CLINICAL APPLICATIONS OF QUINONE-CONTAINING ALKYLATING AGENTS

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1. ABSTRACT

Quinone-containing alkylating agents are a class of chemical agents that have received considerable interest as anticancer drugs. These agents contain a quinone moiety that can be reduced and an alkylating group that can form covalent bonds with a variety of cellular components. The oxidation state of the quinone element can modulate the activity of the alkylating element, and reduction of the quinone is required for activation of the alkylating activity of many of these agents. The quinone element may also contribute to the cytotoxic activity of quinone-containing alkylating agents through the formation of reactive oxygen species during redox cycling. The natural product, mitomycin C, has been the most widely used quinonecontaining alkylating agent in the clinic, but other quinonecontaining alkylating agents like porfiromycin, diaziquone, carbazilquinone, triaziquone and EO9 havealso been used in the clinic for the treatment of cancer. In addition, many other quinone-containing alkylating agents have been tested in preclinical studies and the development of new agents is being actively pursued. This chapter describes the current and past clinical uses of these agents in the treatment of cancer and discusses new agents that are currently in clinical trials.

2. INTRODUCTION

2.1. Historical Perspective

Quinone-containing alkylating agents are a class of chemical agents that have received considerable interest as anticancer drugs since the late 1950s (1). These agents contain a quinone moiety that can be reduced and an alkylating group that can form covalent bonds with a variety of cellular components. Simple alkylating agents were among the first effective agents for the treatment of cancer (2). The nitrogen mustard, HN2, was first tested in the clinic in 1942, and this agent is still used clinically for the treatment of lymphomas (3). Other nitrogen mustards like chlorambucil, melphalan and cyclophosphamide were introduced shortly afterwards and remain important drugs in many treatment regimens (4). Other alkylating agents ontaining methane sulfonates, like busulfan (5), and aziridines, like thiotepa (6) were also early antitumor Putter (7) suggested that combining a agents. benzoquinone with alkylating groups would produce compounds with potent antitumor activity. This suggestion was supported by the studies of Holzer et al (1) who showed that a number of aziridinyl benzoquinones had very good antitumor activity, and by the isolation of the quinone-containing mitomycin antitumor antibiotics (8,9). Mitomycin C was extensively tested in animals (8,10) and humans (11-13) in the late 1950s and early 1960s, and was shown to be an effective clinical agent. These findings prompted a proliferation of synthetic and biological studies of quinone-containing alkylators as potential antitumor agents (14-16). Many of these agents showed good antitumor activity in cell culture or animal models.

Studies of the mechanism of action of mitomycin C demonstrated that this agent required reductive activation, which was followed by the formation of DNA adducts and strand breaks (17,18). This finding was consistent with earlier predictions by Ross (19) that the oxidation state of the quinone group could influence the activity of an adjacent alkylating moiety. Sartorelli et al. (20) used this principle to prepare a series of benzoquinone and naphthoquinone compounds, which had the potential to alkylate after reduction, as potential "bioreductive alkylating agents". These agents were able to bind to DNA and demonstrated antitumor activity (20). These studies lead to increased interest in the development of quinone-containing alkylating agents as antitumor agents and in understanding their mechanism of action.

2.2. Mechanism of Action of Quinone-containing Alkylating Agents

Quinone-containing alkylating agents contain two important structural elements that contribute to their activity. The alkylating element can form covalent bonds to cellular components including proteins, membranes and DNA; however, the interaction with DNA is generally

thought to be the most important contributor to the antitumor activity of these agents (21). This interaction can result in the formation of DNA monoadducts, or in the case of bifunctional alkylating agents, DNA crosslinks (22,23), leading to the induction of apoptosis (24) and cell death (25). The oxidation state of the quinone element can modulate the activity of the alkylating element, and reduction of the quinone is required for activation of the alkylating activity of agents such as mitomycin C (26,27) and EO9 (26,28). The most important activating enzymes for quinone-containing alkylating agents appear to be the one-electron reducing enzyme, NADPH:cytochrome P450 reductase (21,27,29,30), and the two-electron reducing NAD(P)H:quinone oxidoreductase enzyme, (DTdiaphorase) (21,29,31-34). However, not all agents may require reductive activation (35), and reduction of the quinone group by some enzymes may decrease the activity of these agents (36). The quinone element may also contribute to the cytotoxic activity of guinone-containing alkylating agents through the formation of reactive oxygen species during redox cycling. Reactive oxygen species can cellular components, including damage DNA (21,23,37,38), but this seems to be a minor contributor to the antitumor activity of bioreductive agents (39,40).

2.3. Clinical Application of Quinone-containing Alkylating Agents

The natural product, mitomycin C, has been the most widely used quinone-containing alkylating agent in the clinic, but other quinone-containing alkylating agents like porfiromycin, diaziquone, carbazilquinone, triaziquone and EO9 have also been used in the clinic for the treatment of cancer. In addition, many other quinone-containing alkylating agents have been tested in preclinical studies and the development of new agents is being actively pursued. This chapter will describe the current and past clinical uses of these agents in the treatment of cancer and will discuss new agents that are currently in clinical trials.

3. CLINICAL APPLICATIONS OF MITOMYCINS

3.1. History

The mitomycins were first discovered by Hata et al (8) in 1956, and mitomycin C was isolated by Wakaki et al (9) from *Streptomyces caespitosus* in 1958. Four mitomycins occur naturally (Figure 1). All are antibiotics effective against both gram positive and gram negative bacteria (41), but only mitomycin C and porfiromycin have appreciable anticancer activity. Mitomycin C has been used clinically in Japan since the early 1960's (42), but this agent was not approved for general use in North America until 1974 (43). Since then mitomycin C has been used to treat a wide variety of solid tumors, but its utility has been decreased by toxicity. Porfiromycin has also received limited clinical use, and a number of mitomycin analogues have been investigated (42).

3.2. Mechanism of Action

The mitomycins require intracellular activation (26) by one electron reducing enzymes like NADPH:cytochrome P450 reductase (26,39) or NADH:cytochrome b_5 reductase (44), or by two electron reducing enzymes like DT-diaphorase



	0		
	<u>X</u>	<u>Y</u>	<u>Z</u>
Mitomycin A	OCH ₃	OCH ₃	Н
Mitomycin B	OCH ₃	OH	CH_3
Mitomycin C	NH ₂	OCH ₃	Н
Porfiromycin	NH_2	OCH ₃	CH_3
BMY-25282	N=CHN(CH ₃) ₂	OCH ₃	Н
BMS-181174	NH(CH ₂) ₂ SS-C ₆ H ₄ -NO ₂	OCH ₃	Н
KW2149	NH(CH ₂) ₂ SS(CH ₂) ₂ NHCO(CH ₂) ₂ CH(NH ₂)CO ₂ H	OCH ₃	Н
M83	NH-C ₆ H ₄ -OH	OCH ₃	Н

Figure 1. Structures of mitomycins

(21,31) or xanthine dehydrogenase (45); however, NADPH:cytochrome P450 reductase (21,29,30) and DTdiaphorase (28,29,31-34) are the most important activating enzymes. Reduction of the mitomycins activates the aziridine and carbamate alkylating groups and results in the formation of DNA crosslinks which appear to be the primary mechanism for the antitumor effects of these agents (21-23). If oxygen is present, the reduced mitomycin products can redox cycle to form reactive oxygen species. Reactive oxygen species damage cellular components, including DNA (21,23,37,46), but this seems to be a minor contributor to the antitumor activity of these agents (39). The mitomycins are generally more active under hypoxic conditions and can act as radiosensitizers (26,33).

3.3. Mitomycin C 3.3.1. Clinical Applications

Mitomycin C has been used for the clinical treatment of human cancers for more than 25 years. Despite multiple clinical trials, the use and effectiveness of this agent remains confined to a small number of cancer types. Mitomycin C has proved to be most effective in the front line treatment of a small number of solid tumors, such as superficial bladder cancer (47) and gastric (48), pancreatic (49), anal (50) and esophageal (51) carcinomas. It is also used in palliative treatment of advanced cancers or cancers that have become resistant to other forms of therapy, generally in combination regimens with doxorubicin and 5-fluorouracil or with bleomycin and vincristine (43,52,53). However, the activity of this agent in solid tumors and its enhanced effectiveness against hypoxic cells that are resistant to radiation (26) have resulted in continued interest in mitomycin C and the investigation of new approaches to increasing its effectiveness.

Superficial bladder cancer represents approximately 70% of all bladder cancers at time of presentation (47,54). Transurethral resection is the primary treatment for this disease, but tumors will recur in 50% of all patients (54). Thus, patients at highest risk for tumor recurrence generally receive adjuvant intravesical therapy

with anticancer drugs such as mitomycin C, thiotepa or doxorubicin, or with immunotherapy with Bacillus Many clinical studies have Calmette-Guerin (BCG). established the effectiveness of mitomycin C for the treatment of superficial bladder cancer, with complete response rates of from 39% to 77% reported (47). The efficacy of mitomycin C in the adjuvant setting in preventing the recurrence of tumors has also been confirmed (47,55). Drug doses of 20-40 mg by instillation after transurethral resection and then weekly for 6 to 8 weeks (47,56) with, or without monthly maintenance treatments have been used (56,57). Furthermore, recent studies comparing mitomycin C and BCG showed that there was no difference in tumor progression or survival with these two agents, but that BCG might be superior in preventing tumor recurrence (56) possibly at the expense of greater adverse reactions (58).

Adenocarcinoma of the stomach is a highly malignant tumor with a poor prognosis (49). This disease is a major cause of cancer mortality in Japan and other Asian countries, and many patients present with advanced disease which is not curable by surgery alone (48). Mitomycin C has been used extensively as a single agent in the treatment of gastric cancer, especially in Japan with an overall objective response rate of 30% (49). This compares favorably with other single agent therapy with 5-FU or doxorubicin. More recently, mitomycin C has also been extensively used in a number of combination chemotherapy regimens, most notable in the FAM regimen with 5-FU and doxorubicin (48,49,59,60). Mitomycin C is typically given as an *i.v.* bolus at 10 mg/m² on day 1 of a 6 to 10 week cycle. The response rates have been approximately 30% with a small number of complete responders. Although this adjuvant treatment increased the time to progression it did not alter survival time in patients with more advanced disease (49). Thus, mitomycin C regimens provide only limited benefit in gastric cancer.

Pancreatic carcinoma has a similar incidence to gastric cancer in North America and also has a poor prognosis. This disease is highly resistant to chemotherapy but response rates of approximately 25% have been seen with 5-FU or mitomycin C as single agents or with combinations of these agents (49). However, these treatments have little effect on overall survival. More recent trials to investigate the use of preoperative radiation and chemotherapy with 5-FU and mitomycin C to enhance resectability have provided encouraging results (61,62).

Epidermoid carcinomas of the anal region represent only a small proportion of large bowel cancers (63). When these cancers were treated by radical surgery, the 5-year survival rate ranged from 40% to 60% with local relapses being common. More recently, radiotherapy combined with 5-FU and mitomycin C have been extensively used in the treatment of this disease. A number of clinical trials have demonstrated that the concomitant use of radiotherapy and chemotherapy produced a significant improvement in locoregional control and a reduction in the need for surgery (64) and that mitomycin C increased colostomy-free and disease-free survival (63). This regimen has become the standard of care in many centers. Mitomycin C is usually given as an *i.v.* bolus at from 10 to 15 mg/m² on day 1 of treatment, with a second dose given at 6 weeks in some cases.

Although carcinoma of the esophagus is still a relatively uncommon disease in North American, the incidence of this disease is increasing and overall 5-year survival rates are only 20%. While this disease is primarily treated by surgery many studies have demonstrated the utility of preoperative concurrent chemoradiotherapy (65,66). Such treatment produces complete response rates of approximately 25% for both squamous cell carcinoma and adenocarcinoma, and may provide a slight survival advantage (66). The chemotherapy regimens normally contain 5-FU with mitomycin C or cisplatin (51,65,66). While combinations with mitomycin C or cisplatin produced similar results (66), the latter agent is more commonly used.

Numerous studies have shown that mitomycin C, used as a single agent, can produce good response rates in a wide variety of solid tumors (43). Thus, this agent is used in the palliative treatment of a number of advanced cancers. In patients with advanced non-small cell lung cancer, combinations of mitomycin C with cisplatin and vinca alkaloids (67) or cyclophosphamide (68) produced high response rates and a moderate increase in survival time. Similarly, the MMM combination of mitomycin C, mitoxantrone and methotrexate produced good response rates in advanced breast cancer (69), which were comparable to other regimens (70,71). However, these regimens have received only limited use because of their high toxicity.

3.3.2. Clinical Studies

Although the current clinical use of mitomycin C is limited, this agent continues to be a focus of many clinical trials because of its intrinsic activity in many solid tumors and its preferential activity in hypoxic cells (26). A number of clinical trials have used this agent as an adjunct to radiotherapy, particularly in tumors that are routinely

treated by radiotherapy. These trials are based on the premise that mitomycin C may produce a synergistic effect with radiation by acting as a radiosensitizer and by targeting hypoxic cells in the tumor which are resistant to radiation (26,72). For example, two studies using mitomycin C as an adjunct to radiation therapy in patients with squamous cell carcinoma of the head and neck demonstrated a significant improvement in the local regional relapse and recurrence-free survival for patients receiving mitomycin C treatment (73,74). In addition, a study of concomitant radiotherapy with mitomycin C and bleomycin in inoperable head and neck cancer showed that the chemotherapy significantly increased the complete remission rate and disease-free survival in all the patients studied (75). The combination treatment had an even greater effect on the complete remission rate and disease free-survival in patients with oropharvngeal carcinoma and also increased the overall survival in this sub-group of patients from 10% to 38% (75).

Similarly, radiotherapy is standard treatment for women with locally advanced cervical cancer, but this treatment fails to control disease progression within the irradiated field in 40% of women (76). Thus, there has been interest in combining radiation with chemotherapeutic agents that are radiosensitizers or that target hypoxic cells. A number of studies have examined both concomitant and neoadjuvant chemotherapy with radiotherapy in the management of locally advanced (76,77) and advanced (78) cervical carcinoma, using drug combinations of mitomycin C with 5-FU and/or cisplatin. A number of studies have also demonstrated excellent response rates and improved survival using combination therapy with radiation and mitomycin C, cisplatin and vinca alkaloids in stage III nonsmall-cell lung cancer (79,80).

Alternatively, mitomycin С containing chemotherapy regimens have been studied for induction therapy prior to surgery in a number of different types of cancer. For example, various combinations of mitomycin C, 5-FU, vincristine and bleomycin have been used to shrink locally advanced cervical cancer prior to radical hysterectomy (81,82). Preoperative chemotherapy with mitomycin C, vinca alkaloids and cisplatin in advanced non-small-cell lung cancer produced high response rates, high complete resection rates after response to chemotherapy and increased survival rates (83-85). A recent study also demonstrated the feasibility of preoperative chemotherapy with mitomycin C, vinblastine and cisplatin in early stage resectable non-small-cell lung cancer and a phase III trial has now been initiated (86).

Combination chemotherapy regimens containing mitomycin C have also been extensively studied for treatment of a variety of resistant and recurrent cancers. In a small trial, mitomycin C with 5-FU and folinic acid produced a rapid improvement in performance status and a reduction of analgesics in approximately 80% of patients with advanced breast cancer that had failed first- and second-line chemotherapy (87). Mitomycin C with methotrexate, vincristine and a steroid produced a 30% response rate in women with advanced breast cancer who were resistant to doxorubicin or who had relapsed after doxorubicin treatment (88). Mitomycin C with cisplatin and/or doxorubicin also produced approximately 20% response rates in malignant mesothelioma, a disease which has a very poor prognosis (89,90). A 21% response rate was seen with bleomycin, mitomycin C and cisplatin in advanced squamous carcinoma of the uterine cervix (91) and mitomycin C, ifosfamide and cisplatin produced a 56% response rate in inoperable non-small cell lung cancer (92).

3.3.3. Toxicity

Although mitomycin C is an active anticancer drug, its use in the clinic has been severely restricted by its toxicity. Usual dose-limiting toxicity is myelosuppression, which occurs three to four weeks after drug administration, with recovery within eight weeks (43). This toxicity is dose related, increasing with doses >10 to 20 mg/m², but effects are cumulative and can be unpredictable. Generally, thrombocytopenia or leukopenia are most severe, but anemia is also common. The appearance of thrombocytopenia after mitomycin C treatment is frequently delayed. Other common toxicities include anorexia, nausea, vomiting and diarrhea (43).

There are a number of other less common, but potentially serious or fatal, toxicities associated with the use of mitomvcin C. Approximately 5% of patients receiving mitomycin C develop severe pulmonary toxicity including noncardiogenic pulmonary edema, interstitial pneumonitis and pleural effusions (93) leading to progressive respiratory insufficiency and death (43). Corticosteroid therapy results in improvement in the pulmonary symptoms in less severe cases (43) and a temporary improvement in severe cases (93). This toxicity does not appear to be dose-related, but the risk may increase with exposure to high oxygen concentrations or when mitomycin C is combined with other anticancer drugs like bleomycin which can also cause pulmonary toxicity (43). Between 4% and 15% of patients treated with mitomycin C develop a syndrome known by a number of different names including cancer-associated hemolytic-uremic syndrome (C-HUS) (43,94). This syndrome comprises a complex of microangiopathic hemo;ytic anemia, thrombocytopenia and renal failure that may lead to death. The syndrome is most common in patients that receive six to twelve months of mitomycin therapy or total doses >60mg (43,94). In addition, mitomycin C extravasation during administration can cause painful skin ulcerations and some cases of severe congestive heart failure have been reported after mitomycin C treatment in patients previously treated with doxorubicin (43).

3.3.4. Dosage and Drug Delivery

Mitomycin C is normally administered *i.v.* at a single dose of 20 mg/m² or at 2 mg/m²/day over 12 days; however, other forms of drug delivery are used or have been studied for specific tumors (43,95). Treatment is repeated every 6 to 8 weeks provided the leucocyte and platelet counts have recovered sufficiently. This dose may be reduced if mitomycin C is combined with other myelosuppressive agents. The median terminal half-lives of the drug in single-agent and combination chemotherapy is approximately 50 and 42 minutes, respectively (43,96). Metabolism of mitomycin C is greatest in the liver, spleen

and kidney (43) with hepatic metabolism being the most important route of elimination (43,96). In addition, intravesical administration of mitomycin C at doses of 20 to 40 mg is routinely used for the treatment of superficial bladder cancer (47,56).

A number of other methods for more direct delivery of mitomycin C to tumors have been investigated in order to reduce the systemic toxicity of this agent. For example, hepatic intraarterial infusion has been used for liver metastases from breast cancer (97) or gastrointestinal cancers (98).

A variation of this approach has involved hepatic intraarterial administration of mitomycin C and other chemotherapeutic agents combined with cutting off the blood supply to hepatic tumors by embolization (99). A variety of methods have been used to accomplish the vascular occlusion including the use of lipid particles or collagen. This procedure may prolong the transit time of the drugs through the tumor and may also produce ischemic damage. Alternatively, adjuvant intraportal administration of mitomycin C and 5-FU during surgery and during the early postoperative period appeared to reduced the incidence of liver metastasis and increased survival in patients with colorectal cancer (100). Other approaches have included bronchial artery infusion of mitomycin C in patients with non-small cell lung cancer (101) and direct intratumoral injection of mitomycin C adsorbed to activated carbon particles into pancreatic tumors (102). In addition, the combined use of mitomycin C and hyperthermia has also been investigated (103).

3.4. Porfiromycin 3.4.1. Preclinical Studies

Porfiromvcin was first isolated from Streptomyces ardus in 1960 (104) and has been shown to have activity against many experimental tumors (105-107). A number of significant differences have been observed between the activity of this agent and mitomycin C. Porfiromycin is less toxic to tumor cells than mitomycin C under aerobic conditions (105-107), but shows similar (105) or greater activity (106) under hypoxic conditions. Thus, porfiromycin has a greater hypoxic:oxic ratio and has greater preferential toxicity to hypoxic cells than mitomycin C. This is likely due to poorer activation of porfiromycin by DT-diaphorase and a greater dependence of this agent on activation by NADPH:cytochrome P450 reductase compared with mitomycin C (108). Other observed differences include a greater specificity for targeting hypoxic regions of tumors (109) and differences in the spectra of toxic lesions produced by porfiromycin and mitomycin C under aerobic and hypoxic conditions (110).

Based on the greater hypoxic:oxic ratio of porfiromycin compared with mitomycin C, porfiromycin was studied as an adjuvant to radiotherapy. Treatment of EMT6 murine breast tumors *in vivo* with porfiromycin and radiation produced synergistic tumor cell kill but only additive cytotoxicity to marrow CFU-GM (72). In another study, it was shown that EMT6 tumors implanted into old mice had a higher proportion of radioresistant hypoxic cells than tumors implanted in young mice; however, combining porfiromycin with X-rays overcame the radioresistance of EMT6 tumors in the older mice (111).

3.4.2. Clinical Studies

Phase I clinical trials investigated the administration of porfiromycin using multiple doses (112,113) or large intermittent doses (112,114). The major toxicity observed was hematological toxicity, mainly leukopenia and thrombocytopenia (112-114), similar to that observed with mitomycin C (43). These trials also produced objective responses in patients with carcinoma of the cervix, ovary, stomach, head and neck, and colon (114). A number of phase II clinical trials with porfiromycin demonstrated that this agent was useful in disseminated squamous cell carcinoma of the cervix and also showed activity in carcinomas of the lung and head and neck (115). A study comparing the effectiveness of porfiromycin and mitomycin C found that these agents produced comparable results in patients with colorectal carcinoma, gastrointestinal cancer and ovarian cancer (116).

Based on the laboratory studies combining porfiromycin with radiation, porfiromycin in combination with radiation therapy for squamous cell carcinoma of the head and neck was studied in a phase I clinical trial (117). Patients with locally advanced disease and a low probability of cure were treated with standard fractionated daily radiation, and porfiromycin was administered on days 5 and 47 of the course of radiation therapy. This treatment regimen resulted in acceptable acute hematological and nonhematological toxicities, and produced a 5 year disease free survival rate of 32%. Based on these results a phase III trial comparing porforomycin and radiation therapy with radiation therapy alone in squamous cell carcinoma of the head and neck has been initiated.

3.5. Other Mitomycin Analogues 3.5.1. Preclinical Studies

While both mitomycin C and porfiromycin have very good antitumor activity in a variety of solid tumors, their clinical use has been limited by cumulative myelosuppression and C-HUS. Thus, there has been considerable interest in developing mitomycin analogues with decreased toxicity and increased antitumor activity. Most studies have focussed on substitutions at the C7 amino group of mitomycin C or porfiromycin (118). Although many mitomycin analogues have been prepared and have been tested for antitumor activity, only a small number of these compounds have been targeted for further development.

The 7-*N*-(dimethylamino methylene) analogue of mitomycin C, BMY-25282, was more potent in a number of different mouse tumors *in vitro* and *in vivo*, showed greater activity under aerobic conditions and generated more oxygen radicals than the parent compound (119,120). This agent had a lower quinone-containing reduction potential than mitomycin C (121), and was active in human colon carcinoma cells that were resistant to the parent drug because of deficient activation of mitomycin C (122). However, BMY-25282 showed greater toxicity to neonatal rat-heart myocytes than mitomycin C (123) and produced

delayed cardiac toxicity in rats *in vivo* (124). Thus, this agent was not tested in humans.

M83, a 7-*N*-(4-hydroxyphenyl) analogue of mitomycin C, showed greater potency than the parent drug against a number of rodent leukemia, lymphoma and solid tumors *in vivo* (125). This agent also produced lower myelosuppression and leukopenia than mitomycin C and had a markedly increased therapeutic index in these rodent models (125). In contrast, M83 showed similar activitity to mitomycin C in human tumor xenografts (126). However, based on the lower toxicity in the rodent model, M83 was tested in a phase I-II clinical study (127).

The 7-*N*-(2-(4-nitrophenyldithio)ethyl) analogue of mitomycin C, BMS-181174 (formerly known as BMY-25067), was shown to have superior activity against solid tumors in mice compared to mitomycin C (128). In addition, this agent was less neutropenic and thrombocytopenic than mitomycin C (128), and produced only minor renal changes with no cardiac or pulmonary toxicity in animals (124). BMS-181174 was more potent under aerobic conditions than under hypoxia and may produce its cytotoxic effects by a different mechanism than the parent compound (129,130). Based on these considerations BMS-181174 was evaluated in a phase I trial (131).

KW2149, 7-*N*-(2-((2-(gamma-L-glutamylamino) ethyl)dithio)ethyl)mitomycin C (also known as KT6149) is a water soluble mitomycin analogue (132) that showed enhanced antitumor activity against human tumors *in vitro* (133) and in nude mice (134). The mechanism of antitumor activity of this analogue may be similar to that of BMS-181174 (130). KW2149 has not yet been tested in the clinic.

3.5.2. Clinical Studies

To date only M83 and BMS-181174 have been studied in the clinic. A phase I-II study of M83 in 22 patients found that this analogue produced hematological and non-hematological toxicities that were very similar to those seen with mitomycin C and only one objective response was observed (127). Based on this initial study it was suggested that this agent was not superior to the parent drug and no further clinical studies were carried out. A phase I trial with BMS-181174 found that myelosuppression, particularly thrombocytopenia, was the dose limiting toxicity (131). Other toxicities included thrombophlebitis, pneumonitis and possible cardiotoxicity and renal damage. Although this agent showed antitumor activity in previously treated and untreated patients, no further clinical studies are planned.

4. CLINICAL APPLICATIONS OF BENZOQUINONE-CONTAINING ALKYLATING AGENTS

4.1. History

Benzoquinone-containing alkylating agents were amongst the first quinone-containing alkylating



	•	
	<u>R</u> 1	$\underline{\mathbf{R}}_2$
Diaziquone (AZQ)	NHCO ₂ CH ₂ CH ₃	NHCO ₂ CH ₂ CH ₃
Carbazilquinone	CH ₃	CH(OCH ₃)CH ₂ OCONH ₂
Triaziquone	N	Н
BZQ	NHCH ₂ CH ₂ OH	NHCH ₂ CH ₂ OH
RH1	CH ₃	CH ₂ OH

Figure 2. Structures of benzoquinone-containing alkylating agents

agents studied as possible anticancer drugs. Putter (7) suggested that adding alkylating groups to the benzoquinone structure would produce compounds with potent antitumor activity

Holzer et al (1) provided support for this suggestion by demonstrating that a number of aziridinyl benzoquinones had very good antitumor activity. This lead to the synthesis and testing of a large number of benzoquinone analogues having differing alkylating groups (14-16). The majority of these analogues had either an aziridine (1,14-16) or nitrogen mustard (14) alkylating group; however, a variety of other alkylating groups were also investigated (14,15,20). While some of these agents showed good activity in tumor cells or animal models many of them also produced major toxicities, particularly hematological toxicity (14). To date only four benzoquinone-containing alkylating agents, diaziquone (AZQ), carbazilquinone, triaziquone and BZQ, have been used in the clinic, although other agents such as RH1 (135) have received considerable attention recently (Figure 2).

4.2. Diaziquone

4.2.1. Preclinical Studies

Diaziquone was first tested as a potential central nervous system antitumor agent in the early 1960's and was shown to have significant activity in both intracerebral and intraperitoneal mouse leukemia models, as well as activity in solid tumors (136,137). The mechanism of action of diaziquone is still not fully understood but appears to result from DNA alkylation, crosslinking and strand break formation (138). Studies by Gutierrez et al (139) have shown that this agent is reduced by NADPH:cytochrome P450 reductase to the semiguinone species which generates reactive oxygen species by redox cycling in the presence of oxygen. However, diaziquone has also been shown to be a substrate for DT-diaphorase (140,141), and reduction of the quinone by this enzyme resulted in increased DNA crosslinking, oxygen radical formation and cytotoxicity (140-142). It is still not clear which of the two activation pathways and which types of DNA damage are most important for the antitumor activity of this agent (140,141).

4.2.2. Clinical Studies

Phase I studies investigated the use of diaziquone by daily (143) or weekly (144) *i.v.* injection, by monthly

intraarterial infusion (145) or by twice-weekly or weekly intrathecal injection (146). The dose limiting toxicities were generally leukopenia and thrombocytopenia (143-145), while gastrointestinal toxicity (144,145) and headache (146) were also common. Diaziquone was rapidly removed from plasma by a two-compartment opensystem model (143,144), and the agent was found at appreciable levels in the cerebrospinal fluid (146,147). A number of complete and partial responses were seen in patients with meningeal leukemia (146) and malignant astrocytomas (145).

Based on preclinical studies and phase I clinical trials, diaziquone has been studied in the treatment of CNS tumors. A number of clinical trials showed response rates of approximately 20% in patients with high-grade or progressive gliomas treated with diaziquone (148,149) and approximately 25% in patients with astrocytic neoplasms treated with diaziquone and either BCNU or procarbazine (150). However, phase III trials comparing diaziquone to the nitrosoureas, BCNU or PCNU, found that diaziquone was not significantly better than BCNU (151) and was less effective than PCNU (152) in patients with brain tumors.

The activity of diaziquone in a wide variety of leukemias, lymphomas and solid tumors has been extensively studied. This agent produced responses in 30% of patients with relapsed acute nonlymphocytic leukemia (153,154) and in 20% of patients with refractory lymphoma (155,156) or acute myeloid leukemia (157). In contrast, the drug was not active in patients with resistant multiple myeloma (158,159). Diaziquone produced only minor responses in patients with head and neck (160), ovarian (161), lung (162,163), colorectal (164,165), breast (166), cervical (167), uterine (168), gastric (169), pancreatic (170) and renal (171) cancer, and was inactive in soft tissue and bony sarcomas (172) and melanomas (173). Diaziquone has also been extensively evaluated in pediatric tumors. It produced a 10% response rate in recurrent pediatric brain tumors (174,175), but had little effect in other solid tumors (174) or acute leukemias (176). In addition, because diaziquone shows little non-hematopoietic toxicity, it has been investigated for inclusion into bone marrow transplant preparative regimens (177).

Despite showing good activity in recurrent or resistant brain tumors, nonlymphocytic leukemias and lymphomas, diaziquone is not currently in general use. This is likely due to its toxicity and because it did not demonstrate clearly superior effectiveness to existing or newer chemotherapeutic agents.

4.3 Carbazilquinone

4.3.1 Preclinical Studies

Carbazilquinone was first synthesized in 1970 as a mitomycin C analogue having a quinone group and aziridine and carbamate alkylating groups (178). This agent showed good activity in mouse tumors in vivo including plasmacytoma, lymphoid leukemia and Ehrlich carcinoma, and was more effective than mitomycin C in the latter two tumors (179). It also showed good antitumor activity in human primary tumor specimens of urothelial transitional-cell carcinoma (180), lung cancer (181), hepatocellular carcinoma (182), squamous cell carcinoma (183) and gastric cancer (184) in vitro, and in human ovarian (185) and pancreatic (186) tumor xenografts in nude mice. The mechanism of action of carbazilquinone is not completely understood, but it is thought that this agent acts as an alkylating agent (187). The drug is a substrate for both NADH (188) and NADPH-dependent (189) reductive enzymes; generates reactive oxygen species (189); produces DNA strand breaks (190), and is more active under hypoxic conditions (191). Thus. carbazilquinone is likely a bioreductive agent and production of DNA strand breaks through the generation of reactive oxygen species may also contribute to its antitumor activity.

4.3.2 Clinical Studies

Carbazilquinone has been extensively studied in the clinic, primarily in Japan, since the 1970s. Initial studies suggested that this agent had activity as a single agent in gastric, ovarian and hematological cancers (187). Later studies found response rates of from 6% to 30% in lung tumors (192-194), 60% in gastric tumors (195,196) and 40% in ovarian tumors (197)when the agent was combined with other antitumor agents or immunotherapy. Carbazilquinone also produced a 60% response rate when used with doxorubicin and bleomycin in patients with superficial bladder cancer (198), but lower response rates were seen with doxorubicin, mitomycin C, cisplatin or 5-FU in prostate (199) or liver cancer (200). However, this agent appeared to be highly effective in patients with hematological disorders including chronic myelogenous leukemia, lymphomas, multiple myeloma, polycythemia vera and essential thrombocythemia (201-203).

Initial studies indicated that carbazilquinone produced the usual toxicities associated with mitomycin C analogues. Leukopenia, thrombocytopenia and anorexia were the major toxic effects of the drug, but there was no evidence of toxicity to the liver or kidneys (187). However, more recent studies suggest that this agent may promote transformation of some myeloproliferative disorders to acute leukemias (203). In a controlled study 17% of patients with polycythemia vera and 31% of patients with essential thrombocythemia that were treated with carbazilquinone had transformations to acute leukemia and there appeared to be a relationship between the dose of drug received and transformation (203). Thus, there has been little use of this agent in North America or Europe.

4.4 Other Benzoquinone-containing Alkylating Agents 4.4.1 Preclinical Studies

Triaziquone was first synthesized in 1958 (204) and was shown to have activity in a variety of different animal tumors *in vitro* and *in vivo* and in human tumors including Jensen sarcoma (205), Ehrlich mouse carcinomas and GW77 human colon carcinoms (206) and rat Yoshida sarcoma (207). This agent presumably produces its antitumor effects by alkylation of cellular components (208), and has been shown to inhibit DNA and RNA synthesis (209,210). Triaziquone may block synthesis of deoxynucleotide bases (209) and interacts with the plasma membrane (211). More recent studies suggest that this agent is a substrate for both one and two-electron reducing agents and that the cytotoxic activity may result from protein alkylation and oxidative stress (212,213).

BZQ was first synthesized in 1976 (137) and showed good activity against intraperitoneal mouse leukemias and melanoma (214). This agent had only moderate activity in an intracerebral mouse model (214), but showed good activity in human colon carcinoma (215) and human leukemia cells (216) *in vitro*. Mechanistic studies suggested that BZQ was not a substrate for DTdiaphorase (215) and did not produce DNA strand breaks (217). However, it produced DNA crosslinking without reduction and this was enhanced under acidic conditions (218). Thus, the cytotoxicity of this agent appeared to be due to its alkylating activity, and the drug probably does not act as a bioreductive agent (219).

4.4.2 Clinical Studies and Use

Triaziquone was used clinically in the 1960s for the treatment of a number of cancers (220). It was used intravenously in the treatment of leukemias and lymphomas like chronic lymphocytic leukemia and Hodgkin's disease, and for ovarian cancer. The agent was also used as an ointment for basal cell skin cancers. Because of its toxicity to bone marrow and blood vessel walls it has been replaced by more effective agents and has not been used clinically for many years. A more recent study of this agent as an adjuvant to surgery in carcinoma of the cervix found that there was no difference in 5 year survival between the triaziquone treated patients and patients receiving conventional therapy (221).

BZQ has received only minimal investigation in humans. A pharmacokinetic study showed that, like diaziquone, this agent was rapidly removed from the plasma but had much lower plasma protein binding than diaziquone (222).

5. CLINICAL APPLICATIONS OF INDOLOQUINONE-CONTAINING ALKYLATING AGENTS

5.1. History

Following the isolation of mitomycin C and its identification as a potent antibiotic and antitumor agent,



Figure 3. Structure of EO9

there was considerable interest in developing simpler synthetic analogues. Because the mitomycin structure contains the indoloquinone ring system, indoloquinone analogues were synthesized and tested for their biological activity (223). While many of these compounds demonstrated antibiotic activity, most did not show good antitumor activity (223). There was renewed interest in the indoloquinones as antitumor agents in the 1980s with new analogues being prepared (224,225) and tested for biological activity. This interest has continued to the present time with many structure-activity studies aimed at identifying new bioreductive agents being reported (226-229). However, to date only one indoloquinone, EO9, has been tested in the clinic (Figure 3).

5.2. EO9

5.2.1. Preclinical Studies

EO9 was synthesized in 1987 (225) and its antitumor activity has been extensively studied in animal and human tumors (230). EO9 showed very good antitumor activity in many human tumor cell lines *in vitro* and displayed preferential activity in cell lines derived from solid tumors in the NCI human tumor screen, particularly colon, melanoma, central nervous system, renal and nonsmall cell lung tumors (230,231). This agent was also very potent in a number of *in vivo* murine tumors and human tumor xenografts, but showed little activity in leukemias (231). In addition, EO9 produced no significant bone marrow toxicity in mice. However, preclinical studies in animals found that the drug was rapidly eliminated (232).

The mechanism of action of EO9 has also been extensively studied. The agent is reduced by both one and two-electron reducing enzymes (233-235) and produces DNA strand breaks (234) and crosslinks (233,235) following reduction. The two-electron reducing enzyme, DT-diaphorase, appears to be the most important activating enzyme for EO9 under aerobic conditions, and the sensitivity of cells to this agent correlated with the level of the enzyme under these conditions (236,237). EO9 showed enhanced activity under hypoxic conditions; however, DTdiaphorase appeared to decrease the activity of the agent under these conditions (238).

5.2.2. Clinical Studies

Based on the selective activity of EO9 toward solid tumors, its preferential toxicity to hypoxic cells and its low marrow toxicity, this agent was entered into clinical trials. Two phase I studies found that the drug was rapidly eliminated from the plasma following intravenous injection, but did not produce any bone marrow suppression (239, 240). The dose-limiting toxicity was proteinurea (239). Phase II clinical trials with EO9 confirmed the renal toxicity and lack of myelosuppression with the drug; however, the agent showed no antitumor activity in breast, gastric, pancreatic, colorectal or nonsmall cell lung cancer (241, 242). This failure has primarily been attributed to the rapid clearance of the agent from the body.

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7. REFERENCES

1. Holzer, H.: Zum mechanismus der glykolysehemmung durch carcinostatisch wirkende äthylenimin verbindungen. *Biochem Z* 330, 59-72 (1958)

2. W. B. Pratt, R. W. Ruddon, W. D. Ensminger & J. Maybaum: The Anticancer Drugs. Oxford University Press, New York 17-18 (1994)

3. Gilman, A. & F. S. Philips: The biological actions and therapeutic applications of ∃-chloroethyl amines and sulfides. *Science* 103, 409- 415 (1946)

4. W. B. Pratt, R. W. Ruddon, W. D. Ensminger & J. Maybaum: The Anticancer Drugs. Oxford University Press, New York 119-122 (1994)

5. Galton, D. A. G.: The use of myleran and similar agents in chronic leukemias. *Advances Cancer Res* 4, 73-112 (1956)

6. Sykes, M., D. Karnofsky, F. Phillips & J. H. Buchenal: Clinical studies of triethylene-phosphoramide and diethylene-phosphoramide compounds with nitrogen mustard-like activity. *Cancer (Phila)* 6, 142-148 (1953)

7. Pütter, J.: Stoff wechselwirkungen von äthyleniminochinonen. *Arzneimittel-Forsch* 10, 8-12 (1960)

8. Hata, T., Y. Sano, R. Sugawara, A. Matsumae, K. Kanamori, T. Shima & T. Hoshi: Mitomycin, a new antibiotic from *Streptomyces*. *J Antibiot* 9, 141-146 (1956)

9. Wakaki, S., H. Marumo, K. Tomioka, G. Shimizu, E. Kato, H. Kamada, S. Kudo & Y. Fujimoto: Isolation of new fractions of antitumor mitomycins. *Antibiot Chemother* 8, 228-240 (1958)

10. Sugiura, K.: Antitumor activity of mitomycin C. *Cancer Chemother Rep* 13, 51-65 (1961)

11. Sokoloff, B., K. Nakabayashi, K. Enomoto, T. R. Miller, A. Bicknell, L. Bird, W. Trauner, J. Niswonger & G. Renninger: Experimental studies on mitomycin C. *Growth* 23, 109-136 (1959)

12. Sukie, K., T. Takeishi & T. Noguchi: Clinical trials of mitomycin C (antitumor substance). *Chemotherapy* 5, 223-224 (1957)

13. Taguchi, T., S. Shiba, I. Ito, M. Matsui, T. Fujii, M. Yamamoto, S. Sawada, Y. Omukai, E. Yamamoto, K. Horino, A. Terawaki, M. Ueda, T. Uenishi & H. Kontani: Clinical experience with mitomycin C. *Gann* (Suppl) 49, 16-18 (1959)

14. Hayashi, S., H. Ueki & Y. Ueki: Studies on antitumor substances I. Antitumor effect of quinone derivatives containing an alkylating group. *Gann* 54, 381-390 (1963)

15. Nakao, H., M. Arakawa, T. Nakamura & M. Fukushima: Antileukemic agents. II. New 2,5-bis(1-aziridinyl)-*p*-benzoquinone derivatives. *Chem Pharm Bull* (*Tokyo*) 20, 1968-1979 (1972)

16. Yoshimoto, M., H. Miyazawa, H. Nakao, K. Shinkai & M. Arakawa: Quantitative structure-activity relationships in 2,5-bis(1-aziridinyl)-*p*-benzoquinone derivatives against. leukemia L-1210. *J Med Chem* 22, 491-496 (1979)

17. Schwartz, H. S.: Pharmacology of mitomycin C: III. In vitro metabolism by rat liver. *J Pharmacol Exp Ther* 136, 250-258 (1962)

18. Iyer, V. N. & W. Szybalski: Mitomycin and porfiromycin: chemical mechanism of activation and cross-linking of DNA. *Science* 145, 55-58 (1964)

19. W. C. J.Ross: Biological Alkylating Agents. Butterworths, London 170-172 (1962)

20. Lin, A. J., L. A. Cosby & A. C. Sartorelli: Quinones as anticancer agents: potential bioreductive alkylating agents. *Cancer Chemotherapy Rep* Part 24, 23-25 (1974)

21. Riley, R. J. & P. Workman: DT-diaphorase and cancer chemotherapy. *Biochem Pharmacol* 43, 1657-1669 (1992)

22. Tomasz, M., R. Lipman, D. Chowdary, L. Pawlak, G. L. Verdine & K. Nakanishi: Isolation and structure of a covalent cross-link adduct between mitomycin C and DNA. *Science* 235, 1204-1208 (1987)

23. Lown, J. W., A. Begleiter, D. Johnson & A. R. Morgan: Studies related to antitumor antibiotics. PartV. Reaction of mitomycin C with DNA examined by ethidium fluorescence assay. *Can J Biochem* 54, 110-119 (1976)

24. Sun, X. M. & D. Ross: Quinone-induced apoptosis in human colon adenocarcinoma cells via DT-diaphorase mediated bioactivation. *Chem Biol Interact* 100, 267-276 (1996)

25. Tomasz, M. & Y. Palom: The mitomycin bioreductive antitumor agents: Cross-linking and alkylation of DNA as the molecular basis of their activity. *Pharmacol Ther* 76, 73-87 (1997)

26. Workman, P. & I. J. Stratford: The experimental development of bioreductive drugs and their role in cancer therapy. *Cancer Metastasis Rev* 12, 73-82 (1993)

27. Pan, S. S., P. A. Andrews, C. J. Glover & N. R. Bachur: Reductive activation of mitomycin C and mitomycin C metabolites catalyzed by NADPH-cytochrome P-450 reductase and xanthine oxidase. *J Biol Chem* 259, 959-966 (1984)

28. Bailey, S. M., M. D. Wyatt, F. Friedlos, J. A. Hartley, R. J. Knox, A. D. Lewis & P. Workman: Involvement of DT-diaphorase (EC 1.6.99.2) in the DNA cross-linking and sequence selectivity of the bioreductive anti-tumour agent EO9. *Br J Cancer* 76, 1596-1603 (1997)

29. Keyes, S. R., P. M. Fracasso, D. C. Heimbrook, S. Rockwell, S. G. Sligar & A. C. Sartorelli: Role of NADPH:cytochrome c reductase and DT-diaphorase in the biotransformation of mitomycin C1. *Cancer Res* 44, 5638-5643 (1984)

30. Hoban, P. R., M. I. Walton, C. N. Robson, J. Godden, I. J. Stratford, P. Workman, A. L. Harris & I. D. Hickson: Decreased NADPH:cytochrome P-450 reductase activity and impaired drug activation in a mammalian cell line resistant to mitomycin C under aerobic but not hypoxic conditions. *Cancer Res* 50, 4692-4697 (1990)

31. Ross, D., D. Siegel, H. Beall, A. S. Prakash, R. T. Mulcahy & N. W. Gibson: DT-diaphorase in activation and detoxification of quinones. *Cancer Metastasis Rev* 12, 83-101 (1993)

32. Begleiter, A., E. Robotham, G. Lacey & M. K. Leith: Increased sensitivity of quinone resistant cells to mitomycin C. *Cancer Lett* 45, 173-176 (1989)

33. Begleiter, A., E. Robotham & M. K. Leith: Role of NAD(P)H:(quinone acceptor) oxidoreductase (DT-diaphorase) in activation of mitomycin C under hypoxia. *Mol Pharmacol* 41, 677-682 (1992)

34. Begleiter, A. & M. K. Leith: Role of NAD(P)H:(quinone acceptor) oxidoreductase (DT-diaphorase) in activation of mitomycin C under acidic conditions. *Mol Pharmacol* 44, 210-215 (1993)

35. Begleiter, A., M. K. Leith & S. S. Pan: Mechanisms for the modulation of alkylating activity by the quinone group in quinone alkylating agents. *Mol Pharmacol* 40, 454-458 (1991)

36. Begleiter, A. & M. K. Leith: Activity of quinone alkylating agents in quinone-resistant cells. *Cancer Res* 50, 2872-2876 (1990)

37. Doroshow, J. H.: Role of hydrogen peroxide and hydroxyl radical formation in the killing of Ehrlich tumor cells by anticancer quinones. *Proc Natl Acad Sci (USA)* 83, 4514-4518 (1986)

38. Begleiter, A.: Studies on the mechanism of action of quinone antitumor agents. *Biochem Pharmacol* 34, 2629-2636 (1985)

39. Rockwell, S., A. C. Sartorelli, M. Tomasz & K. A. Kennedy: Cellular pharmacology of quinone bioreductive

alkylating agents. *Cancer Metastasis Rev* 12, 165-176 (1993)

40. Begleiter, A.: The contribution of alkylation to the activity of quinone antitumor agents. *Can J Physiol Pharmacol* 64, 581-585 (1986)

41. W. Szybalski & V. N. Iyer: Antibiotics I. In: The Mitomycins and Porfiromycins. Eds: Gottlieb, D. & Shaw, P.D., Springer-Verlag, New York 211 (1967)

42. W. A. Remers: Mitomycin C and analog development. In: Mitomycin C Current Status and New Developments. Eds: Carter, S. K. & Crooke, S. T., Academic Press, New York 27-32 (1979)

43. Doll, D. C., R. B. Weiss & B. F. Issell: Mitomycin: ten years after approval for marketing. *J Clin Oncol* 3, 276-286 (1985)

44. Hodnick, W. F. & A. C. Sartorelli: Reductive activation of mitomycin C by NADH:cytochrome b5 reductase. *Cancer Res* 53, 4907-4912 (1993)

45. Gustafson, D. L. & C. A. Pritsos: Bioactivation of mitomycin C by xanthine dehydrogenase from EMT6 mouse mammary carcinoma tumors. *J Natl Cancer Inst* 84, 1180-1185 (1992)

46. Dusre, L., J. M. Covey, C. Collins & B. K. Sinha: DNA damage, cytotoxicity and free radical formation by mitomycin C in human cells. *Chem Biol Interact* 71, 63-78 (1989)

47. Crawford, E. D.: Diagnosis and treatment of superficial bladder cancer: an update. *Semin Urol Oncol* 14, 1-9 (1996)

48. Schnall, S. & J. S. Macdonald: Mitomycin therapy in gastric cancer. *Oncology (Basel)* 50 Suppl 1, 70-77 (1993)

49. Kelsen, D.: The use of chemotherapy in the treatment of advanced gastric and pancreas cancer. *Semin Oncol* 21, 58-66 (1994)

50. Cummings, B. J., T. J. Keane, B. O'Sullivan, C. S. Wong & C. N. Catton: Mitomycin in anal canal carcinoma. *Oncology (Basel)* 50 Suppl 1, 63-69 (1993)

51. Coia, L.: The use of mitomycin in esophageal cancer. *Oncology (Basel)* 50 Suppl 1, 53-62 (1993)

52. Miyamoto, T.: Recent results of using a sequential combination of bleomycin and mitomycin C in the treatment of metastatic cervical cancer. *Recent Results Cancer Res* 70, 211-217 (1980)

53. Miller, T. P., L. J. McMahon & R. B. Livingston: Extensive adenocarcinoma and large cell undifferentiated carcinoma of the lung treated with 5-FU, vincristine, and mitomycin C (FOMi). *Cancer Treat Rep* 64, 1241-1245 (1980) 54. Soloway, M. S. & P. E. Perito: Superficial bladder cancer: diagnosis, surveillance and treatment. *J Cell Biochem* Suppl 16I, 120-127 (1992)

55. Solsona, E., I. Iborra, J. V. Ricos, J. L. Monros, J. Casanova & R. Dumont: Effectiveness of a single immediate mitomycin C instillation in patients with low risk superficial bladder cancer: short and long-term followup. *J Urol* 161, 1120-1123 (1999)

56. Malmstroem, P. U., H. Wijkstroem, C. Lundholm, K. Wester, C. Busch, B. J. Norlen & Swedish Norwegian Bladder Cancer Study Group: 5-year followup of a randomized prospective study comparing mitomycin C and bacillus Calmette-Guerin in patients with superficial bladder carcinoma. *J Urol* 161, 1124-1127 (1999)

57. Hamdy, F. C., K. J. Hastie, R. Kerry & J. L. Williams: Mitomycin-C in superficial bladder cancer. Is long-term maintenance therapy worthwhile after initial treatment? *Br J Urol* 71, 183-186 (1993)

58. Huben, R. P.: Intravesical chemotherapy versus immunotherapy for superficial bladder cancer. *Semin Urol Oncol* 14, 17-22 (1996)

59. Lise, M., D. Nitti, A. Marchet, T. Sahmoud, M. Buyse, N. Duez, M. Fiorentino, S. J. dos, R. Labianca, P. Rougier & et al: Final results of a phase III clinical trial of adjuvant chemotherapy with the modified fluorouracil, doxorubicin, and mitomycin regimen in resectable gastric cancer. *J Clin Oncol* 13, 2757-2763 (1995)

60. Kim, N. K., Y. S. Park, D. S. Heo, C. Suh, S. Y. Kim, K. C. Park, Y. K. Kang, D. B. Shin, H. T. Kim, H. J. Kim & et al: A phase III randomized study of 5-fluorouracil and cisplatin versus 5-fluorouracil, doxorubicin, and mitomycin C versus 5-fluorouracil alone in the treatment of advanced gastric cancer. *Cancer* 71, 3813-3818 (1993)

61. Coia, L., J. Hoffman, R. Scher, J. Weese, L. Solin, L. Weiner, B. Eisenberg, A. Paul & G. Hanks: Preoperative chemoradiation for adenocarcinoma of the pancreas and duodenum. *Int J Radiat Oncol Biol Phys* 30, 161-167 (1994)

62. Hoffman, J. P., S. Lipsitz, T. Pisansky, J. L. Weese, L. Solin & A. B. Benson: Phase II trial of preoperative radiation therapy and chemotherapy for patients with localized, resectable adenocarcinoma of the pancreas: an Eastern Cooperative Oncology Group Study. *J Clin Oncol* 16, 317-323 (1998)

63. Flam, M., M. John, T. F. Pajak, N. Petrelli, R. Myerson, S. Doggett, J. Quivey, M. Rotman, H. Kerman, L. Coia & K. Murray: Role of mitomycin in combination with fluorouracil and radiotherapy, and of salvage chemoradiation in the definitive nonsurgical treatment of epidermoid carcinoma of the anal canal: Results of a phase III randomized intergroup study. *J Clin Oncol* 14, 2527-2539 (1996)

64. Bartelink, H., F. Roelofsen, F. Eschwege, P. Rougier, J. F. Bosset, D. G. Gonzalez, D. Peiffert, G. M. van & M. Pierart: Concomitant radiotherapy and chemotherapy is superior to radiotherapy alone in the treatment of locally advanced anal cancer: results of a phase III randomized trial of the European Organization for Research and Treatment of Cancer Radiotherapy and Gastrointestinal Cooperative Groups. *J Clin Oncol* 15, 2040-2049 (1997)

65. Smith, T. J., L. M. Ryan, H. O. Douglass Jr, D. G. Haller, Y. Dayal, J. Kirkwood, D. C. Tormey, A. J. Schutt, J. Hinson & B. Sischy: Combined chemoradiotherapy vs. radiotherapy alone for early stage squamous cell carcinoma of the esophagus: a study of the Eastern Cooperative Oncology Group. *Int J Radiat Oncol Biol Phys* 42, 269-276 (1998)

66. Ajani, J. A.: Current status of new drugs and multidisciplinary approaches in patients with carcinoma of the esophagus. *Chest* 113, 112S-119S (1998)

67. Spain, R. C.: The case for mitomycin in non-small cell lung cancer. *Oncology (Basel)* 50 Suppl 1, 35-52 (1993)

68. Cartei, G., F. Cartei, A. Cantone, D. Causarano, G. Genco, A. Tobaldin, G. Interlandi & T. Giraldi: Cisplatincyclophosphamide-mitomycin combination chemotherapy with supportive care versus supportive care alone for treatment of metastatic non-small-cell lung cancer. *J Natl Cancer Inst* 85, 794-800 (1993)

69. Hortobagyi, G. N.: Mitomycin: Its evolving role in the treatment of breast cancer. *Oncology (Basel)* 50 Suppl 1, 1-8 (1993)

70. Jodrell, D. I., I. E. Smith, J. L. Mansi, M. C. Pearson, G. Walsh, S. Ashley, H. D. Sinnett & J. A. McKinna: A randomised comparative trial of mitozantrone/methotrexate/mitomycin C (MMM) and cyclophosphamide/methotrexate/5 FU (CMF) in the treatment of advanced breast cancer. *Br J Cancer* 63, 794-798 (1991)

71. Powles, T. J., A. L. Jones, I. R. Judson, J. R. Hardy & S. E. Ashley: A randomised trial comparing combination chemotherapy using mitomycin C, mitozantrone and methotrexate (3M) with vincristine, anthracycline and cyclophosphamide (VAC) in advanced breast cancer. *Br J Cancer* 64, 406-410 (1991)

72. Rockwell, S., S. R. Keyes & A. C. Sartorelli: Preclinical studies of porfiromycin as an adjunct to radiotherapy. *Radiat Res* 116, 100-113 (1988)

73. Weissberg, J. B., Y. H. Son, R. J. Papac, C. Sasaki, D. B. Fischer, R. Lawrence, S. Rockwell, A. C. Sartorelli & J. J. Fischer: Randomized clinical trial of mitomycin c as an adjunct to radiotherapy in head and neck cancer. *Int J Radiat Oncol Biol Phys* 17, 3-9 (1989)

74. Haffty, B. G., Y. H. Son, R. Papac, C. T. Sasaki, J. B. Weissberg, D. Fischer, S. Rockwell, A. C. Sartorelli & J. J.

Fischer: Chemotherapy as an adjunct to radiation in the treatment of squamous cell carcinoma of the head and neck: Results of the Yale mitomycin randomized trials. *J Clin Oncol* 15, 268-276 (1997)

75. Zakotnik, B., L. Smid, M. Budihna, H. Lesnicar, E. Soba, L. Furlan & M. Zargi: Concomitant radiotherapy with mitomycin C and bleomycin compared with radiotherapy alone in inoperable head and neck cancer: Final report. *Int J Radiat Oncol Biol Phys* 41, 1121-1127 (1998)

76. Tattersall, M. H.: Concomitant and neoadjuvant chemotherapy in conjunction with radiotherapy in the management of locally advanced cervical cancer. *J Natl Cancer Inst Monogr* 101-103 (1996)

77. John, M., M. Flam, R. Caplan, M. Rotman, J. Quivey, A. Steinfeld & A. Russell: Final results of a phase II chemoradiation protocol for locally advanced cervical cancer: RTOG 85-15. *Gynecol Oncol* 61, 221-226 (1996)

78. Souhami, L., R. A. Gil, S. E. Allan, P. C. Canary, C. M. Araujo, L. H. Pinto & T. R. Silveira: A randomized trial of chemotherapy followed by pelvic radiation therapy in stage IIIB carcinoma of the cervix. *J Clin Oncol* 9, 970-977 (1991)

79. Kubota, K., K. Furuse, M. Kawahara, N. Kodama, M. Yamamoto, M. Ogawara, S. Negoro, N. Masuda, M. Takada, K. Matsui & et al: Role of radiotherapy in combined modality treatment of locally advanced non-small-cell lung cancer. *J Clin Oncol* 12, 1547-1552 (1994)

80. Furuse, K., K. Kubota, M. Kawahara, N. Kodama, M. Ogawara, M. Akira, S. Nakajima, M. Takada, Y. Kusunoki, S. Negoro & et al: Phase II study of concurrent radiotherapy and chemotherapy for unresectable stage III non-small-cell lung cancer. Southern Osaka Lung Cancer Study Group. *J Clin Oncol* 13, 869-875 (1995)

81. Dottino, P. R., S. C. Plaxe, A. M. Beddoe, C. Johnston & C. J. Cohen: Induction chemotherapy followed by radical surgery in cervical cancer. *Gynecol Oncol* 40, 7-11 (1991)

82. Nguyan, H. N. & S. R. Nordqvist: Chemotherapy of advanced and recurrent cervical carcinoma. *Semin Surg Oncol* 16, 247-250 (1999)

83. Folman, R. S.: Experience with mitomycin in the treatment of non-small cell lung cancer. *Oncology (Basel)* 50 Suppl 1, 24-30 (1993)

84. Martini, N., M. G. Kris, B. J. Flehinger, R. J. Gralla, M. S. Bains, M. E. Burt, R. Heelan, P. M. McCormack, K. M. Pisters, J. R. Rigas & et al: Preoperative chemotherapy for stage IIIa (N2) lung cancer: the Sloan-Kettering experience with 136 patients. *Ann Thorac Surg* 55, 1365-1373 (1993)

85. Pisters, K. M., M. G. Kris, R. J. Gralla, M. B. Zaman, R. T. Heelan & N. Martini: Pathologic complete response in advanced non-small-cell lung cancer following preoperative chemotherapy: implications for the design of future non-small-cell lung cancer combined modality trials. *J Clin Oncol* 11, 1757-1762 (1993)

86. de Boer, R. H., I. E. Smith, U. Pastorini, M. E. R. O'Brien, F. Ramage, S. Ashley & P. Goldstraw: Preoperative chemotherapy in early stage resectable nonsmall-cell lung cancer: a randomized feasibility study justifying a multicentre phase III trial. *Br J Cancer* 79, 1514-1518 (1999)

87. Francini, G., R. Petrioli, A. Aquino & S. Gonnelli: Advanced breast cancer treatment with folinic acid, 5fluorouracil, and mitomycin C. *Cancer Chemother Pharmacol* 32, 359-364 (1993)

88. Tashiro, H. & Y. Nomura: Mitomycin C, methotrexate, and vincristine with medroxyprogesterone acetate or prednisolone for doxorubicin resistant advanced breast cancer - A randomized control study. *Anticancer Res* 15, 2229-2237 (1995)

89. Chahinian, A. P., K. Antman, M. Goutsou, J. M. Corson, Y. Suzuki, C. Modeas, J. E. Herndon, J. Aisner, R. R. Ellison, L. Leone & et al: Randomized phase II trial of cisplatin with mitomycin or doxorubicin for malignant mesothelioma by the Cancer and Leukemia Group B. *J Clin Oncol* 11, 1559-1565 (1993)

90. Pennucci, M. C., A. Ardizzoni, P. Pronzato, M. Fioretti, C. Lanfranco, A. Verna, G. Giorgi, A. Vigani, C. Frola & R. Rosso: Combined cisplatin, doxorubicin, and mitomycin for the treatment of advanced pleural mesothelioma - A phase II FONICAP trial. *Cancer* 79, 1897-1902 (1997)

91. Picozzi, Jr. V. J., B. I. Sikic, R. W. Carlson, M. Koretz & S. C. Ballon: Bleomycin, mitomycin, and cisplatin therapy for advanced squamous carcinoma of the uterine cervix: A phase II study of the Northern California Oncology Group. *Cancer Treat Rep* 69, 903-905 (1985).

92. Cullen, M. H., R. Joshi, A. D. Chetiyawardana & C. M. Woodroffe: Mitomycin, ifosfamide and *cis*-platin in non-small cell lung cancer: Treatment good enough to compare. *Br J Cancer* 58, 359-361 (1998).

93. Castro, M., M. H. Veeder, J. A. Mailliard, H. D. Tazelaar & J. R. Jett: A prospective study of pulmonary function in patients receiving mitomycin. *Chest* 109, 939-944 (1996)

94. Lesesne, J. B., N. Rothschild, B. Erickson, S. Korec, R. Sisk, J. Keller, M. Arbus, P. V. Woolley, L. Chiazze, P. S. Schein & et al: Cancer-associated hemolytic-uremic syndrome: analysis of 85 cases from a national registry. *J Clin Oncol* 7, 781-789 (1989)

95. V. H. Baker: The development of an acute intermittent schedule-mitomycin C. In: Mitomycin C Current Status and New Developments. Eds: Carter, S. K. & Crooke, S. T., Academic Press, New York 77-82 (1979)

96. den Hartigh, J., J. G. McVie, W. J. van Oort & H. M. Pinedo: Pharmacokinetics of mitomycin C in humans. *Cancer Res* 43, 5017-5021 (1983)

97. Arai, Y., Y. Sone, Y. Inaba, Y. Ariyoshi & C. Kido: Hepatic arterial infusion chemotherapy for liver metastases from breast cancer. *Cancer Chemother Pharmacol* 33 Suppl, S142-S144 (1994)

98. Taton, G., G. Ghanem, P. Pandin, J. Goldzarian, C. Thaysse, L. Bertin, B. Ickx, A. Medhi, A. Salari, J. L. Vanherweghem & J. P. Lambilliotte: First results of a clinical pilot study on intraarterial chemotherapy with haemofiltration of locally advanced gastrointestinal cancers. *Acta Chir Belg* 96, 206-210 (1996)

99. Tellez, C., A. B. Benson, M. T. Lyster, M. Talamonti, J. Shaw, M. A. Braun, A. A. Nemcek, Jr. & R. L. Vogelzang: Phase II trial of chemoembolization for the treatment of metastatic colorectal carcinoma to the liver and review of the literature. *Cancer* 82, 1250-1259 (1998)

100. Long-term results of single course of adjuvant intraportal chemotherapy for colorectal cancer. Swiss Group for Clinical Cancer Research (SAKK). *Lancet* 345, 349-353 (1995)

101. Shimizu, E., Y. Nakamura, J. Mukai, K. Tani, T. Yamashita, F. Hojo, Y. Hashimoto & T. Ogura: Pharmacokinetics of bronchial artery infusion of mitomycin in patients with non-small cell lung cancer. *Eur J Cancer* 27, 1046-1048 (1991)

102. Moriai, T., T. T. Makino & K. Ishii: Intratumoral treatment of pancreatic cancer with mitomycin-C adsorbed to activated carbon particles. A clinical trial on 15 cases. *Anticancer Res* 9, 1799-1804 (1989)

103. Jacquet, P., A. D. Stephens, A. M. Averbach, D. Chang, S. E. Ettinghausen, R. R. Dalton, M. A. Steves & P. H. Sugarbaker: Analysis of morbidity and mortality in 60 patients with peritoneal carcinomatosis treated by cytoreductive surgery and heated intraoperative intraperitoneal chemotherapy. *Cancer* 77, 2622-2629 (1996)

104. C. DeBoer, A. Dietz, N. E. Lummis & et al: Porfiromycin, a new antibiotic. I. Discovery and biological activities. Antimicrobial Agents Annual. Plenum Press, New York 17-22 (1960)

105. Keyes, S. R., S. Rockwell & A. C. Sartorelli: Porfiromycin as a bioreductive alkylating agent with selective toxicity to hypoxic EMT6 tumor cells in vivo and in vitro. *Cancer Res* 45, 3642-3645 (1985)

106. Marshall, R. S. & A. M. Rauth: Oxygen and exposure kinetics as factors influencing the cytotoxicity of porfiromycin, a mitomycin C analogue, in Chinese hamster ovary cells. *Cancer Res* 48, 5655-5659 (1988) 107. Pan, S. S., S. A. Akman, G. L. Forrest, C. Hipsher & R. Johnson: The role of NAD(P)H:quinone oxidoreductase in mitomycin C- and porfiromycin-resistant HCT 116 human colon-cancer cells. *Cancer Chemother Pharmacol* 31, 23-31 (1992)

108. Belcourt, M. F., W. F. Hodnick, S. Rockwell & A. C. Sartorelli: Exploring the mechanistic aspects of mitomycin antibiotic bioactivation in Chinese hamster ovary cells overexpressing NADPH:cytochrome C (P-450) reductase and DT-diaphorase. *Adv Enzyme Regul* 38, 111-133 (1998)

109. Keyes, S. R., S. Rockwell, K. A. Kennedy & A. C. Sartorelli: Distribution of porfiromycin in EMT6 solid tumors and normal tissues of BALB/c mice. *J Natl Cancer Inst* 83, 632-637 (1991)

110. Hughes, C. S., C. G. Irvin & S. Rockwell: Effect of deficiencies in DNA repair on the toxicity of mitomycin C and porfiromycin to CHO cells under aerobic and hypoxic conditions. *Cancer Commun* 3, 29-35 (1991)

111. Rockwell, S., C. S. Hughes, S. R. Keyes, A. C. Sartorelli & K. A. Kennedy: Porfiromycin as an adjunct to radiotherapy in young and old mice. *Exp Gerontol* 28, 281-293 (1993)

112. Foley, H. T., B. I. Shnider, G. L. Gold, P. I. Matias, J. Colsky & S. P. Miller: Phase 1 studies of porfiromycin (NSC-56410). *Cancer Chemother Rep* 51, 283-293 (1967)

113. Grage, T. B., A. J. Weiss, W. Wilson & V. Reynolds: Phase I studies of porfiromycin (NSC--56410) in solid tumors. *J Surg Oncol* 7, 415-420 (1975)

114. Izbicki, R., M. Al Sarraf, M. L. Reed, C. B. Vaughn & V. K. Vaitkevicius: Further clinical trials with porfiromycin (NSC-56410) (large intermittent doses). *Cancer Chemother Rep* 56, 615-624 (1972)

115. Panettiere, F. J., R. W. Talley, J. Torres & M. Lane: Porfiromycin in the management of epidermoid and transitional cell cancer: a phase II study. *Cancer Treat Rep* 60, 907-911 (1976)

116. Baker, L. H., R. M. Izbicki & V. K. Vaitkevicius: Phase II study of profiromycin vs mitomycin-C utilizing acute intermittent schedules. *Med Pediatr Oncol* 2, 207-213 (1976)

117. Haffty, B. G., Y. H. Son, L. D. Wilson, R. Papac, D. Fischer, S. Rockwell, A. C. Sartorelli, D. Ross, C. T. Sasaki & J. J. Fischer: Bioreductive alkylating agent porfiromycin in combination with radiation therapy for the management of squamous cell carcinoma of the head and neck. *Radiat Oncol Invest* 5, 235-245 (1997)

118. Stringfellow, D. A. & J. E. Schurig: The search for more active and less toxic mitomycin and etoposide analogs. *Cancer Treat Rev* 14, 291-295 (1987)

119. Bradner, W. T., W. C. Rose, J. E. Schurig, A. P. Florczyk, J. B. Huftalen & J. J. Catino: Antitumor activity

and toxicity in animals of BMY-25282, a new mitomycin derivative. *Cancer Res* 45, 6475-6481 (1985)

120. Pritsos, C. A. & A. C. Sartorelli: Generation of reactive oxygen radicals through bioactivation of mitomycin antibiotics. *Cancer Res* 46, 3528-3532 (1986)

121. Xu, B. H., V. Gupta & S. V. Singh: Mechanism of differential sensitivity of human bladder cancer cells to mitomycin C and its analogue. *Br J Cancer* 69, 242-246 (1994)

122. Chakrabarty, S., Y. J. Danels, B. H. Long, J. K. Willson & M. G. Brattain: Circumvention of deficient activation in mitomycin C-resistant human colonic carcinoma cells by the mitomycin C analogue BMY25282. *Cancer Res* 46, 3456-3458 (1986)

123. Dorr, R. T., N. G. Shipp, J. D. Liddil, B. S. Iyengar, K. R. Kunz & W. A. Remers: Cardiotoxicity of mitomycin A, mitomycin C, and seven N7 analogs in vitro. *Cancer Chemother Pharmacol* 31, 1-5 (1992)

124. Bregman, C. L., R. A. Buroker, W. T. Bradner, R. S. Hirth & H. Madissoo: Cardiac, renal, and pulmonary toxicity of several mitomycin derivatives in rats. *Fundam Appl Toxicol* 13, 46-64 (1989)

125. Imai, R. & M. Morimoto: Comparative antitumor activities of 7-N-(p-hydroxyphenyl)mitomycin C (M-83) and mitomycin C. *J Antibiot Tokyo* 36, 559-565 (1983)

126. Asanuma, F.: Antitumor activity and pharmacokinetics of 7-N-(p-hydroxyphenyl)mitomycin C in human tumor xenografts transplanted into nude mice. *J Antibiot Tokyo* 38, 401-410 (1985)

127. Meguro, S., T. Nagata, K. Yokoyama, T. Chinen, T. Kobayashi & et al: Phase I-II study of 7-N-(P-hydroxyphenyl)-mitomycin C (KW2083, M83). *Gan To Kagaku Ryoho* 11, 1818-1822 (1984)

128. Bradner, W. T., W. C. Rose, J. E. Schurig & A. P. Florczyk: Antitumor activity and toxicity in animals of N-7(2-(4-nitrophenyldithio) ethyl) mitomycin C (BMY-25067). *Invest New Drugs* 8 Suppl 1, S1-S7 (1990)

129. Rockwell, S., B. Kemple & M. Kelley: Cytotoxicity of BMS-181174. Effects of hypoxia, dicoumarol, and repair deficits. *Biochem Pharmacol* 50, 1239-1243 (1995)

130. Wang, S. & H. Kohn: Studies on the mode of action of mitomycin C(7) aminoethylene disulfides (BMS-181174 and KW-2149): reactivity of 7-N-(mercaptoethyl)mitomycin C. *J Med Chem* 42, 788-790 (1999)

131. Macaulay, V. M., K. J. O'Byrne, J. A. Green, P. A. Philip, L. McKinley, F. P. LaCreta, B. Winograd, T. S. Ganesan, A. L. Harris & D. C. Talbot: Phase I study of the mitomycin C analogue BMS-181174. *Br J Cancer* 77, 2020-2027 (1998)

132. Kono, M., Y. Saitoh, M. Kasai, A. Sato, K. Shirahata, M. Morimoto & T. Ashizawa: Synthesis and antitumor activity of a novel water soluble mitomycin analog; 7-N-(2-((2-(gamma-L-

glutamylamino)ethyl)dithio)ethyl)mitomycin C. *Chem Pharm Bull Tokyo* 37, 1128-1130 (1989)

133. Ohe, Y., K. Nakagawa, Y. Fujiwara, Y. Sasaki, K. Minato, M. Bungo, S. Niimi, N. Horichi, M. Fukuda & N. Saijo: In vitro evaluation of the new anticancer agents KT6149, MX-2, SM5887, menogaril, and liblomycin using cisplatin- or adriamycin-resistant human cancer cell lines. *Cancer Res* 49, 4098-4102 (1989)

134. Kubota, T., T. Inada, S. Inoue, M. Kuzuoka, Y. Arisawa, A. Suto, S. Kodaira, K. Ishibiki & O. Abe: Antitumor activity of 7-N-(2-((2-(gamma-L-glutamylamino)ethyl)dithio)ethyl) mitomycin C (KW2149) against human tumor xenografts serially transplanted into nude mice. *Jpn J Clin Oncol* 19, 216-221 (1989)

135. Winski, S. L., R. H. Hargreaves, J. Butler & D. Ross: A new screening system for NAD(P)H:quinone oxidoreductase (NQO1)-directed antitumor quinones: identification of a new aziridinylbenzoquinone, RH1, as a NQO1-directed antitumor agent. *Clin Cancer Res* 4, 3083-3088 (1998)

136. Khan, A. H. & J. S. Driscoll: Potential central nervous system antitumor agents. Aziridinylbenzoquinones. 1. *J Med Chem* 19, 313-317 (1976)

137. Chou, F., A. H. Khan & J. S. Driscoll: Potential central nervous system antitumor agents. Aziridinylbenzoquinones. 2. *J Med Chem* 19, 1302-1308 (1976)

138. King, C. L., W. N. Hittelman & T. L. Loo: Induction of DNA strand breaks and cross-links by 2,5-diaziridinyl-3,6-bis(carboethoxyamino)-1,4-benzoquinone in Chinese hamster ovary cells. *Cancer Res* 44, 5634-5637 (1984)

139. Gutierrez, P. L., R. D. Friedman & N. R. Bachur: Biochemical activation of AZQ (3,6-diaziridinyl-2,5bis(carboethoxyamino)-1,4-benzoquinone) to its free radical species. *Cancer Treat Rep* 66, 339-342 (1982)

140. Siegel, D., N. W. Gibson, P. C. Preusch & D. Ross: Metabolism of diaziquone by NAD(P)H:(quinone acceptor) oxidoreductase (DT-diaphorase): role in diaziquoneinduced DNA damage and cytotoxicity in human colon carcinoma cells. *Cancer Res* 50, 7293-7300 (1990)

141. Fisher, G. R., J. Donis & P. L. Gutierrez: Reductive metabolism of diaziquone (AZQ) in the S9 fraction of MCF-7 cells. II. Enhancement of the alkylating activity of AZQ by NAD(P)H: quinone-acceptor oxidoreductase (DT-diaphorase). *Biochem Pharmacol* 44, 1625-1635 (1992)

142. Fisher, G. R., L. H. Patterson & P. L. Gutierrez: A comparison of free radical formation by quinone antitumour agents in MCF-7 cells and the role of NAD(P)H

(quinone-acceptor) oxidoreductase (DT-diaphorase). Chem Biol Interact 88, 137-153 (1993)

143. Griffin, J. P., R. A. Newman, J. J. McCormack & I. H. Krakoff: Clinical and clinical pharmacologic studies of aziridinylbenzoquinone. *Cancer Treat Rep* 66, 1321-1325 (1982)

144. Schilcher, R. B., J. D. Young, L. P. Leichman, C. D. Haas & L. H. Baker: Phase I evaluation and pharmacokinetics of aziridinylbenzoquinone using a weekly intravenous schedule. *Cancer Res* 43, 3907-3911 (1983)

145. Greenberg, H. S., W. D. Ensminger, P. B. Layton, S. Gebarski, M. Meyer, B. Chaffee, J. F. Bender & L. A. Grillo: Phase I-II evaluation of intra-arterial diaziquone for recurrent malignant astrocytomas. *Cancer Treat Rep* 70, 353-357 (1986)

146. Berg, S. L., F. M. Balis, S. Zimm, R. F. Murphy, J. Holcenberg, J. Sato, G. Reaman, P. Steinherz, A. Gillespie, K. Doherty & et al: Phase I/II trial and pharmacokinetics of intrathecal diaziquone in refractory meningeal malignancies. *J Clin Oncol* 10, 143-148 (1992)

147. Bachur, N. R., J. M. Collins, J. A. Kelley, E. D. Van, R. S. Kaplan & M. Whitacre: Diaziquone, 2,5-diaziridinyl-3,6-biscarboethoxyamino-1,4-benzoquinone, plasma and cerebrospinal fluid kinetics. *Clin Pharmacol Ther* 31, 650-655 (1982)

148. Curt, G. A., J. A. Kelley, C. V. Kufta, B. H. Smith, P. L. Kornblith, R. C. Young & J. M. Collins: Phase II and pharmacokinetic study of aziridinylbenzoquinone (2,5-diaziridinyl-3,6-bis(carboethoxyamino)-1,4-benzoquinone, diaziquone, NSC 182986) in high-grade gliomas. *Cancer Res* 43, 6102-6105 (1983)

149. Haid, M., J. D. Khandekar, M. Christ, C. M. Johnson, S. J. Miller, G. Y. Locker, J. M. Merrill, H. Reisel, A. Hatfield, V. Lanzotti & et al: Aziridinylbenzoquinone in recurrent, progressive glioma of the central nervous system. A Phase II study by the Illinois Cancer Council. *Cancer* 56, 1311-1315 (1985)

150. Schold, Jr., S. C., M. S. Mahaley, Jr., N. A. Vick, H. S. Friedman, P. C. Burger, E. R. DeLong, R. E. Albright, Jr., D. E. Bullard, J. D. Khandekar, J. G. Cairncross & et al: Phase II diaziquone-based chemotherapy trials in patients with anaplastic supratentorial astrocytic neoplasms. *J Clin Oncol* 5, 464-471 (1987)

151. Schold, Jr., S. C., J. E. Herndon, P. C. Burger, E. C. Halperin, N. A. Vick, J. G. Cairncross, D. R. Macdonald, E. J. Dropcho, R. Morawetz, D. D. Bigner & et al: Randomized comparison of diaziquone and carmustine in the treatment of adults with anaplastic glioma. *J Clin Oncol* 11, 77-83 (1993)

152. Malkin, M. G., S. B. Green, D. P. Byar, T. A. Strike, P. C. Burger, F. S. Vogel, D. A. Pistenmaa, M. S. Mahaley, Jr., J. Ransohoff, W. R. Shapiro, J. Mealey, Jr., J. T. Robertson, R. G. Selker & J. C. Van Gilder: Superiority of PCNU over AZQ in the treatment of primary brain tumors: Results of a prospective randomized trial (81-20) by the Brain Tumor Study Group. *J Neuro-Oncol* 22, 55-65 (1994)

153. Lee, E. J., E. D. Van, M. J. Egorin, M. S. Nayar, P. Shulman & C. A. Schiffer: Diaziquone given as a continuous infusion is an active agent for relapsed adult acute nonlymphocytic leukemia. *Blood* 67, 182-187 (1986)

154. Schulman, P., R. Davis, E. Lee, J. Ellerton & H. Staszweski: Phase II study of continuous-infusion diaziquone in relapsed/refractory acute nonlymphocytic leukemia. *Cancer Treat Rep* 71, 755-757 (1987)

155. McLaughlin, P., F. Cabanillas, A. Y. Bedikian & G. P. Bodey: Phase II trial of aziridinylbenzoquinone (AZQ) in patients with refractory lymphoma. *Cancer Treat Rep* 67, 507-508 (1983)

156. Case, Jr., D. C. & D. M. Hayes: Phase II study of aziridinylbenzoquinone in refractory lymphoma. *Cancer Treat Rep* 67, 993-996 (1983)

157. Lyman, G. H., W. R. Vogler & M. Raney: Continuous-infusion diaziquone in acute myeloid leukemia: a Southeastern Cancer Study Group trial. *Cancer Treat Rep* 71, 309-310 (1987)

158. Vinciguerra, V., K. Anderson & O. R. McIntyre: Diaziquone for resistant multiple myeloma. Cancer and Leukemia Group B. *Cancer Treat Rep* 69, 331-332 (1985)

159. Stuckey, W. J., J. Crowley, L. H. Baker, N. R. Larrimer, K. H. Hanson, J. D. Bonnet & L. A. White, Jr.: Phase II trial of diaziquone in patients with refractory and relapsing multiple myeloma: a Southwest Oncology Group Study. *Cancer Treat Rep* 71, 1095-1096 (1987)

160. Muniz, F., G. E. Velez, J. A. Neidhart, J. F. Bender, M. Becker & L. A. Grillo: A phase II trial of diaziquone in patients with head and neck cancer. *Cancer Chemother Pharmacol* 19, 265-268 (1987)

161. Slayton, R. E., J. A. Blessing, G. P. Sutton, H. D. Homesley & R. Park: Phase II clinical trial of diaziquone in the treatment of patients with epithelial ovarian cancer: a Gynecologic Oncology Group Study. *Cancer Treat Rep* 70, 309-310 (1986)

162. Abeloff, M. D., D. M. Finkelstein, A. Y. Chang, F. J. Camacho, R. H. Creech & D. S. Ettinger: Phase II study of aclarubicin and diaziquone in the treatment of advanced small cell bronchogenic carcinoma (EST 4581): an Eastern Cooperative Oncology Group Study. *Cancer Treat Rep* 69, 451-452 (1985)

163. Fuks, J. Z., J. Aisner, E. D. Van, M. Levitt, D. C. Ihde, D. Carney & P. H. Wiernik: Phase II trial of aziridinylbenzoquinone (AZQ) in patients with refractory small cell carcinoma of the lung. *Am J Clin Oncol* 6, 171-173 (1983) 164. Shildt, R., R. L. Stephens, V. P. Subramanian, L. H. Baker, W. S. Fletcher, R. M. O'Bryan & J. D. McCracken: Phase II trial of diaziquone in advanced large bowel carcinoma in previously treated and untreated patients: a Southwest Oncology Group Study. *Cancer Treat Rep* 69, 709-710 (1985)

165. Bedikian, A. Y., J. R. Stroehlein, D. A. Karlin, J. Korinek & G. P. Bodey: Phase II clinical evaluation of AZQ in colorectal cancer. *Am J Clin Oncol* 5, 535-537 (1982)

166. Martino, S., V. Ratanatharathorn, B. A. Samal, A. Singhakowinta & C. Haas: Treatment of metastatic breast cancer with AZQ: a phase II trial. *Cancer Invest* 6, 289-291 (1988)

167. Slayton, R. E., J. A. Blessing, M. Rettenmaier & H. Ball: A phase II clinical trial of diaziquone (AZQ) in the treatment of patients with recurrent adenocarcinoma and adenosquamous carcinoma of the cervix. A Gynecologic Oncology Group study. *Invest New Drugs* 7, 337-340 (1989)

168. Slayton, R. E., J. A. Blessing & P. D. Clarke: A phase II trial of diaziquone (AZQ) in mixed mesodermal sarcomas of the uterus. A Gynecologic Oncology Group study. *Invest New Drugs* 9, 93-94 (1991)

169. Pugh, R. P., T. Fleming, J. T. Guy, J. K. Weick & J. H. Ward: Phase II study of diaziquone in untreated advanced gastric carcinoma. A Southwest Oncology Group Study. *Am J Clin Oncol* 12, 11-13 (1989)

170. Tilchen, E. J., T. Fleming, G. Mills, N. Oishi, J. D. Bonnett, R. B. Natale, G. Harker & J. Coltman-CA: Phase II evaluation of diaziquone in pancreatic carcinoma: a Southwest Oncology Group Study. *Cancer Treat Rep* 71, 1309-1310 (1987)

171. Stephens, R. L., R. Kirby, E. D. Crawford, R. Bukowski, S. E. Rivkin & R. M. O'Bryan: High dose AZQ in renal cancer. A Southwest Oncology Group phase II study. *Invest New Drugs* 4, 57-59 (1986)

172. Chan, C., A. Bartolucci, D. Brenner, C. Presant, E. Davila, J. Carpenter, F. A. Greco, G. Clamon & J. Moore: Phase II trial of diaziquone in anthracycline-resistant adult soft tissue and bone sarcoma patients: a Southeastern Cancer Study Group Trial. *Cancer Treat Rep* 70, 427-428 (1986)

173. Host, H., R. Joss, H. Pinedo, U. Bruntsch, F. Cavalli, G. Renard, G. M. van & M. Rozencweig: Phase II trial of diaziquone (AZQ) in advanced malignant melanoma. *Eur J Cancer Clin Oncol* 19, 295-298 (1983)

174. Castleberry, R. P., A. H. Ragab, C. P. Steuber, B. Kamen, S. Toledano, K. Starling, D. Norris, P. Burger & J. P. Krischer: Aziridinylbenzoquinone (AZQ) in the treatment of recurrent pediatric brain and other malignant solid tumors. A Pediatric Oncology Group phase II study. *Invest New Drugs* 8, 401-406 (1990)

175. Ettinger, L. J., N. Ru, M. Krailo, K. S. Ruccione, W. Krivit & G. D. Hammond: A phase II study of diaziquone in children with recurrent or progressive primary brain tumors: a report from the Childrens Cancer Study Group. *J Neurooncol* 9, 69-76 (1990)

176. Ettinger, L. J., M. Krailo, K. S. Ruccione, W. Krivit & G. D. Hammond: A phase II study of diaziquone in childhood leukemia: a report from the Children's Cancer Study Group. *Am J Pediatr Hematol Oncol* 10, 18-22 (1988)

177. Stiff, P. J., R. S. McKenzie, L. D. Potempa, K. Albain, D. Koch, E. Braud, V. K. Bansal, M. K. Weidner, V. J. Lanzotti, H. G. Chun & et al: A phase I trial of high-dose diaziquone and autologous bone marrow transplantation: an Illinois Cancer Council study. *J Clin Oncol* 9, 1487-1494 (1991)

178. Arakawa, M., T. Aoki & H. Nakao: Effect of carbazilquinone on lymphoid leukemia L-1210. *Gann* 61, 485-493 (1970)

179. Okada, N. & M. Arakawa: Comparative studies on the antitumor effect on intravenous administration of carbazilquinone and mitomycin-C. *Gann* 67, 805-812 (1976)

180. Naito, K., H. Hisazumi, S. Mihara, T. Asari, K. Kobashi, T. Amano & T. Uchibayashi: Chemosensitivity study of urological malignancies using a novel dyeexclusion method. *Cancer Chemother Pharmacol* 20 Suppl, S1-S5 (1987)

181. Mitsudomi, T., S. Kaneko, M. Tateishi, T. Yano, T. Ishida, S. Kohnoe, Y. Maehara & K. Sugimachi: Chemosensitivity testing of human lung cancer tissues using the succinate dehydrogenase inhibition test. *Anticancer Res* 10, 987-990 (1990)

182. Kanematsu, T., Y. Maehara, T. Matsumata, K. Shirabe, K. Akazawa & K. Sugimachi: Human hepatocellular carcinoma sensitivity to antitumor drugs assayed using the succinate dehydrogenase inhibition test. *Oncology* 48, 34-38 (1991)

183. Kusumoto, T., Y. Sakaguchi, Y. Maehara, T. Nakashima, M. Furusawa & K. Sugimachi: Comparison of in vitro anticancer chemosensitivity between human squamous cell carcinoma and adenocarcinoma. *Oncology* 49, 343-346 (1992)

184. Baba, H., H. Takeuchi, S. Inutsuka, M. Yamamoto, K. Endo, S. Ohno, Y. Maehara & K. Sugimachi: Clinical value of SDI test for predicting effect of postoperative chemotherapy for patients with gastric cancer. *Semin Surg Oncol* 10, 140-144 (1994)

185. Nakata, H.: Experimental combination chemotherapy with vincristine and carboquone in human ovarian cancer transplanted into nude mice. *Nippon Sanka Fujinka Gakkai Zasshi* 35, 1991-1998 (1983)

186. Imai, S., Y. Nio, T. Shiraishi, T. Manabe & T. Tobe: In vivo inhibitory effect of anticancer agents on human pancreatic cancer xenografts transplanted in nude mice. *Anticancer Res* 11, 657-664 (1991)

187. Saito, T., S. Oira, A. Wakui, M. Yokoyama & H. Takahashi: Clinical experience with a new anticancer agent, carbazilquinone (NSC-134679). *Cancer Chemother Rep* 57, 447-457 (1973)

188. Tamura, Y. & S. Sato: Stimulation of a reconstituted, microsomal NADH oxidase system by carboquone, a quinoid anticancer chemical. *Gann* 68, 353-356 (1977)

189. Komiyama, T., T. Kikuchi & Y. Sugiura: Generation of hydroxyl radical by anticancer quinone drugs, carbazilquinone, mitomycin C, aclacinomycin A and adriamycin, in the presence of NADPH-cytochrome P-450 reductase. *Biochem Pharmacol* 31, 3651-3656 (1982)

190. Maehara, Y., H. Anai, Y. Sakaguchi, T. Kusumoto, Y. Emi & K. Sugimachi: Detection of DNA strand breaks in HeLa cells in vitro and in mouse sarcoma 180 cells in vivo induced by an alkylating agent, carboquone, using in situ nick translation. *Oncology* 47, 282-286 (1990)

191. Kohnoe, S., Y. Emi, I. Takahashi, M. Yoshida, Y. Maehara & K. Sugimachi: Hypoxia and acidity increase the cytotoxicity of mitomycin C and carboquone to human tumor cells in vitro. *Anticancer Res* 11, 1401-1404 (1991)

192. Kobayashi, K., M. Hino, S. Kurane, T. Yano, H. Niitani, Y. Yamano, K. Hasegawa & E. Tuboi: A comparative randomized phase II study of CDDP, CDDP-carboquone (CQ) and CDDP-etoposide as second-line chemotherapy in small cell lung cancer (SCLC). *Gan To Kagaku Ryoho* 16, 207-212 (1989)

193. Nakarai, I., T. Ueno, A. Hayashi & A. Shinoda: Combination therapy of high-dose carboquone and OK-432 in unresectable non-small cell lung cancer. *Gan To Kagaku Ryoho* 13, 3508-3512 (1986)

194. Nakao, I., Y. Ohashi, K. Ito, T. Yokoyama, I. Nishi, T. Kanko & T. Saito: Combination chemotherapy of nonsmall cell lung cancer (NSCLC) with cisplatin (CDDP), carboquone (CQ) and prednisolone (PDN)(PPQ therapy). *Gan To Kagaku Ryoho* 17, 217-220 (1990)

195. Sakata, Y., Y. Yoshida, Y. Komatsu, K. Sugawara, S. Nishimura & K. Kikuchi: MQF-OK therapy in advanced terminal stomach cancer--with special reference to a comparison with MFC therapy. *Gan To Kagaku Ryoho* 9, 109-115 (1982)

196. Nasu, K., I. Nakarai, M. Horiuchi & A. Shinoda: Clinical study of combination chemotherapy using carboquone, cisplatin, UFT and OK-432 in 7 cases of advanced gastric cancers and a relapsed gastric cancer. *Gan To Kagaku Ryoho* 15, 2410-2413 (1988)

197. Inoue, S., O. Fukuda, C. Tajima, T. Tokunaga, M. Nakayama & M. Maeyama: Combined administration of futraful and esquinone for the treatment of advanced

ovarian cancer. Gan To Kagaku Ryoho 9, 1226-1230 (1982)

198. Moriyama, M., Y. Kubota, T. Miura, T. Shuin & S. Noguchi: The effect of intravesical chemotherapy on superficial urinary bladder cancer. *Hinyokika Kiyo* 29, 351-355 (1983)

199. Ito, H.: Experimental study and clinical application of a new combination chemotherapy with cis-platinum, adriamycin and carboquone in patients with advanced prostate cancer. *Nippon Ika Daigaku Zasshi* 62, 456-468 (1995)

200. Sakata, Y., Y. Komatsu, S. Takagi, S. Saitoh, T. Itoh, H. Suzuki, K. Tsushima, D. Sasaki & Y. Yoshida: Randomized controlled study of mitomycin C/carboquone/5-fluorouracil/OK-432 (MQ-F-OK) therapy and mitomycin C/5-fluorouracil/doxorubicin (FAM) therapy against advanced liver cancer. *Cancer Chemother Pharmacol* 23 Suppl, S9-12 (1989)

201. Uzuka, Y., Y. Saito, H. Takahashi & M. Komatsu: Carboquone therapy for hematologic neoplasms. *Tohoku J Exp Med* 138, 151-160 (1982)

202. Akashi, M., S. Sakamoto, M. Ohta, S. Kitagawa, M. Yoshida, M. Saito, F. Takaku & Y. Miura: Treatment of multiple myeloma with carboquone-prednisolone. *Eur J Haematol* 42, 265-269 (1989)

203. Higuchi, T., S. Okada, H. Mori, H. Niikura, M. Omine & H. Terada: Leukemic transformation of polycythemia vera and essential thrombocythemia possibly associated with an alkylating agent. *Cancer* 75, 471-477 (1995)

204. Gauss, W.: Über die umsetzung einiger alkoxybenzochinone mit äthylenimin. *Chem Ber* 91, 2216-2222 (1958)

205. Wust, G. P. & K. J. Matthes: In vitro measurement of 3H thymidine uptake in Jensen sarcoma under the effect of cytostatics using fluid scintillation spectrometry. *Z Krebsforsch Klin Onkol* 73, 204-214 (1970)

206. Seidel, H. J.: Estimation of tumor sensitivity in vitro. II. Studies with the solid Ehrlich carcinoma of the mouse and the GW 77, a heterotransplanted human carcinoma. *Z Krebsforsch Klin Onkol* 74, 131-140 (1970)

207. Hartleib, J.: Study about the growth of two tumors of the solid Yoshida sarcoma of the rat in the same animals as well as about the reactions of tumors of different sizes to treatment with 2,3,5-triethyleneimino-1,4-benzoquinone. *Z Krebsforsch* 81, 1-6 (1974)

208. Linford, J. H.: 2,3,5-Tris-ethylenimino-1,4-benzoquinone (Trenimon): some chemical and biological properties. *Chem Biol Interact* 6, 149-168 (1973)

209. Kummer, D. & H. D. Ochs: Differenciation of reaction mechanisms of alkylating cytostatics on Ehrlich ascitescarcinoma- and lymphatic leucemic cells. *Z Krebsforsch Klin Onkol* 73, 315-328 (1970) 210. Puschendorf, B., H. Wolf & H. Grunicke: Effect of the alkylating agent trisethyleneiminobenzoquinone (trenimon) on the template activity of chromatin and DNA in RNA and DNA polymerase systems. *Biochem Pharmacol* 20, 3039-3050 (1971)

211. Ihlenfeldt, M., G. Gantner, M. Harrer, B. Puschendorf, H. Putzer & H. Grunicke: Interaction of the alkylating antitumor agent 2,3,5-tris(ethyleneimino)benzoquinone with the plasma membrane of Ehrlich ascites tumor cells. *Cancer Res* 41, 289-293 (1981)

212. Silva, J. M., D. N. Rao & P. J. O'Brien: Modulation of trenimon-induced cytotoxicity by DT-diaphorase in isolated rat hepatocytes under aerobic versus hypoxic conditions. *Cancer Res* 52, 3015-3021 (1992)

213. Silva, J. M. & P. J. O'Brien: Molecular mechanisms of trenimon-induced cytotoxicity in resistant L5178Y/HBM10 cells. *Int J Radiat Oncol Biol Phys* 22, 639-642 (1992)

214. Driscoll, J. S., L. Dudeck, G. Congleton & R. I. Geran: Potential CNS antitumor agents VI: aziridinylbenzoquinones III. *J Pharm Sci* 68, 185-188 (1979)

215. Gibson, N. W., J. A. Hartley, J. Butler, D. Siegel & D. Ross: Relationship between DT-diaphorase-mediated metabolism of a series of aziridinylbenzoquinones and DNA damage and cytotoxicity. *Mol Pharmacol* 42, 531-536 (1992)

216. Lea, J. S., H. J. Garner, J. Butler, B. M. Hoey & T. H. Ward: The lack of correlation between toxicity and free radical formation of two diaziridinyl benzoquinones. *Biochem Pharmacol* 37, 2023-2025 (1988)

217. Ward, T. H., J. Butler, H. Shahbakhti & J. T. Richards: Comet assay studies on the activation of two diaziridinylbenzoquinones in K562 cells. *Biochem Pharmacol* 53, 1115-1121 (1997)

218. Hartley, J. A., M. Berardini, M. Ponti, N. W. Gibson, A. S. Thompson, D. E. Thurston, B. M. Hoey & J. Butler: DNA cross-linking and sequence selectivity of aziridinylbenzoquinones: a unique reaction at 5'-GC-3' sequences with 2,5-diaziridinyl-1,4-benzoquinone upon reduction. *Biochemistry* 30, 11719-11724 (1991)

219. Butler, J., B. M. Hoey & T. H. Ward: The alkylation of DNA in vitro by 2,5-bis(2-hydroxyethylamino)-3,6-diaziridinyl-1,4-benzoquinone (BZQ)--I. *Biochem Pharmacol* 38, 923-927 (1989)

220. G. Brulé, S. J. Eckhardt, T. C. Hall & A. Winkler: Drug Therapy of Cancer. World Health Organization, Geneva (1973)

221. Gasparri, F., P. Periti, G. Scarselli, T. Mazzei, L. Savino, F. Branconi & R. G. Dancygier: Perioperative chemotherapy in the treatment of carcinoma of the uterine cervix. *Chemioterapia* 5, 44-52 (1986)

222. Betteridge, R. F., A. G. Bosanquet & E. D. Gilby: Pharmacokinetics of 2,5-diaziridinyl-3,6-bis(2hydroxyethylamino)-1,4-benzoquinone (BZG, NSC 224070) during a phase I clinical trial. Eur J Cancer 26, 107-112 (1990)

223. Weiss, M. J., G. S. Redin, G. R. Allen, A. C. Dornbush, H. L. Lindsay, J. F. Poletto, W. A. Remers, R. H. Roth & A. E. Sloboda: The mitomycin antibiotics. Synthetic studies. XXII. Antibacterial structure-activity relationships in the indologuinone series. *J Med Chem* 11, 742-745 (1968)

224. Iyengar, B. S., W. A. Remers & J. J. Catino: New 2substituted indoloquinone mitomycin analogues. *J Med Chem* 32, 1866-1872 (1989)

225. Oosterveen, E. A. & W. N. Speckamp: Mitomycin analogues 1. Indoloquinones as potent bisalkylating agents. *Tetrahedron* 43, 255-262 (1987)

226. Bailey, S. M., N. Suggett, M. I. Walton & P. Workman: Structure-activity relationships for DTdiaphorase reduction of hypoxic cell directed agents: indoloquinones and diaziridinyl benzoquinones. *Int J Radiat Oncol Biol Phys* 22, 649-653 (1992)

227. Naylor, M. A., M. Jaffar, J. Nolan, M. A. Stephens, S. Butler, K. B. Patel, S. A. Everett, G. E. Adams & I. J. Stratford: 2-Cyclopropylindoloquinones and their analogues as bioreductively activated antitumor agents: structure-activity in vitro and efficacy in vivo. *J Med Chem* 40, 2335-2346 (1997)

228. Beall, H. D., S. Winski, E. Swann, A. R. Hudnott, A. S. Cotterill, N. O'Sullivan, S. J. Green, R. Bien, D. Siegel, D. Ross & C. J. Moody: Indolequinone antitumor agents: correlation between quinone structure, rate of metabolism by recombinant human NAD(P)H:quinone oxidoreductase, and in vitro cytotoxicity. *J Med Chem* 41, 4755-4766 (1998)

229. Phillips, R. M., M. A. Naylor, M. Jaffar, S. W. Doughty, S. A. Everett, A. G. Breen, G. A. Choudry & I. J. Stratford: Bioreductive activation of a series of indolequinones by human DT-diaphorase: structure-activity relationships. *J Med Chem* 42, 4071-4080 (1999)

230. Smitskamp-Wilms, E., H. R. Hendriks & G. J. Peters: Development, pharmacology, role of DT-Diaphorase and prospects of the indoloquinone EO9. *General Pharmacol* 27, 421-429 (1996)

231. Hendriks, H. R., P. E. Pizao, D. P. Berger, K. L. Kooistra, M. C. Bibby, E. Boven, H. C. Dreef van der Meulen, R. E. Henrar, H. H. Fiebig, J. A. Double & et al: EO9: a novel bioreductive alkylating indoloquinone with preferential solid tumour activity and lack of bone marrow toxicity in preclinical models. *Eur J Cancer* 29A, 897-906 (1993)

232. Kaye, S. B., P. Workman, M. A. Graham, J. Cassidy & D. Jodrell: Pharmacokinetics and early clinical studies of selected new drugs. *Cancer Surv* 17, 371-396 (1993)

233. Maliepaard, M., A. Wolfs, S. E. Groot, N. J. De Mol & L. H. M. Janssen: Indoloquinone EO9: DNA interstrand cross-linking upon reduction by DT-diaphorase or xanthine oxidase. *Br J Cancer* 71, 836-839 (1995)

234. Walton, M. I., P. J. Smith & P. Workman: The role of NAD(P)H: quinone reductase (EC 1.6.99.2, DT-diaphorase) in the reductive bioactivation of the novel indoloquinone antitumor agent EO9. *Cancer Commun* 3, 199-206 (1991)

235. Bailey, S. M., M. D. Wyatt, F. Friedlos, J. A. Hartley, R. J. Knox, A. D. Lewis & P. Workman: Involvement of DT-diaphorase (EC 1.6.99.2) in the DNA cross-linking and sequence selectivity of the bioreductive anti-tumour agent EO9. *Br J Cancer* 76, 1596-1603 (1997)

236. Smitskamp-Wilms, E., G. J. Peters, H. M. Pinedo, J. Van Ark-Otte & G. Giaccone: Chemosensitivity to the indoloquinone EO9 is correlated with DT- diaphorase activity and its gene expression. *Biochem Pharmacol* 47, 1325-1332 (1994)

237. Plumb, J. A., M. Gerritsen, R. Milroy, P. Thomson & P. Workman: Relative importance of DT-diaphorase and hypoxia in the bioactivation of EO9 by human lung tumor cell lines. *Int J Radiat Oncol Biol Phys* 29, 295-299 (1994)

238. Plumb, J. A., M. Gerritsen & P. Workman: DTdiaphorase protects cells from the hypoxic cytotoxicity of indoloquinone EO9. *Br J Cancer* 70, 1136-1143 (1994)

239. Schellens, J. H. M., A. S. T. Planting, B. A. C. Van Acker, W. J. Loos, M. De Boer-Dennert, M. E. L. Van der Burg, I. Koier, R. T. Krediet, G. Stoter & J. Verweij: Phase I and pharmacologic study of the novel indoloquinone bioreductive alkylating cytotoxic drug E09. *J Natl Cancer Inst* 86, 906-912 (1994)

240. McLeod, H. L., M. A. Graham, S. Aamdal, A. Setanoians, Y. Groot & B. Lund: Phase I pharmacokinetics and limited sampling strategies for the bioreductive alkylating drug E09. *Eur J Cancer (A)* 32A, 1518-1522 (1996)

241. Dirix, L. Y., F. Tonnesen, J. Cassidy, R. Epelbaum, W. W. T. Huinink, N. Pavlidis, R. Sorio, T. Gamucci, I. Wolff, A. Te Velde, J. Lan & J. Verweij: EO9 phase II study in advanced breast, gastric, pancreatic and colorectal carcinoma by the EORTC early clinical studies group. *Eur J Cancer (A)* 32A, 2019-2022 (1996)

242. Pavlidis, N., A. R. Hanauske, T. Gamucci, J. Smyth, M. Lehnert, A. te Velde, J. Lan & J. Verweij: A randomized phase II study with two schedules of the novel indoloquinone EO9 in non-small-cell lung cancer: a study of the EORTC Early Clinical Studies Group (ECSG). *Ann Oncol* 7, 529-531 (1996)

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