

The anaplastic lymphoma kinase as an oncogene in solid tumors

Claudia Voena^{1,2,3}, Silvia Peola^{1,2}, Roberto Chiarle^{1,2,3,4}

¹Department of Molecular Biotechnology and Health Sciences, University of Torino, Torino, Italy, ²Center for Experimental Research and Medical Studies (CERMS), Città della Salute e della Scienza, Torino, Italy,

³Department of Pathology, Children's Hospital and Harvard Medical School, Boston, USA, ⁴European Research Initiative on ALK-related malignancies (ERIA)

TABLE OF CONTENTS

1. Abstract
2. Introduction
3. ALK translocations in solid tumors
 - 3.1. ALK rearrangements in non-small cell lung cancer
 - 3.2. ALK rearrangements in other solid tumors
4. ALK point mutations and overexpression in solid tumors
 - 4.1. Neuroblastoma
 - 4.2. ALK as an oncogene in anaplastic thyroid cancer and rhabdomyosarcoma
5. Targeted therapies for ALK-rearranged solid tumors
6. Conclusions
7. Acknowledgements
8. References

1. ABSTRACT

Twenty years ago anaplastic lymphoma kinase (ALK) was discovered in anaplastic large cell lymphoma (ALCL), but the interest in ALK as an oncogene grew only in recent years when ALK rearrangements were reported as recurrent genetic lesions in lung carcinoma and activating single point mutations were described in neuroblastoma. In this review we will describe the main features of ALK-rearranged solid tumors, with particular emphasis to NSCLC and neuroblastoma. We will discuss the numerous *in vitro* and *in vivo* studies that confirmed ALK as the “driver” oncogene in these tumors and the achievements in clinical settings with ALK inhibitors that validated ALK as a therapeutic target. We will finally end with the description of putative innovative therapeutic approaches that are on going to overcome acquired resistance that invariably occurs in crizotinib treated NSCLC patients or intrinsic resistance to crizotinib therapy reported in neuroblastoma.

2. INTRODUCTION

Anaplastic lymphoma kinase (ALK) is described as the “driver” oncogene in a variety of human cancer, both hematological and solid tumors. ALK was originally cloned and identified in anaplastic large cell lymphoma (ALCL) in 1994 as the result of a recurrent chromosomal translocation, t(2;5; p23;q35) (1). ALK is a receptor tyrosine kinase that belongs to the insulin receptor superfamily. In invertebrates, the role of ALK and its ligands (jelly belly and hesitation behavior-

(HEN-1)) is clearly defined (2, 3). In *Drosophila melanogaster* ALK is required for survival and is involved in the development of the visual system (4), in visceral muscle patterning (5, 6) and in protecting neural progenitors during nutrient restriction (7). In contrast, in mammals, ALK is considered an orphan receptor and its physiological role is only partially elucidated. Indeed, the biological relevance of two putative ligands for ALK, pleiotrophin and midkine, is still unclear (8, 9). Recently, heparin was proposed to be the ligand for ALK (10). In mammals, ALK is expressed during embryonic and neonatal development in specific regions of the nervous system, whereas in adults it is restricted to some neurons in the central nervous system at barely detectable levels (11-13). In line with these observations, *Alk* knock-out mice are viable and without evident tissue abnormalities showing a very mild phenotype, mostly related to behavioral control (14, 15).

In recent years, ALK rearrangements have been described in other tumors, including non-small cell lung carcinoma (NSCLC), inflammatory myofibroblastic tumor (IMT), diffuse large B-cell lymphoma (DLBCL), renal cell carcinoma (RCC), colon carcinoma, breast and thyroid tumors (16, 17). In addition, activating point mutations and gene amplifications have been reported in neuroblastoma (NB) and anaplastic thyroid cancer (ATC) (18-21) (Figure 1). ALK chromosomal rearrangements generate fusion proteins that invariably contain the

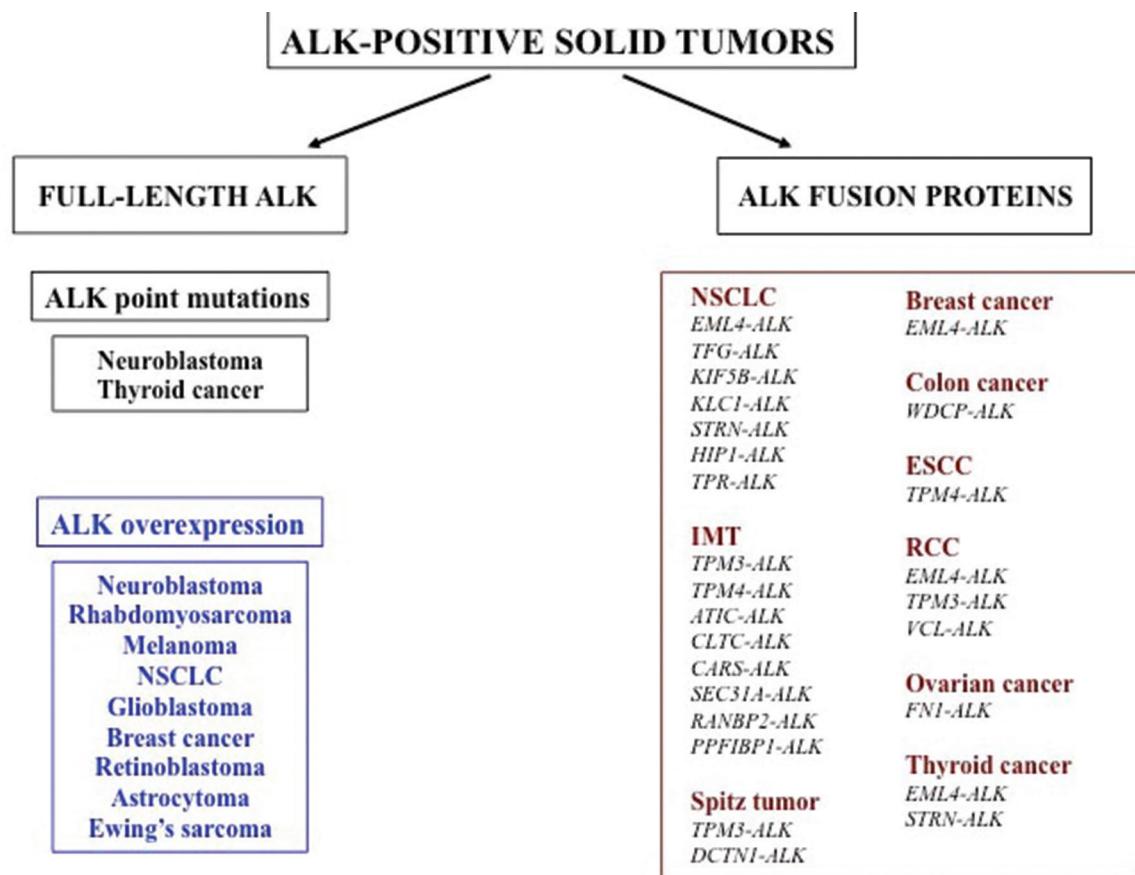


Figure 1. Schematic representation of ALK alterations in solid tumors. NSCLC: non-small cell lung carcinoma; IMT: inflammatory myofibroblastic tumor; RCC: renal cell carcinoma; ESCC: esophageal squamous cell carcinoma

intracytoplasmic signaling portion of ALK fused to a partner that contributes a dimerization domain. Spontaneous homodimerization induces cross-phosphorylation of ALK, and triggers the constitutive tyrosine kinase activity that generates oncogenic signals. The common feature of all ALK fusions is the aberrant activation of ALK downstream signaling and, as reported for ALCL, in the majority of ALK-rearranged tumors the ablation of the ALK signaling leads to growth arrest and cell death (16, 22). The transforming properties and the signaling triggered by ALK have been extensively characterized for NPM-ALK in ALCL. Several downstream pathways are activated by NPM-ALK, with a broad range of signals that lead to increased cell proliferation, survival, motility, and cytoskeletal rearrangements (22). In transgenic mice (23), as well as in lymphoma-derived cell lines, ALK oncogenic signals are mediated by a series of key molecules and pathways, including Stat3 (24, 25), phosphatidyl inositol 3-Kinase (PI3K) (26), Ras/MAPK/ERK, Shp2 (27), p130Cas, (28), PLC γ (29) and Src (30). Similar pathways are activated in ALK-driven lung cancers and neuroblastoma.

3. ALK TRANSLOCATIONS IN SOLID TUMORS

3.1. ALK rearrangements in non-small cell lung cancer

In 2007 ALK rearrangements were identified in a small subset of NSCLC patients as a result of an inversion within chromosome 2, inv 2(2p21;p23) (31, 32). First reports described ALK rearrangements in 6-7% of NSCLC patients, but following studies reported different percentages depending on the population studied, since it is more frequent in the Asiatic population than in the Western population. To date the overall incidence of ALK rearrangements in NSCLC is reported approximately as 5-6% of worldwide NSCLC cases (33). ALK-rearranged NSCLC are mainly adenocarcinoma characterized by the presence of signet-ring cells with abundant intracellular mucin (34, 35) and are prevalent in young non- or light-smoker patients (33, 36). In contrast to EGFR mutations, NSCLC harboring ALK rearrangements do not show any ethnic/racial differences (37). Excluding few rare exceptions, ALK rearrangements are mutually exclusive with other frequent oncogenic mutations found in NSCLC, such as EGFR and KRAS mutations (37-40).

The inversion within the short arm of chromosome 2 generates a fusion between the 5' region of echinoderm-microtubule-associated protein-like 4 (EML4) gene and the 3' region of the ALK gene that creates a new constitutively active ALK fusion protein, EML4-ALK. EML4-ALK contains the entire intracytoplasmic portion of ALK and the N-terminal portion of EML4 whose coiled-coil domain provides the oligomerization domain for the ligand-independent activation of EML4-ALK (32). To date, multiple variants of EML4-ALK have been described in NSCLC patients due to different breakpoints in the EML4 gene (occurring at exons 2, 6, 13, 14, 15, 18, 20) (33). All EML4-ALK variants are fully active and transforming because they retain the same portion of ALK that includes the kinase domain (33). As described for ALK-rearranged ALCL, additional ALK translocations have also been reported in NSCLC involving different partners, including TRK-fused gene (TFG-ALK), kinesin family member 5B (KIF5B-ALK), kinesin light chain 1 (KLC1-ALK), striatin (STRN-ALK), huntingtin interacting protein 1 (HIP1-ALK) and translocated promoter region (TPR-ALK) (17, 31, 41-44). In contrast to ALCL, all ALK fusions in NSCLC are localized in the cytoplasm where they trigger proliferative and survival downstream pathways (16). Numerous *in vitro* and *in vivo* studies have demonstrated the transforming properties of ALK translocations and the strong addiction to the tyrosine activity of ALK of ALK-rearranged NSCLC. Indeed, NSCLC cells depend on EML4-ALK for growth and survival and are sensitive to ALK inhibition (45, 46) regardless the type of translocation or EML4-ALK variant. However, some *in vitro* observations reported a different sensitivity to ALK inhibitors for some EML4-ALK variants likely related to their protein stability (47). The generation of constitutive and conditional ALK-rearranged transgenic (Tg) mice helped to elucidate the biology of ALK-rearranged NSCLC and provided a useful model for preclinical studies (48, 49). ALK driven NSCLC mouse models developed hundreds of adenocarcinoma nodules in both lungs with a very short latency period, and with 100% penetrance and showed dramatic responses to ALK inhibitors, TAE684 and crizotinib. Recently, the CRISPR/Cas9 technology has been exploited to induce EML4-ALK rearrangement *in vivo* and to generate mouse models of EML4-ALK driven lung cancer (50, 51). All derived tumors expressed the EML4-ALK fusion protein, displayed histopathological features of human ALK-driven NSCLC and responded to crizotinib treatment (50). Interestingly, tumors were strongly positive for the alveolar type II marker surfactant protein C (SP-C) and completely negative for the Clara cell marker CCSP (also known as CC10), thus definitively validating previous mouse models of ALK-rearranged NSCLC in which EML4-ALK expression was forced in type II alveolar epithelial cells by the use of SP-C promoter.

3.2. ALK rearrangements in other solid tumors

The first evidence of ALK involvement in the pathogenesis of IMT dates back to 1999 when the first ALK

fusion protein was described in these tumors (52). So far, approximately 50% of IMT harbors ALK rearrangements that lead to different ALK fusion proteins, all sharing the ALK kinase domain, such as TPM3- and TPM4-ALK, ATIC-ALK, CLTC-ALK, CARS-ALK, RANBP2-ALK and SEC31L1-ALK (53-59). In contrast to ALK-driven NSCLC, in which ALK fusions are unique, most of the fusions in IMT have been described in ALCL (60). IMT is frequently diagnosed in young patients and is now categorized as a mesenchymal tumor derived from myofibroblasts, but it was initially considered an "inflammatory pseudo-tumor" because of the presence of a rich inflammatory infiltrate. The discovery of ALK-rearrangements supported the etiology of a low-grade mesenchymal neoplasm, even though the prognostic relevance of ALK rearrangement in IMT is still controversial. Nonetheless ALK-directed therapy is emerging as a highly effective treatment option for a subset of patients with IMT with more aggressive disease, although acquired resistance has been already reported (61, 62).

After the discovery of ALK rearrangements in NSCLC and the promising clinical benefits obtained with the treatment with ALK kinase inhibitors, many efforts have been directed to detect ALK genetic lesions in other solid tumors. To date the list of ALK fusion proteins in other epithelial tumors, although in isolated cases, includes TPM3-ALK, EML4-ALK and vinculin (VCL)-ALK in poor outcome RCC (63-65), EML4-ALK in breast cancer (66), C2orf44-ALK in colon carcinoma, recently reported as WD repeat and coiled coil containing protein (WDCP)-ALK, (39) (67), fibronectin (FN1)-ALK in ovarian cancer (68), EML4-ALK and STRN-ALK in thyroid carcinoma (17), TPM3-ALK and dynactin 1 (DCTN1)-ALK in Spitz tumors(69) and it will likely grow in the future. A recent report on ALK-rearranged anaplastic thyroid cancer demonstrated that STRN-ALK is transforming and tumorigenic *in vivo* and that blockade of its kinase activity with specific ALK inhibitors arrests tumor growth *in vivo* (70). Therefore, although in most of these tumors the role of ALK needs to be fully elucidated, as well as their addiction to ALK, these tumors might benefit from ALK inhibition.

4. ALK POINT MUTATIONS AND OVEREXPRESSION

4.1. Neuroblastoma

Neuroblastoma is the most common extracranial malignant tumor of childhood accounting for 15% of pediatric oncology deaths (71). To date the amplification of the MYCN oncogene on chromosome 2 (2p24) is the most frequent lesion, occurring in about 20-25% of cases and has been used for the stratification of neuroblastoma patients being associated with poor prognosis (72). Recently, the discovery of activating single point mutations of ALK tyrosine kinase receptor in both hereditary and sporadic neuroblastoma allowed

a better stratification of patients and provided the basis for a targeted therapy in these tumors (18-21). Different non-synonymous mutations mostly within the tyrosine kinase domain of ALK have been reported so far in primary tumors (73). The most common mutations were the substitution of the phenylalanine at codon 1174 with different aminoacids (F1174L/C/I) only in sporadic cases, the R1275Q and the F1245C in both familial and sporadic neuroblastoma. Other mutations were detected only in familial cases of neuroblastoma, such as R1192P and G1128A. Interestingly, F1174L was preferentially associated with MYCN amplification in neuroblastoma and was also detected in crizotinib resistant IMT (62, 74). Besides point mutations, ALK overexpression has been detected in the majority of neuroblastoma and has been associated to a worse prognosis in patients (75, 76). Mutations and overexpression induce a constitutive ligand-independent activation of the ALK receptor, with differential transforming ability in various cell types (Schulte et al., 2012). ALK mutations display different phosphorylation status and activate downstream pathways in a mutation-dependent manner, i.e. F1174L preferentially activated STAT3 and AKT, whereas R1275Q efficiently activated AKT1/2 and ERK1/2 (18-21). Knock-down of ALK or inhibition of ALK kinase activity by small molecules in neuroblastoma mutant cell lines led to growth arrest and apoptosis and further proved the “addiction” to mutated ALK of neuroblastoma tumors (18-21, 77). Two different transgenic mouse models of neuroblastoma harboring ALK F1174L mutation under Dbh or Th promoters shed light on the role of ALK mutations in neuroblastoma pathogenesis and provided important clues for the therapeutic testing of ALK inhibitors in neuroblastoma (78, 79). Only one mouse model developed tumors similar to neuroblastoma in presence of ALK F1174L, but with a very long latency and low penetrance (79). In both models, cooperation with MYCN accelerated neuroblastoma formation and led to development of high penetrant and aggressive tumors. The fact that these models developed tumors only when ALKF1174L and MYCN are co-expressed demonstrate that ALK alone is not enough to initiate tumorigenesis in neuroblastoma and need the cooperation with MYCN to develop tumors. Interestingly, both models showed intrinsic resistance to crizotinib treatment in agreement with *in vitro* assays that reported a different sensitivity to crizotinib for the ALK mutant form F1174L in neuroblastoma cells (78-80). However, using other ALK inhibitors or a combination treatment with mTor inhibitor a partial response in terms of tumor regression was observed thus providing evidence that ALK could be a good target also in MYCN-amplified tumors. A transgenic neuroblastoma model developed in zebrafish confirmed the cooperative role of wild-type or mutated ALK (F1174L) in the pathogenesis of neuroblastoma. As observed in Tg mice, neither ALK wt nor ALK F1174L alone developed tumors (81). In recently generated knock-in mice for both F1174L and R1275Q (murine F1178L and

R1279Q, respectively), a prolonged neurogenesis of the sympathetic ganglia has been observed, but not tumor development (82). Also in this context both ALK mutations accelerated and increased MYCN tumor formation and penetrance, with more aggressive behavior in presence of F1174L mutation. These new knock-in models will help to define more precisely the role of ALK in neuroblastoma and will likely represent a better platform for the screening of alternative targeted approaches.

4.2. ALK as an oncogene in anaplastic thyroid cancer and rhabdomyosarcoma

Missense mutations of ALK receptor have been reported in thyroid cancer both in cell lines and primary tumors (83). These mutations increased the kinase activity of ALK and were fully transforming both *in vitro* and *in vivo* assays, but the proof of ALK “addiction” is still missing in this type of tumors. Recently, a recurrent STRN-ALK rearrangement was found in approx. 2% of thyroid cancers, with possibly higher prevalence in poorly differentiated cancers (17, 70, 84).

A recent wide-genome screening identified frequent copy number gain of ALK in rhabdomyosarcoma, an aggressive form of soft tissue sarcoma in children and adolescents with very poor prognosis (85). Copy number gain was associated with high level of ALK protein expression. In contrast to neuroblastoma, ALK copy number gain was not associated with MYCN amplification that is frequently observed in these tumors. ALK copy number gain correlated with more aggressive and metastatic forms of rhabdomyosarcoma, but its clinical relevance as a potential therapeutic target has not yet been assessed (85, 86).

5. TARGETED THERAPIES FOR ALK-REARRANGED SOLID TUMORS

The findings of recurring rearrangements of ALK in NSCLC prompted the development and the use of ALK inhibitors in the clinic. Crizotinib, a potent ATP-competitive inhibitor of c-Met and ALK, which was developed as a MET inhibitor before the identification of EML4-ALK in NSCLC, was rapidly tested in phase I-II clinical trials (87, 88). Due to the exceptionally high rate of clinical responses (tumor regression in nearly 60% of patients), that further supported the “driver” role of ALK fusions in NSCLC harboring ALK rearrangements, crizotinib received in 2011 an accelerated approval from the US Food and Drug Administration (FDA) to treat ALK-rearranged NSCLC. Despite a high rate of response, only a minimal extension in overall survival was achieved in ALK-rearranged NSCLC patients treated with crizotinib because most of the patients developed resistance to the drug (89, 90). Three general mechanisms of acquired drug resistance to crizotinib in NSCLC have been described: (i) activating ALK mutations, spread throughout the tyrosine kinase domain, that affect the drug interaction regions, (ii) ALK

amplification that leads to overexpression of the EML4-ALK fusion and (iii) activation of alternative compensatory signaling pathways, or so-called bypass track activation, where other RTKs or downstream molecules compensate for the inhibited ALK signaling (36, 62, 91-93). Several next-generation ALK inhibitors have been developed to overcome crizotinib resistance in NSCLC patients. The most notable second-generation ALK inhibitors currently in clinical trials are ceritinib (LDK-378, recently approved by FDA), alectinib (CH542802, already approved in Japan), AP26113 (dual ALK and EGFR inhibitor) and PF-06463922(94). Remarkably, in a recent clinical trial ceritinib showed activity in NSCLC patients that relapsed under treatment with crizotinib and prolonged progression free-survival by 7 months, but it is unclear whether the effects were due only to better pharmacodynamics of ceritinib or to a real increased activity against the ALK kinase (95). Nonetheless, acquired resistance to ceritinib has already been described in NSCLC patients (96). Mechanisms of resistance to crizotinib or next-generation ALK inhibitors will be discussed more in details in a parallel review in this issue (Farina et al.).

The encouraging results of crizotinib treatment in NSCLC provided a strong rationale for targeting ALK in neuroblastoma. In accordance with the *in vitro* observations, the results of phase I clinical trial in neuroblastoma highlighted the differential sensitivity to ALK kinase inhibition compared to the sensitivity observed in ALK-rearranged tumors (80, 97). Therefore, the intrinsic resistance to ALK inhibitors is a main issue in the treatment of neuroblastoma patients and is emerging as the main mechanism that underlies the lack of tumor responsiveness and relapse in neuroblastoma (98).

Several alternative approaches to treat ALK-driven tumors are currently under investigation, such as next-generation ALK inhibitors or combination therapy aimed to inhibit ALK together with the inhibition of downstream molecules or the so called bypass tracks

A promising treatment in ALK-rearranged NSCLC is represented by inhibition of the heat shock protein-90 (HSP-90) that is a chaperone molecule necessary for the correct folding, stabilization and degradation of many oncogenes, including EML4-ALK. HSP-90 inhibition has shown a therapeutic effect in terms of tumor regression and stabilization of the disease in both mouse models and ALK-rearranged NSCLC patients (49, 99). Importantly, the HSP-90 inhibitor, ganetespib overcame multiple forms of crizotinib resistance *in vitro* consistent with the activity seen in a patient with crizotinib-resistant NSCLC (100). Based on these findings, targeting HSP-90 in ALK-rearranged tumors might be a suitable alternative to small molecule inhibitors in the clinical setting.

Recently, immunotherapy has showed promising efficacy in some solid tumors, including

NSCLC (101, 102). In this context, the ALK protein has many features of an ideal tumor oncoantigen that can be exploited for the generation of a cancer vaccine: specificity, strong immunogenicity and absolute requirement for tumor maintenance (22, 103, 104). Indeed, DNA-based ALK vaccine induced a specific immune response in pre-clinical mouse models of ALK-rearranged lymphoma and prevented lymphoma growth (105). More recently, the same ALK vaccine was successfully tested in mouse models of ALK-driven NSCLC and delayed tumor progression in the lungs (Voena et al, personal communication). Remarkably, ALK vaccine was highly efficient against the most common crizotinib ALK mutants found in NSCLC. Although not yet tested, these encouraging results in preclinical models of ALK-driven ALCL and NSCLC suggest that ALK-vaccine might also be an optional therapy for ALK+ neuroblastoma.

In addition, ALK-directed immunotherapy with monoclonal antibodies targeting the ALK full-length receptor was reported to induce an antibody-dependent cell mediated cytotoxicity (ADCC) against neuroblastoma cells (106, 107). This approach might represent a therapeutic option for neuroblastoma that express both wild-type or mutated ALK receptor.

6. CONCLUSIONS

The identification of ALK rearrangements in different tumor types have unveiled the ALK locus as a "hot spot" for genetic alterations in the genome of various solid tumors. Future understanding of mechanisms that underlie ALK translocations will likely shed light on ALK mediated tumor initiation and transformation in different tissues. Experimental and clinical evidences throughout the years have validated ALK as therapeutic target in NSCLC and neuroblastoma and have established the basis for ALK-targeted treatment in other ALK-rearranged tumors. Despite the great expectations originated by the identification of a novel specific target for pharmacological therapies in ALK-rearranged tumors, clinical successes have been partial and transient due to acquired or intrinsic drug resistance, urging for the search of additional innovative therapeutic strategies. Future efforts should be then directed to the comprehension of ALK-driven transformation and tumor maintenance in different tumors to clarify ALK oncogene addiction and to find new targets or molecular mechanisms for the development of future therapeutic strategies.

7. ACKNOWLEDGEMENTS

The work has been supported by grants FP7 ERC-2009-StG (Proposal No. 242965 - "Lunely") to RC; Associazione Italiana per la Ricerca sul Cancro (AIRC) grant IG-12023 to RC; Koch Institute/DFCC Bridge Project Fund to RC; Ellison Foundation Boston to RC;

International Association for Cancer Research (AICR) grant 12-0216 to RC. The present review was concerted inside the European Research Initiative of ALK-related malignancies (ERIA) (<http://www.erialcl.net>).

8. REFERENCES

1. S. W. Morris, M. N. Kirstein, M. B. Valentine, K. G. Dittmer, D. N. Shapiro, D. L. Saltman and A. T. Look: Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. *Science* 263, 1281-4 (1994)
DOI: 10.1126/science.8122112
2. T. Ishihara, Y. Iino, A. Mohri, I. Mori, K. Gengyo-Ando, S. Mitani and I. Katsura: HEN-1, a secretory protein with an LDL receptor motif, regulates sensory integration and learning in *Caenorhabditis elegans*. *Cell* 109, 639-49 (2002)
DOI: 10.1016/S0092-8674(02)00748-1
3. H. H. Lee, A. Norris, J. B. Weiss and M. Frasch: Jelly belly protein activates the receptor tyrosine kinase Alk to specify visceral muscle pioneers. *Nature* 425, 507-12 (2003)
DOI: 10.1038/nature01916
4. E. Bazigou, H. Apitz, J. Johansson, C. E. Loren, E. M. Hirst, P. L. Chen, R. H. Palmer and I. Salecker: Anterograde Jelly belly and Alk receptor tyrosine kinase signaling mediates retinal axon targeting in *Drosophila*. *Cell* 128, 961-75 (2007)
DOI: 10.1016/j.cell.2007.02.024
5. C. E. Loren, C. Englund, C. Grabbe, B. Hallberg, T. Hunter and R. H. Palmer: A crucial role for the Anaplastic lymphoma kinase receptor tyrosine kinase in gut development in *Drosophila melanogaster*. *EMBO Rep* 4, 781-6 (2003)
DOI: 10.1038/sj.embo.embor897
6. C. Englund, C. E. Loren, C. Grabbe, G. K. Varshney, F. Deleuil, B. Hallberg and R. H. Palmer: Jeb signals through the Alk receptor tyrosine kinase to drive visceral muscle fusion. *Nature* 425, 512-6 (2003)
DOI: 10.1038/nature01950
7. L. Y. Cheng, A. P. Bailey, S. J. Leevers, T. J. Ragan, P. C. Driscoll and A. P. Gould: Anaplastic lymphoma kinase spares organ growth during nutrient restriction in *Drosophila*. *Cell* 146, 435-47 (2011)
DOI: 10.1016/j.cell.2011.06.040
8. G. E. Stoica, A. Kuo, A. Aigner, I. Sunitha, B. Souttou, C. Malerczyk, D. J. Caughey, D. Wen, A. Karavanov, A. T. Riegel and A. Wellstein: Identification of anaplastic lymphoma kinase as a receptor for the growth factor pleiotrophin. *J Biol Chem* 276, 16772-9 (2001)
DOI: 10.1074/jbc.M010660200
9. G. E. Stoica, A. Kuo, C. Powers, E. T. Bowden, E. B. Sale, A. T. Riegel and A. Wellstein: Midkine binds to anaplastic lymphoma kinase (ALK) and acts as a growth factor for different cell types. *J Biol Chem* 277, 35990-8 (2002)
DOI: 10.1074/jbc.M205749200
10. P. B. Murray, I. Lax, A. Reshetnyak, G. F. Ligon, J. S. Lillquist, E. J. Natoli, Jr., X. Shi, E. Folta-Stogniew, M. Gunel, D. Alvarado and J. Schlessinger: Heparin is an activating ligand of the orphan receptor tyrosine kinase ALK. *Sci Signal* 8, ra6 (2015)
DOI: 10.1126/scisignal.2005916
11. E. Vernersson, N. K. Khoo, M. L. Henriksson, G. Roos, R. H. Palmer and B. Hallberg: Characterization of the expression of the ALK receptor tyrosine kinase in mice. *Gene Expr Patterns* 6, 448-61 (2006)
DOI: 10.1016/j.modgep.2005.11.006
12. S. W. Morris, C. Naeve, P. Mathew, P. L. James, M. N. Kirstein, X. Cui and D. P. Witte: ALK, the chromosome 2 gene locus altered by the t(2;5) in non-Hodgkin's lymphoma, encodes a novel neural receptor tyrosine kinase that is highly related to leukocyte tyrosine kinase (LTK). *Oncogene* 14, 2175-88 (1997)
DOI: 10.1038/sj.onc.1201062
13. T. Iwahara, J. Fujimoto, D. Wen, R. Cupples, N. Bucay, T. Arakawa, S. Mori, B. Ratzkin and T. Yamamoto: Molecular characterization of ALK, a receptor tyrosine kinase expressed specifically in the nervous system. *Oncogene* 14, 439-49 (1997)
DOI: 10.1038/sj.onc.1200849
14. J. G. Bilsland, A. Wheeldon, A. Mead, P. Znamenskiy, S. Almond, K. A. Waters, M. Thakur, V. Beaumont, T. P. Bonnert, R. Heavens, P. Whiting, G. McAllister and I. Munoz-Sanjuan: Behavioral and neurochemical alterations in mice deficient in anaplastic lymphoma kinase suggest therapeutic potential for psychiatric indications. *Neuropsychopharmacology* 33, 685-700 (2008)
DOI: 10.1038/sj.npp.1301446

15. A. W. Lasek, J. Gesch, F. Giorgetti, V. Kharazia and U. Heberlein: Alk is a transcriptional target of LMO4 and ERalpha that promotes cocaine sensitization and reward. *J Neurosci* 31, 14134-41 (2011)
DOI: 10.1523/JNEUROSCI.3415-11.2011
16. B. Hallberg and R. H. Palmer: Mechanistic insight into ALK receptor tyrosine kinase in human cancer biology. *Nat Rev Cancer* 13, 685-700 (2013)
DOI: 10.1038/nrc3580
DOI: 10.1038/nrc3614
17. G. Perot, I. Soubeyran, A. Ribeiro, B. Bonhomme, F. Savagner, N. Boutet-Bouzamondo, I. Hostein, F. Bonichon, Y. Godbert and F. Chibon: Identification of a recurrent STRN/ALK fusion in thyroid carcinomas. *PLoS One* 9, e87170 (2014)
DOI: 10.1371/journal.pone.0087170
18. R. E. George, T. Sanda, M. Hanna, S. Frohling, W. Luther, 2nd, J. Zhang, Y. Ahn, W. Zhou, W. B. London, P. McGrady, L. Xue, S. Zozulya, V. E. Gregor, T. R. Webb, N. S. Gray, D. G. Gilliland, L. Diller, H. Greulich, S. W. Morris, M. Meyerson and A. T. Look: Activating mutations in ALK provide a therapeutic target in neuroblastoma. *Nature* 455, 975-8 (2008)
DOI: 10.1038/nature07397
19. I. Janoueix-Lerosey, D. Lequin, L. Brugieres, A. Ribeiro, L. de Pontual, V. Combaret, V. Raynal, A. Puisieux, G. Schleiermacher, G. Pierron, D. Valteau-Couanet, T. Frebourg, J. Michon, S. Lyonnet, J. Amiel and O. Delattre: Somatic and germline activating mutations of the ALK kinase receptor in neuroblastoma. *Nature* 455, 967-70 (2008)
DOI: 10.1038/nature07398
20. Y. P. Mosse, M. Laudenslager, L. Longo, K. A. Cole, A. Wood, E. F. Attiyeh, M. J. Laquaglia, R. Bennett, J. E. Lynch, P. Perri, G. Laureys, F. Speleman, C. Kim, C. Hou, H. Hakonarson, A. Torkamani, N. J. Schork, G. M. Brodeur, G. P. Tonini, E. Rappaport, M. Devoto and J. M. Maris: Identification of ALK as a major familial neuroblastoma predisposition gene. *Nature* 455, 930-5 (2008)
DOI: 10.1038/nature07261
21. Y. Chen, J. Takita, Y. L. Choi, M. Kato, M. Ohira, M. Sanada, L. Wang, M. Soda, A. Kikuchi, T. Igarashi, A. Nakagawara, Y. Hayashi, H. Mano and S. Ogawa: Oncogenic mutations of ALK kinase in neuroblastoma. *Nature* 455, 971-4 (2008)
DOI: 10.1038/nature07399
22. R. Chiarle, C. Voena, C. Ambrogio, R. Piva and G. Inghirami: The anaplastic lymphoma kinase in the pathogenesis of cancer. *Nat Rev Cancer* 8, 11-23 (2008)
DOI: 10.1038/nrc2291
23. R. Chiarle, J. Z. Gong, I. Guasparri, A. Pesci, J. Cai, J. Liu, W. J. Simmons, G. Dhall, J. Howes, R. Piva and G. Inghirami: NPM-ALK transgenic mice spontaneously develop T-cell lymphomas and plasma cell tumors. *Blood*, 101, 1919-27 (2003)
DOI: 10.1182/blood-2002-05-1343
24. R. Chiarle, W. J. Simmons, H. Cai, G. Dhall, A. Zamo, R. Raz, J. G. Karras, D. E. Levy and G. Inghirami: Stat3 is required for ALK-mediated lymphomagenesis and provides a possible therapeutic target. *Nat Med* 11, 623-9 (2005)
DOI: 10.1038/nm1249
25. A. Zamo, R. Chiarle, R. Piva, J. Howes, Y. Fan, M. Chilosi, D. E. Levy and G. Inghirami: Anaplastic lymphoma kinase (ALK) activates Stat3 and protects hematopoietic cells from cell death. *Oncogene* 21, 1038-47 (2002)
DOI: 10.1038/sj.onc.1205152
26. A. Slupianek, M. Nieborowska-Skorska, G. Hoser, A. Morrione, M. Majewski, L. Xue, S. W. Morris, M. A. Wasik and T. Skorski: Role of phosphatidylinositol 3-kinase-Akt pathway in nucleophosmin/anaplastic lymphoma kinase-mediated lymphomagenesis. *Cancer Res* 61, 2194-9 (2001)
doi not found
27. C. Voena, C. Conte, C. Ambrogio, E. Boeri Erba, F. Boccalatte, S. Mohammed, O. N. Jensen, G. Palestro, G. Inghirami and R. Chiarle: The tyrosine phosphatase Shp2 interacts with NPM-ALK and regulates anaplastic lymphoma cell growth and migration. *Cancer Res* 67, 4278-86 (2007)
DOI: 10.1158/0008-5472.CAN-06-4350
28. C. Ambrogio, C. Voena, A. D. Manazza, R. Piva, L. Riera, L. Barberis, C. Costa, G. Tarone, P. Defilippi, E. Hirsch, E. Boeri Erba, S. Mohammed, O. N. Jensen, G. Palestro, G. Inghirami and R. Chiarle: p130Cas mediates the transforming properties of the anaplastic lymphoma kinase. *Blood* 106, 3907-16 (2005)
DOI: 10.1182/blood-2005-03-1204

29. R. Y. Bai, P. Dieter, C. Peschel, S. W. Morris and J. Duyster: Nucleophosmin-anaplastic lymphoma kinase of large-cell anaplastic lymphoma is a constitutively active tyrosine kinase that utilizes phospholipase C-gamma to mediate its mitogenicity. *Mol Cell Biol* 18, 6951-61 (1998)
doi not found
30. D. Cussac, C. Greenland, S. Roche, R. Y. Bai, J. Duyster, S. W. Morris, G. Delsol, M. Allouche and B. Payrastre: Nucleophosmin-anaplastic lymphoma kinase of anaplastic large-cell lymphoma recruits, activates, and uses pp60c-src to mediate its mitogenicity. *Blood* 103, 1464-71 (2004)
DOI: 10.1182/blood-2003-04-1038
31. K. Rikova, A. Guo, Q. Zeng, A. Possemato, J. Yu, H. Haack, J. Nardone, K. Lee, C. Reeves, Y. Li, Y. Hu, Z. Tan, M. Stokes, L. Sullivan, J. Mitchell, R. Wetzel, J. Macneill, J. M. Ren, J. Yuan, C. E. Bakalarski, J. Villen, J. M. Kornhauser, B. Smith, D. Li, X. Zhou, S. P. Gygi, T. L. Gu, R. D. Polakiewicz, J. Rush and M. J. Comb: Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell* 131, 1190-203 (2007)
DOI: 10.1016/j.cell.2007.11.025
32. M. Soda, Y. L. Choi, M. Enomoto, S. Takada, Y. Yamashita, S. Ishikawa, S. Fujiwara, H. Watanabe, K. Kurashina, H. Hatanaka, M. Bando, S. Ohno, Y. Ishikawa, H. Aburatani, T. Niki, Y. Sohara, Y. Sugiyama and H. Mano: Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 448, 561-6 (2007)
DOI: 10.1038/nature05945
33. T. Sasaki, S. J. Rodig, L. R. Chirieac and P. A. Janne: The biology and treatment of EML4-ALK non-small cell lung cancer. *Eur J Cancer* 46, 1773-80 (2010)
DOI: 10.1016/j.ejca.2010.04.002
34. K. Inamura, K. Takeuchi, Y. Togashi, K. Nomura, H. Ninomiya, M. Okui, Y. Satoh, S. Okumura, K. Nakagawa, M. Soda, Y. L. Choi, T. Niki, H. Mano and Y. Ishikawa: EML4-ALK fusion is linked to histological characteristics in a subset of lung cancers. *J Thorac Oncol* 3, 13-7 (2008)
DOI: 10.1097/JTO.0b013e31815e8b60
35. A. Yoshida, K. Tsuta, H. Nakamura, T. Kohno, F. Takahashi, H. Asamura, I. Sekine, M. Fukayama, T. Shibata, K. Furuta and H. Tsuda: Comprehensive histologic analysis of ALK-rearranged lung carcinomas. *Am J Surg Pathol* 35, 1226-34 (2011)
DOI: 10.1097/PAS.0b013e3182233e06
36. A. T. Shaw and J. A. Engelman: ALK in lung cancer: past, present, and future. *J Clin Oncol* 31, 1105-11 (2013)
DOI: 10.1200/JCO.2012.44.5353
37. J. F. Gainor, A. M. Varghese, S. H. Ou, S. Kabraji, M. M. Awad, R. Katayama, A. Pawlak, M. Mino-Kenudson, B. Y. Yeap, G. J. Riely, A. J. Iafrate, M. E. Arcila, M. Ladanyi, J. A. Engelman, D. Dias-Santagata and A. T. Shaw: ALK rearrangements are mutually exclusive with mutations in EGFR or KRAS: an analysis of 1,683 patients with non-small cell lung cancer. *Clin Cancer Res* 19, 4273-81 (2013)
DOI: 10.1158/1078-0432.CCR-13-0318
38. R. Govindan, L. Ding, M. Griffith, J. Subramanian, N. D. Dees, K. L. Kanchi, C. A. Maher, R. Fulton, L. Fulton, J. Wallis, K. Chen, J. Walker, S. McDonald, R. Bose, D. Ornitz, D. Xiong, M. You, D. J. Dooling, M. Watson, E. R. Mardis and R. K. Wilson: Genomic landscape of non-small cell lung cancer in smokers and never-smokers. *Cell* 150, 1121-34 (2012)
DOI: 10.1016/j.cell.2012.08.024
39. D. Lipson, M. Capelletti, R. Yelensky, G. Otto, A. Parker, M. Jarosz, J. A. Curran, S. Balasubramanian, T. Bloom, K. W. Brennan, A. Donahue, S. R. Downing, G. M. Frampton, L. Garcia, F. Juhn, K. C. Mitchell, E. White, J. White, Z. Zwirko, T. Peretz, H. Nechushtan, L. Soussan-Gutman, J. Kim, H. Sasaki, H. R. Kim, S. I. Park, D. Ercan, C. E. Sheehan, J. S. Ross, M. T. Cronin, P. A. Janne and P. J. Stephens: Identification of new ALK and RET gene fusions from colorectal and lung cancer biopsies. *Nat Med* 18, 382-4 (2012)
DOI: 10.1038/nm.2673
40. K. Takeuchi, M. Soda, Y. Togashi, R. Suzuki, S. Sakata, S. Hatano, R. Asaka, W. Hamanaka, H. Ninomiya, H. Uehara, Y. Lim Choi, Y. Satoh, S. Okumura, K. Nakagawa, H. Mano and Y. Ishikawa: RET, ROS1 and ALK fusions in lung cancer. *Nat Med* 18, 378-81 (2012)
DOI: 10.1038/nm.2658
41. K. Takeuchi, Y. L. Choi, Y. Togashi, M. Soda, S. Hatano, K. Inamura, S. Takada, T. Ueno,

- Y. Yamashita, Y. Satoh, S. Okumura, K. Nakagawa, Y. Ishikawa and H. Mano: KIF5B-ALK, a novel fusion oncokinase identified by an immunohistochemistry-based diagnostic system for ALK-positive lung cancer. *Clin Cancer Res* 15, 3143-9 (2009)
DOI: 10.1158/1078-0432.CCR-08-3248
42. Y. Togashi, M. Soda, S. Sakata, E. Sugawara, S. Hatano, R. Asaka, T. Nakajima, H. Mano and K. Takeuchi: KLC1-ALK: a novel fusion in lung cancer identified using a formalin-fixed paraffin-embedded tissue only. *PLoS One* 7, e31323 (2012)
DOI: 10.1371/journal.pone.0031323
43. Y. L. Choi, M. E. Lira, M. Hong, R. N. Kim, S. J. Choi, J. Y. Song, K. Pandy, D. L. Mann, J. A. Stahl, H. E. Peckham, Z. Zheng, J. Han, M. Mao and J. Kim: A novel fusion of TPR and ALK in lung adenocarcinoma. *J Thorac Oncol* 9, 563-6 (2014)
DOI: 10.1097/JTO.0000000000000093
44. D. D. Fang, B. Zhang, Q. Gu, M. Lira, Q. Xu, H. Sun, M. Qian, W. Sheng, M. Ozeck, Z. Wang, C. Zhang, X. Chen, K. X. Chen, J. Li, S. H. Chen, J. Christensen, M. Mao and C. C. Chan: HIP1-ALK, a novel ALK fusion variant that responds to crizotinib. *J Thorac Oncol* 9, 285-94 (2014)
DOI: 10.1097/JTO.0000000000000087
45. B. Solomon, K. D. Wilner and A. T. Shaw: Current status of targeted therapy for anaplastic lymphoma kinase-rearranged non-small cell lung cancer. *Clin Pharmacol Ther* 95, 15-23 (2014)
DOI: 10.1038/cpt.2013.200
46. U. McDermott, A. J. Iafrate, N. S. Gray, T. Shioda, M. Classon, S. Maheswaran, W. Zhou, H. G. Choi, S. L. Smith, L. Dowell, L. E. Ulkus, G. Kuhlmann, P. Greninger, J. G. Christensen, D. A. Haber and J. Settleman: Genomic alterations of anaplastic lymphoma kinase may sensitize tumors to anaplastic lymphoma kinase inhibitors. *Cancer Res* 68, 3389-95 (2008)
DOI: 10.1158/0008-5472.CAN-07-6186
47. J. M. Heuckmann, H. Balke-Want, F. Malchers, M. Peifer, M. L. Sos, M. Koker, L. Meder, C. M. Lovly, L. C. Heukamp, W. Pao, R. Koppers and R. K. Thomas: Differential Protein Stability and ALK Inhibitor Sensitivity of EML4-ALK Fusion Variants. *Clin Cancer Res* 18, 4682-90 (2012)
DOI: 10.1158/1078-0432.CCR-11-3260
48. M. Soda, S. Takada, K. Takeuchi, Y. L. Choi, M. Enomoto, T. Ueno, H. Haruta, T. Hamada, Y. Yamashita, Y. Ishikawa, Y. Sugiyama and H. Mano: A mouse model for EML4-ALK-positive lung cancer. *Proc Natl Acad Sci U S A* 105, 19893-7 (2008)
DOI: 10.1073/pnas.0805381105
49. Z. Chen, T. Sasaki, X. Tan, J. Carretero, T. Shimamura, D. Li, C. Xu, Y. Wang, G. O. Adelman, M. Capelletti, H. J. Lee, S. J. Rodig, C. Borgman, S. I. Park, H. R. Kim, R. Padera, J. A. Marto, N. S. Gray, A. L. Kung, G. I. Shapiro, P. A. Janne and K. K. Wong: Inhibition of ALK, PI3K/MEK, and HSP90 in murine lung adenocarcinoma induced by EML4-ALK fusion oncogene. *Cancer Res* 70, 9827-36 (2010)
DOI: 10.1158/0008-5472.CAN-10-1671
50. D. Maddalo, E. Manchado, C. P. Concepcion, C. Bonetti, J. A. Vidigal, Y. C. Han, P. Ogrodowski, A. Crippa, N. Rekhtman, E. de Stanchina, S. W. Lowe and A. Ventura: *In vivo* engineering of oncogenic chromosomal rearrangements with the CRISPR/Cas9 system. *Nature* (2014)
DOI: 10.1038/nature13902
51. R. B. Blasco, E. Karaca, C. Ambrogio, T. C. Cheong, E. Karayol, V. G. Minero, C. Voena and R. Chiarle: Simple and Rapid *In vivo* Generation of Chromosomal Rearrangements using CRISPR/Cas9 Technology. *Cell Rep* 9, 1219-27 (2014)
DOI: 10.1016/j.celrep.2014.10.051
52. C. A. Griffin, A. L. Hawkins, C. Dvorak, C. Henkle, T. Ellingham and E. J. Perlman: Recurrent involvement of 2p23 in inflammatory myofibroblastic tumors. *Cancer Res* 59, 2776-80 (1999)
doi not found
53. B. Lawrence, A. Perez-Atayde, M. K. Hibbard, B. P. Rubin, P. Dal Cin, J. L. Pinkus, G. S. Pinkus, S. Xiao, E. S. Yi, C. D. Fletcher and J. A. Fletcher: TPM3-ALK and TPM4-ALK oncogenes in inflammatory myofibroblastic tumors. *Am J Pathol* 157, 377-84 (2000)
DOI: 10.1016/S0002-9440(10)64550-6
54. M. Debiec-Rychter, P. Marynen, A. Hagemeijer and P. Pauwels: ALK-ATIC fusion in urinary bladder inflammatory myofibroblastic tumor.

- Genes Chromosomes Cancer* 38, 187-90 (2003)
DOI: 10.1002/gcc.10267
55. J. A. Bridge, M. Kanamori, Z. Ma, D. Pickering, D. A. Hill, W. Lydiatt, M. Y. Lui, G. W. Colleoni, C. R. Antonescu, M. Ladanyi and S. W. Morris: Fusion of the ALK gene to the clathrin heavy chain gene, CLTC, in inflammatory myofibroblastic tumor. *Am J Pathol* 159, 411-5 (2001)
DOI: 10.1016/S0002-9440(10)61711-7
56. Z. Ma, D. A. Hill, M. H. Collins, S. W. Morris, J. Sumegi, M. Zhou, C. Zuppan and J. A. Bridge: Fusion of ALK to the Ran-binding protein 2 (RANBP2) gene in inflammatory myofibroblastic tumor. *Genes Chromosomes Cancer* 37, 98-105 (2003)
DOI: 10.1002/gcc.10177
57. L. V. Debelenko, D. C. Arthur, S. D. Pack, L. J. Helman, D. S. Schrump and M. Tsokos: Identification of CARS-ALK fusion in primary and metastatic lesions of an inflammatory myofibroblastic tumor. *Lab Invest* 83, 1255-65 (2003)
DOI: 10.1097/01.LAB.0000088856.49388.EA
58. I. Panagopoulos, T. Nilsson, H. A. Domanski, M. Isaksson, P. Lindblom, F. Mertens and N. Mandahl: Fusion of the SEC31L1 and ALK genes in an inflammatory myofibroblastic tumor. *Int J Cancer* 118, 1181-6 (2006)
DOI: 10.1002/ijc.21490
59. A. S. Patel, K. M. Murphy, A. L. Hawkins, J. S. Cohen, P. P. Long, E. J. Perlman and C. A. Griffin: RANBP2 and CLTC are involved in ALK rearrangements in inflammatory myofibroblastic tumors. *Cancer Genet Cytogenet* 17, 107-14 (2007)
DOI: 10.1016/j.cancergenryo.2007.04.004
60. Z. Tothova and A. J. Wagner: Anaplastic lymphoma kinase-directed therapy in inflammatory myofibroblastic tumors. *Curr Opin Oncol* 24, 409-13 (2012)
DOI: 10.1097/CCO.0b013e328354c155
61. J. E. Butrynski, D. R. D'Adamo, J. L. Hornick, P. Dal Cin, C. R. Antonescu, S. C. Jhanwar, M. Ladanyi, M. Capelletti, S. J. Rodig, N. Ramaiya, E. L. Kwak, J. W. Clark, K. D. Wilner, J. G. Christensen, P. A. Janne, R. G. Maki, G. D. Demetri and G. I. Shapiro: Crizotinib in ALK-rearranged inflammatory myofibroblastic tumor. *N Engl J Med* 363, 1727-33 (2010)
DOI: 10.1056/NEJMoa1007056
62. T. Sasaki, K. Okuda, W. Zheng, J. Butrynski, M. Capelletti, L. Wang, N. S. Gray, K. Wilner, J. G. Christensen, G. Demetri, G. I. Shapiro, S. J. Rodig, M. J. Eck and P. A. Janne: The neuroblastoma-associated F1174L ALK mutation causes resistance to an ALK kinase inhibitor in ALK-translocated cancers. *Cancer Res* 70, 10038-43 (2010)
DOI: 10.1158/0008-5472.CAN-10-2956
63. L. V. Debelenko, S. C. Raimondi, N. Daw, B. R. Shivakumar, D. Huang, M. Nelson and J. A. Bridge: Renal cell carcinoma with novel VCL-ALK fusion: new representative of ALK-associated tumor spectrum. *Mod Pathol* 24, 430-42 (2011)
DOI: 10.1038/modpathol.2010.213
64. A. Marino-Enriquez, W. B. Ou, C. B. Weldon, J. A. Fletcher and A. R. Perez-Atayde: ALK rearrangement in sickle cell trait-associated renal medullary carcinoma. *Genes Chromosomes Cancer* 50, 146-53 (2011)
DOI: 10.1002/gcc.20839
65. W. R. Sukov, J. C. Hodge, C. M. Lohse, M. K. Akre, B. C. Leibovich, R. H. Thompson and J. C. Cheville: ALK alterations in adult renal cell carcinoma: frequency, clinicopathologic features and outcome in a large series of consecutively treated patients. *Mod Pathol* 25, 1516-25 (2012)
DOI: 10.1038/modpathol.2012.107
66. E. Lin, L. Li, Y. Guan, R. Soriano, C. S. Rivers, S. Mohan, A. Pandita, J. Tang and Z. Modrusan: Exon array profiling detects EML4-ALK fusion in breast, colorectal, and non-small cell lung cancers. *Mol Cancer Res* 7, 1466-76 (2009)
DOI: 10.1158/1541-7786.MCR-08-0522
67. N. Yokoyama and W. T. Miller: Molecular characterization of WDCP, a novel fusion partner for the anaplastic lymphoma tyrosine kinase ALK. *Biomed Rep* 3, 9-13 (2015)
doi not found
68. H. Ren, Z. P. Tan, X. Zhu, K. Crosby, H. Haack, J. M. Ren, S. Beausoleil, A. Moritz, G. Innocenti, J. Rush, Y. Zhang, X. M. Zhou, T. L. Gu, Y. F. Yang and M. J. Comb: Identification of anaplastic lymphoma kinase as a potential therapeutic target in ovarian cancer. *Cancer Res* 72, 3312-23 (2012)
DOI: 10.1158/0008-5472.CAN-11-3931
69. T. Wiesner, J. He, R. Yelensky, R. Esteve-Puig, T. Botton, I. Yeh, D. Lipson, G. Otto, K.

- Brennan, R. Murali, M. Garrido, V. A. Miller, J. S. Ross, M. F. Berger, A. Sparatta, G. Palmedo, L. Cerroni, K. J. Busam, H. Kutzner, M. T. Cronin, P. J. Stephens and B. C. Bastian: Kinase fusions are frequent in Spitz tumours and spitzoid melanomas. *Nat Commun* 5, 3116 (2014)
DOI: 10.1038/ncomms4116
70. L. M. Kelly, G. Barila, P. Liu, V. N. Evdokimova, S. Trivedi, F. Panebianco, M. Gandhi, S. E. Carty, S. P. Hodak, J. Luo, S. Dacic, Y. P. Yu, M. N. Nikiforova, R. L. Ferris, D. L. Altschuler and Y. E. Nikiforov: Identification of the transforming STRN-ALK fusion as a potential therapeutic target in the aggressive forms of thyroid cancer. *Proc Natl Acad Sci U S A* 111, 4233-8 (2014)
DOI: 10.1073/pnas.1321937111
71. J. M. Maris: Recent advances in neuroblastoma. *N Engl J Med* 362, 2202-11 (2010)
DOI: 10.1056/NEJMra0804577
72. J. M. Maris, M. D. Hogarty, R. Bagatell and S. L. Cohn: Neuroblastoma. *Lancet* 369, 2106-20 (2007)
DOI: 10.1016/S0140-6736(07)60983-0
73. S. Ogawa, J. Takita, M. Sanada and Y. Hayashi: Oncogenic mutations of ALK in neuroblastoma. *Cancer Sci* 102, 302-8 (2011)
DOI: 10.1111/j.1349-7006.2010.01825.x
74. S. De Brouwer, K. De Preter, C. Kumps, P. Zabrocki, M. Porcu, E. M. Westerhout, A. Lakeman, J. Vandesompele, J. Hoebeeck, T. Van Maerken, A. De Paepe, G. Laureys, J. H. Schulte, A. Schramm, C. Van Den Broecke, J. Vermeulen, N. Van Roy, K. Beiske, M. Renard, R. Noguera, O. Delattre, I. Janoueix-Lerosey, P. Kognier, T. Martinsson, A. Nakagawara, M. Ohira, H. Caron, A. Eggert, J. Cools, R. Versteeg and F. Speleman: Meta-analysis of neuroblastomas reveals a skewed ALK mutation spectrum in tumors with MYCN amplification. *Clin Cancer Res* 16, 4353-62 (2010)
DOI: 10.1158/1078-0432.CCR-09-2660
75. L. Passoni, L. Longo, P. Collini, A. M. Coluccia, F. Bozzi, M. Podda, A. Gregorio, C. Gambini, A. Garaventa, V. Pistoia, F. Del Gross, G. P. Tonini, M. Cheng, C. Gambacorti-Passerini, A. Anichini, F. Fossati-Bellani, M. Di Nicola and R. Luksch: Mutation-independent anaplastic lymphoma kinase overexpression in poor prognosis neuroblastoma patients. *Cancer Res* 69, 7338-46 (2009)
DOI: 10.1158/0008-5472.CAN-08-4419
76. J. H. Schulte, H. S. Bachmann, B. Brockmeyer, K. Depreter, A. Oberthur, S. Ackermann, Y. Kahlert, K. Pajtler, J. Theissen, F. Westermann, J. Vandesompele, F. Speleman, F. Berthold, A. Eggert, B. Brors, B. Hero, A. Schramm and M. Fischer: High ALK receptor tyrosine kinase expression supersedes ALK mutation as a determining factor of an unfavorable phenotype in primary neuroblastoma. *Clin Cancer Res* 17, 5082-92 (2011)
DOI: 10.1158/1078-0432.CCR-10-2809
77. D. Di Paolo, C. Ambrogio, F. Pastorino, C. Brignole, C. Martinengo, R. Carosio, M. Loi, G. Pagnan, L. Emionite, M. Cilli, D. Ribatti, T. M. Allen, R. Chiarle, M. Ponzoni and P. Perri: Selective therapeutic targeting of the anaplastic lymphoma kinase with liposomal siRNA induces apoptosis and inhibits angiogenesis in neuroblastoma. *Mol Ther* 19, 2201-12 (2011)
DOI: 10.1038/mt.2011.142
78. T. Berry, W. Luther, N. Bhatnagar, Y. Jamin, E. Poon, T. Sanda, D. Pei, B. Sharma, W. R. Vetharoy, A. Hallsworth, Z. Ahmad, K. Barker, L. Moreau, H. Webber, W. Wang, Q. Liu, A. Perez-Atayde, S. Rodig, N. K. Cheung, F. Raynaud, B. Hallberg, S. P. Robinson, N. S. Gray, A. D. Pearson, S. A. Eccles, L. Chesler and R. E. George: The ALK(F1174L) mutation potentiates the oncogenic activity of MYCN in neuroblastoma. *Cancer Cell* 22, 117-30 (2012)
DOI: 10.1016/j.ccr.2012.06.001
79. L. C. Heukamp, T. Thor, A. Schramm, K. De Preter, C. Kumps, B. De Wilde, A. Odersky, M. Peifer, S. Lindner, A. Spruessel, F. Pattyn, P. Mestdagh, B. Menten, S. Kuhfittig-Kulle, A. Kunkele, K. Konig, L. Meder, S. Chatterjee, R. T. Ullrich, S. Schulte, J. Vandesompele, F. Speleman, R. Buttner, A. Eggert and J. H. Schulte: Targeted expression of mutated ALK induces neuroblastoma in transgenic mice. *Sci Transl Med* 4, 141ra91 (2012)
doi not found
80. S. C. Bresler, A. C. Wood, E. A. Haglund, J. Courtright, L. T. Belcastro, J. S. Plegaria, K. Cole, Y. Toporovskaya, H. Zhao, E. L.

- Carpenter, J. G. Christensen, J. M. Maris, M. A. Lemmon and Y. P. Mosse: Differential inhibitor sensitivity of anaplastic lymphoma kinase variants found in neuroblastoma. *Sci Transl Med* 3, 108ra114 (2011)
doi not found
81. S. Zhu, J. S. Lee, F. Guo, J. Shin, A. R. Perez-Atayde, J. L. Kutok, S. J. Rodig, D. S. Neuberg, D. Helman, H. Feng, R. A. Stewart, W. Wang, R. E. George, J. P. Kanki and A. T. Look: Activated ALK collaborates with MYCN in neuroblastoma pathogenesis. *Cancer Cell* 21, 362-73 (2012)
DOI: 10.1016/j.ccr.2012.02.010
82. A. Cazes, L. Lopez-Delisle, K. Tsarovina, C. Pierre-Eugene, K. De Preter, M. Peuchmaur, A. Nicolas, C. Provost, C. Louis-Brennertot, R. Daveau, C. Kumps, I. Cascone, G. Schleiermacher, A. Prignon, F. Speleman, H. Rohrer, O. Delattre and I. Janoueix-Lerosey: Activated Alk triggers prolonged neurogenesis and Ret upregulation providing a therapeutic target in ALK-mutated neuroblastoma. *Oncotarget* 5, 2688-702 (2014)
doi not found
83. A. K. Murugan and M. Xing: Anaplastic thyroid cancers harbor novel oncogenic mutations of the ALK gene. *Cancer Res* 71, 4403-11 (2011)
DOI: 10.1158/0008-5472.CAN-10-4041
84. A. Chou, S. Fraser, C. W. Toon, A. Clarkson, L. Sioson, M. Farzin, C. Cussigh, A. Aniss, C. O'Neill, N. Watson, R. J. Clifton-Bligh, D. L. Learoyd, B. G. Robinson, C. I. Selinger, L. W. Delbridge, S. B. Sidhu, S. A. O'Toole, M. Sywak and A. J. Gill: A Detailed Clinicopathologic Study of ALK-translocated Papillary Thyroid Carcinoma. *Am J Surg Pathol* (2014)
DOI: 10.1097/PAS.0000000000000368
85. J. C. van Gaal, U. E. Flucke, M. H. Roeffen, E. S. de Bont, S. Sleijfer, A. M. Mavinkurve-Groothuis, A. J. Suurmeijer, W. T. van der Graaf and Y. M. Versleijen-Jonkers: Anaplastic lymphoma kinase aberrations in rhabdomyosarcoma: clinical and prognostic implications. *J Clin Oncol* 30, 308-15 (2012)
DOI: 10.1200/JCO.2011.37.8588
86. P. Bonvini, A. Zin, R. Alaggio, B. Pawel, G. Bisogno and A. Rosolen: High ALK mRNA expression has a negative prognostic significance in rhabdomyosarcoma. *Br J Cancer* 109, 3084-91 (2013)
DOI: 10.1038/bjc.2013.653
87. E. L. Kwak, Y. J. Bang, D. R. Camidge, A. T. Shaw, B. Solomon, R. G. Maki, S. H. Ou, B. J. Dezube, P. A. Janne, D. B. Costa, M. Varella-Garcia, W. H. Kim, T. J. Lynch, P. Fidias, H. Stubbs, J. A. Engelmann, L. V. Sequist, W. Tan, L. Gandhi, M. Mino-Kenudson, G. C. Wei, S. M. Shreeve, M. J. Ratain, J. Settleman, J. G. Christensen, D. A. Haber, K. Wilner, R. Salgia, G. I. Shapiro, J. W. Clark and A. J. Iafrate: Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 363, 1693-703 (2010)
DOI: 10.1056/NEJMoa1006448
88. D. R. Camidge, Y. J. Bang, E. L. Kwak, A. J. Iafrate, M. Varella-Garcia, S. B. Fox, G. J. Riely, B. Solomon, S. H. Ou, D. W. Kim, R. Salgia, P. Fidias, J. A. Engelmann, L. Gandhi, P. A. Janne, D. B. Costa, G. I. Shapiro, P. Lorusso, K. Ruffner, P. Stephenson, Y. Tang, K. Wilner, J. W. Clark and A. T. Shaw: Activity and safety of crizotinib in patients with ALK-positive non-small-cell lung cancer: updated results from a phase 1 study. *Lancet Oncol* 13, 1011-9 (2012)
DOI: 10.1016/S1470-2045(12)70344-3
89. Y. L. Choi, M. Soda, Y. Yamashita, T. Ueno, J. Takashima, T. Nakajima, Y. Yatabe, K. Takeuchi, T. Hamada, H. Haruta, Y. Ishikawa, H. Kimura, T. Mitsudomi, Y. Tanio and H. Mano: EML4-ALK mutations in lung cancer that confer resistance to ALK inhibitors. *N Engl J Med* 363, 1734-9 (2010)
DOI: 10.1056/NEJMoa1007478
90. A. T. Shaw, D. W. Kim, K. Nakagawa, T. Seto, L. Crino, M. J. Ahn, T. De Pas, B. Besse, B. J. Solomon, F. Blackhall, Y. L. Wu, M. Thomas, K. J. O'Byrne, D. Moro-Sibilot, D. R. Camidge, T. Mok, V. Hirsh, G. J. Riely, S. Iyer, V. Tassell, A. Polli, K. D. Wilner and P. A. Janne: Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med* 368, 2385-94 (2013)
DOI: 10.1056/NEJMoa1214886
91. R. Katayama, A. T. Shaw, T. M. Khan, M. Mino-Kenudson, B. J. Solomon, B. Halmos, N. A. Jessop, J. C. Wain, A. T. Yeo, C. Benes, L. Drew, J. C. Saeh, K. Crosby, L. V. Sequist, A. J. Iafrate and J. A. Engelmann: Mechanisms of acquired crizotinib resistance in ALK-rearranged lung cancers. *Sci Transl Med* 4, 120ra17 (2012)
doi not found

92. C. Voena and R. Chiarle: The battle against ALK resistance: successes and setbacks. *Expert Opin Investig Drugs* 21, 1751-4 (2012)
DOI: 10.1517/13543784.2012.717930
93. C. Voena, F. Di Giacomo, E. Panizza, L. D'Amico, F. E. Boccalatte, E. Pellegrino, M. Todaro, D. Recupero, F. Tabbo, C. Ambrogio, C. Martinengo, L. Bonello, R. Pulito, J. Hamm, R. Chiarle, M. Cheng, B. Ruggeri, E. Medico and G. Inghirami: The EGFR family members sustain the neoplastic phenotype of ALK+ lung adenocarcinoma via EGR1. *Oncogenesis*, 2, e43 (2013).
DOI: 10.1038/oncsis.2013.7
94. M. M. Awad and A. T. Shaw: ALK Inhibitors in Non-Small Cell Lung Cancer: Crizotinib and Beyond. *Clin Adv Hematol Oncol* 12, 429-39 (2014)
doi not found
95. A. T. Shaw, D. W. Kim, R. Mehra, D. S. Tan, E. Felip, L. Q. Chow, D. R. Camidge, J. Vansteenkiste, S. Sharma, T. De Pas, G. J. Riely, B. J. Solomon, J. Wolf, M. Thomas, M. Schuler, G. Liu, A. Santoro, Y. Y. Lau, M. Goldwasser, A. L. Boral and J. A. Engelman: Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med* 370, 1189-97 (2014)
DOI: 10.1056/NEJMoa1311107
96. L. Friboulet, N. Li, R. Katayama, C. C. Lee, J. F. Gainor, A. S. Crystal, P. Y. Michellys, M. M. Awad, N. Yanagitani, S. Kim, A. C. Pferdekamper, J. Li, S. Kasibhatla, F. Sun, X. Sun, S. Hua, P. McNamara, S. Mahmood, E. L. Lockerman, N. Fujita, M. Nishio, J. L. Harris, A. T. Shaw and J. A. Engelman: The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discov* 4, 662-73 (2014)
doi not found
97. Y. P. Mosse, M. S. Lim, S. D. Voss, K. Wilner, K. Ruffner, J. Laliberte, D. Rolland, F. M. Balis, J. M. Maris, B. J. Weigel, A. M. Ingle, C. Ahern, P. C. Adamson and S. M. Blaney: Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: a Children's Oncology Group phase 1 consortium study. *Lancet Oncol* 14, 472-80 (2013)
DOI: 10.1016/S1470-2045(13)70095-0
98. E. L. Carpenter and Y. P. Mosse: Targeting ALK in neuroblastoma--preclinical and clinical advancements. *Nat Rev Clin Oncol* 9, 391-9 (2012)
DOI: 10.1038/nrclinonc.2012.72
99. L. V. Sequist, S. Gettinger, N. N. Senzer, R. G. Martins, P. A. Janne, R. Lilienbaum, J. E. Gray, A. J. Iafrate, R. Katayama, N. Hafeez, J. Sweeney, J. R. Walker, C. Fritz, R. W. Ross, D. Grayzel, J. A. Engelman, D. R. Borger, G. Paez and R. Natale: Activity of IPI-504, a novel heat-shock protein 90 inhibitor, in patients with molecularly defined non-small-cell lung cancer. *J Clin Oncol* 28, 4953-60 (2010)
DOI: 10.1200/JCO.2010.30.8338
100. J. Sang, J. Acquaviva, J. C. Friedland, D. L. Smith, M. Sequeira, C. Zhang, Q. Jiang, L. Xue, C. M. Lovly, J. P. Jimenez, A. T. Shaw, R. C. Doebele, S. He, R. C. Bates, D. R. Camidge, S. W. Morris, I. El-Hariry and D. A. Proia: Targeted inhibition of the molecular chaperone Hsp90 overcomes ALK inhibitor resistance in non-small cell lung cancer. *Cancer Discov* 3, 430-43 (2013)
DOI: 10.1158/2159-8290.CD-12-0440
101. J. R. Brahmer, S. S. Tykodi, L. Q. Chow, W. J. Hwu, S. L. Topalian, P. Hwu, C. G. Drake, L. H. Camacho, J. Kauh, K. Odunsi, H. C. Pitot, O. Hamid, S. Bhatia, R. Martins, K. Eaton, S. Chen, T. M. Salay, S. Alaparthi, J. F. Gross, A. J. Korman, S. M. Parker, S. Agrawal, S. M. Goldberg, D. M. Pardoll, A. Gupta and J. M. Wigginton: Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med* 366, 2455-65 (2012)
DOI: 10.1056/NEJMoa1200694
102. S. L. Topalian, F. S. Hodi, J. R. Brahmer, S. N. Gettinger, D. C. Smith, D. F. McDermott, J. D. Powderly, R. D. Carvajal, J. A. Sosman, M. B. Atkins, P. D. Leming, D. R. Spigel, S. J. Antonia, L. Horn, C. G. Drake, D. M. Pardoll, L. Chen, W. H. Sharfman, R. A. Anders, J. M. Taube, T. L. McMiller, H. Xu, A. J. Korman, M. Jure-Kunkel, S. Agrawal, D. McDonald, G. D. Kollia, A. Gupta, J. M. Wigginton and M. Sznol: Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med* 366, 2443-54 (2012)
DOI: 10.1056/NEJMoa1200690
103. L. Passoni and C. Gambacorti-Passerin: ALK a novel lymphoma-associated tumor antigen for vaccination strategies. *Leuk Lymphoma* 44, 1675-81 (2003)
DOI: 10.1080/1042819031000099625

104. C. Mastini, C. Martinengo, G. Inghirami and R. Chiarle: Anaplastic lymphoma kinase: an oncogene for tumor vaccination. *J Mol Med* 87, 669-77 (2009)
DOI: 10.1007/s00109-009-0460-5
105. R. Chiarle, C. Martinengo, C. Mastini, C. Ambrogio, V. D'Escamard, G. Forni and G. Inghirami: The anaplastic lymphoma kinase is an effective oncoantigen for lymphoma vaccination. *Nat Med* 14, 676-80 (2008)
DOI: 10.1038/nm1769
106. C. Moog-Lutz, J. Degoutin, J. Y. Gouzi, Y. Frobert, N. Brunet-de Carvalho, J. Bureau, C. Creminon and M. Vigny: Activation and inhibition of anaplastic lymphoma kinase receptor tyrosine kinase by monoclonal antibodies and absence of agonist activity of pleiotrophin. *J Biol Chem* 280, 26039-48 (2005)
DOI: 10.1074/jbc.M501972200
107. E. L. Carpenter, E. A. Haglund, E. M. Mace, D. Deng, D. Martinez, A. C. Wood, A. K. Chow, D. A. Weiser, L. T. Belcastro, C. Winter, S. C. Bresler, S. Asgharzadeh, R. C. Seeger, H. Zhao, R. Guo, J. G. Christensen, J. S. Orange, B. R. Pawel, M. A. Lemmon and Y. P. Mosse: Antibody targeting of anaplastic lymphoma kinase induces cytotoxicity of human neuroblastoma. *Oncogene* 31, 4859-67 (2012)
DOI: 10.1038/onc.2011.647
DOI: 10.1038/onc.2012.208

Harvard Medical School, Enders 1116.1., 300 Longwood Ave, Boston, MA 02115, Tel: 617-919-2662, Fax: 617-730-0148, E-mail: roberto.chiarle@childrens.harvard.edu

Abbreviations: ALK: anaplastic lymphoma kinase; NSCLC: non-small cell lung carcinoma; IMT: inflammatory myofibroblastic tumor; DLBCL: diffuse large B-cell lymphoma; RCC: renal cell carcinoma; NB: neuroblastoma; ATC: anaplastic thyroid cancer; HEN-1: hesitation behavior-1; NPM: nucleophosmin; EML4: echinoderm-microtubule-associated protein-like 4; TFG: TRK-fused gene; KIF5B: kinesin family member 5B; KLC1: kinesin light chain 1; STRN: striatin; HIP1: huntingtin interacting protein 1; TPR: translocated promoter region; DCTN1: dynactin 1

Key Words: ALK-rearrangements, Non-Small Cell Lung Cancer, ALK mutations, Neuroblastoma, Solid tumors, Targeted therapy, Review

Send correspondence to: Roberto Chiarle, Department of Pathology, Children's Hospital Boston, Associate Professor in Pathology,