumatic injury through its secretion from ameboid microglia in the rat brain (43-45). Furthermore IL-1-containing microglia were found to be 30-fold more abundant in Down syndrome brain and 6-fold more abundant in Alzheimer's disease brain as compared to control brain (46). Interestingly, this induction was restricted to resident microglia in brain regions that normally develop mature neuritic plaques, but not in cerebellum where mostly diffuse plaques are seen. (47-48). Furthermore IL-1 was shown to upregulate APP-mRNA expression in endothelial cells through a protein kinase C dependent pathway that was targeted to a 180bp region in the APP-promoter (49). IL-1 has recently been shown to upregulate the translational efficiency of APP-mRNA in astrocytes via a stem-loop structure in the 5'UTR of the APP-transcript (50).

IL-6 is a multifunctional cytokine and the other main pro-inflammatory signaling molecule linked to Alzheimer's disease pathology. It is the principal inducer of plasma proteins during the hepatic acute phase response—a homeostatic mechanism to limit protease degradation in the aftermath of injury, trauma, or infection (51). Elevated levels of IL-6 have been found in Alzheimer's brain and cerebrospinal fluid (52-55), and this induction appears to occur concurrently with early stages of senile plaque formation (56). The findings from *in vivo* studies on the effect of overexpression of IL-6 in the brain is contradictory. Astrocyte-targeted IL-6 overexpression has been shown to cause marked neurodegeneration and learning impairments (57-58), while IL-6 expression under the control of the neuronal-specific enolase promoter led to astrogliosis in the absence of neurodegeneration or behavioral changes (59).

Pathogenic pathway of Alzheimer's disease

TNF- α is another major pro-inflammatory cytokine found in the brain. Increased levels of TNF- α has been reported after head injury (60). The functional significance of TNF- α in terms neurodegeneration/protection is unclear. Cytotoxic effects on oligodendrocytes in experimental models of autoimmune diseases have been described (61-62). However, TNF- α receptor knockout mice displayed suppressed microglial activation, greater neuronal loss and larger cortical infarct area than normal mice, suggesting a neuroprotective role for TNF- α against seizure and ischemia-induced damage (63). TNF- α -positive cells have not been observed in Alzheimer's disease brain (64-65), and serum measurements are conflicting (66-67).

Astrocyte expression of TGF- β_1 , an anti-inflammatory cytokine, has recently been linked to beta-amyloid deposition in the cerebral blood vessels and meninges (68), that resemble the cerebral amyloid angiopathy (CAA) which is frequently found in Alzheimer's pathology (69-70). Previous transgenic experiments have demonstrated that this cytokine dose-dependently upregulates extracellular matrix proteins, which might be the mechanistic link to the amyloid deposition (71). TGF- β_1 has also been identified in subsets of amyloid plaques (72).

Various other proteins such as complement proteins (73), acute phase reactants (52) and proteoglycans (74) have been demonstrated in amyloid plaques, in most cases with immunohistochemistry. The significance of these markers has previously been extensively reviewed (75). These plaque-associated proteins are, like ACT, apoE and IL-1, mainly derived from microglia and astrocytes, cell categories which play important functions in regeneration and immune functions of the central nervous system.

5. EPIDEMIOLOGY

Epidemiological studies on the use of non-steroidal anti-inflammatory drugs (NSAID) and on traumatic head injury provide further evidence for inflammatory mechanisms as being important for Alzheimer's disease. Besides the currently-established risk factors for Alzheimer's disease—age, apoE genotype, and family history of dementia—there is growing evidence for a previous history of head injury as a contributor to the disease development. These findings come from retrospective case-control studies (76-78) as well as prospective incidence studies (79). Furthermore an Alzheimer-like pathology, dementia pugilistica, can be generated by the repeated head injury that boxers experience (80-82). A mechanistic link is indicated by morphological studies showing that suspected contributors to Alzheimer's neuropathology such as IL-1- α -positive microglia (83) and beta-amyloid deposition (84) are observed in the acute phase of head trauma in human brain. Other studies suggest an interaction with a known genetic risk factor for Alzheimer's disease by showing that the trauma-generated beta-amyloid deposition is dependent upon the presence of the apoE4 allele (85).

A number of pharmaco-epidemiological studies support the routine use of NSAID as therapeutic of Alzheimer's disease. Initially two reports suggested that patients suffering from inflammatory diseases such as rheumatoid arthritis had a reduced incidence of Alzheimer's disease (86-87). The explanation for the results, as suggested by the authors, was that the anti-inflammatory drugs routinely used by these patients exerted a protective effect against the development of Alzheimer's disease. Alternative interpretations of the findings were possible since the studies were prone to methodological problems such as selection bias and/or under-reported frequency of Alzheimer's disease among the rheumatoid patients by the clinicians. However most probably anti-inflammatory drugs exert a truly protective effect since these early studies have been reproduced in larger experimental settings (88-90) and by alternative methods such as co-twin control studies (91). Indeed, initial clinical trials have demonstrated that the inflammatory drug indomethacin exerted beneficial effects on Alzheimer patients with respect to cognitive decline (92). Larger prospective studies are currently underway to test whether routine use of anti-inflammatory agents by Alzheimer patients will have significant therapeutic benefits.

6. PATHOLOGICAL CHAPERONES THE ACCELERATOR HYPOTHESIS

A unifying hypothesis for the Alzheimer's disease pathology would be helpful in order to get a comprehensive view of the various findings so far discussed. Currently, the leading model for the etiology of Alzheimer's disease is the amyloid hypothesis/amyloid cascade hypothesis (13, 93-94) which states that overproduction or insufficient clearance of beta-amyloid fosters the deposition and formation of amyloid plaques as the central event of the pathology. The deposition then causes tau phosphorylation, tangle formation, subsequent neuritic degeneration, and ultimately the clinical symptomatology. One major pitfall with this hypothesis is its inability to explain the apparent region-specificity of Alzheimer's disease. Furthermore the density of amyloid plaques is not well correlated with cognitive decline (95). Most probably the latter is due to the fact that amyloid plaques are neuropathologically heterogeneous and that only subtypes such as the paired helical filament (PHF)-containing neuritic plaques are detrimental to neuronal survival. Thus it seems likely that additional mechanisms e.g. cofactors released downstream of the initial beta-amyloid aggregation are essential for disease development. Gliosis is a prominent feature of Alzheimer's pathology, with both activated microglia and reactive astrocytes being clearly visible in the periphery surrounding the amyloid core structure (19, 96-98). These changes have often been viewed simply as a secondary tissue response to the ongoing amyloidosis without any etiologic significance. Here we reiterate and extend our model of the pathogenic pathway in Alzheimer's disease which postulates that cytokines and amyloid associated proteins released by these cells accelerate the beta-amyloid aggregation-transforming and stabilizing the amyloid filaments to form cores of mature senile plaques (21-23, 47, 99, 16).

Pathogenic pathway of Alzheimer's disease

The initial step in this pathogenic pathway is the accumulation of beta-amyloid peptides into amorphous deposits. There is an apparent consistency in the *in vitro* fibrillogenesis studies (100-101), the *in vivo* data from Alzheimer's disease brain (102-103), and the temporal examination of Down syndrome brains (104-105), in that the longer beta-amyloid peptide(1-42) is the most amyloidogenic and earliest deposited. This initial deposition probably acts as seed for further deposition of shorter and less amyloidogenic peptides, such as beta-amyloid(1-40) (106).

The second step in the pathway is the reactive expression of inflammation-related proteins in association with amyloid deposits. There is histopathological evidence that IL-1 (107) as well as IL-6 (56) - immunoreactivity in the cerebral cortex is preferentially associated with early stages of plaque evolution when the beta-amyloid aggregation is mostly of the diffuse type, rather than mature plaques with the classic congophilic dense core. Microglial activation and proliferation in amyloid plaque-forming areas has also been described in the APP_{sw} transgenic strain (108). Perhaps some beta-amyloid aggregate with a low level of the characteristic beta-sheet structure observed in the diffuse deposits is sufficient to induce the microglial activation and cytokine production.

The third step of the cascade is the production and release of amyloid-associated proteins that accelerate amyloid filament formation—the pathological chaperones. It has long been known that the transcriptional induction of the peripheral acute-phase response in the liver is mediated by IL-1 and IL-6 (109). Results from our lab demonstrated that the ACT-overexpression in astrocytes of Alzheimer's disease brain is likely caused by cytokines as well. Specifically ACT mRNA was 3-5-fold upregulated by IL-1 in mixed glial cells of human fetal origin. Interestingly, the ACT mRNA expression was found to be both species- and brain region-dependent. The rat gene homologue of ACT, contrapsin, was not induced by either IL-1 or IL-6. Furthermore human cerebral cortical, but not cerebellar or brain stem mixed glial cultures were able to spontaneously express IL-1 and ACT, suggesting that the mechanisms for the region specificity of Alzheimer's disease pathology may reflect differences in glial cell functioning (47-48). Whether the apoE gene is inducible in a corresponding fashion, and if so by which putative regulatory factors is largely unknown, although overexpression of apoE mRNA was early demonstrated in Alzheimer's disease brain (110). Certainly the synthesis of apoE mRNA is increased following peripheral nerve injury (111-113) and entorhinal-cortex lesion—a model for reactive synaptogenesis and compensatory reinnervation of the hippocampus (114-115). Association between a polymorphic site in the apoE promoter and Alzheimer's dementia also stresses the importance of regulation of apoE gene expression in the disease process (116).

Another link between IL-1 and Alzheimer pathology is the ability of IL-1 to increase the translational efficiency of the amyloid precursor protein (APP) in astrocytes by means of a stem-loop structure in the 5'UTR

of the APP-transcript (50). Thus astrocytes in the vicinity of a developing amyloid plaques, overproduce not only the amyloid associated proteins apoE and ACT, but also the APP protein (and therefore the beta-amyloid as well). Indeed, increased APP protein levels have been detected *in vivo* following IL-1 injection into the rat brain parenchyma (117).

Given the close association of apoE and ACT with the Alzheimer amyloid deposits, it was reasonable to determine whether these proteins influenced the formation of amyloid filaments. Results from our and other labs have demonstrated that beta-amyloid filaments form much more rapidly in the presence of the amyloid-associated proteins ACT or apoE (99, 118-120). Instead of days, filaments formed in a matter of hours and grew to very great lengths. Particularly interesting was the fact that apoE4, the apolipoprotein E isoform which is linked to late-onset familial Alzheimer's disease (40), was the most effective amyloid promoting factor (99). There is now an abundant literature on the role of apoE and ACT as amyloid promoting factors. The concept of pathological chaperones (a term coined by Wisniewski and Frangione, 34) is not yet fully established, since some *in vitro* studies are partly contradictory (121-123). However the discrepancies are probably due to methodological differences in terms of the amyloid peptide used (beta-amyloid(1-40) versus beta-amyloid(1-42)), the quality and purity of the beta-amyloid peptide preparations, and the molar ratio of beta-amyloid to the chaperone used. Indeed, a recent *in vivo* study has unequivocally demonstrated that apoE is an amyloid promoting factor. Specifically, mice strains carrying zero, one, or two copies of the mouse apoE gene on the background of the PDAPP-transgenic mouse strain showed an apoE dose dependent increase in beta-amyloid deposition in cerebral cortex and the hippocampus, strongly supporting the chaperone concept. The beta-amyloid peptide was relatively harmless and essentially unable to polymerize efficiently into amyloid filaments in the absence of apoE (124). The demonstration of accelerated cerebrovascular amyloid deposition achieved by crossing a GFAP/TGF-beta₁-low-expressing mouse strain with the PDAPP-transgenic strain also supports the idea that cofactors are involved in amyloid deposition, in this case possibly mediated via expression of extracellular matrix proteins (68). It has recently been clinically shown that the ACT/A-allele, which is a suspected genetic risk factor for Alzheimer disease (125), is dose-dependently associated with higher load of amyloid angiopathy in Alzheimer's patients, as measured by Congo Red staining (126). Thus increased secretion of ACT might promote amyloid deposition in the cerebral vessels and meninges as well as in the brain parenchyma.

The mature amyloid deposits induce tangle formation, neuritic degeneration and ultimately neuronal loss. These final steps in the Alzheimer pathogenic pathway remain rather undefined, since the mechanisms for neurodegeneration and neuronal death in Alzheimer's disease are essentially unknown. Mature amyloid filaments could exert direct neurotoxic effect as demonstrated *in vitro* (127-128, 16), although no neuronal loss was observed in

Pathogenic pathway of Alzheimer's disease

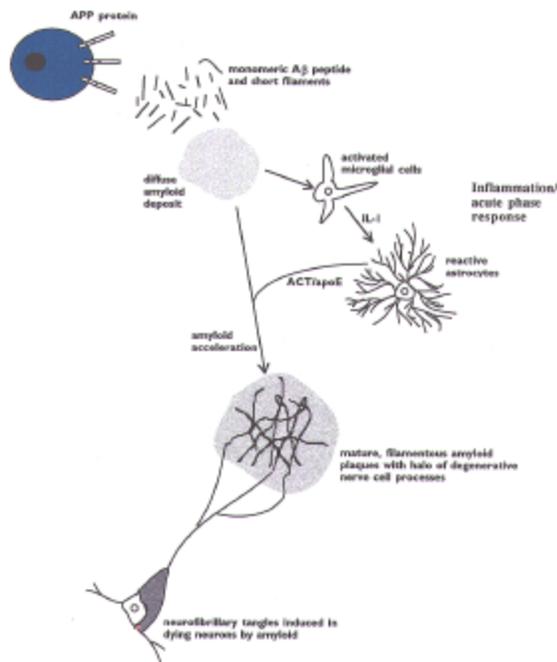


Figure 1. Pathogenic pathway for Alzheimer's Disease. The earliest and most widespread pathological change in Alzheimer's disease is the accumulation of beta-amyloid peptides into amorphous deposits. The deposition induces an inflammatory reaction and consequently an acute phase response involving IL-1 and IL-6 in microglial cells of the brain parenchyma and TGF- β_1 in perivascular astrocytes. The cytokines trigger release of various secretory products from the surrounding astrocytes that catalyze the transformation of diffuse beta-amyloid in amorphous deposits into mature amyloid filaments. For example astrocyte production of beta-amyloid and pathological chaperones such as ACT and apoE accelerate the polymerization of beta-amyloid peptides into filaments. Extracellular matrix proteins may stabilize the amyloid structures. Finally neurons of the human brain respond to the glial activation and the mature amyloid deposits by tangle formation, subsequent neuritic degeneration and ultimately cell death.

the PDAPP mice in spite of heavy congophilic amyloid load (129). The necessity for intraneuronally directed beta-amyloid deposition has been suggested, however this hypothesis remains unproven and speculative (130-131). Perhaps there is a species difference in the way human neurons respond to amyloid deposits and the mounted gliosis, since tangle formation, morphologically similar to those of Alzheimer's disease is human specific. Such an hypothesis could be tested by grafting primary human fetal neurons into any of the to-date-available transgenic strains producing congophilic plaques, or by mating a human tau transgenic mouse with the PDAPP mouse. A recently – performed *in vivo* study supports this view by showing differential neurotoxic effects of beta-amyloid in the primate and rat brain respectively (Yankner, personal comm.).

The pathogenic pathway for Alzheimer's disease shown in "figure 1" provides an explanation for the experimental and epidemiological findings linking the disease development to injury and inherent inflammatory mechanisms of the central nervous system. Furthermore it exposes several potential targets for drug discovery. For instance most of the detrimental effects of gliosis would be inhibited at an early stage if microglial activation in general, or that induced by beta-amyloid filaments, were suppressed (132-133). Substances that lower the biosynthesis of cytokines or the pathological chaperones and/or inhibit the actions of cytokines, such as IL-1 or IL-6 receptor antagonists or ICE-inhibitors could be valuable drug candidates. Designed ligands that stabilize the recently identified stem-loop structure in the 5'UTR of the APP-mRNA and thereby mediate translational inhibition of the amyloid precursor protein in astrocytes should possibly lower beta-amyloid production and fibril formation. Furthermore, ligands that specifically block the interaction of beta-amyloid with the pathological chaperones, apoE or ACT, would have the ability to significantly retard the polymerization process (16). These putative therapeutic strategies would all have the potential to significantly delay the progress of Alzheimer's disease.

7. ACKNOWLEDGEMENT

Postdoctoral fellowships from the STINT- foundation and Riksbankens Jubileumsfond, following a donation from Erik Rönnberg were greatly acknowledged (L.N.).

8. REFERENCES

- Goate A, MC. Chartier-Harlin, M. Mullan, J. Brown, F. Crawford, L. Fidani, L. Giuffra, A. Haynes, N. Irving, L. James, R. Mant, P. Newton, K. Rooke, P. Roges, C. Talbot, M. Pericak-Vance, A. Roses, R. Williamson, M. Rosser, M. Owen & J. Hardy Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* 349, 704-706 (1991)
- Mullan M., F. Crawford, K. Axelman, H. Houlden, L. Lilius, B. Winblad & L Lannfelt: A pathogenic mutation for probable Alzheimer's disease in the APP gene at the N-terminus of beta-amyloid. *Nat Genet* 1, 345-347 (1992)
- Levy-Lahad E, EM. Wijsman, E. Nemens, L. Anderson, KAB. Goddard, JL. Weber, TD. Bird & GD. Schellenberg: A familial Alzheimer's disease locus on chromosome 1. *Science* 269, 970-973 (1995)
- Levy-Lahad E, W. Wasco, P. Poorkaj, DM. Romano, J. Oshima, WH. Pettingell, CE. Yu, PD. Jondro, SD. Schmidt, K. Wang, AC. Crowley, YH. Fu, SY. Guenette, D. Galas, E. Nemens, EM. Wijsman, TD. Bird, GD. Schellenberg & RE. Tanzi.: Candidate gene for the chromosome 1 familial Alzheimer's disease locus. *Science* 269, 973-976 (1995)
- Li J, J. Ma & H. Potter: Identification and expression analysis of a potential familial Alzheimer's disease gene on chromosome 1 related to AD3. *Proc Natl Acad Sci USA* 92,

Pathogenic pathway of Alzheimer's disease

12180-12184 (1995)

6. Rogaev E, R. Sherrington, EA. Rogaeva, G. Levesque, M. Ikeda, Y. Liang, H. Chi, C. Lin, K. Holman, T. Tsuda, L. Mar, S. Sorbi, B. Nacmias, S. Piacentini, L. Amaducci, I. Chumakov, D. Cohen, L. Lannfelt, PE. Fraser, JM. Rommens & PH. St George-Hyslop: Familial Alzheimer's disease in kindreds with missense mutations in a gene on chromosome 1 related to the Alzheimer's disease type 3 gene. *Nature* 376, 775-778 (1995)

7. Sherrington R, EI. Rogaev, Y. Liang, EA. Rogaeva, G. Levesque, M. Ikeda, H. Chi, C. Lin, G. Li, K. Holman, T. Tsuda, L. Mar, JF. Foncin, AC. Bruni, MP. Montesi, S. Sorbi, I. Rainero, L. Pinessi, L. Nee, I. Chumakov, D. Pollen, A. Brookes, P. Sanseau, RJ. Polinsky, W. Wasco, HAR. Da Silva, JL. Haines, MA. Pericak-Vance, RE. Tanzi, AD. Roses, RE. Fraser, JM. Rommens & PH. St. George-Hyslop: Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* 375, 754-760 (1995)

8. Glenner G & CW. Wong: Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochem Biophys Res Commun* 120, 885-890 (1984)

9. Masters C, G. Simms, NA. Weinman, G. Multhaup, BL. McDonald & K. Beyreuther: Amyloid core protein in Alzheimer disease and Down syndrome. *Proc Natl Acad Sci USA* 82, 4245-4249 (1985)

10. Citron M, T. Oltersdorf, C. Haass, L. McConlogue, AY. Hung, P. Seubert, C. Vigo-Pelfrey, I. Lieberburg & DJ. Selkoe: Mutation of the beta-amyloid precursor protein in familial Alzheimer's disease increases beta-protein production. *Nature* 360, 672-674 (1992)

11. Suzuki N, TT. Cheung, XD. Cai, A. Odaka, L. Otvos Jr., C. Eckman, TE. Golde & SG. Younkin: An increased percentage of long amyloid beta protein secreted by familial amyloid beta protein precursor (beta APP717) mutants. *Science* 264, 1336-1340 (1994)

12. Scheuner D, C. Eckman, M. Jensen, X. Song, M. Citron, N. Suzuki, TD. Bird, J. Hardy, M. Hutton, W. Kukull, E. Larson, E. Levy-Lahad, M. Viitanen, E. Peskind, P. Poorkaj, G. Schellenberg, R. Tanzi, W. Wasco, L. Lannfelt, D. Selkoe & S. Younkin: Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat Med.* 2, 864-870 (1996)

13. Hardy J. & D. Allsop: Amyloid deposition as the central event in the aetiology of Alzheimer's. *Trends Pharmacol Sci* 12, 383-388 (1991)

14. Citron M, TS. Diehl, A. Capell, C. Haass, DB. Teplow & DJ. Selkoe: Inhibition of amyloid beta-protein production in neural cells by the serine protease inhibitor AEBSPF. *Neuron* 17, 171-179 (1996)

15. Lansbury P: Inhibition of amyloid formation—a strategy to delay the onset of Alzheimer's disease. *Curr Opin Chem Biol* 1, 260-267 (1997)

16. Ma J, HB. Brewer Jr. & H. Potter: Alzheimer Abeta neurotoxicity: promotion by antichymotrypsin, ApoE4; inhibition by Abeta-related peptides. *Neurobiol Aging* 17, 773-780 (1996)

17. Tischer E & B. Cordell: Beta-amyloid precursor protein. Location of transmembrane domain and specificity of gamma-secretase cleavage. *J Biol Chem* 271, 21914-21919 (1996)

18. Koo E, SS. Sisodia, DR. Archer, LJ. Martin, A. Weidemann, K. Beyreuther, P. Fischer, CL. Masters & DL. Price: Precursor of amyloid protein in Alzheimer disease undergoes fast anterograde axonal transport. *Proc Natl Acad Sci USA* 87, 1561-1565 (1990)

19. Alzheimer A: Über eine eigenartige Erkrankung der Hirnrinde. *Allg Z Psychiatr Psych-Gerichtl Med* 64, 146-148 (1907).

20. Eikelenboom P. & FC. Stam: Immunoglobulins and complement factors in senile plaques: an immunoperoxidase study. *Acta Neuropathol (Berlin)* 57, 239-242 (1982)

21. Abraham C, D. Selkoe & H. Potter: Immunohistochemical identification of the serine protease inhibitor alpha₁-antichymotrypsin in the brain amyloid deposits of Alzheimer's disease. *Cell* 52, 487-501 (1988)

22. Potter H: The involvement of astrocytes and acute phase response in the amyloid deposition of Alzheimer's disease. *Prog Brain Res* 94, 447-458 (1992)

23. Potter H, RB. Nelson, S. Das, R. Siman, US. Kayyali & D. Dressler: The involvement of proteases, protease inhibitors, and an acute phase response in Alzheimer's disease. *Ann NY Acad Sci* 674, 161-173 (1992)

24. Abraham C, T. Shirahama & H. Potter: Alpha 1-antichymotrypsin is associated solely with amyloid deposits containing the beta-protein. Amyloid and cell localization of alpha-1-antichymotrypsin. *Neurobiol Aging* 11, 123-129 (1990).

25. Pasternack J, CR. Abraham, BJ. Van Dyke, H. Potter & SG. Younkin: Astrocytes in Alzheimer's disease gray matter express alpha₁-antichymotrypsin mRNA. *Am J Pathol* 135, 827-834 (1989)

26. Koo E, CR. Abraham, H. Potter, LC. Cork & DL. Price: Developmental expression of alpha 1-antichymotrypsin in brain may be related to astroglia. *Neurobiol Aging* 12, 495-501 (1991)

27. Matsubara E, M. Amari, M. Shoji, Y. Harigaya, H. Yamaguchi, K. Okamoto & S. Hirai: Serum concentration of alpha 1-antichymotrypsin is elevated in patients with

Pathogenic pathway of Alzheimer's disease

senile dementia of the Alzheimer type. *Prog Clin Biol Res* 317, 707-714 (1989)

28. Matsubara E, S. Hirai, M. Amari, M. Shoji, H. Yamaguchi, K. Okamoto, K. Ishiguro, Y. Harigaya & K. Wakabayashi: Alpha 1-antichymotrypsin as a possible biochemical marker for Alzheimer-type dementia. *Ann Neurol* 28, 561-567 (1990)

29. Lieberman J., L. Schleissner, KH. Tachiki & AS. Kling: Serum alpha 1-antichymotrypsin level as a marker for Alzheimer-type dementia. *Neurobiol Aging* 16, 747-753 (1995)

30. Licastro F, MC. Morini, E. Polazzi & LJ. Davis: Increased serum alpha 1-antichymotrypsin in patients with probable Alzheimer's disease: an acute phase reactant without the peripheral acute phase response. *J Neuroimmunol* 57, 71-75 (1995)

31. Brugge K, R. Katzman, LR. Hill, LA. Hansen & T. Saitoh: Serological alpha 1-antichymotrypsin in Down's syndrome and Alzheimer's disease. *Ann Neurol* 32, 193-197 (1992)

32. Hinds T, WA. Kukull, G. Van Belle, GD. Schellenberg, EC. Villacres & EB. Larson: Relationship between serum alpha 1-antichymotrypsin and Alzheimer's disease. *Neurobiol Aging* 15, 21-27 (1994)

33. Namba Y, M. Tomonaga, H. Kawasaki, E. Otomo & K. Ikeda: Apolipoprotein E immunoreactivity in cerebral amyloid deposits and neurofibrillary tangles in Alzheimer's disease and kuru plaque amyloid in Creutzfeldt- Jakob disease. *Brain Res* 541, 163-166 (1991)

34. Wisniewski T. & B. Frangione: Apolipoprotein E: a pathological chaperone protein in patients with cerebral and systemic amyloid. *Neurosci Lett* 135, 235-238 (1992)

35. Corder E, AM. Saunders, WJ. Strittmatter, DE. Schmechel, PC. Gaskell, GW. Small, AD. Roses & JL. Haines: Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261, 921-923 (1993)

36. Strittmatter W, AM. Saunders, D. Schmechel, M. Pericak-Vance, J. Enghild, GS. Salvesen & AD. Roses: Apolipoprotein E: high avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer's disease. *Proc Natl Acad Sci USA* 90, 1977-1981 (1993)

37. Poirier J, J. Davignon, D. Bouthillier, S. Kogan, P. Bertrand & S. Gauthier: Apolipoprotein E polymorphism and Alzheimer's disease. *Lancet* 342, 697-699 (1993)

38. Poirier J: Apolipoprotein E in animal models of CNS injury and in Alzheimer's disease. *Trends Neurosci* 17, 525-530 (1994)

39. Näslund J, J. Thyberg, LO. Tjernberg, C. Wernstedt, AR. Karlström, N. Bogdanovic, SE. Gandy, L. Lannfelt, L. Terenius & C. Nordstedt: Characterization of stable complexes involving apolipoprotein E and amyloid beta peptide in Alzheimer's disease brain. *Neuron* 15, 219-228 (1995)

40. Strittmatter W & AD. Roses: Apolipoprotein E and Alzheimer's disease. *Ann Rev Neurosci* 19, 53-77 (1996)

41. Strijbos P. & NJ. Rothwell: Interleukin-1 attenuates excitatory amino acid induced neurodegeneration *in vitro*: involvement of nerve growth factor. *J Neurosci* 15, 3468-3474 (1995)

42. Schwartz L & CE. Milligan: Cold thoughts of death: the role of ICE proteases in neuronal cell death. *Trends Neurosci* 19, 555-562 (1996)

43. Giulian D. & LB. Lachman: Interleukin-1 stimulation of astroglial proliferation after brain injury. *Science* 228, 497-499 (1985)

44. Giulian D, TJ. Baker, LC. Shih & LB. Lachman: Interleukin 1 of the central nervous system is produced by ameboid microglia. *J Exp Med* 164, 594-604 (1986)

45. Giulian D, J. Woodward, DG. Young, JF. Krebs & LB. Lachman: Interleukin-1 injected into mammalian brain stimulates astrogliosis and neovascularization. *J Neurosci* 8, 2485-2490 (1988)

46. Griffin W, LC. Stanley, C. Ling, L. White, V. MacLeod, LJ. Perrot, CL. White III & C. Araoz: Brain interleukin-1 and S100 immunoreactivity elevated in Down's syndrome and Alzheimer's disease. *Proc Natl Acad Sci USA* 86, 7611-7615 (1989)

47. Das S. & H. Potter: Expression of the Alzheimer amyloid-promoting factor antichymotrypsin is induced in human astrocytes by IL-1. *Neuron* 14, 447-456 (1995)

48. Sheng J, RE. Mrak & WS. Griffin: Microglial interleukin-1 alpha expression in brain regions in Alzheimer's disease: correlation with neuritic plaque distribution. *Neuropathol Appl Neurobiol* 21, 290-301 (1995)

49. Goldgaber D, HW. Harris, T. Hla, T. Maciag, RJ. Donnelly, JS. Jacobsen, MP. Vitek & DC. Gajdusek: Interleukin 1 regulates synthesis of amyloid-beta-protein precursor mRNA in human endothelial cells. *Proc Natl Acad Sci USA* 86, 7606-7610 (1989)

50. Rogers J, L. Nilsson, J. McPhee, SS. Zhan, H. Potter & L. Leiter: Translational regulation of amyloid precursor protein in astrocytes by the inflammatory cytokine IL-1. *Submitted*

51. Baumann H & J. Gauldie: The acute phase response. *Immunol Today* 15, 74-80 (1994)

Pathogenic pathway of Alzheimer's disease

52. Bauer J, S. Strauss, U. Schreiter-Gasser, U. Genter, P. Schlegel, I. Witt, B. Volk & M. Berger: Interleukin-6 and alpha₂-macroglobulin indicate an acute-phase state in Alzheimer's disease cortex. *FEBS Lett* 285, 111-114 (1991)
53. Blum-Degen D, T. Muller, W. Kuhn, M. Gerlach, H. Przuntek & P. Riederer: Interleukin-1 beta and interleukin-6 are elevated in the cerebrospinal fluid of Alzheimer's and de novo Parkinson's disease patients. *Neurosci Lett* 202, 17-20 (1995)
54. Strauss S, J. Bauer, U. Ganter, U. Jonas, M. Berger & B. Volk: Detection of interleukin-6 and alpha 2-macroglobulin immunoreactivity in cortex and hippocampus of Alzheimer's disease patients. *Lab Invest* 66, 223-30 (1992)
55. Wood J, PL. Wood, R. Ryan, NR. Graff-Radford, C. Pilapil, Y. Robitaille & R. Quirion: Cytokine indices in Alzheimer's temporal cortex: no changes in mature IL-1 beta or IL-1RA but increases in the associated acute phase proteins IL-6, alpha 2-macroglobulin and C-reactive protein. *Brain Res* 629, 245-252 (1993)
56. Huell M, S. Strauss, B. Volk, M. Berger & J. Bauer. Interleukin-6 is present in early stages of plaque formation and is restricted to the brains of Alzheimer's disease patients. *Acta Neuropathol* 89, 544-551 (1995)
57. Campbell I, CR. Abraham, E. Masliah, P. Kemper, JD. Inglis, MB. Oldstone & L. Mucke: Neurologic disease induced in transgenic mice by cerebral overexpression of interleukin 6. *Proc Nat Acad Sci USA* 90, 10061-11065 (1993)
58. Heyser C, E. Masliah, A. Samimi, IL. Campbell & LH. Gold: Progressive decline in avoidance learning paralleled by inflammatory neurodegeneration in transgenic mice expressing interleukin 6 in the brain. *Proc Natl Acad Sci USA* 94, 1500-1505 (1997)
59. Fattori E, D. Lazzaro, P. Musiani, A. Modesti, T. Alonzi & G. Ciliberto: IL-6 expression in neurons of transgenic mice causes reactive astrocytosis and increase in ramified microglial cells but no neuronal damage. *Eur J Neurosci* 7, 2441-2449 (1995)
60. Goodman J, CS. Robertson, RG. Grossman & RK. Narayan: Elevation of tumor necrosis factor in head injury. *J Neuroimmunol* 30, 213-217 (1990)
61. Louis J, E. Magal, S. Takayama & S. Varon: CNTF protection of oligo-dendrocytes against natural and tumor necrosis factor-induced death. *Science* 259, 689-692 (1993)
62. Morganti-Kossmann M, T. Kossmann & SM. Wahl: Cytokines and neuropathology. *Trends Pharmacol Sci* 13, 286-291 (1992)
63. Bruce A, W. Boling, MS. Kindy, J. Peschon, PJ. Kraemer, MK. Carpenter, FW. Holtzberg & MP. Mattson: Altered neuronal and microglial responses to excitotoxic and ischemic brain injury in mice lacking TNF receptors. *Nat Med* 20, 788-94 (1996)
64. Hofman F, DR. Hinton, K. Johnson & JE Merrill: Tumor necrosis factor identified in multiple sclerosis brain. *J Exp Med* 170, 607-612 (1989)
65. Selmaj K, CS. Raine, B. Cannella & CF. Brosnan: Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. *J Clin Invest* 87, 949-954 (1991)
66. Fillit H, WH. Ding, L. Buee, J. Kalman, L. Altstiel, B. Lawlor & G. Wolf-Klein: Elevated circulating tumor necrosis factor levels in Alzheimer's disease. *Neurosci Lett* 129, 318-320 (1991)
67. Cacabelos R, XA. Alvarez, A. Franco-Maside, L. Fernandez-Novoa & J. Caamano: Serum tumor necrosis factor (TNF) in Alzheimer's disease and multi-infarct dementia. *Meth Find Exp Clin Pharmacol* 16, 29-35 (1994)
68. Wyss-Coray T, E. Masliah, M. Mallory, L. McConlogue, K. Johnson-Wood, C. Lin & L. Mucke: Amyloidogenic role of cytokine TGF-beta1 in transgenic mice and in Alzheimer's disease. *Nature* 389, 603-606 (1997)
69. Glenner G, JH. Henry & S. Fujihara: Congophilic angiopathy in the pathogenesis of Alzheimer's degeneration. *Ann de Pathol* 1, 120-129 (1981)
70. Vinters H: Cerebral amyloid angiopathy. A critical review. *Stroke* 18, 311-324 (1987)
71. Wyss-Coray T, L. Feng, E. Masliah, MD. Ruppe, HS. Lee, SM. Toggas, EM. Rockenstein & L. Mucke: Increased central nervous system production of extracellular matrix components and development of hydrocephalus in transgenic mice overexpressing transforming growth factor-beta 1. *Am J Pathol* 147, 53-67 (1995)
72. Van der Wal EA, F. Gomez-Pinilla & CW. Cotman: Transforming growth factor-beta 1 is in plaques in Alzheimer and Down pathologies. *Neuroreport* 4, 69-72 (1993)
73. Rozemuller J, P. Eikelenboom, FC. Stam, K. Beyreuther & CL. Masters: A4 protein in Alzheimer's disease: primary and secondary cellular events in extracellular amyloid deposition. *J Neuropathol Exp Neurol* 48, 674-691 (1989)
74. Snow A, H. Mar, D. Nochlin, RT. Sekiguchi, K. Kimata, Y. Koike & TN. Wight: Early accumulation of heparin sulfate in neurons and in the amyloid-protein-containing lesions of Alzheimer's disease and Down's syndrome. *Am J Pathol* 137, 1253-1270 (1990)
75. McGeer P & EG. McGeer: The inflammatory response system of the brain: implications for therapy of Alzheimer and other neurodegenerative diseases. *Brain Res Brain Res*

Pathogenic pathway of Alzheimer's disease

Rev 21, 195-218 (1995)

76. Heyman A, WE. Wilkinson, JA. Stafford, MJ. Helms, AH. Sigmon & T. Weinberg: Alzheimer's disease: a study of epidemiological aspects. *Ann Neurol* 15, 335-341 (1984)

77. Mortimer J, LR. French, JT. Hutton & LM. Schuman: Head injury as a risk factor for Alzheimer's disease. *Neurology* 35, 264-267 (1985)

78. Mortimer, J, CM. van Duijn, V. Chandra, L. Fratiglioni, AB. Graves, A. Heyman, AF. Jorm, E. Kokmen, K. Kondo, WA. Rocca, SL. Shalat & H. Soininen: Head trauma as a risk factor for Alzheimer's disease: a collaborative reanalysis of case-control studies. EURODEM Risk Factors Research Group. *Int J Epidemiol* 20 Suppl 2, S28-35 (1991)

79. Schofield P, M. Tang, K. Marder, K. Bell, G. Dooneief, M. Chun, M. Sano, Y. Stern & R. Mayeux: Alzheimer's disease after remote head injury: an incidence study. *J Neurol Neurosurg Psychiatry* 62, 119-124 (1997)

80. Corsellis J, CJ. Bruton & D. Freeman-Browne: The aftermath of boxing. *Psychol Med* 3, 270-303 (1973)

81. Roberts GW: Immunohistochemistry of neurofibrillary tangles in dementia pugilistica and Alzheimer's disease: evidence for common genesis. *Lancet* 2, 1456-1458 (1988)

82. Roberts GW, D. Allsop & C. Bruton: The occult aftermath of boxing. *J Neurol Neurosurg Psychiatry* 53, 373-378 (1990)

83. Griffin W, JG. Sheng, SM. Gentleman, DI. Graham, RE. Mrak & GW. Roberts: Microglial interleukin-1 alpha expression in human head injury correlations with neuronal and neuritic beta-amyloid precursor protein expression. *Neurosci Lett* 176, 133-136 (1994)

84. Roberts GW, SM. Gentleman, A. Lynch & PI. Graham: Ab4 amyloid protein deposition in the brain after head trauma. *Lancet* 338, 1422-1423 (1991)

85. Nicoll J, GW. Roberts & DI. Graham: Apolipoprotein E epsilon 4 allele is associated with deposition of amyloid beta-protein following head injury. *Nat Med.* 1, 135-137 (1995)

86. Jenkinson M, MR. Bliss, AT. Brain & DL. Scott: Rheumatoid arthritis and senile dementia of the Alzheimer's type. *Br J Rheumatol* 28, 86-88 (1989)

87. McGeer P, E. McGeer, J. Rogers & J. Sibley: Anti-inflammatory drugs and Alzheimer disease. *Lancet* 335, 1037 (1990)

88. Andersen K, LJ. Launer, A. Ott, A.W. Hoes, M.M. Breteler & A. Hofman: Do nonsteroidal anti-inflammatory drugs decrease the risk for Alzheimer's disease? The Rotterdam Study. *Neurology* 45, 1441-1445 (1995)

89. Rich J, DX. Rasmusson, MF. Folstein, KA. Carson, C. Kawas & J. Brandt: Nonsteroidal anti-inflammatory drugs in Alzheimer's disease. *Neurology* 45, 51-55 (1995)

90. McGeer P, M. Schulzer & EG. McGeer: Arthritis and anti-inflammatory agents as possible protective factors for Alzheimer's disease: a review of 17 epidemiologic studies. *Neurology* 47, 425-432 (1996)

91. Breitner J, BA. Gau, KA. Welsh, BL. Plassman, WM. McDonald, MJ. Helms & JC. Anthony: Inverse association of anti-inflammatory treatments and Alzheimer's disease: initial results of a co-twin control study. *Neurology* 44, 227-232 (1994)

92. Rogers J, LC. Kirby, SR. Hempelman, DL. Berry, PL. McGeer, AW. Kaszniak, J. Zaluski, M. Cofield, L. Mansukhani, P. Willson, & F. Kogan: Clinical trials of indomethacin in Alzheimer's disease. *Neurology* 43, 1609-1611 (1993)

93. Selkoe D: Cell biology of the amyloid beta-protein precursor and the mechanism of Alzheimer's disease. *Annu Rev Cell Biol* 10, 373-403 (1994)

94. Selkoe D: Alzheimer's disease: genotypes, phenotypes, and treatments. *Science.* 275, 630-631 (1997)

95. Terry RD, E. Masliah, DP. Salmon, N. Butters, R. DeTeresa, R. Hill, LA. Hansen & R. Katzman: Physical basis of cognitive alterations in Alzheimer's disease: synapse loss is the major correlate of cognitive impairment. *Ann Neurol* 30, 572-580 (1991)

96. Wegiel J. & HM. Wisniewski: The complex of microglial cells and amyloid star in three-dimensional reconstruction. *Acta Neuropathol* 81, 116-124 (1990)

97. Wisniewski H. & J. Wegiel: Spatial relationships between astrocytes and classical plaque components. *Neurobiol Aging* 12, 593-600 (1991)

98. McGeer P, S. Itagaki, H. Tago & EG. McGeer: Reactive microglia in patients with senile dementia of the Alzheimer type are positive for the histocompatibility glycoprotein HLA-DR. *Neurosci Lett* 79, 195-200 (1987)

99. Ma J, A. Yee, HB. Brewer, S. Das & H. Potter: Amyloid-associated proteins alpha₁-antichymotrypsin and apolipoprotein E promote assembly of Alzheimer beta-protein into filaments. *Nature* 372, 92-94 (1994)

100. Burdick D, B. Soreghan, M. Kwon, J. Kosmoski, M. Knauer, A. Henschen, J. Yates, C. Cotman & C. Glabe: Assembly and aggregation properties of synthetic Alzheimer's A4/beta amyloid peptide analogs. *J Biol Chem* 267, 546-554 (1992)

101. Jarrett J, EP. Berger & PT. Lansbury Jr: The carboxy terminus of the beta amyloid protein is critical for the seeding of amyloid formation: implications for the pathogenesis of Alzheimer's disease. *Biochemistry* 32,

Pathogenic pathway of Alzheimer's disease

4693-4697 (1993)

102. Roher A, JD. Lowenson, S. Clarke, AS. Woods, RJ. Cotter, E. Gowing & MJ. Ball: Beta-Amyloid-(1-42) is a major component of cerebrovascular amyloid deposits: implications for the pathology of Alzheimer disease. *Proc Natl Acad Sci USA* 90, 10836-10840 (1993)

103. Iwatsubo T, A. Odaka, N. Suzuki, H. Mizusawa, N. Nukina & Y. Ihara: Visualization of A beta 42(43) and A beta 40 in senile plaques with end-specific A beta monoclonals: evidence that an initially deposited species is A beta 42(43). *Neuron* 13, 45-53 (1994)

104. Iwatsubo T, DM. Mann, A. Odaka, N. Suzuki & Y. Ihara: Amyloid beta protein (A beta) deposition: A beta 42(43) precedes A beta 40 in Down syndrome. *Ann Neurol* 37, 294-299 (1995)

105. Lemere C, JK. Blusztajn, H. Yamaguchi, T. Wisniewski, TC. Saido & DJ. Selkoe: Sequence of deposition of heterogeneous amyloid beta-peptides and APO E in Down syndrome: implications for initial events in amyloid plaque formation. *Neurobiol Dis* 3, 16-32 (1996)

106. Jarrett J & PT. Lansbury Jr. Seeding "one-dimensional crystallization" of amyloid: a pathogenic mechanism in Alzheimer's disease and scrapie? *Cell* 73, 1055-1058 (1993)

107. Griffin W, JG. Sheng, GW. Roberts & RE. Mrak: Interleukin-1 expression in different plaque types in Alzheimer's disease: significance in plaque evolution. *J Neuropathol Exp Neurol* 54, 276-281 (1995)

108. Frautschy S, FS. Yang, M. Irizarry, B. Hyman, TC. Saido, K. Hsiao & GM. Cole: Microglial response to amyloid plaques in APPsw transgenic mice. *Am J Pathol* 152, 307-317 (1998)

109. Baumann H, C. Richards & J. Gaudie: Interaction among hepatocyte-stimulating factors, interleukin 1, and glucocorticoids for regulation of acute phase plasma proteins in human hepatoma (HepG2) cells. *J Immunol* 139, 4122-8 (1987)

110. Diedrich JF, H. Minnigan, RI. Carp, JN. Whitaker, R. Race, W. Frey II & AT. Haase: Neuropathological changes in scrapie and Alzheimer's disease are associated with increased expression of apolipoprotein E and cathepsin D in astrocytes. *J Virol* 65, 4759-4768 (1991)

111. Müller H, PJ. Gebicke-Harter, DH. Hangen & EM. Shooter: A specific 37,000-dalton protein that accumulates in regenerating but not in nonregenerating mammalian nerves. *Science* 228, 499-501 (1985)

112. Ignatius M, PJ. Gebicke-Harter, JH. Skene, JW. Schilling, KH. Weisgraber, RW. Mahley & EM Shooter: Expression of apolipoprotein E during nerve degeneration and regeneration. *Proc Natl Acad Sci USA* 83, 1125-1129 (1986)

113. Snipes GJ, CB. McGuire, JJ. Norden & JA. Freeman: Nerve injury stimulates the secretion of apolipoprotein E by nonneuronal cells. *Proc Natl Acad Sci USA* 83, 1130-1134 (1986)

114. Poirier J, M. Hess, PC. May & CE. Finch: Cloning of hippocampal poly(A) RNA sequences that increase after entorhinal cortex lesion in adult rat. *Brain Res Mol Brain Res* 9, 191-195 (1991)

115. Poirier J, A. Baccichet, D. Dea & S. Gauthier: Cholesterol synthesis and lipoprotein reuptake during synaptic remodelling in hippocampus in adult rats. *Neuroscience* 55, 81-90 (1993)

116. Bullido M, MJ. Artiga, M. Recuero, I. Sastre, MA. Garcia, J. Aldudo, C. Lendon, SW. Han, JC. Morris, A. Frank, J. Vazquez, A. Goate & F. Valdivieso: A polymorphism in the regulatory region of ApoE associated with risk for Alzheimer's dementia. *Nat Genet* 18, 69-71 (1998)

117. Sheng J, K. Ito, RD. Skinner, RE. Mrak, CR. Rovnaghi, LJ. v Eldik, & WST. Griffin: *In vivo* and *in vitro* evidence supporting a role for the inflammatory cytokine interleukin-1 as a driving force in Alzheimer pathogenesis. *Neurobiol Aging* 17, 761-766 (1996)

118. Sanan D, KH. Weisgraber, SJ. Russell, RW. Mahley, D. Huang, A. Saunders, D. Schmechel, T. Wisniewski, B. Frangione, AD. Roses & WJ. Strittmatter: Apolipoprotein E associates with beta-amyloid peptide of Alzheimer's disease to form novel monofibrils. *J Clin Invest* 94, 860-869 (1994)

119. Wisniewski T, EM. Castaño, A. Golabek, T. Vogel, & B. Frangione: Acceleration of Alzheimer's fibril formation by apolipoprotein E *in vitro*. *Am J Pathol* 145, 1030-1035 (1994)

120. Janciauskiene S, S. Eriksson & HT. Wright: A specific structural interaction of Alzheimer's peptide A beta 1-42 with alpha 1-antichymotrypsin. *Nat Struct Biol* 3, 668-671 (1996)

121. Eriksson S, S. Janciauskiene & L. Lannfelt: Alpha₁-antichymotrypsin regulates Alzheimer beta-amyloid peptide fibril formation. *Proc Natl Acad Sci USA* 92, 2313-2317 (1995).

122. Evans KC, EP. Berger CG. Cho, KH. Weisgraber & PT. Lansbury: Apolipoprotein E is a kinetic but not a thermodynamic inhibitor of amyloid formation: Implications for the pathogenesis and treatment of Alzheimer's disease. *Proc Natl Acad Sci USA* 92, 763-767 (1995)

123. Fraser P, JT. Nguyen, DR. McLachlan, CR. Abraham & A. Kirschner: Alpha₁-antichymotrypsin binding to Alzheimer Abeta peptides is sequence specific and induces fibril disaggregation *in vitro*. *J Neurochem* 61, 298-305 (1993)

Pathogenic pathway of Alzheimer's disease

124. Bales K, T. Verina, RC. Dodel, YS. Du, L. Altstiel, M. Bender, P. Hyslop, EM. Johnstone, SP. Little, DJ. Cummins, P. Piccardo, B. Ghetti & SM. Paul.: Lack of apolipoprotein E dramatically reduces amyloid beta-peptide deposition. *Nat Genet* 17, 263-264 (1997)

125. Kamboh M, DK. Sanghera, RE. Ferrell & ST. DeKosky: ApoE4-associated Alzheimer's disease risk is modified by alpha1-antichymotrypsin polymorphism. *Nat Genet* 10, 486-488 (1995)

126. Yamada M, N. Sodeyama, Y. Itoh, N. Suematsu, E. Otomo, M. Matsushita & H. Mizusawa: Association of the alpha1- antichymotrypsin polymorphism with cerebral amyloid angiopathy. *Ann Neurol* in press. (1998)

127. Yankner B.: Mechanisms of neuronal degeneration in Alzheimer's disease. *Neuron* 16, 921-932 (1996)

128. Busciglio J, A. Lorenzo, J. Yeh & BA. Yankner: Beta-amyloid fibrils induce tau phosphorylation and loss of microtubule binding. *Neuron* 14, 879-888 (1995)

129. Irizarry M, F. Soriano, M. McNamara, KJ. Page, D. Schenk, D. Games & BT. Hyman: Abeta deposition is associated with neuropil changes, but not with overt neuronal loss in the human amyloid precursor protein V717F (PDAPP) transgenic mouse. *J Neurosci* 17, 7053-7059 (1997)

130. Wertkin AM, RS. Turner, SJ. Pleasure, TE. Golde, SG. Younkin, JQ. Trojanowski & VM. Lee: Human neurons derived from a teratocarcinoma cell line express solely the 695-amino acid amyloid precursor protein and produce intracellular beta-amyloid or A4 peptides. *Proc Natl Acad Sci USA* 90, 9513-9517 (1993)

131. Tienari P, N. Ida, E. Ikonen, M. Simons, A. Weidemann, G. Multhaup, CL. Masters CG, Dotti & K. Beyreuther: Intracellular and secreted Alzheimer beta-amyloid species are generated by distinct mechanisms in cultured hippocampal neurons. *Proc Natl Acad Sci USA* 94, 4125-4130 (1997)

132. El Khoury J, SE. Hickman, CA. Thomas, L. Cao, SC. Silverstein & JD. Loike: Scavenger receptor-mediated adhesion of microglia to beta-amyloid fibrils. *Nature* 382, 716-719 (1996)

133. Yan SD, X. Chen, J. Fu, M. Chen, H. Zhu, A. Roher, T. Slattery, L. Zhao, M. Nagashima, J. Morser, A. Migheli, P. Nawroth, D. Stern & AM. Schmidt: RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. *Nature* 382, 685-91 (1996)

Key Words: Alzheimer's Disease, Alpha₁-Antichymotrypsin, Apolipoprotein E, Beta-Amyloid, Chaperone, Cytokine, Gene Expression, Inflammation

Send correspondence to: Huntington Potter, Department of Neurobiology, Harvard Medical School, Boston, MA 02115, USA, Tel: (617)-432-0751, Fax: (617)-734-7557, E-mail: hpotter@warren.med.harvard.edu