THE SIGNIFICANCE OF AUTOIMMUNITY IN THE PATHOGENESIS OF CHAGAS HEART DISEASE

Juan S. Leon and David M. Engman

Departments of Microbiology-Immunology and Pathology, Feinberg Medical School of Northwestern University, Chicago, IL 60611

TABLE OF CONTENTS

1. Abstract

- 2. Introduction
- 3. Chagas heart disease is caused by persistent T. cruzi and T. cruzi-specific immunity
- 4. Chagas heart disease is caused by autoimmunity
- 5. Potential mechanisms of T. cruzi-induced autoimmunity
- 6. Conclusions
- 7. Acknowledgements
- 8. References

1. ABSTRACT

Chagas heart disease develops in approximately one-third of individuals infected with the protozoan parasite Trypanosoma cruzi. Among the many possible mechanisms responsible for this illness, an autoimmune mechanism has received much experimental support during the past several decades. Initial observations of the absence or near absence of parasites from the inflamed tissues suggested the autoimmunity hypothesis, and the finding of measurable autoimmune responses in humans and experimental animals has bolstered this idea. The rigorous testing of the hypothesis has been difficult, primarily because other mechanisms are likely to play a role during active infection, particularly immunity to parasite antigens that may persist in the infected animal. While the role autoimmunity plays in disease pathogenesis is not known, it is clear that autoimmune responses are induced during infection of some humans and animals. A number of mechanisms may explain the induction of autoimmunity during T. cruzi infection, including parasite-induced polyclonal lymphocyte activation, molecular mimicry, bystander activation, and presentation of cryptic self epitopes. The genetic makeup of both the parasite and host are also critical to the outcome of infection. The autoimmune hypothesis deserves further exploration, while public health interventions should focus on control of the insects that transmit the parasite, development of parasiticidal drugs and vaccines, and testing of blood products, which have become an important threat of new infections.

2. INTRODUCTION

Chagas heart disease (CHD) is caused by the protozoan *Trypanosoma cruzi* and develops in roughly 30% of infected individuals (1). CHD encompasses both acute and chronic manifestations of myocarditis and may take months to decades to develop. After nearly a century of investigation, the exact mechanism of CHD is unclear and

under debate. Why only some individuals are susceptible to CHD, why there is such variability in disease manifestations, why it takes so long for chronic CHD to develop, and what causes CHD are some of the many questions investigators and physicians have addressed. There are at least six proposed mechanisms for CHD pathogenesis. CHD may be caused by: (i) anti-*T. cruzi* immune responses to persistent parasites or parasite antigens; (ii) autoimmunity induced by *T. cruzi*; (iii) microvascular spasm; (iv) ischemia; (v) chronic eosinophilia; and (vi) direct toxicity of the parasite (reviewed in (2-4)). These mechanisms may be interrelated. This review summarizes our views on the first and second hypotheses – that CHD is caused by anti-*T. cruzi* immunity and persistent parasite or parasite antigens (reviewed in (5-8)) and that CHD is an autoimmune disease (reviewed in (3, 9-13)).

The hypothesis that Chagas disease is an autoimmune disease arose from early observations of cardiac pathology and the discovery of antibodies cross-reactive to T. cruzi and host in patients with chronic CHD. Histologic analysis indicated that chronic CHD patients had cardiac inflammation and fibrosis in the apparent absence of T. cruzi suggesting that these inflammatory lesions were not caused by parasitized tissue as previously believed (14). Later, antibodies specific for host proteins, which could be blocked by T. cruzi antigens, were found in chronic CHD patients (15). This report was later retracted for methodological concerns (16), but it encouraged other groups to research autoantibodies in patients with CHD (reviewed in (13)). If inflammation was not associated with parasitized tissue and the immune system was targeting host antigens, what caused these inflammatory processes? The concept of Chagas disease as an autoimmune disease was thus put forth by Santos-Buch and Teixeira in 1974(17).

An autoimmune disease is defined as tissue inflammation or cellular damage caused by an immune reaction against self antigens (autoimmunity) that results when mechanisms responsible for mantaining immunologic self-tolerance fail (18). Autoimmunity may be present in the host in the absence of tissue inflammation or disease. In fact, autoimmunity may be the effect and not the cause of tissue damage. To prove that CHD is an autoimmune disease, tissue inflammation must be shown to be directly caused by autoimmunity, regardless of the initiating agent.

The public health and clinical significance of whether CHD is an autoimmune disease are serious. If CHD is an autoimmune disease, then the treatments for the disease must address autoimmune mechanisms. Therapies solely directed against the parasite may not prevent autoimmune destruction and should be developed to eliminate the parasite and reduce autoimmune damage. In addition, vaccine candidates would have to be screened to confirm that they do not induce an autoimmune disease. Thus, anti-*T. cruzi* chemotherapy and *T. cruzi* vaccines would safeguard against the potential for autoimmune sequelae.

3. CHAGAS HEART DISEASE IS CAUSED BY PERSISTENT T. CRUZI AND T. CRUZI-SPECIFIC IMMUNITY

The hypothesis that chronic Chagas disease is due to residual parasites or antigens and accompanying anti-*T. cruzi* immunity has four supporting lines of evidence: (i) *T. cruzi* or *T. cruzi* DNA/antigens can be detected in chronic lesions; (ii) *T. cruzi* or *T. cruzi* DNA/antigens can sometimes colocalize with inflammation; (iii) inflammation is targeted against *T. cruzi* and has been shown to cause damage; and (iv) elimination of *T. cruzi* reduces disease severity. Each of these points is covered briefly here but are explored in depth in references (3, 6-8).

As discussed above, examination of cardiac tissue from chronic Chagas patients by early clinicians revealed tissue inflammation and fibrosis occuring in the apparent absence of T. cruzi (14). However, with the advent of more sensitive techniques. such as immunohistochemistry (19-23), whole tissue PCR (24-26), in situ hybridization (27), and in situ PCR (28), parasite antigens and DNA have been shown to persist in the tissues of infected humans and experimental animals. An important point to highlight is that these techniques cannot distinguish between live parasites and residual parasite antigens or DNA. One report using beta-galactosidaseexpressing transgenic T. cruzi demonstrated that live parasites could be detected in mice 10 months post infection (29). Interestingly, the number of live parasites was far smaller than estimated by other techniques (23, 30), suggesting that antigens from destroyed T. cruzi may remain in tissues for a long time and provoke ongoing inflammatory processes (29).

The second line of evidence is that *T. cruzi* can sometimes colocalize with inflammation. Colocalization often means "present in the same section" because, to date, there are no reports on the correlation between localization of *T. cruzi* antigens and physical proximity to inflammatory

lesions. Several reports showed that inflammation is often found in cardiac sections with T. cruzi detected by immunohistochemistry (19-23) or in situ PCR (28). Sections with high severity of myocarditis are significantly associated with the presence of the parasite (22). At the same time, there are many sections with mild or moderate inflammation in which T. cruzi antigens are not detected (21, 22, 29). There are also reports that state that the severity of inflammation is not associated with the presence of the parasite (19, 26). Interestingly, the degree of inflammation sometimes seems disproportionate to the quantity of T. cruzi in the tissue (23, 29). These observations could support either the parasite persistence hypothesis or the autoimmunity hypothesis. The inflammation could be due to (i) the presence of parasite antigens from nearby disintegrating T. cruzi; (ii) autoimmunity against neighboring cells; (iii) tissue ischemia and subsequent inflammation; (iv) an overly aggressive anti-parasite inflammatory response to the chronic presence of the parasite, among other hypotheses (29). In summary, T. cruzi antigens are associated with severely inflamed tissue but there is not a high association between T. cruzi antigens and the presence of inflammation, because of limitations in the specific assay or biologic reasons.

In support of the third line of evidence, immune responses have been shown to target *T. cruzi* and mediate destruction of infected cells *in vitro*. There is a wealth of reports in the literature on this topic (reviewed in (7, 31)), including destruction by antibody-dependent cellular cytotoxicity (32, 33), cytotoxic T lymphocytes (34-36), and Th1 CD4⁺ lymphocytes (37), among others.

Lastly, elimination of *T. cruzi* by chemotherapy reduces disease in certain instances. Administration of anti-*T. cruzi* chemotherapy during acute disease generally protects mice from parasitosis, mortality and disease (reviewed in (38)). Administration of chemotherapy during the chronic phase has been reported to reverse lesion development in experimental animals (39). In human disease, there is no consensus on the efficacy of chemotherapy on cardiomyopathy (40-45) and so it is difficult to ask the question of whether elimination of *T. cruzi* eliminates disease.

Although it might seem obvious that the presence of parasite antigen in tissue would lead to the development of parasite-specific immunity and consequent tissue inflammation, this has not been proven. The mere coexistence of parasite antigen or DNA and infiltrating mononuclear cells does not address the antigen specificities of the lymphocytes and certainly does not speak to the inflammatory potential on a per cell basis. The one experiment that may address this in part was conducted by Kumar *et al.* (37) in which *T. cruzi* expressing ovalbumin was used for infection. Ovalbumin-specific CD4⁺ lymphocytes, adoptively transferred into animals infected with the ovalbumin transgenic *T. cruzi*, accumulated in the parasitized tissue as determined by immunohistochemical staining with a clonotype-specific antibody. In our view, this result strongly suggests that the transferred lymphocytes detected their cognate antigen expressed by the trypanosome. It will be important to follow up on this finding perhaps by testing for ovalbumin-specific cells that expand naturally upon infection with the transgenic parasite (*i.e.*, not via adoptive transfer) or by developing a number of *T. cruzi* antigen-specific T cell clones and clonotype specific antibodies that can be used to assess antigenspecific T cell infiltration and effector function using wildtype parasites.

In conclusion, there is strong evidence in support of the hypothesis that Chagas disease can be explained by persistent parasite and anti-parasite immunity. At the same time, this does not exclude a role for autoimmunity or other mechanisms in disease (8).

4. CHAGAS HEART DISEASE IS CAUSED BY AUTOIMMUNITY

The hypothesis that CHD is an autoimmune disease should be divided into two separate questions: (i) whether autoimmunity can be induced by infection with T. *cruzi*, and (ii) whether T. *cruzi* induced autoimmunity is pathogenic. The difference between these two questions is, in our view, the main source of confusion in the literature debating the hypothesis that CHD is an autoimmune disease.

The first hypothesis, whether autoimmunity can be induced by infection with T. cruzi, is supported and well documented. Infection with T. cruzi in humans and experimental animals induces both humoral and cellular autoimmunity to host antigens including cardiac, skeletal, and nervous antigens, among others (reviewed in (3)). Infection with T. cruzi induces autoantibodies to myosin (46, 47), beta adrenergic receptor (reviewed in (48)), cytoskeletal and microtubule associated proteins (49), nervous tissue proteins (50), ribosomal P proteins (51, 52), and a novel mammalian protein, Cha (53). On the cellular side, infection with T. cruzi induces T cells specific for heart homogenate (54, 55), neuronal antigens (56), cardiac (47, 57) and skeletal myosin (58, 59), and Cha antigen (53). Therefore, infection with T. cruzi induces both humoral and cellular autoimmunity.

The second hypothesis, whether *T. cruzi* induced autoimmunity is *pathogenic*, is not as well documented. To date, there is no direct evidence that *T. cruzi* induced autoantibodies cause disease. Many of these target proteins have ubiquitous expression which do not support the heart specific disease in CHD patients. In addition, many of the targets are intracellular so it is difficult to devise a mechanism by which autoantibodies cause disease if their targets are inaccessible. Furthermore, the evidence that these autoantibodies are more prevalent in patients with CHD than in asymptomatic, *T. cruzi* infected individuals, is scarce (46, 60; reviewed in (3)) and contested (3). On the other hand, antibodies from infected patients affected the cell signaling and contraction of cardiac myocytes (reviewed in (48)) and lysed myocytes through antibody dependent cytotoxicity *in vitro* (61). In addition, immunization with *T. cruzi* cruzipain induced autoantibodies to myosin, conduction abnormalities, and IgG deposition (62). The authors suggested that these autoantibodies are pathogenic because of the association with the observed conduction abnormalities in mice.

There is some direct evidence supporting a contribution of cellular autoimmunity to CHD. It has been reported that splenocytes from chronically infected mice can lyse syngeneic myoblasts *in vitro* (63, 64) or induce inflammation in sciatic nerves (56) and hearts when adoptively transferred to naïve recipient mice (63). Immunization of mice with *T. cruzi* ribosomal proteins (65) or cruzipain induced cardiac conduction abnormalities (58, 62). Cruzipain immunization also induced skeletal myositis accompanied by production of myosin autoantibodies and autoreactive T cells. Since no live parasites were used in the ribosomal or cruzipain protein immunization experiments, the induction of autoreactive cells and cardiac damage was thought to be due to a molecular mimicry

mechanism. Transfer of CD4⁺ T cells from chronically infected mice mediated the rejection of implanted syngeneic newborn hearts (54), but if a different combination of mouse and parasite strain was used, this did not occur (30). Perhaps the conflict in these results was due to the difference in susceptibility to autoimmunity of host strains infected with certain parasite strains (discussed below). Strikingly, suppressing autoimmunity to heart enriched for myosin reduced cardiac antigens inflammation, fibrosis and myosin autoantibody production in chronically infected mice (66). Though the autoimmune disease control used in this study does not agree with previously published results (67), the result still suggests that cardiac autoimmunity significantly contributes to chronic experimental CHD.

Two criticisms are often used against the hypothesis that CHD is an autoimmune disease. The first is that immunosuppressants, which generally relieve symptoms of autoimmune diseases, exacerbate mortality and disease in Chagas patients. The best examples of this are Chagas heart transplant recipients receiving immunosuppressants, and Chagas patients infected with HIV. For the record, the largest multicenter study on CHD heart transplant recipients concluded that CHD patients have no difference in mortality compared to heart transplant recipients suffering from idiopathic dilated cardiomyopathy or ischemic cardiomyopathy (68). In addition, the presence of the parasite confounds the question of whether autoimmunity contributes to disease, since suppressing host immunity results in an increased proliferation of parasites and therefore disease. The second criticism states that autoimmunity does not contribute to CHD because T. cruzi chemotherapy alone reduces clinical disease in humans and experimental animals. However, there is no consensus on the efficacy of chemotherapy on human CHD (40-45). Unless chemotherapy completely eliminates disease, any residual disease can be explained by additional mechanisms. In experimental models of T. cruzi infection, chemotherapy given *immediately* after infection reduces and sometimes eliminates cardiac disease

(reviewed in (38)). Because *T. cruzi* is the trigger for autoimmunity, elimination of this trigger in the *acute* disease phase could potentially eliminate the induction of autoimmunity, making the analysis of the contribution of autoimmunity irrelevant.

In summary, infection with *T. cruzi* induces autoantibodies and autoreactive T cells to a variety of autoantigens. Though there are some reports on the pathogenic potential of this autoimmunity, CHD has not yet been shown to be an autoimmune disease. Adoptive transfer and immunization experiments with parasite proteins do not necessarily recapitulate events in infected mice and so no conclusion can yet be made about whether *T. cruzi*-induced autoimmunity directly contributes to tissue damage in human Chagas disease, or for that matter, in infected mice.

5. POTENTIAL MECHANISMS OF T. CRUZI-INDUCED AUTOIMMUNITY

A number of mechanisms may explain how *T*. *cruzi* induces autoimmunity in a host which is normally tolerant to its own antigens. These are reviewed in some detail in ref. (10) and are briefly summarized here.

Polyclonal activation involves the antigenindependent stimulation of self-reactive lymphocytes. Polyclonal activators stimulate a large percentage of both T cells and B cells, irrespective of antigen specificity and, in some cases, by interacting with surface molecules other than antigen receptors. The most common example of a polyclonal activator is lipopolysaccharide, which induces production of a wide repertoire of acute autoantibodies in mice (69). These autoantibodies have weak affinities and are often of the IgM isotype. Certain strains of *T. cruzi* have been shown to possess polyclonal activators, suggesting that this mechanism may possibly induce autoimmunity during infection (70).

In **molecular mimicry**, autoimmunity results from a "misdirected" immune response. The immune response is first directed to a parasite antigen and "crossreacts" with a host antigen that resembles it, causing autoimmunity. Putative cross-reactive proteins of host and *T. cruzi* have been identified. These include *T. cruzi* ribosomal P proteins and their mammalian homologues (51, 52), *T. cruzi* shed acute phase antigen and the human Cha autoantigen (53), *T. cruzi* B13 and cardiac myosin (46) and *T. cruzi* cruzipain with skeletal myosin (58). Furthermore, autoimmunity can result from immunization with a *T. cruzi* antigen (58, 65).

The third mechanism, **cryptic epitope**, posits that *T. cruzi* infection leads to release of previously sequestered epitopes or, alternatively, that the inflammatory environment induced by *T. cruzi* alters antigen processing and presentation such that novel self epitopes are generated. Because the immune system is not tolerant to the novel epitopes, immunity develops rapidly (71). In support

of this hypothesis, antigen processing and presentation was altered after *in vitro* treatment with interferon gamma (reviewed in (72)). Although not formally demonstrated in *T. cruzi* infection, this mechanism is thought to be involved in the pathogenesis of rheumatoid arthritis, systemic lupus erythematosus, and other autoimmune diseases (73).

In the mechanism of **bystander activation**, *T*. cruzi infection causes tissue destruction and release of host antigens. The excess levels of host antigens released in a proinflammatory environment may stimulate autoreactive cells, initiating autoimmunity. Evidence for this hypothesis includes the observation that cardiac autoimmunity can result from many, varied types of insults to the heart, including infection with viruses and parasites, heart surgery and heart transplantation (74-76). In addition, the autoantigen that initiates bystander autoimmunity may not be the major target autoantigen throughout the course of disease. In this process, called "epitope spreading," autoimmunity against one epitope develops, causing damage to tissue(s) containing that epitope. This damage then results in the release of additional self antigens, the processing and presentation of which induces the stimulation of autoimmunity of additional epitope specificity(ies). Interestingly, the initial responses may wane due to immunoregulation, leading to "waves" of autoimmunity targeted to different epitopes (77).

In addition, there is the question of the time at which T. cruzi-induced autoimmunity develops. It may be induced immediately after the initial contact of the parasite with the host, during the acute phase of disease (47, 78, 79). This early autoimmunity likely results from tissue damage caused by the parasite and/or molecular mimicry. The polyclonal, polyspecific nature of the autoantibody response supports the former hypothesis. Autoimmunity may also develop later in the disease course (61, 64). Persistent, chronic inflammation may be necessary to overcome the threshold of cardiac damage or produce the proper inflammatory environment for the stimulation and expansion of autoreactive cells. These mechanisms are not exclusive of each other and combinations of these mechanisms may play a role in the induction of autoimmunity during *T. cruzi* infection.

Finaly, immunogenetic factors of the host and genetic characteristics of the parasite may also influence the induction of autoimmunity. Certain strains of mice are susceptible to *T. cruzi* induced autoimmunity, while others are resistant (47). There is also evidence that *T. cruzi* genetics determine the induction of autoimmunity. Infection of mice with the Brazil strain of *T. cruzi* induces anti-heart antibodies whereas infection with the Guayas strain of *T. cruzi* does not (80, 81)

6. CONCLUSIONS

It is clear that autoimmunity can be induced by *T*. *cruzi* infection in humans and experimental animals (Figure 1), although, to date, there is no proof that autoimmunity directly contributes to disease pathogenesis. However, as discussed above, it is only presumed and not proven that *in*

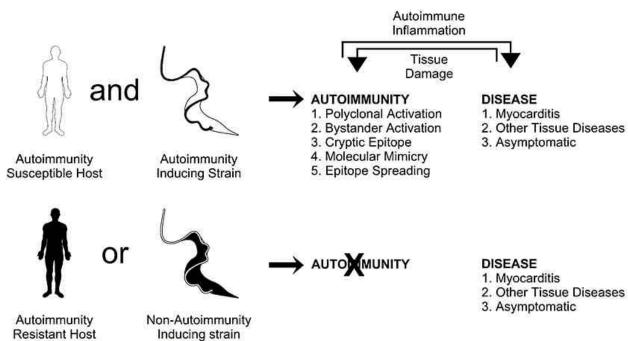


Figure 1. Model of autoimmunity induction in Chagas heart disease. Host infection with *T. cruzi* may induce autoimmunity and/or disease depending on host immunogenetics and parasite genetics. Infection involving an "autoimmunity susceptible" host and "autoimmunity virulent" parasite clone (open figures, top) may lead to pathology via direct or indirect (*.e.*, bystander activation after tissue damage) mechanisms. Infection involving an "autoimmunity resistant" host or "autoimmunity avirulent" parasite (filled figures, bottom) may lead to disease only through mechanisms other than autoimmunity.

vivo anti-T. cruzi immunity is responsible for inflammation and damage present in the myocardium. In our opinion, the mechanism(s) that underlie the pathogenesis of Chagas disease are as varied as the outcome of human infection and the factors that determine which mechanisms may participate are only beginning to be understood. Why is it that less than half of all infected people develop clinical disease? In what fraction of individuals with Chagas heart disease does autoimmunity exist and in what fraction of those is the autoimmunity pathogenic and not merely phenomenologic? Various mechanisms may explain how an infectious organism can break immunologic self tolerance. To convince skeptics that T. cruzi-induced autoimmunity contributes to the pathology of Chagas disease additional evidence is required. Anti-T. cruzi chemotherapy and vaccine trials must be pursued regardless of whether CHD is proven to have an autoimmune component to pathogenesis.

7. ACKNOWLEDGEMENTS

We thank Drs. Kegiang Wang and Randal Tibbetts for their contributions to the research. Much of the work was supported by grants and fellowships from the National Institutes of Health and the American Heart Association.

8. REFERENCES

1. Moncayo, A.: Progress towards interruption of transmission of Chagas disease. *Mem Inst Oswaldo Cruz* 94 Suppl 1, 401-404 (1999)

2. Tanowitz, H. B., L. V. Kirchhoff, D. Simon, S. A. Morris, L. M. Weiss & M. Wittner: Chagas' disease. *Clin Microbiol Rev* 5, 400-419 (1992)

3. Kierszenbaum, F.: Chagas' disease and the autoimmunity hypothesis. *Clin Microbiol Rev* 12, 210-223 (1999)

4. Kierszenbaum, F.: Chronic chagasic tissue lesions in the absence of *Trypanosoma cruzi*: a proposed mechanism. *Parasitol Today* 12, 414-415 (1996)

5. Levin, M. J.: In chronic Chagas heart disease, don't forget the parasite. *Parasitol Today* 12, 415-416 (1996)

6. Tarleton, R. L.: Parasite persistence in the aetiology of Chagas disease. *Int J Parasitol* 31, 550-554 (2001)

7. Tarleton, R. L.: *Trypanosoma cruzi* and Chagas disease: cause and effect. Pages 107-115 in S. Black and J. R. Seed, eds. *World Class Parasites: American Trypanosomiasis*. Kluwer Academic Publishers, New York. (2002)

8. Tarleton, R. L. & L. Zhang: Chagas disease etiology: autoimmunity or parasite persistence? *Parasitol Today* 15, 94-99 (1999)

9. Kalil, J. & E. Cunha-Neto: Autoimmunity in Chagas disease cardiomyopathy: fulfilling the criteria at last? *Parasitol Today* 12, 396-399 (1996)

10. Leon, J. S. & D. M. Engman: Autoimmunity in Chagas heart disease. *Int J Parasitol* 31, 554-560 (2001)
11. Leon, J. S. & D. M. Engman: The contribution of autoimmunity to Chagas disease? Pages 97-106 in S. Black

and J. R. Seed, eds. *World Class Parasites: American Trypanosomiasis*. Kluwer Academic Publishers, New York. (2002)

12. Engman, D. M. & J. S. Leon: Pathogenesis of Chagas heart disease: role of autoimmunity. *Acta Trop* 81, 123-132. (2002)

13. Kierszenbaum, F.: Autoimmunity in Chagas' disease. J Parasitol 72, 201-211 (1986)

14. Torres, C. M.: Sobre a anatomia patologica da doenca de Chagas. *Mem Inst Oswaldo Cruz* 36, 391-404 (1941)

15. Cossio, P. M., C. Diez, A. Szarfman, E. Kreutzer, B. Candiolo & R. M. Arana: Chagasic cardiopathy. Demonstration of a serum gamma globulin factor which reacts with endocardium and vascular structures. *Circulation* 49, 13-21 (1974)

16. Khoury, E. L., C. Diez, P. M. Cossio & R. M. Arana: Heterophil nature of EVI antibody in *Trypanosoma cruzi* infection. *Clin Immunol Immunopathol* 27, 283-288. (1983)

17. Santos-Buch, C. A. & A. R. Teixeira: The immunology of experimental Chagas' disease. 3. Rejection of allogeneic heart cells in vitro. *J Exp Med* 140, 38-53 (1974)

18. Abbas, A. K., A. H. Lichtman & J. S. Pober. 2000. *Cellular and Molecular Immunology*. W B Saunders Co, Philadelphia.

19. Palomino, S. A., V. D. Aiello & M. L. Higuchi: Systematic mapping of hearts from chronic chagasic patients: the association between the occurrence of histopathological lesions and *Trypanosoma cruzi* antigens. *Ann Trop Med Parasitol* 94, 571-579. (2000)

20. Anez, N., H. Carrasco, H. Parada, G. Crisante, A. Rojas, C. Fuenmayor, N. Gonzalez, G. Percoco, R. Borges, P. Guevara & J. L. Ramirez: Myocardial parasite persistence in chronic chagasic patients. *Am J Trop Med Hyg* 60, 726-732 (1999)

21. Reis, M. M., L. Higuchi Mde, L. A. Benvenuti, V. D. Aiello, P. S. Gutierrez, G. Bellotti & F. Pileggi: An *in situ* quantitative immunohistochemical study of cytokines and IL- 2R+ in chronic human chagasic myocarditis: correlation with the presence of myocardial *Trypanosoma cruzi* antigens. *Clin Immunol Immunopathol* 83, 165-172. (1997)

22. Bellotti, G., E. A. Bocchi, A. V. de Moraes, M. L. Higuchi, M. Barbero-Marcial, E. Sosa, A. Esteves-Filho, R. Kalil, R. Weiss, A. Jatene & F. Pileggi: *In vivo* detection of *Trypanosoma cruzi* antigens in hearts of patients with chronic Chagas' heart disease. *Am Heart J* 131, 301-307 (1996)

23. Ben Younes-Chennoufi, A., M. Hontebeyrie-Joskowicz, V. Tricottet, H. Eisen, M. Reynes & G. Said: Persistence of *Trypanosoma cruzi* antigens in the inflammatory lesions of chronically infected mice. *Trans R Soc Trop Med Hyg* 82, 77-83 (1988)

24. Brandariz, S., A. Schijman, C. Vigliano, P. Arteman, R. Viotti, C. Beldjord & M. J. Levin: Detection of parasite DNA in Chagas' heart disease. *Lancet* 346, 1370-1371. (1995)

25. Vago, A. R., A. M. Macedo, S. J. Adad, D. D. Reis & R. Correa-Oliveira: PCR detection of *Trypanosoma cruzi* DNA in oesophageal tissues of patients with chronic digestive Chagas' disease. *Lancet* 348, 891-892. (1996)

26. Olivares-Villagomez, D., T. L. McCurley, C. L. Vnencak-Jones, R. Correa-Oliveira, D. G. Colley & C. E. Carter: Polymerase chain reaction amplification of three different *Trypanosoma cruzi* DNA sequences from human chagasic cardiac tissue. *Am J Trop Med Hyg* 59, 563-570 (1998)

27. Lane, J. E., D. Olivares-Villagomez, C. L. Vnencak-Jones, T. L. McCurley & C. E. Carter: Detection of *Trypanosoma cruzi* with the polymerase chain reaction and *in situ* hybridization in infected murine cardiac tissue. *Am J Trop Med Hyg* 56, 588-595 (1997)

28. Zhang, L. & R. L. Tarleton: Parasite persistence correlates with disease severity and localization in chronic Chagas' disease. *J Infect Dis* 180, 480-486 (1999)

29. Buckner, F. S., A. J. Wilson & W. C. Van Voorhis: Detection of live *Trypanosoma cruzi* in tissues of infected mice by using histochemical stain for beta-galactosidase. *Infect Immun* 67, 403-409. (1999)

30. Tarleton, R. L., L. Zhang & M. O. Downs: "Autoimmune rejection" of neonatal heart transplants in experimental Chagas disease is a parasite-specific response to infected host tissue. *Proc Natl Acad Sci USA* 94, 3932-3937 (1997)

31. DosReis, G. A.: Cell-mediated immunity in experimental *Trypanosoma cruzi* infection. *Parasitol Today* 13, 335-342 (1997)

32. Tambourgi, D. V., T. L. Kipnis & W. Dias da Silva: *Trypanosoma cruzi:* antibody-dependent killing of bloodstream trypomastigotes by mouse bone marrow-derived mast cells and by mastocytoma cells. *Exp Parasitol* 68, 192-201. (1989)

33. Velge, P., J. P. Kusnierz, A. Ouaissi, B. Marty, B. N. Pham & A. Capron: *Trypanosoma cruzi*: infection of T lymphocytes and their destruction by antibody-dependent cell-mediated cytotoxicity. *Eur J Immunol* 21, 2145-2152. (1991)

34. Low, H. P., M. A. Santos, B. Wizel & R. L. Tarleton: Amastigote surface proteins of *Trypanosoma cruzi* are targets for CD8⁺ CTL. *J Immunol* 160, 1817-1823. (1998)

35. Nickell, S. P., G. A. Stryker & C. Arevalo: Isolation from *Trypanosoma cruzi*-infected mice of CD8⁺, MHC-

restricted cytotoxic T cells that lyse parasite-infected target cells. *J Immunol* 150, 1446-1457. (1993)

36. Wizel, B., M. Palmieri, C. Mendoza, B. Arana, J. Sidney, A. Sette & R. Tarleton: Human infection with *Trypanosoma cruzi* induces parasite antigen- specific cytotoxic T lymphocyte responses. *J Clin Invest* 102, 1062-1071 (1998)

37. Kumar, S. & R. L. Tarleton: Antigen-specific Th1 but not Th2 cells provide protection from lethal *Trypanosoma cruzi* infection in mice. *J Immunol* 166, 4596-4603 (2001)

38. Urbina, J. A.: Specific treatment of Chagas disease: current status and new developments. *Curr Opin Infect Dis* 14, 733-741. (2001)

39. Segura, M. A., E. Molina de Raspi & M. A. Basombrio: Reversibility of muscle and heart lesions in chronic, *Trypanosoma cruzi* infected mice after late trypanomicidal treatment. *Mem Inst Oswaldo Cruz* 89, 213-216. (1994)

40. Lauria-Pires, L., M. S. Braga, A. C. Vexenat, N. Nitz, A. Simoes-Barbosa, D. L. Tinoco & A. R. Teixeira: Progressive chronic Chagas heart disease ten years after treatment with anti-*Trypanosoma cruzi* nitroderivatives. *Am J Trop Med Hyg* 63, 111-118. (2000)

41. Bahia-Oliveira, L. M., J. A. Gomes, J. R. Cancado, T. C. Ferrari, E. M. Lemos, Z. M. Luz, M. C. Moreira, G. Gazzinelli & R. Correa-Oliveira: Immunological and clinical evaluation of chagasic patients subjected to chemotherapy during the acute phase of *Trypanosoma cruzi* infection 14- 30 years ago. *J Infect Dis* 182, 634-638. (2000)

42. Inglessis, I., H. A. Carrasco, N. Anez, C. Fuenmayor, H. Parada, J. A. Pacheco & H. R. Carrasco: [Clinical, parasitological and histopathologic follow-up studies of acute Chagas patients treated with benznidazole]. *Arch Inst Cardiol Mex* 68, 405-410. (1998)

43. Parada, H., H. A. Carrasco, N. Anez, C. Fuenmayor & I. Inglessis: Cardiac involvement is a constant finding in acute Chagas' disease: a clinical, parasitological and histopathological study. *Int J Cardiol* 60, 49-54. (1997)

44. Fabbro De Suasnabar, D., E. Arias, M. Streiger, M. Piacenza, M. Ingaramo, M. Del Barco & N. Amicone: Evolutive behavior towards cardiomyopathy of treated (nifurtimox or benznidazole) and untreated chronic chagasic patients. *Rev. Inst. Med. Trop. Sao Paulo* 42, 99-109. (2000)

45. Viotti, R., C. Vigliano, H. Armenti & E. Segura: Treatment of chronic Chagas' disease with benznidazole: clinical and serologic evolution of patients with long-term follow-up. *Am Heart J* 127, 151-162 (1994)

46. Cunha-Neto, E., M. Duranti, A. Gruber, B. Zingales, I. de Messias, N. Stolf, G. Bellotti, M. E. Patarroyo, F. Pilleggi & J. Kalil: Autoimmunity in Chagas' disease cardiomyopathy: biological relevance of a cardiac myosin-specific epitope crossreactive to an immunodominant *Trypanosoma cruzi* antigen. *Proc Natl Acad Sci USA* 92, 3541-3545 (1995)

47. Leon, J. S., L. M. Godsel, K. Wang & D. M. Engman: Cardiac myosin autoimmunity in acute Chagas heart disease. *Infect Immun* 69, 5643-5649 (2001)

48. Sterin-Borda, L. & E. Borda: Role of neurotransmitter autoantibodies in the pathogenesis of chagasic peripheral dysautonomia. *Ann NY Acad Sci* 917, 273-280 (2000)

49. Kerner, N., P. Liegeard, M. J. Levin & M. Hontebeyrie-Joskowicz: *Trypanosoma cruzi*: antibodies to a MAP-like protein in chronic Chagas' disease cross-react with mammalian cytoskeleton. *Exp Parasitol* 73, 451-459 (1991)

50. Van Voorhis, W. C., L. Schlekewy & H. L. Trong: Molecular mimicry by *Trypanosoma cruzi*: the Fl-160 epitope that mimics mammalian nerve can be mapped to a 12-amino acid peptide. *Proc Natl Acad Sci USA* 88, 5993-5997 (1991)

51. Mesri, E. A., G. Levitus, M. Hontebeyrie-Joskowicz, G. Dighiero, M. H. Van Regenmortel & M. J. Levin: Major *Trypanosoma cruzi* antigenic determinant in Chagas' heart disease shares homology with the systemic lupus erythematosus ribosomal P protein epitope. *J Clin Microbiol* 28, 1219-1224 (1990)

52. Levitus, G., M. Hontebeyrie-Joskowicz, M. H. Van Regenmortel & M. J. Levin: Humoral autoimmune response to ribosomal P proteins in chronic Chagas heart disease. *Clin Exp Immunol* 85, 413-417. (1991)

53. Girones, N., C. I. Rodriguez, E. Carrasco-Marin, R. F. Hernaez, J. L. de Rego & M. Fresno: Dominant T- and B-cell epitopes in an autoantigen linked to Chagas' disease. *J Clin Invest* 107, 985-993. (2001)

54. Ribeiro dos Santos, R., M. A. Rossi, J. L. Laus, J. S. Silva, W. Silvino & J. Mengels: Anti-CD4 abrogates rejection and reestablishes long-term tolerance to syngeneic newborn hearts grafted in mice chronically infected with *Trypanosoma cruzi. J Exp Med* 175, 29-39 (1992)

55. Mosca, W. & J. Plaja: Delayed hypersensitivity to heart antigens in Chagas' disease as measured by *in vitro* lymphocyte stimulation. *J Clin Microbiol* 14, 1-5. (1981)

56. Hontebeyrie-Joskowicz, M., G. Said, G. Milon, G. Marchal & H. Eisen: $L3T4^+$ T cells able to mediate parasite-specific delayed-type hypersensitivity play a role in the pathology of experimental Chagas' disease. *Eur J Immunol* 17, 1027-1033 (1987)

57. Cunha-Neto, E., V. Coelho, L. Guilherme, A. Fiorelli, N. Stolf & J. Kalil: Autoimmunity in Chagas' disease: identification of cardiac myosin-B13 *Trypanosoma cruzi* protein crossreactive T cell clones in heart lesions of a chronic Chagas' cardiomyopathy patient. *J Clin Invest* 98, 1709-1712 (1996)

58. Giordanengo, L., R. Fretes, H. Diaz, R. Cano, A. Bacile, E. Vottero-Cima & S. Gea: Cruzipain induces autoimmune response against skeletal muscle and tissue damage in mice. *Muscle Nerve* 23, 1407-1413 (2000)

59. Rizzo, L. V., E. Cunha-Neto & A. R. Teixeira: Autoimmunity in Chagas' disease: specific inhibition of reactivity of CD4⁺ T cells against myosin in mice chronically infected with *Trypanosoma cruzi*. *Infect Immun* 57, 2640-2644 (1989)

60. Girones, N., C. I. Rodriguez, B. Basso, J. M. Bellon, S. Resino, M. A. Munoz-Fernandez, S. Gea, E. Moretti & M. Fresno: Antibodies to an epitope from the Cha human autoantigen are markers of Chagas' disease. *Clin Diagn Lab Immunol* 8, 1039-1043. (2001)

61. Laguens, R. P., P. C. Meckert & J. G. Chambo: Antiheart antibody-dependent cytotoxicity in the sera from mice chronically infected with *Trypanosoma cruzi*. *Infect Immun* 56, 993-997 (1988)

62. Giordanengo, L., C. Maldonado, H. W. Rivarola, D. Iosa, N. Girones, M. Fresno & S. Gea: Induction of antibodies reactive to cardiac myosin and development of heart alterations in cruzipain-immunized mice and their offspring. *Eur J Immunol* 30, 3181-3189. (2000)

63. Laguens, R. P., P. M. Cabeza Meckert & J. G. Chambo: Immunologic studies on a murine model of Chagas disease. *Medicina (Buenos Aires)* 49, 197-202 (1989)

64. Acosta, A. M. & C. A. Santos-Buch: Autoimmune myocarditis induced by *Trypanosoma cruzi*. *Circulation* 71, 1255-1261 (1985)

65. Motran, C. C., R. E. Fretes, F. M. Cerban, H. W. Rivarola & E. Vottero de Cima: Immunization with the C-terminal region of *Trypanosoma cruzi* ribosomal P1 and P2 proteins induces long-term duration cross-reactive antibodies with heart functional and structural alterations in young and aged mice. *Clin Immunol* 97, 89-94. (2000)

66. Pontes-De-Carvalho, L., C. C. Santana, M. B. Soares, G. G. Oliveira, E. Cunha-Neto & R. Ribeiro-Dos-Santos: Experimental chronic Chagas' disease myocarditis is an autoimmune disease preventable by induction of immunological tolerance to myocardial antigens. *J Autoimmun* 18, 131-138. (2002)

67. Godsel, L. M., K. Wang, B. A. Schodin, J. S. Leon, S. D. Miller & D. M. Engman: Prevention of autoimmune myocarditis through the induction of antigen-specific peripheral immune tolerance. *Circulation* 103, 1709-1714 (2001)

68. Bocchi, E. A. & A. Fiorelli: The paradox of survival results after heart transplantation for cardiomyopathy caused by *Trypanosoma cruzi*. First Guidelines Group for Heart Transplantation of the Brazilian Society of Cardiology. *Ann Thorac Surg* 71, 1833-1838. (2001)

69. Granholm, N. A. & T. Cavallo: Autoimmunity, polyclonal B-cell activation and infection. *Lupus* 1, 63-74. (1992)

70. Minoprio, P.: Parasite polyclonal activators: new targets for vaccination approaches? *Int J Parasitol* 31, 588-591 (2001)

71. Lanzavecchia, A.: How can cryptic epitopes trigger autoimmunity? *J Exp Med* 181, 1945-1948. (1995)

72. York, I. A., A. L. Goldberg, X. Y. Mo & K. L. Rock: Proteolysis and class I major histocompatibility complex antigen presentation. *Immunol Rev* 172, 49-66 (1999)

73. Warnock, M. G. & J. A. Goodacre: Cryptic T-cell epitopes and their role in the pathogenesis of autoimmune diseases. *Br J Rheumatol* 36, 1144-1150. (1997)

74. de Scheerder, I. K., M. L. de Buyzere, J. R. Delanghe, D. L. Clement & R. J. Wieme: Anti-myosin humoral immune response following cardiac injury. *Autoimmunity* 4, 51-58 (1989)

75. Fedoseyeva, E. V., F. Zhang, P. L. Orr, D. Levin, H. J. Buncke & G. Benichou: De novo autoimmunity to cardiac myosin after heart transplantation and its contribution to the rejection process. *J Immunol* 162, 6836-6842 (1999)

76. Neu, N., S. W. Craig, N. R. Rose, F. Alvarez & K. W. Beisel: Coxsackievirus induced myocarditis in mice: cardiac myosin autoantibodies do not cross-react with the virus. *Clin Exp Immunol* 69, 566-574 (1987)

77. Vanderlugt, C. L. & S. D. Miller: Epitope spreading in immune-mediated diseases: implications for immunotherapy. *Nat Rev Immunol* 2, 85-95. (2002)

78. Grauert, M. R., M. Houdayer & M. Hontebeyrie-Joskowicz: *Trypanosoma cruzi* infection enhances polyreactive antibody response in an acute case of human Chagas' disease. *Clin Exp Immunol* 93, 85-92 (1993)

79. Ternynck, T., C. Bleux, J. Gregoire, S. Avrameas & C. Kanellopoulos-Langevin: Comparison between autoantibodies arising during *Trypanosoma cruzi* infection in mice and natural autoantibodies. *J Immunol* 144, 1504-1511 (1990)

80. Tibbetts, R. S., T. S. McCormick, E. C. Rowland, S. D. Miller & D. M. Engman: Cardiac antigen-specific autoantibody production is associated with cardiomyopathy in *Trypanosoma cruzi*-infected mice. *J Immunol* 152, 1493-1499 (1994)

81. Rowland, E., H. Luo & T. McCormick: Infection characteristics of an Ecuadorian *Trypanosoma cruzi* strain with reduced virulence. *J Parasitol* 81, 123-126 (1995)

Key Words: Heart, Infection, Parasite, Myocarditis, Autoimmunity, Chagas Disease, *Trypanosoma cruzi*, Review

Send correspondence to: David M. Engman, Feinberg Medical School of Northwestern University, Department of Pathology, Ward Building 6-175, 303 E. Chicago Ave., Chicago, IL 60611, Tel: 312-503-1288, Fax: 312-503-1265, email: d-engman@northwestern.edu