Effect of sulfated glycosaminoglycans on tumor invasion and metastasis

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TABLE OF CONTENTS

- 1. Abstract
- 2. Introduction
- 3. Glycosaminoglycans and cell proliferation
- 4. Glycosaminoglycans and tumor invasion
- 5. Glycosaminoglycans and hematogenous metastasis
- 6. Therapeutic use of glycosaminoglycans from marine invertebrates in cancer and metastasis
- 7. Acknowledgements
- 9. Reference

1. ABSTRACT

Metastasis is the most devastating aspect of the tumor, being the main cause of morbidity and mortality in cancer patients. The events that lead to tumor invasion and metastasis depend fundamentally on the initial aquisition of a mesenchymal phenotype by the primary carcinoma, which provides the necessary machinary for invasion, intravasation, vascular transport, extravasation and tumor colonization. These events are orguestrated by different growth factors, proteoglycans and adhesion molecules, acting at the surface of cells. The anticoagulant heparin binds several of these molecules and can regulate the interactions that occur during tumor invasion and metastasis. For example, heparin modulates the binding of FGF-2 to its tyrosine kinase receptor during tumor proliferation, and the binding of growth factors involved in epithelial to mesenchymal transition during tumor invasion. It also binds P-selectin on activated platelets, preventing tumor cell-platelet interaction during hematogeneous metastasis. In this review, we discuss the role of sulfated glycosaminoglycans during tumor invasion and metastasis, and the possible therapeutic use of heparin analogs on cancer treatment.

2. INTRODUCTION

In nature, polysaccharides are produced as highly functional polymers with magnificent structural diversity. Sulfated polysaccharides abound in the Animal Kingdom and the glycosaminoglycans are one of the most abundant and best studied sulfated polysaccharides in vertebrates. Specific structural motifs in the chain enable these glycans to interact with several biomolecules involving in distinct biological roles. Recent advances in developmental biology and cancer biology have revealed numerous pivotal roles of these polymers in diverse biological processes, such as regulatory functions in development (1-3), axonal growth (4-6), angiogenesis (7-9), microbial pathogenesis (10, 11) and anticoagulation (12-14). Heparin is the best known glycosaminoglycan and its anticoagulant activity was the earliest pharmacological effect attributed to these molecules (15). However, several reports have shown highly relevant effects of heparin in other processes such as inflammation (16-18) and metastasis (19-25), angiogenesis (26, 27) and cell-cell adhesion (28). Although most of the work published focus on the effects of heparin or heparan sulfate, chondroitin sulfate and dermatan sulfate have also been shown to have similar effects (29-31). Here, we

discuss the effects of sulfated glycosaminoglycans in cancer, focusing on invasion and metastasis of tumor cells.

According to the WHO (World Health Organization), cancer is the leading cause of death in all parts of the world (32) and metastasis is the most devastating aspect of the disease. "Metastasis" is a Greek word meaning "changing the place". The expression "tumor metastasis" was first used in 1829 by the French physician Joseph Claude Recamier in his treatise "Recherches du Cancer", referring to the movement of cells from a primary tumor to distant organs. Metastasis is a complex multistep process in which tumor cells acquire the ability of colonizing distant sites. Although the primary tumor can release millions of cells per day into the circulation (33), only ~0.01% of the cells that enter the circulation have the potential to form secondary tumors, and are called cancer initiating cells (34, 35). The cascade of events involved in metastasis includes: (1) the proliferation of transformed (malignant) cells to form a small tumor mass, which can be supported by simple diffusion of nutrients; (2) the induction of its own vascularization (angiogenesis); (3) detachment of tumor cells from the tumor and invasion through the basement membrane or to the surrounding stroma; (4) entering of cells into the blood circulation; (5) evasion from the immune system; (6) arrest of tumor cells in the capillary beds of distant organs; (7) invasion of the surrounding parenchyma and (8) proliferation of cells to form a tumor mass in a secondary organ.

Recent advances on cancer glycobiology have raised increasing interest in the role of several glycoconjugates on different steps of the metastatic cascade. Sulfated glycosaminoglycans, such as heparin, heparan sulfate, dermatan sulfate and chondroitin sulfate have been widely studied on different molecular events of tumor dissemination, such as proliferation (36), recruitment of leukocytes (37), invasion (38), and survival in blood circulation (21). The relevance of glycosaminoglycans in each one of those mechanisms is discussed below.

3. GLYCOSAMINOGLYCANS AND CELL PROLIFERATION

One of the hallmarks of cancer cells is their uncontrolled proliferation. Multiples genes and pathways are differently regulated, resulting in an altered cell cycle. In general, competent growth factors such as PDGF (Platelet-Derived Growth Factor) and FGF (Fibroblast Growth Factor) (39, 40), as well as EGF (Epidermal Growth Factor) and IGF-1 (Insulin-like Growth Factor) (41, 42) are required for a quiescent cell to enter in the G1 phase. The growth factor-growth factor receptor signaling usually culminates in the modulation of cyclins, specifically those from the cyclin D family that induce the phosphorylation of retinoblastoma protein (RbP), leading the cell to enter into the S-phase (43-45). Some of the signaling cascades initiated by growth factors are modulated by proteoglycans and/or glycosaminoglycans. Several studies have shown an increased expression of the proteoglycan decorin gene during the quiescence of cells in

organogenesis and tissue differentiation (46, 47). For example, De Luca et al. (1996) observed an arrest of decorin-expressing tumor cells at the G1 phase that concomitantly expressed the cyclin-dependent kinase inhibitor p21_{waf1} (48). It has also been shown that decorin directly interacts with Epidermal Growth Factor Receptor (EGFR) evoking a profound downregulation of the receptor, and inhibition of its downstream signaling activity (49-51). Albeit the signaling effect of decorin is attributed to the core protein, dermatan sulfate chains of decorin have been shown to interact with Transforming Growth Factor-b (TGF-b) (52) and FGF-2, acting as a co-receptor, and inducing cell mitogenic signaling (53). In melanoma, chondroitin sulfate proteoglycans control cell proliferation by enhancing focal adhesion kinase and activating the extracellular signal-regulated kinase (54).

Membrane heparan sulfate proteoglycans (HSPGs), such as Syndecans are classic mediators of cell growth and differentiation, acting by interacting and modulating different growth factors activities. Extracellular HSPGs can also bind to soluble ligands, increasing their local concentration and modulating ligand-receptors encounters (55). HSPGs markedly enhance signaling of FGF-2 and Hepatocyte Growth Factor (HGF) (see (56) for review). In particular, FGF-2 completely depends on heparan sulfate to transduce an intracellular signal through the formation of ternary complex HSPG-FGF-2-FGFR (57-60). Due to the HS-dependence of those growth factors, heparin is likely to be a good tool for inhibiting the growth factor-induced cell signaling. In fact, several works have shown that heparin shows anti-proliferative effects, acting as a competitor for HSPGs on the cell surface. Although the anti-cancer effects of heparin have been documented for decades (8, 21, 22, 61-63), the mechanisms involved in cell proliferation and tumor growth still need to be fully understood.

Apoptosis is a key process in cancer development and progression (64, 65). The ability of cancer cells to escape apoptosis and continue to proliferate is a hallmark of tumor cell and one of the major targets in cancer therapy (65). Sulfated glycosaminoglycans actively participate in the regulation of the apoptotic process (66, 67) by interacting with different growth factors. For instance, expression of glypican-3 is frequently silenced in mesotheliomas, ovarian and breast cancer (68-70). Reexpression of glypican-3 in tumor mammary cells increased the susceptibility to apoptosis, by modulation of Wnt, PKB/Akt and p38MAPK (71, 72). Modifications on glycosaminoglycans chains are likely to be involved in different susceptibility of cancer cells to apoptosis. Sulfatase-2 (SULF2), an extracellular 6-O-endosulfatase, which modifies the sulfation pattern of extracellular heparan sulfate chais, induces an increase in phosphorylation of the anti-apoptotic Akt kinase substrate GSK3, leading to a reduction of the apoptotic index in human hepatocellular carcinoma cells. (73).

Syndecan-1 has been pointed as a powerful tumor suppressor. Syndecan-1 sheded from cell surface acts by reducing cell growth and inducing apoptosis of

myeloma cells in vitro (74). Morover, the expression of Syndecan-1 on myeloma cells is associated with a reduction of mortality in a mouse model of tumor (75). In tumors of epithelial cell lineage, syndecan-1 is usually downregulated, contributing to progression of malignancy. In contrast, overexpression of syndecan-1 is also related to unfavorable phenotypes in ovarian, breast, pancreatic and endometrial cancer (76-78). However, how syndecan-1 effectively affects tumor progression is still unknown. Recent work has shown that syndecan-1 ectodomain was effective in inducing apoptosis and inhibiting the growth of low expressing syndecan-1 prostate cells, through a PDK1/Akt/Bad signaling pathway (79). Analogs of syndecan-1 ectodomain, designed by conjugation of glycosaminoglycan chains with carbodiimide, are shown to inhibit myeloma and breast cancer cells viability by inducing apoptosis, without any apparent toxicity to the adjacent normal tissue (80). Unfractionated heparin was also shown to induce cell apoptosis, affecting cell growth in four oral squamous cell carcinoma cell lines, through suppression of Akt pathway (81). The low molecular weight heparin, dalteparin, was shown to induce lung adenocarcinoma A549 cells to early apoptosis (82). Despite interesting results, the lack of mechanisms to explain the effects of exogenous glycosaminoglycans and some contradictory data, since heparin did not affect apoptosis in several colon carcinoma cells (83, 84) and glioma (85), makes the use of glycosaminoglycan as antiapoptotic drugs a subject of discussion.

5. GLYCOSAMINOGLYCANS AND TUMOR INVASION

The ability of cancer cells to metastasize first depends on their ability to overcome local adhesive forces, migrate and invade the surrounding tissue. Cell migration is a result of increased cell motility, a highly coordinated multifactorial process, requiring continuous formation and disassembly of matrix adhesions (86-89). About 80% of the cancers are carcinomas that acquire a migratory phenotype through the epithelial-to-mesenchymal transition (EMT) program (90) (Figure 1A). EMT is a complex process whereby polarized epithelial cells transform into apolar fibroblastoid-like cells. A reduction of cell-cell adherence via a switching of cadherins (down regulation of Ecadherin and up regulation of N-cadherin) and acquisition of migratory capability are important changes observed upon EMT. In 2001, Zajchowski et al. (91) found that vimentin and other mesenchymal gene products, such as integrin a3, osteonectin, thrombospondin-1 and collagen I and VI were part of the gene signature predicting invasiveness of human breast cancer cells. Several well known invasive cell lines, such as BT-549, Hs578T, MDA-MB-157, MDA-MB-231, MDA-MB-435, MDA-MB-436, posses a basal/mesenchymal genotype (92). The activation of transcription factors leading to initiation of EMT and consequently metastasis may be triggered by many extraand intracellular signals. The "scatter factor" or HGF was the first factor observed to induce EMT (93, 94) and since then, many growth factors such as EGF, VEGF, TGF-b, Wnt, SDF and PGE2 have been shown to induce EMT in vitro (95). The binding of some growth factors to their

receptors on cell surface usually requires the presence of cell surface proteoglycans carrying heparan sulfate and/or chondroitin sulfate (HSPG and/or CSPG) (96, 97). It has been shown that either HS/heparin or dermatan sulfate bind to HGF with high affinity and play role as cofactors in vitro. Although the glycosaminoglycan region that is recognized by the growth factor remains to be elucidated, it has recently been shown that a tri-saccharide containing a single iduronic acid, flanked by monosulfated hexosamines, is enough to bind to HGF with high affinity (98). Dermatan sulfates isolated from ascidians, primitive chordate marine invertebrates have been shown to bind HGF and regulates its activity, enhancing cell signaling (99, 100). HGF stimulates skeletal muscle proliferation, differentiation and migration in synergy with FGF-2, in a DS-depending manner (101). Thus, it is possible that DSs and other glycosaminoglycans could influence the EMT process (Figure 1B). However, how dermatan sulfate and other glycosaminoglycans are involved in cancer-related signaling pathways still requires extensive investigation.

Invasion of tumor cells into surrounding tissues requires not only an increased mobility, but also a "degrading activity" on the extracellular matrix to allow migration of the cells. This activity is provided by secretion of degrading enzymes, such as matrix metaloproteases (MMPs) and heparanases (102, 103). Heparanase is a HSdegrading endoglycosidase preferentially expressed in human tumors. Its overexpression is strongly associated with an invasive phenotype and highly correlated with the metastatic potential of cancer cells (104). Heparanase cleaves HS at low sulfated regions, contributing to structural changes of the extracellular matrix and basement membrane underling epithelial and endothelial cells, which is fundamental for the invasion of cancer cells. HS fragments generated by heparanase cleavage are usually still of appreciable size and can act as signaling molecules for cancer and stromal cells, enhancing proliferation and migration (105-107). Due to the important role of heparanase in tumor invasion and metastasis, a variety of inhibitory molecules have been developed, including neutralizing antibodies, modified species of heparin (108, 109) as well as many polyanionic molecules, such as suramin and laminaran (110, 111).

6. GLYCOSAMINOGLYCANS AND HEMATOGENOUS METASTASIS

The surface of carcinoma cells shows altered glycosylation patterns (112-114), expressing highly branched or sialylated oligosaccharides, especially fucosylated glycans containing sialyl-Lewis^X and sialyl-Lewis^a. The presence of these oligosaccharides in tumor cells is directly associated with a poor prognosis because of tumor progression and metastatic spread (112) (113) (115). The sialyl-Lewis^X-oligosaccharides from the carcinoma cells are ligands of the 3 members of the selectin family of cell adhesion molecules. E-, P- and L-selectins are vascular receptors for certain normal glycoproteins that contain Sialyl-Lewis^{x,a} found on leucocytes and endothelium (116). Therefore, by mediating the interactions of tumor cells with

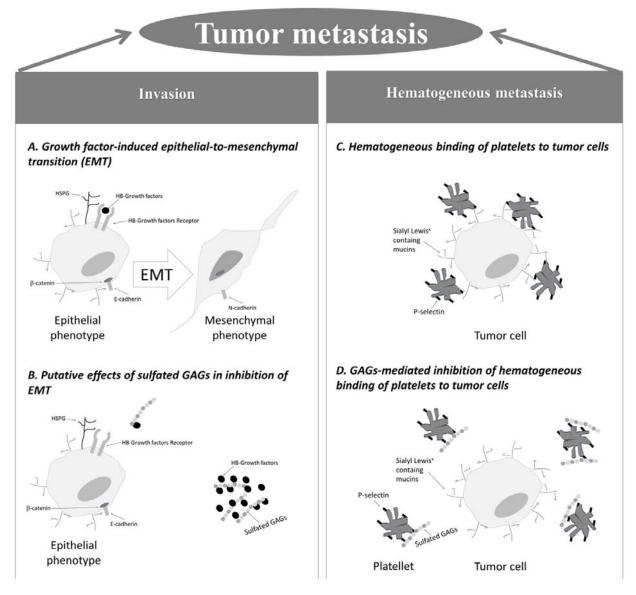


Figure 1. Putative effects of sulfated glycosaminoglycans on tumor metastasis.

platelets and endothelium, the selectins participate in metastatic spread of tumor (Figure 1C) (112) (113) (115).

Studies from several groups have indicated that tumor metastasis in experimental animals can be inhibited by heparin (117) (118) (119). Some clinical studies have also shown a benefic effect of heparin in some types of human cancer (120) (121) (122). However, the antimetastatic effect of heparin is not related to its anticoagulant action (117) (123), but to the heparin's ability to inhibit the interaction of tumor cells with platelets (Figure 1D). Heparin inhibits the binding of P- and L-selectin to their natural ligands (sialyl Lewis^{x,a}-rich oligosaccharides) (124). The inhibition of L- and P-selectin requires the presence of 6-O-sulfated glucosamine residues in the heparin molecule (125). Therefore, the heparin-mediated mechanism of metastasis attenuation involves the inhibition of the interaction of sialyl-Lewis^{x,a}-rich

oligosaccharides on the surface of tumor cells and Pselectin on platelets (21). In the presence of heparin, tumor cells loose the protection conferred by platelets becoming susceptible to the potentially cytotoxic action of immune effector cells, which leads to the inhibition of metastasis. A single intravascular injection of heparin promotes the immediate attenuation of the interaction of tumor cell-platelet, with a marked reduction of metastasis 6 weeks after the initial steps of the metastatic cascade. The antimetastatic activity of heparin could not be demonstrated in P- or L-selectin-deficient mice, indicating the involvement of selectins in the antimetastatic activity of heparin (22). Although heparin might influence several steps of metastatic cascade, it is clear that the ability of heparin in inhibiting the binding of tumor cells to Pselectin can explain the clinical evidence that administration of heparin to cancer patients might influence survival (126).

Table 1. Anti-tumor activity of invertebrate glycosaminoglycans

Phylum	Family	Species	Gag (Disaccharide units)	Biological Activity	References
Echinodermata	Holothuridae	Holothuria grisea	oversulfated CS	anti P- and L-selectin inhibitory activity;	(18)
		(Ludwigothurea grisea)	(GlcA-GalNAc4S,6S) (GlcA3S	antimetastatic activity;	
			residues)		
			(sulfated fucose branches)		
Chordata	Ascidiidae	Phallusia nigra	oversulfated DS	HGF-SF binding activity;	(13, 127),
		(Ascida nigra)	(IdoA2S-GalNAc6S) (80%)	anti P- and L-selectin inhibitory activity;	(110)
			(IdoA-GalNAc6S)	antimetastatic activity	
		Styela plicata	oversulfated DS	HGF-SF binding activity	(13, (13)
			(IdoA2S-GalNAc4S) (70%)	anti P- and L-selectin inhibitory activity;	(110)
			(IdoA-GalNAc4S)	antimetastatic activity	

However, due to its high hemorrhagic risk the therapeutic use of heparin is still restricted. In addition, it has been reported that the dose required for P-selectin inhibition is higher than that required to achieve anticoagulation, which increases the chances of bleeding and prevents the clinical used of heparin to treat tumor metastasis. The use of low molecular weight heparin, which has a lower bleeding effect, is not an adequate alternative due to its weak inhibitory effect on the selectins. Our laboratory isolated and characterized various heparin analogs from marine invertebrates (Table 1). Several of these compounds, such as dermatan sulfates with different degrees of sulfation from the ascidians Styela plicata and Ascidia nigra (13) (127) and a fucosylated chondroitin sulfate from the sea cucumber Ludwiguthurea grisea (18) possess a high anti-selectin activity. Initial studies indicated that, after intravenous administration to animals, these compounds were able to inhibit P- and L-selectin-mediated events, such as inflammation and tumor metastasis (18), without producing significant bleeding. Therefore, the invertebrate glycans represent a potential therapeutic alternative to mammalian heparin to treat inflammation and tumor metastasis. More recently, the fucosylated chondroitin sulfate was shown to be absorbed after oral administration in rats, which is a great advantage in comparison to heparin, that needs to be administrated intravenous or subcutaneously (128). The antimetastatic effect of orally administrated fucosylated chondroitin sulfate remains to be tested.

7. THERAPEUTIC USE OF GLYCOSAMINOGLYCANS FROM MARINE INVERTEBRATES IN CANCER AND METASTASIS

Metastatic disease is responsible for most cancerassociated deaths and EMT along with hematogeneous metastasis, are critical steps in cancer progression. Therefore, inhibition of these events could be an effective approach to reduce the metastatic disease. Heparin has been show to modulate EMT-associated growth factors and also to inhibit P-selectin, leading to attenuation of metastasis. Previously, we showed that unique dermatan sulfates from the ascidians Styela plicata and Pallusia nigra (Table 1), by 2,4-O-sulfated and 2,6-O-sulfated composed disaccharide units, respectively, bind with high affinity (KD of ~ 33nM) to the EMT-related growth factor, HGF, modulating its MET-dependent intracellular signaling (99, 100). More recently, we demonstrated that the 2,6disulfated dermatan sulfate from P.nigra inhibits the TGFβ-mediated migration of human mammary cells in a wound-healing cell migration assay (personal observation,

not published) and also the binding of tumor cells to Pselectin, with IC₅₀ value 2-fold lower than that of unfractionated heparin (129). To investigate the effect of the ascidian DSs in vivo, mice were injected with LS180 cells 10 minutes after treatment with PBS, UFH (1 mg/mouse) or ascidian dermatan sulfate (100 μg/mouse) and the presence of platelets-tumor cells aggregates in the lung microvasculature was evaluated. The ascidian DSs inhibited the adhesion of platelets to tumor cells in vivo in a concentration 10-fold lower than heparin. Metastasis in mice was also investigated in a long-term metastasis model, which showed that both ascidian DSs drastically reduced metastasis of murine colon carcinoma cells. Such effects were shown to be P-selectindependent. In conclusion, ascidian DSs can inhibit cell migration and reduce hematogeneous metastasis by EMTand P-selectin mediated events and could be used therapeutically to prevent tumor invasion and metastasis.

Similar to the ascidian dermatan sulfate, several heparin analogs obtained from different marine invertebrate sources were described (for review, see (130)). Some of these compounds have been extensively studied in terms of structure, biological activity and mechanism of action, and evaluated in pre-clinical experiments in rodent animals with promising results. The most critical question related to therapeutics from natural sources is the technical and economic possibility to obtain very large quantities of the compounds in a constant and ecologically correct manner. Most of the invertebrate glycans can be obtained at a reasonable yield (about 0.5% of the dry weight, comparing to 0.022 % from pig intestinal mucosa (125), by procedures similar to those already employed in the preparation of pharmaceutical heparin. Several species of mollusks and sea cucumbers, including those containing high quantities of heparin analogs, have been successfully cultivated in different parts of the world (131, 132). Therefore, the conditions required to use marine invertebrates as source of natural therapeutic compounds have already been established. What is necessary now is a cooperative effort from scientists of related areas to specifically adapt and optimize current methodologies.

8. ACKNOWLEDGMENTS

EOK and MSGP contributed equally to this work. MSGP is a research fellow from CNPq and is supported by grants from Mizutani Foundation for Glycoscience, CNPq and FAPERJ. EOK is supported by a PhD Fellowship from CNPq.

9. REFERENCE

- 1. S. Mizuguchi, T. Uyama, H. Kitagawa, K. H. Nomura, K. Dejima, K. Gengyo-Ando, S. Mitani, K. Sugahara and K. Nomura: Chondroitin proteoglycans are involved in cell division of Caenorhabditis elegans. *Nature*, 423(6938), 443-8 (2003)
- 2. G. K. Dhoot, M. K. Gustafsson, X. Ai, W. Sun, D. M. Standiford and C. P. Emerson, Jr.: Regulation of Wnt signaling and embryo patterning by an extracellular sulfatase. *Science*, 293(5535), 1663-6 (2001)
- 3. U. Hacker, K. Nybakken and N. Perrimon: Heparan sulphate proteoglycans: the sweet side of development. *Nat Rev Mol Cell Biol*, 6(7), 530-41 (2005)
- 4. C. H. Chau, D. K. Shum, H. Li, J. Pei, Y. Y. Lui, L. Wirthlin, Y. S. Chan and X. M. Xu: Chondroitinase ABC enhances axonal regrowth through Schwann cell-seeded guidance channels after spinal cord injury. *FASEB J*, 18(1), 194-6 (2004)
- 5. E. J. Bradbury, L. D. Moon, R. J. Popat, V. R. King, G. S. Bennett, P. N. Patel, J. W. Fawcett and S. B. McMahon: Chondroitinase ABC promotes functional recovery after spinal cord injury. *Nature*, 416(6881), 636-40 (2002)
- 6. D. Carulli, T. Laabs, H. M. Geller and J. W. Fawcett: Chondroitin sulfate proteoglycans in neural development and regeneration. *Curr Opin Neurobiol*, 15(1), 116-20 (2005)
- 7. I. Vlodavsky, O. Goldshmidt, E. Zcharia, R. Atzmon, Z. Rangini-Guatta, M. Elkin, T. Peretz and Y. Friedmann: Mammalian heparanase: involvement in cancer metastasis, angiogenesis and normal development. *Semin Cancer Biol*, 12(2), 121-9 (2002)
- 8. B. Casu, M. Guerrini, A. Naggi, M. Perez, G. Torri, D. Ribatti, P. Carminati, G. Giannini, S. Penco, C. Pisano, M. Belleri, M. Rusnati and M. Presta: Short heparin sequences spaced by glycol-split uronate residues are antagonists of fibroblast growth factor 2 and angiogenesis inhibitors. *Biochemistry*, 41(33), 10519-28 (2002)
- 9. R. V. Iozzo: Basement membrane proteoglycans: from cellar to ceiling. *Nat Rev Mol Cell Biol*, 6(8), 646-56 (2005)
- 10. J. Liu, Z. Shriver, R. M. Pope, S. C. Thorp, M. B. Duncan, R. J. Copeland, C. S. Raska, K. Yoshida, R. J. Eisenberg, G. Cohen, R. J. Linhardt and R. Sasisekharan: Characterization of a heparan sulfate octasaccharide that binds to herpes simplex virus type 1 glycoprotein D. *J Biol Chem*, 277(36), 33456-67 (2002)
- 11. K. Mardberg, E. Trybala, F. Tufaro and T. Bergstrom: Herpes simplex virus type 1 glycoprotein C is necessary for efficient infection of chondroitin sulfate-expressing gro2C cells. *J Gen Virol*, 83(Pt 2), 291-300 (2002)
- 12. D. M. Tollefsen: Vascular dermatan sulfate and heparin cofactor II. *Prog Mol Biol Transl Sci*, 93, 351-72 (2010)

- 13. M. S. Pavao, P. A. Mourao, B. Mulloy and D. M. Tollefsen: A unique dermatan sulfate-like glycosaminoglycan from ascidian. Its structure and the effect of its unusual sulfation pattern on anticoagulant activity. *J Biol Chem*, 270(52), 31027-36 (1995)
- 14. U. Lindahl, M. Kusche, K. Lidholt and L. G. Oscarsson: Biosynthesis of heparin and heparan sulfate. *Ann NY Acad Sci*, 556, 36-50 (1989)
- 15. D. L. Rabenstein: Heparin and heparan sulfate: structure and function. *Nat Prod Rep*, 19(3), 312-31 (2002)
- 16. N. M. Melo-Filho, C. L. Belmiro, R. G. Goncalves, C. M. Takiya, M. Leite, Jr., M. S. Pavao and P. A. Mourao: Fucosylated chondroitin sulfate attenuates renal fibrosis in animals submitted to unilateral ureteral obstruction: a P-selectin-mediated event? *Am J Physiol Renal Physiol*, 299(6), F1299-307 (2010)
- 17. C. L. Belmiro, M. T. Castelo-Branco, L. M. Melim, A. Schanaider, C. Elia, K. Madi, M. S. Pavao and H. S. de Souza: Unfractionated heparin and new heparin analogues from ascidians (chordate-tunicate) ameliorate colitis in rats. *J Biol Chem*, 284(17), 11267-78 (2009)
- 18. L. Borsig, L. Wang, M. C. Cavalcante, L. Cardilo-Reis, P. L. Ferreira, P. A. Mourao, J. D. Esko and M. S. Pavao: Selectin blocking activity of a fucosylated chondroitin sulfate glycosaminoglycan from sea cucumber. Effect on tumor metastasis and neutrophil recruitment. *J Biol Chem*, 282(20), 14984-91 (2007)
- 19. M. Belting, L. Borsig, M. M. Fuster, J. R. Brown, L. Persson, L. A. Fransson and J. D. Esko: Tumor attenuation by combined heparan sulfate and polyamine depletion. *Proc Natl Acad Sci U S A*, 99(1), 371-6 (2002)
- 20. L. Borsig: Antimetastatic activities of modified heparins: selectin inhibition by heparin attenuates metastasis. *Semin Thromb Hemost*, 33(5), 540-6 (2007)
- 21. L. Borsig, R. Wong, J. Feramisco, D. R. Nadeau, N. M. Varki and A. Varki: Heparin and cancer revisited: mechanistic connections involving platelets, P-selectin, carcinoma mucins, and tumor metastasis. *Proc Natl Acad Sci U S A*, 98(6), 3352-7 (2001)
- 22. N. Hostettler, A. Naggi, G. Torri, R. Ishai-Michaeli, B. Casu, I. Vlodavsky and L. Borsig: P-selectin- and heparanase-dependent antimetastatic activity of non-anticoagulant heparins. *FASEB J*, 21(13), 3562-72 (2007)
- 23. H. Laubli and L. Borsig: Heparins attenuate cancer metastasis: are selectins the link? *Cancer Invest*, 27(5), 474-81 (2009)
- 24. D. Liu, Z. Shriver, Y. Qi, G. Venkataraman and R. Sasisekharan: Dynamic regulation of tumor growth and metastasis by heparan sulfate glycosaminoglycans. *Semin Thromb Hemost*, 28(1), 67-78 (2002)

- 25. J. M. Trowbridge and R. L. Gallo: Dermatan sulfate: new functions from an old glycosaminoglycan. *Glycobiology*, 12(9), 117R-25R (2002)
- 26. S. A. Mousa and L. J. Petersen: Anti-cancer properties of low-molecular-weight heparin: preclinical evidence. *Thromb Haemost*, 102(2), 258-67 (2009)
- 27. M. Presta, D. Leali, H. Stabile, R. Ronca, M. Camozzi, L. Coco, E. Moroni, S. Liekens and M. Rusnati: Heparin derivatives as angiogenesis inhibitors. *Curr Pharm Des*, 9(7), 553-66 (2003)
- 28. S. Miyamoto, H. Yagi, F. Yotsumoto, S. Horiuchi, T. Yoshizato, T. Kawarabayashi, M. Kuroki and E. Mekada: New approach to cancer therapy: heparin binding-epidermal growth factor-like growth factor as a novel targeting molecule. *Anticancer Res*, 27(6A), 3713-21 (2007)
- 29. G. M. Smith and C. Strunz: Growth factor and cytokine regulation of chondroitin sulfate proteoglycans by astrocytes. *Glia*, 52(3), 209-18 (2005)
- 30. J. Tapon-Bretaudiere, D. Chabut, M. Zierer, S. Matou, D. Helley, A. Bros, P. A. Mourao and A. M. Fischer: A fucosylated chondroitin sulfate from echinoderm modulates *in vitro* fibroblast growth factor 2-dependent angiogenesis. *Mol Cancer Res.* 1(2), 96-102 (2002)
- 31. K. R. Taylor and R. L. Gallo: Glycosaminoglycans and their proteoglycans: host-associated molecular patterns for initiation and modulation of inflammation. *FASEB J*, 20(1), 9-22 (2006)
- 32. D. F. Zech, S. Grond, J. Lynch, D. Hertel and K. A. Lehmann: Validation of World Health Organization Guidelines for cancer pain relief: a 10-year prospective study. *Pain*, 63(1), 65-76 (1995)
- 33. L. Weiss, E. Grundmann, J. Torhorst, F. Hartveit, I. Moberg, M. Eder, C. M. Fenoglio-Preiser, J. Napier, C. H. Horne, M. J. Lopez and *et al.*: Haematogenous metastatic patterns in colonic carcinoma: an analysis of 1541 necropsies. *J Pathol*, 150(3), 195-203 (1986)
- 34. M. L. Butler, R. L. Van Heertum and S. K. Teplick: Metastatic malignant melanoma of the esophagus: a case report. *Gastroenterology*, 69(6), 1334-7 (1975)
- 35. E. Mayhew and D. Glaves: Quantitation of tumorigenic disseminating and arrested cancer cells. *Br J Cancer*, 50(2), 159-66 (1984)
- 36. C. Mundhenke, K. Meyer, S. Drew and A. Friedl: Heparan sulfate proteoglycans as regulators of fibroblast growth factor-2 receptor binding in breast carcinomas. *Am J Pathol*, 160(1), 185-94 (2002)
- 37. K. Zen, D. Q. Liu, L. M. Li, C. X. Chen, Y. L. Guo, B. Ha, X. Chen, C. Y. Zhang and Y. Liu: The heparan sulfate proteoglycan form of epithelial CD44v3 serves as a

- CD11b/CD18 counter-receptor during polymorphonuclear leukocyte transepithelial migration. *J Biol Chem*, 284(6), 3768-76 (2009)
- 38. B. U. Pauli, D. E. Schwartz, E. J. Thonar and K. E. Kuettner: Tumor invasion and host extracellular matrix. *Cancer Metastasis Rev*, 2(2), 129-52 (1983)
- 39. E. Oleszak: Inhibition of mitogenic activity of PDGF, EGF, and FGF by interferon-gamma. *Exp Cell Res*, 179(2), 575-80 (1988)
- 40. S. Higgins, S. H. Wong, M. Richner, C. L. Rowe, D. F. Newgreen, G. A. Werther and V. C. Russo: Fibroblast growth factor 2 reactivates G1 checkpoint in SK-N-MC cells via regulation of p21, inhibitor of differentiation genes (Id1-3), and epithelium-mesenchyme transition-like events. *Endocrinology*, 150(9), 4044-55 (2009)
- 41. Y. C. Yeh, E. R. Burns, J. Yeh and H. W. Yeh: Synergistic effects of endothelin-1 (ET-1) and transforming growth factor alpha (TGF-alpha) or epidermal growth factor (EGF) on DNA replication and G1 to S phase transition. *Biosci Rep.*, 11(3), 171-80 (1991)
- 42. D. R. Clemmons and L. I. Gardner: A factor contained in plasma is required for IGF binding protein-1 to potentiate the effect of IGF-I on smooth muscle cell DNA synthesis. *J Cell Physiol*, 145(1), 129-35 (1990)
- 43. K. V. Ramana, R. Tammali and S. K. Srivastava: Inhibition of aldose reductase prevents growth factor-induced G1-S phase transition through the AKT/phosphoinositide 3-kinase/E2F-1 pathway in human colon cancer cells. *Mol Cancer Ther*, 9(4), 813-24 (2010)
- 44. C. S. Bryant, S. Kumar, S. Chamala, J. Shah, J. Pal, M. Haider, S. Seward, A. M. Qazi, R. Morris, A. Semaan, M. A. Shammas, C. Steffes, R. B. Potti, M. Prasad, D. W. Weaver and R. B. Batchu: Sulforaphane induces cell cycle arrest by protecting RB-E2F-1 complex in epithelial ovarian cancer cells. *Mol Cancer*, 9, 47 (2010)
- 45. S. Elangovan, T. C. Hsieh and J. M. Wu: Growth inhibition of human MDA-mB-231 breast cancer cells by delta-tocotrienol is associated with loss of cyclin D1/CDK4 expression and accompanying changes in the state of phosphorylation of the retinoblastoma tumor suppressor gene product. *Anticancer Res*, 28(5A), 2641-7 (2008)
- 46. A. Mauviel, M. Santra, Y. Q. Chen, J. Uitto and R. V. Iozzo: Transcriptional regulation of decorin gene expression. Induction by quiescence and repression by tumor necrosis factor-alpha. *J Biol Chem*, 270(19), 11692-700 (1995)
- 47. T. Scholzen, M. Solursh, S. Suzuki, R. Reiter, J. L. Morgan, A. M. Buchberg, L. D. Siracusa and R. V. Iozzo: The murine decorin. Complete cDNA cloning, genomic organization, chromosomal assignment, and expression during organogenesis and tissue differentiation. *J Biol Chem*, 269(45), 28270-81 (1994)

- 48. A. De Luca, M. Santra, A. Baldi, A. Giordano and R. V. Iozzo: Decorin-induced growth suppression is associated with up-regulation of p21, an inhibitor of cyclindependent kinases. *J Biol Chem*, 271(31), 18961-5 (1996)
- 49. R. V. Iozzo, D. K. Moscatello, D. J. McQuillan and I. Eichstetter: Decorin is a biological ligand for the epidermal growth factor receptor. *J Biol Chem*, 274(8), 4489-92 (1999)
- 50. M. Santra, I. Eichstetter and R. V. Iozzo: An antioncogenic role for decorin. Down-regulation of ErbB2 leads to growth suppression and cytodifferentiation of mammary carcinoma cells. *J Biol Chem*, 275(45), 35153-61 (2000)
- 51. G. Csordas, M. Santra, C. C. Reed, I. Eichstetter, D. J. McQuillan, D. Gross, M. A. Nugent, G. Hajnoczky and R. V. Iozzo: Sustained down-regulation of the epidermal growth factor receptor by decorin. A mechanism for controlling tumor growth *in vivo. J Biol Chem*, 275(42), 32879-87 (2000)
- 52. A. Hildebrand, M. Romaris, L. M. Rasmussen, D. Heinegard, D. R. Twardzik, W. A. Border and E. Ruoslahti: Interaction of the small interstitial proteoglycans biglycan, decorin and fibromodulin with transforming growth factor beta. *Biochem J*, 302 (Pt 2), 527-34 (1994)
- 53. S. F. Penc, B. Pomahac, T. Winkler, R. A. Dorschner, E. Eriksson, M. Herndon and R. L. Gallo: Dermatan sulfate released after injury is a potent promoter of fibroblast growth factor-2 function. *J Biol Chem*, 273(43), 28116-21 (1998)
- 54. J. Yang, M. A. Price, C. L. Neudauer, C. Wilson, S. Ferrone, H. Xia, J. Iida, M. A. Simpson and J. B. McCarthy: Melanoma chondroitin sulfate proteoglycan enhances FAK and ERK activation by distinct mechanisms. *J Cell Biol*, 165(6), 881-91 (2004)
- 55. R. Sasisekharan, Z. Shriver, G. Venkataraman and U. Narayanasami: Roles of heparan-sulphate glycosaminoglycans in cancer. *Nat Rev Cancer*, 2(7), 521-8 (2002)
- 56. V. P. Eswarakumar, I. Lax and J. Schlessinger: Cellular signaling by fibroblast growth factor receptors. *Cytokine Growth Factor Rev.*, 16(2), 139-49 (2005)
- 57. A. Mansukhani, P. Dell'Era, D. Moscatelli, S. Kornbluth, H. Hanafusa and C. Basilico: Characterization of the murine BEK fibroblast growth factor (FGF) receptor: activation by three members of the FGF family and requirement for heparin. *Proc Natl Acad Sci U S A*, 89(8), 3305-9 (1992)
- 58. A. Yayon, M. Klagsbrun, J. D. Esko, P. Leder and D. M. Ornitz: Cell surface, heparin-like molecules are required for binding of basic fibroblast growth factor to its high affinity receptor. *Cell*, 64(4), 841-8 (1991)
- 59. L. Pellegrini: Role of heparan sulfate in fibroblast growth factor signalling: a structural view. *Curr Opin Struct Biol*, 11(5), 629-34 (2001)

- 60. Y. Zhang, J. Li, C. Partovian, F. W. Sellke and M. Simons: Syndecan-4 modulates basic fibroblast growth factor 2 signaling *in vivo*. *Am J Physiol Heart Circ Physiol*, 284(6), H2078-82 (2003)
- 61. L. Borsig: Heparin as an inhibitor of cancer progression. *Prog Mol Biol Transl Sci*, 93, 335-49 (2010)
- 62. J. L. Stevenson, A. Varki and L. Borsig: Heparin attenuates metastasis mainly due to inhibition of P- and L-selectin, but non-anticoagulant heparins can have additional effects. *Thromb Res*, 120 Suppl 2, S107-11 (2007)
- 63. Y. Zhang, J. C. Rassa, M. E. deObaldia, L. M. Albritton and S. R. Ross: Identification of the receptor binding domain of the mouse mammary tumor virus envelope protein. *J Virol*, 77(19), 10468-78 (2003)
- 64. A. Ashkenazi: Targeting death and decoy receptors of the tumour-necrosis factor superfamily. *Nat Rev Cancer*, 2(6), 420-30 (2002)
- 65. D. Hanahan and R. A. Weinberg: The hallmarks of cancer. *Cell*, 100(1), 57-70 (2000)
- 66. X. Z. Wu and D. Chen: Effects of sulfated polysaccharides on tumour biology. *West Indian Med J*, 55(4), 270-3 (2006)
- 67. G. M. Campo, A. Avenoso, S. Campo, A. D'Ascola, P. Traina, D. Sama and A. Calatroni: Glycosaminoglycans modulate inflammation and apoptosis in LPS-treated chondrocytes. *J Cell Biochem*, 106(1), 83-92 (2009)
- 68. H. Lin, R. Huber, D. Schlessinger and P. J. Morin: Frequent silencing of the GPC3 gene in ovarian cancer cell lines. *Cancer Res*, 59(4), 807-10 (1999)
- 69. S. S. Murthy, T. Shen, A. De Rienzo, W. C. Lee, P. C. Ferriola, S. C. Jhanwar, B. T. Mossman, J. Filmus and J. R. Testa: Expression of GPC3, an X-linked recessive overgrowth gene, is silenced in malignant mesothelioma. *Oncogene*, 19(3), 410-6 (2000)
- 70. Y. Y. Xiang, V. Ladeda and J. Filmus: Glypican-3 expression is silenced in human breast cancer. *Oncogene*, 20(50), 7408-12 (2001)
- 71. C. Buchanan, I. Stigliano, H. M. Garay-Malpartida, L. Rodrigues Gomes, L. Puricelli, M. C. Sogayar, E. Bal de Kier Joffe and M. G. Peters: Glypican-3 reexpression regulates apoptosis in murine adenocarcinoma mammary cells modulating PI3K/Akt and p38MAPK signaling pathways. *Breast Cancer Res Treat*, 119(3), 559-74 (2010)
- 72. A. D. Gonzalez, M. Kaya, W. Shi, H. Song, J. R. Testa, L. Z. Penn and J. Filmus: OCI-5/GPC3, a glypican encoded by a gene that is mutated in the Simpson-Golabi-Behmel overgrowth syndrome, induces

- apoptosis in a cell line-specific manner. J Cell Biol, 141(6), 1407-14 (1998)
- 73. J. P. Lai, D. S. Sandhu, C. Yu, C. D. Moser, C. Hu, A. M. Shire, I. Aderca, L. M. Murphy, A. A. Adjei, S. Sanderson and L. R. Roberts: Sulfatase 2 protects hepatocellular carcinoma cells against apoptosis induced by the PI3K inhibitor LY294002 and ERK and JNK kinase inhibitors. *Liver Int*, 30(10), 1522-8 (2010)
- 74. M. Mali, H. Andtfolk, H. M. Miettinen and M. Jalkanen: Suppression of tumor cell growth by syndecan-1 ectodomain. *J Biol Chem*, 269(45), 27795-8 (1994)
- 75. M. V. Dhodapkar, E. Abe, A. Theus, M. Lacy, J. K. Langford, B. Barlogie and R. D. Sanderson: Syndecan-1 is a multifunctional regulator of myeloma pathobiology: control of tumor cell survival, growth, and bone cell differentiation. *Blood*, 91(8), 2679-88 (1998)
- 76. E. J. Davies, F. H. Blackhall, J. H. Shanks, G. David, A. T. McGown, R. Swindell, R. J. Slade, P. Martin-Hirsch, J. T. Gallagher and G. C. Jayson: Distribution and clinical significance of heparan sulfate proteoglycans in ovarian cancer. *Clin Cancer Res*, 10(15), 5178-86 (2004)
- 77. M. Barbareschi, P. Maisonneuve, D. Aldovini, M. G. Cangi, L. Pecciarini, F. Angelo Mauri, S. Veronese, O. Caffo, A. Lucenti, P. D. Palma, E. Galligioni and C. Doglioni: High syndecan-1 expression in breast carcinoma is related to an aggressive phenotype and to poorer prognosis. *Cancer*, 98(3), 474-83 (2003)
- 78. J. R. Conejo, J. Kleeff, A. Koliopanos, K. Matsuda, Z. W. Zhu, H. Goecke, N. Bicheng, A. Zimmermann, M. Korc, H. Friess and M. W. Buchler: Syndecan-1 expression is up-regulated in pancreatic but not in other gastrointestinal cancers. *Int J Cancer*, 88(1), 12-20 (2000)
- 79. Y. Hu, H. Sun, R. T. Owens, Z. Gu, J. Wu, Y. Q. Chen, J. T. O'Flaherty and I. J. Edwards: Syndecan-1-dependent suppression of PDK1/Akt/bad signaling by docosahexaenoic acid induces apoptosis in prostate cancer. *Neoplasia*, 12(10), 826-36 (2010)
- 80. C. Y. Pumphrey, A. M. Theus, S. Li, R. S. Parrish and R. D. Sanderson: Neoglycans, carbodiimide-modified glycosaminoglycans: a new class of anticancer agents that inhibit cancer cell proliferation and induce apoptosis. *Cancer Res*, 62(13), 3722-8 (2002)
- 81. K. Ueda, S. Inoue, Y. Zhang, T. Kutsuna, K. Noto, N. Arai and M. Noguchi: Heparin induces apoptosis through suppression of AKt in oral squamous cell carcinoma cells. *Anticancer Res*, 29(4), 1079-88 (2009)
- 82. X. Chen, W. Xiao, X. Qu and S. Zhou: The effect of dalteparin, a kind of low molecular weight heparin, on lung adenocarcinoma A549 cell line *in vitro*. *Cancer Invest*, 26(7), 718-24 (2008)

- 83. G. Chatzinikolaou, D. Nikitovic, A. Berdiaki, A. Zafiropoulos, P. Katonis, N. K. Karamanos and G. N. Tzanakakis: Heparin regulates colon cancer cell growth through p38 mitogen-activated protein kinase signalling. *Cell Prolif*, 43(1), 9-18 (2010)
- 84. Y. Uzun, E. Akdogan, F. Ozdemir and E. Ovali: The effects of heparin on DLD-1 colon cancer cell line. *Bratisl Lek Listy*, 110(1), 3-6 (2009)
- 85. M. Balzarotti, F. Fontana, C. Marras, A. Boiardi, D. Croci, E. Ciusani and A. Salmaggi: *In vitro* study of low molecular weight heparin effect on cell growth and cell invasion in primary cell cultures of high-grade gliomas. *Oncol Res*, 16(5), 245-50 (2006)
- 86. G. Kirfel, A. Rigort, B. Borm and V. Herzog: Cell migration: mechanisms of rear detachment and the formation of migration tracks. *Eur J Cell Biol*, 83(11-12), 717-24 (2004)
- 87. K. T. Chan, C. L. Cortesio and A. Huttenlocher: Integrins in cell migration. *Methods Enzymol*, 426, 47-67 (2007)
- 88. P. Friedl and K. Wolf: Proteolytic interstitial cell migration: a five-step process. *Cancer Metastasis Rev*, 28(1-2), 129-35 (2009)
- 89. K. Wolf and P. Friedl: Mapping proteolytic cancer cell-extracellular matrix interfaces. *Clin Exp Metastasis*, 26(4), 289-98 (2009)
- 90. D. S. Micalizzi, S. M. Farabaugh and H. L. Ford: Epithelial-mesenchymal transition in cancer: parallels between normal development and tumor progression. *J Mammary Gland Biol Neoplasia*, 15(2), 117-34 (2010)
- 91. D. A. Zajchowski, M. F. Bartholdi, Y. Gong, L. Webster, H. L. Liu, A. Munishkin, C. Beauheim, S. Harvey, S. P. Ethier and P. H. Johnson: Identification of gene expression profiles that predict the aggressive behavior of breast cancer cells. *Cancer Res*, 61(13), 5168-78 (2001)
- 92. H. Hugo, M. L. Ackland, T. Blick, M. G. Lawrence, J. A. Clements, E. D. Williams and E. W. Thompson: Epithelial--mesenchymal and mesenchymal--epithelial transitions in carcinoma progression. *J Cell Physiol*, 213(2), 374-83 (2007)
- 93. L. Tamagnone, S. Artigiani, H. Chen, Z. He, G. I. Ming, H. Song, A. Chedotal, M. L. Winberg, C. S. Goodman, M. Poo, M. Tessier-Lavigne and P. M. Comoglio: Plexins are a large family of receptors for transmembrane, secreted, and GPI-anchored semaphorins in vertebrates. *Cell*, 99(1), 71-80 (1999)
- 94. R. Montesano, K. Matsumoto, T. Nakamura and L. Orci: Identification of a fibroblast-derived epithelial morphogen as hepatocyte growth factor. *Cell*, 67(5), 901-8 (1991)

- 95. S. Barr, S. Thomson, E. Buck, S. Russo, F. Petti, I. Sujka-Kwok, A. Eyzaguirre, M. Rosenfeld-Franklin, N. W. Gibson, M. Miglarese, D. Epstein, K. K. Iwata and J. D. Haley: Bypassing cellular EGF receptor dependence through epithelial-to-mesenchymal-like transitions. *Clin Exp Metastasis*, 25(6), 685-93 (2008)
- 96. G. Raab and M. Klagsbrun: Heparin-binding EGF-like growth factor. *Biochim Biophys Acta*, 1333(3), F179-99 (1997)
- 97. K. Itoh and S. Y. Sokol: Heparan sulfate proteoglycans are required for mesoderm formation in Xenopus embryos. *Development*, 120(9), 2703-11 (1994)
- 98. J. A. Deakin, B. S. Blaum, J. T. Gallagher, D. Uhrin and M. Lyon: The binding properties of minimal oligosaccharides reveal a common heparan sulfate/dermatan sulfate-binding site in hepatocyte growth factor/scatter factor that can accommodate a wide variety of sulfation patterns. *J Biol Chem*, 284(10), 6311-21 (2009)
- 99. K. R. Catlow, J. A. Deakin, Z. Wei, M. Delehedde, D. G. Fernig, E. Gherardi, J. T. Gallagher, M. S. Pavao and M. Lyon: Interactions of hepatocyte growth factor/scatter factor with various glycosaminoglycans reveal an important interplay between the presence of iduronate and sulfate density. *J Biol Chem*, 283(9), 5235-48 (2008)
- 100. K. Catlow, J. A. Deakin, M. Delehedde, D. G. Fernig, J. T. Gallagher, M. S. Pavao and M. Lyon: Hepatocyte growth factor/scatter factor and its interaction with heparan sulphate and dermatan sulphate. *Biochem Soc Trans*, 31(2), 352-3 (2003)
- 101. J. Villena and E. Brandan: Dermatan sulfate exerts an enhanced growth factor response on skeletal muscle satellite cell proliferation and migration. *J Cell Physiol*, 198(2), 169-78 (2004)
- 102. I. Vlodavsky, A. Eldor, A. Haimovitz-Friedman, Y. Matzner, R. Ishai-Michaeli, O. Lider, Y. Naparstek, I. R. Cohen and Z. Fuks: Expression of heparanase by platelets and circulating cells of the immune system: possible involvement in diapedesis and extravasation. *Invasion Metastasis*, 12(2), 112-27 (1992)
- 103. X. Li and J. F. Wu: Recent developments in patent anti-cancer agents targeting the matrix metalloproteinases (MMPs). *Recent Pat Anticancer Drug Discov*, 5(2), 109-41 (2010)
- 104. E. Cohen, I. Doweck, I. Naroditsky, O. Ben-Izhak, R. Kremer, L. A. Best, I. Vlodavsky and N. Ilan: Heparanase is overexpressed in lung cancer and correlates inversely with patient survival. *Cancer*, 113(5), 1004-11 (2008)
- 105. L. Fux, N. Feibish, V. Cohen-Kaplan, S. Gingis-Velitski, S. Feld, C. Geffen, I. Vlodavsky and N. Ilan: Structure-function approach identifies a COOH-terminal domain that mediates heparanase signaling. *Cancer Res*, 69(5), 1758-67 (2009)

- 106. V. Cohen-Kaplan, I. Doweck, I. Naroditsky, I. Vlodavsky and N. Ilan: Heparanase augments epidermal growth factor receptor phosphorylation: correlation with head and neck tumor progression. *Cancer Res*, 68(24), 10077-85 (2008)
- 107. V. Cohen-Kaplan, I. Naroditsky, A. Zetser, N. Ilan, I. Vlodavsky and I. Doweck: Heparanase induces VEGF C and facilitates tumor lymphangiogenesis. *Int J Cancer*, 123(11), 2566-73 (2008)
- 108. T. M. Niers, C. P. Klerk, M. DiNisio, C. J. Van Noorden, H. R. Buller, P. H. Reitsma and D. J. Richel: Mechanisms of heparin induced anti-cancer activity in experimental cancer models. Crit Rev Oncol Hematol, 61(3), 195-207 (2007)
- 109. J. Gao, L. Su, R. Qin, Q. Chang, T. Huang and Y. Feng: Transfection of antisense oligodeoxynucleotide inhibits heparanase gene expression and invasive ability of human pancreatic cancer cell in vitro. J Huazhong Univ Sci Technolog Med Sci, 26(1), 72-4 (2006)
- 110. O. Goldshmidt, E. Zcharia, R. Abramovitch, S. Metzger, H. Aingorn, Y. Friedmann, V. Schirrmacher, E. Mitrani and I. Vlodavsky: Cell surface expression and secretion of heparanase markedly promote tumor angiogenesis and metastasis. Proc Natl Acad Sci U S A, 99(15), 10031-6 (2002)
- 111. M. Nakajima, A. DeChavigny, C. E. Johnson, J. Hamada, C. A. Stein and G. L. Nicolson: Suramin. A potent inhibitor of melanoma heparanase and invasion. J Biol Chem, 266(15), 9661-6 (1991)
- 112. J. W. Dennis and S. Laferte: Tumor cell surface carbohydrate and the metastatic phenotype. Cancer Metastasis Rev, 5(3), 185-204 (1987)
- 113. S. Hakomori: Tumor malignancy defined by aberrant glycosylation and sphingo(glyco)lipid metabolism. *Cancer Research*, 56(23), 5309-18 (1996)
- 114. Y. S. Kim, J. Gum, Jr. and I. Brockhausen: Mucin glycoproteins in neoplasia. *Glycoconj J*, 13(5), 693-707 (1996)
- 115. R. Kannagi: Carbohydrate-mediated cell adhesion involved in hematogenous metastasis of cancer. *Glycoconj J*, 14(5), 577-84 (1997)
- 116. G. S. Kansas and F. M. Pavalko: The cytoplasmic domains of E- and P-selectin do not constitutively interact with alpha-actinin and are not essential for leukocyte adhesion. *J Immunol*, 157(1), 321-5 (1996)
- 117. L. R. Zacharski, W. G. Henderson, F. R. Rickles, W. B. Forman, C. J. Cornell, Jr., R. J. Forcier, R. L. Edwards, E. Headley, S. H. Kim, J. F. O'Donnell and *et al.*: Effect of warfarin anticoagulation on survival in carcinoma of the lung, colon, head and neck, and prostate. Final report of VA Cooperative Study #75. *Cancer*, 53(10), 2046-52 (1984)

- 118. H. Engelberg: Actions of heparin that may affect the malignant process. *Cancer*, 85(2), 257-72 (1999)
- 119. M. Hejna, M. Raderer and C. C. Zielinski: Inhibition of metastases by anticoagulants. *J Natl Cancer Inst*, 91(1), 22-36 (1999)
- 120. L. P. Fielding, R. Hittinger, R. H. Grace and J. S. Fry: Randomised controlled trial of adjuvant chemotherapy by portal-vein perfusion after curative resection for colorectal adenocarcinoma. *Lancet*, 340(8818), 502-6 (1992)
- 121. B. Lebeau, C. Chastang, J. M. Brechot, F. Capron, B. Dautzenberg, C. Delaisements, M. Mornet, J. Brun, J. P. Hurdebourcq and E. Lemarie: Subcutaneous heparin treatment increases survival in small cell lung cancer. "Petites Cellules" Group. *Cancer*, 74(1), 38-45 (1994)
- 122. D. Nitti, J. Wils, T. Sahmoud, D. Curran, M. L. Couvreur, M. Lise, H. Rauschecker, J. G. dos Santos, W. Stremmel and F. Roelofsen: Final results of a phase III clinical trial on adjuvant intraportal infusion with heparin and 5-fluorouracil (5-FU) in resectable colon cancer (EORTC GITCCG 1983-1987). European Organization for Research and Treatment of Cancer. Gastrointestinal Tract Cancer Cooperative Group. *Eur J Cancer*, 33(8), 1209-15 (1997)
- 123. C. C. Zielinski and M. Hejna: Warfarin for cancer prevention. *N Engl J Med*, 342(26), 1991-3 (2000)
- 124. A. Koenig, K. Norgard-Sumnicht, R. Linhardt and A. Varki: Differential interactions of heparin and heparan sulfate glycosaminoglycans with the selectins. Implications for the use of unfractionated and low molecular weight heparins as therapeutic agents. *J Clin Invest*, 101(4), 877-89 (1998)
- 125. L. Wang, J. R. Brown, A. Varki and J. D. Esko: Heparin's anti-inflammatory effects require glucosamine 6-O-sulfation and are mediated by blockade of L- and P-selectins. *J Clin Invest*, 110(1), 127-36 (2002)
- 126. D. Y. Lee, K. Park, S. K. Kim, R. W. Park, I. C. Kwon, S. Y. Kim and Y. Byun: Antimetastatic effect of an orally active heparin derivative on experimentally induced metastasis. *Clin Cancer Res*, 14(9), 2841-9 (2008)
- 127. M. S. Pavao, K. R. Aiello, C. C. Werneck, L. C. Silva, A. P. Valente, B. Mulloy, N. S. Colwell, D. M. Tollefsen and P. A. Mourao: Highly sulfated dermatan sulfates from Ascidians. Structure versus anticoagulant activity of these glycosaminoglycans. *J Biol Chem*, 273(43), 27848-57 (1998)
- 128. R. J. Fonseca and P. A. Mourao: Fucosylated chondroitin sulfate as a new oral antithrombotic agent. *Thromb Haemost*, 96(6), 822-9 (2006)
- 129. E. O. Kozlowski, M. S. G.; Borsig, L.: Ascidian dermatan sulfates attenuate metastasis, inflammation and thrombosis by inhibition of P-selectin. *Submitted* (2011)

- 130. E. O. Kozlowski; Gomes, A. M.; Silva, C. F. S.; Pavão, M. S. G.: Structure and biological activities of glycosaminoglycan analogs from marine invertebrates: New therapeutic agents? *In: Pavao, M.S.G. (ed) Glycans in therapeutics and disease, 1st edn. Springer, Heidelberg, In Press.* (2011)
- 131. S. Alban: From heparins to factor Xa inhibitors and beyond. *Eur J Clin Invest*, 35 Suppl 1, 12-20 (2005)
- 132. J. G. Martin, M. Gupta, Y. Xu, S. Akella, J. Liu, J. S. Dordick and R. J. Linhardt: Toward an artificial Golgi: redesigning the biological activities of heparan sulfate on a digital microfluidic chip. *J Am Chem Soc*, 131(31), 11041-8 (2009)
- **Key Words:** Glycosaminoglycans, Metastasis, Epithelial-To-Mesenchymal Transition, Hematogeneous Metastasis, Review
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