

Review

Maturation, Morphology, and Function: The Decisive Role of Intestinal Flora on Microglia: A Review

Lichao Liu¹, Fan Tong¹, Huanhuan Li¹, Yawen Bin¹, Peng Ding¹, Ling Peng¹, Zhiwei Liu¹, Xiaorong Dong^{1,*}¹Cancer Center, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, 430022 Wuhan, Hubei, China*Correspondence: xiaorongdong@hust.edu.cn (Xiaorong Dong)

Academic Editor: Giovanna Traina

Submitted: 12 October 2022 Revised: 16 January 2023 Accepted: 19 January 2023 Published: 9 May 2023

Abstract

Recent studies have shown that the gut microbiota regulates intestinal function and maintains intestinal homeostasis, as well as interacting with the central nervous system to affect brain function and human behavior. Microglia are the most common immune cell type in the central nervous system during homeostasis. These cells play an important role in immune surveillance by responding to infections and other pathological conditions. Microglia also play a major role in maintaining brain homeostasis in both developing and adult mice by phagocytosing cell debris and regulating the formation of neural networks. The specific signaling pathways and cytokines that control the maturation and activation of microglia are currently not fully established. However, research on germ-free (GF) mice and specific pathogen-free (SPF) mice indicate that gut microbiota have important interactions with microglia. Here, we review the latest research findings on how gut microbiota can affect the morphology, maturation, phenotype and function of microglia. We also discuss recent advances in the gut microbiota-microglia-disease axis.

Keywords: gut microbiota; microglia; gut-brain axis; neurodegenerative diseases; neuroinflammation

1. Introduction

Interaction between gut microbiota and the human body, especially the central nervous system, has been demonstrated in numerous studies. These includes studies of the gut-brain axis, as well as animal models of microbiota and brain diseases. Gut microbiota can have bidirectional communications with the brain, thereby affecting behaviors such as social interaction [1], depression-like behavior [2] and anxiety-like behavior [2]. Gut microbiota also show close relationships with the pathological course of several neurological inflammatory diseases. The four main ways in which gut microbiota can affect host brain activity include the vagus nerve pathway [3], the neurotransmitter pathway [4], the neuroendocrine pathway [5], and the immune pathway [6]. The mechanism by which hosts regulate their gut microbiota is thought to be *via* miRNA control of bacterial gene expression [7], with the miRNAs originating from the host epithelium.

Microglia arise from the embryonic yolk sac and are the most common immune cells residing in the brain parenchyma. They are involved in a wide variety of processes in the central nervous system, including defense against infection, mediating inflammation, and phagocytosis of cell debris. Recent studies have found that microglia play important roles in neurodevelopment in addition to traditional macrophage functions. These include: (1) regulation of neuronal cell programmed death [8], (2) promotion of axon formation, synapse generation, and fasciculation of

axons [9,10], and (3) stripping of excess synapses from developing neurons in order to form functional neuronal circuits [11]. Such important functions mean that microglia also play key roles in several diseases of the nervous system, including Alzheimer's disease (AD) [12], Parkinson's disease (PD) [13], multiple sclerosis (MS) [14] and glioma [15].

2. Gut Microbiota Affect the Morphology of Microglia

Microglia have a variety of shapes, including ramified-, amoeboid-, hypertrophic-, rod-, dystrophic-, and satellite-type [16]. The morphology of microglia is thought to be closely related to their function. Rod-shaped microglia participate in the detachment of presynaptic membrane from its corresponding postsynaptic membrane [17]. Amoeboid-shaped microglia exist mainly in the developing central nervous system and during some acute infections of the central nervous system, where they are thought to be involved in myelination and in certain inflammatory processes, respectively [18]. Compared with specific pathogen-free (SPF) mice, the microglia of germ-free (GF) mice have a ramified-type morphology, with longer protrusions and more branch and end points [19]. Moreover, they do not show a normal immune response to Lipopolysaccharide (LPS) or to viral infection. Highly branched microglia are also seen in mice models after long-term treatment with broad-spectrum antibiotics. Researchers have



also found that microglia morphology in a mouse model of Alzheimer disease (AD) is affected by the microbial population in a gender-specific manner [20]. In male AD mice treated with broad-spectrum antibiotics, the microglia are clustered around amyloid plaques and show smaller cell bodies, longer tree-like branches, and more branching points. On the other hand, the microglia in female AD mice show a larger cell body, and no significant changes in the tree-like branches and branch points. The pathological process in male AD mice was also found to be attenuated after treatment with broad-spectrum antibiotics. In a mouse model that combined malnutrition with iterative exposure to fecal commensals, the microglia showed a smaller cell volume and territory, but no significant change in the number of endpoints and branch points [21]. Thus, gut microbiota has an important influence on the morphology of microglia. Although the function of microglia appears to be closely related to their morphology, the specific mechanism involved is unclear and requires further research.

3. Gut Microbiota Affect the Maturation of Microglia

Microglia originate from the yolk sac of the embryo. They enter the brain when the first neurons are generated at around embryonic day 9.5 in mice (E9.5), and self-renew in adulthood. Erny *et al.* [19] found that gut microbiota is an important factor in the maturation and gene expression of microglia. RNA sequencing analysis revealed that microglia RNA transcripts from GF mice are significantly different to those of SPF mice.

Some genes related to microglia activation and immune function are down-regulated in GF mice compared to SPF mice, including *Mapk8*, *Fcgr2 β* , *IL-1 α* , *Ly86*, *Cd86*, *Hif1 α* , *B2m*, *Stat1* and *JAK3* [19]. In contrast, TRIM family members (such as *Trim30 α* , involved in pathogen-recognition and regulation of transcriptional pathways in host defense), inhibitors of transcription (such as *Nfkb α* , encoding I κ B α), and the transcription and survival factors *Spf1* (encoding PU.1) and *Csf1r*, were up-regulated [19]. Genes that are often up-regulated in mature microglial cells are involved in the activation of cell proliferation (*Iqgap1*, *Ddit4*), stimulation of the cell cycle (*Cdk9* and *Ccnd3*), and inhibition of autophagy (*Bcl2*) [19]. The expression of *CSF1R*, *F4/80* and *CD31* decreased significantly during the maturation of microglia. Under GF conditions, however, the percentage of *CSF1R*⁺, *F4/80*⁺ and *CD31*⁺ microglia cells increased significantly, together with the mean fluorescence intensity of *CSF1R* and *F4/80* [19]. The above studies show that the maturation of microglia is clearly slowed in the absence of microbiota. Furthermore, Morgan *et al.* [22] reported that the effect of gut microbiota on microglia maturation during embryonic development showed gender dimorphism. Only 19 genes were found to be differentially expressed in microglia between GF mice and SPF mice at E14.5. The influence of gut microbiota shows

more gender specificity close to birth. Compared to SPF mice embryos, 1216 genes were differentially expressed in microglia from E18.5 GF male mice, but only 20 in microglia from E18.5 GF female mice. In adults, 433 genes were differentially expressed in microglia from female GF mice and only 26 in male GF mice compared with SPF mice. The differentially expressed genes in microglia from males were mainly related to translation and metabolism. In microglia from females, inflammation-related genes were down-regulated, while genes involved in the regulation of transcription were up-regulated.

Investigators have also studied Altered Schaedler Flora mice (ASF mice, mouse model with 8 strains of bacteria) in addition to SPF mice [19]. Although the ASF mice and SPF mice were made to have the same microbial load, their microglia were still very different in terms of morphology, function and maturity. This study showed that single or several different bacterial populations do not help microglia to mature, and that maturation requires complex and diverse microbial populations. By transplanting the microbiota of SPF mice, the immature microglia phenotype of GF mice can essentially return to a normal state and full maturity can be reached in adulthood. Adult SPF mice show an immature microglia phenotype following treatment with broad-spectrum antibiotics. Therefore, the involvement of microbiota on the maturation of microglia continues from embryonic development through to adulthood. The above research findings confirm that microglia in GF mice are in an immature state, but can return to a normal state after the intestinal microbiota is restored. This suggests a close connection between the maturation state of microglia and the intestinal microbiota, although the specific mechanism requires further in-depth study.

4. Gut Microbiota Affect the Microglia Phenotype

Similar to peripheral macrophages, microglia have different states under different stimulating factors. Researchers have simplified these into the M1 and M2 phenotypes in order to distinguish two different polarization states [23–25]. Polarization to M1 phenotype after stimulation with IFN- γ , TNF or TLR, and to M2 phenotype after stimulation with IL-4, IL-10 or IL-13. They can also convert to each other. The M1 phenotype is considered to be an anti-tumor or inflammation-promoting phenotype with a classical activation pathway. This microglia phenotype also appears to secrete more TNF- β , IL-1 β , superoxide, nitric oxide and active oxygen. The M2 phenotype results from *Alternative Activation* and *Acquired Deactivation*, both of which are believed to have anti-inflammatory, tissue repair and reconstruction properties. Under GF conditions, microglia were not polarized to a specific phenotype. M1 and M2 phenotype-related genes are only slightly different, whereas most differentially expressed genes are located in the M0 cluster, indicating that gut microbiota

plays an important role in regulating the microglia phenotype [19]. In a mouse model of postoperative cognitive impairment [26], prebiotic (galacto-oligosaccharide) treatment was found to affect the intestinal microbiota, which in turn affected the phenotype of microglia. Expression of the M1-type markers of microglia (*iNOS*, *CD6*, *CD32*, etc.) are generally increased in this mouse model, and may be related to the occurrence and development of postoperative cognitive impairment. Following treatment with galacto-oligosaccharide, the concentration of *bifidobacterial* in the mouse intestine increased significantly, while the M1 microglia markers were significantly down-regulated. Similar findings were made in mouse models of intracerebral hemorrhage (ICH) [27], with metformin shown to regulate the phenotype of microglia and thus improve neuroinflammation after ICH. Following metformin treatment, the microglia ratio of Iba⁺ decreased whereas the microglia ratio of *ARG-1*⁺, the M2 phenotype marker, increased [27]. Metformin was also found to improve the intestinal flora disorder caused by ICH, with a decrease in the ratio of *Firmicutes* and *Bacteroidetes* [27]. Finally, broad-spectrum antibiotics and fecal bacteria transplantation were used to prove the effects of metformin were dependent on intestinal flora [27]. Similar observations have been reported in other disease models, including AD [28,29] and PD [30,31].

The use of M1/M2 phenotypes helps to simplify our understanding of the functional heterogeneity of microglia. However, it remains controversial whether the the M1/2 phenotype concept, originally applied to peripheral macrophages, can also be applied to microglia [32]. Furthermore, three subsets (M2a, M2b, and M2c) were observed in M2 phenotype macrophages. M2a and M2c macrophages inhibit inflammation and promote tissue repair, while M2b macrophages have both proinflammatory and anti-inflammatory functions [33,34]. Whether these phenotypes also exist in microglia is still unknown. Nevertheless, a close relationship clearly exists between gut microbiota and the phenotype of microglia, thus providing an interesting framework to study the specific mechanism of the microbiota-gut-brain axis.

5. Gut Microbiota Regulates the Function of Microglia

The function of microglia is clearly affected by its morphology, phenotype and maturity, with the gut microbiota being closely linked to these influencing factors. Most studies on the association between intestinal microbiota and microglia function focus on the regulation of neuroinflammation by microglia. *Lactobacillus plantarum* MA2 isolated from traditional Tibetan kefir grains can improve cognitive disorder and anxiety-like behavior in a mouse model of AD, as well as reduce the deformation of neurons and $\alpha\beta$ accumulation in the brain [35]. The mechanism underlying this is the inhibition of microglia activation through the *TLR4/MYD88/NLRP3* signaling pathway, thereby reducing

secretion of the inflammatory factors IL-1 β and IL-18. A mouse model of postoperative emotional disorder showed disorder of the gut microbiota, with activated microglia and increased expression of the proinflammatory cytokines IL-6 and TNF- α [36]. These symptoms could be alleviated by antibiotic treatment. The gut microbiota was also found to upregulate the antigen presentation ability of microglia in a mouse model of virus-induced nerve injury [37]. The more severe symptoms of encephalic hepatitis virus infection in GF mice were found not to be caused by T-cell defects, but rather by external factors. Further research confirmed there was reduced expression of *MHC-II* and the co-stimulatory molecules *CD86* and *CD40* in microglia in the GF state, and that broad-spectrum antibiotics in SPF mice produced similar results. This study demonstrated that homeostasis of the gut microbiota is a key factor in maintaining the antigen presenting function of microglia. The lack of dietary fiber in a mouse model of cognitive impairment led to the disruption of intestinal epithelial barrier integrity and disorder of the gut microbiota, with decreased *Bacteroides* and increased *Proteobacteria* [38]. A lack of dietary fiber also results in activation of microglia, polarization to the M1 type, and increased production of the proinflammatory factors TNF- α , IL-6, and IL-1 β . There was also an increased number of activated microglia gathered around postsynaptic density protein, indicating activation of microglia-mediated synaptic phagocytosis. Finally, cognitive impairment was found to occur after an imbalance of intestinal microbiota, indicating that intestinal microbiota can regulate the synaptic phagocytosis of microglia. Nicotinamide n-oxide, a metabolite of intestinal flora, was recently found to improve the symptoms of herpes simplex encephalitis by activating mitochondrial autophagy of microglia [39]. The effects of intestinal microbiota on microglia function can also be seen in the pathological processes of many other diseases.

6. Gut Microbiota-Related Substances and Microglia

At present, the possible mechanisms of gut microbiota and its related products entering the central nervous system to regulate microglia mainly include direct and indirect ways. In the direct pathway, gut microbiota transmits information to the brain through stimulation of vagus nerve. In ischemic stroke, stimulation of the vagus nerve has been found to modulate microglia M1/M2 type activation [40]. Vagotomy may have the effect of delaying the progression of AD pathology [41]. In addition, metabolites of the intestinal flora including short-chain fatty acids, secondary bile acids and TMAO have modulatory effects on neurodegenerative diseases [42–44]. These products are characterized by low molecular weight and high lipid solubility, so they can cross the blood-brain barrier to the central nervous system. In the indirect way, the gut microbiota promotes the production of pro-inflammatory cytokines such as IL-1 α , IL-1 β , TNF-

α and IL-6 through pathogen-associated molecular patterns (PAMPs) [45]. These cytokines stimulate the development of CD4+ Th1 and TH7 cells [46]. Meanwhile, various neurotransmitters such as norepinephrine, dopamine, acetylcholine, and 5-hydroxytryptamine can be secreted to affect the organism [4]. In addition, GF mice have been found to have Foxp3 + Tregs lymphocyte defects [47] suggesting that gut microbiota can indirectly affect the immune status of central nervous system (CNS) by acting on the peripheral immune system (Fig. 1).

Microglia express a variety of innate immune-related pattern recognition receptors, including Toll-like receptors, NOD-like receptors, and scavenger receptors, which are essential for their recognition and response to PAMPs of gut microbiota. The transcription products of almost all Toll-like receptors are expressed in microglia, including TLR1-9, but not all TLRs are translated into proteins [48]. Only TLR1,2,3,4 and 9 have been found to be expressed at the protein level in primary rodent and human microglia cell lines [48]. TLR4 responds to bacterial LPS and promotes microglia paracrine activation [49], while TLR2 activation promotes antagonism against Gram-positive bacteria in the CNS [50,51]. TLRs, upon receiving LPS stimulation, pass through their intracellular part to promote cell signaling, induce microglia to polarize toward M1 type and secrete proinflammatory factors such as THF- α , IL1- β , and IL-12 [52,53]. NOD-like receptors are intracellular pattern recognition receptors located in the cell and play a role in natural immunity by forming inflammatory vesicles. Inflammatory vesicles within microglia have been found to play a role in *S. aureus* and prion infections [54,55].

The intestinal microbiota can anaerobically ferment undigested food in the colon into extremely diverse metabolites. The intestinal barrier, composed of intestinal epithelial cells, allows these metabolites to enter the cells and to regulate the immune system and disease [56].

Short-Chain Fatty Acids (SCFAs)

The gut microbiota can ferment undigested complex carbohydrates into organic acids, gases and short-chain fatty acids (SCFAs) through a series of complex enzymatic reactions. SCFAs include acetate, propionate and butyrate. Their concentration in the intestine is determined by the composition of the intestinal microbiota and the dietary fiber content of food. SCFAs have been found to regulate the immune status in humans and the development of multiple diseases [42,57–59].

Erym *et al.* [19] first reported that SCFAs produced by the gut microbiota could influence the maturation and function of microglia in the central nervous system. They found that dietary supplementation with SCFAs could promote partial re-maturation of immature microglia in the brains of GF mice and mice treated with broad-spectrum antibiotics. The expression of *CSFR1*, a marker of immature microglia, was reduced after supplementation with SCFA. In

a follow up study, these authors studied the mechanism by which acetate drives microglia maturation and functional changes [60]. Comprehensive analysis of mitochondrial function revealed selective complex II damage in the microglia mitochondria of GF mice. Acetate supplementation was found to restore defective mitochondrial function in microglia by replenishing tricarboxylic acid cycle intermediates. Intestinal-derived acetate labeled with C¹⁴ was converted to acetyl-CoA and stored in the mitochondria. The lack of acetate was speculated to result in the conversion of citric acid to acetyl-CoA in GF mice, causing oxaloacetate to increase which in turn caused subsequent functional impairment of complex II. In Thy1-aSYn mice (a mouse model in which human alpha-synuclein is overexpressed under the Thy1 promoter in many central and peripheral cells type including neurons, SCFAs resulted in significant microglia activation [61]. The PD phenotype could be prevented under GF conditions or by treatment with broad-spectrum antibiotics. Feeding a SCFA mix to a GF mouse model of PD resulted in microglia activation, accompanied by α -synuclein aggregation and motor deficits. Interestingly, another study found that sodium butyrate can cause degradation of α -synuclein via the PI3K/Akt/mTOR pathway [62].

Several studies have shown that SCFAs can cross the blood-brain barrier [63,64], suggesting they may act directly on microglia to affect their functions. SCFAs can be bound by specific receptors, such as Free Fatty Acid Receptor 2 and 3 (FFAR2, FFAR3) [65]. Although FFAR2 is not expressed in any adult brain cells including microglia and endothelial cells, *FFAR2* gene-deficient mice have similar microglia to GF mice [19,66]. This suggests that SCFAs can also bind to FFAR2 expressed by peripheral myeloid cells or lymphocytes (e.g., spleen or intestinal macrophages), thus indirectly sending signals to the central nervous system and thereby regulating the maturation and function of microglia. However, there is currently no evidence showing that brain cells express FFAR2 during early embryonic development. More research is therefore needed to understand the mechanism by which SCFAs affect microglia.

7. Gut Microbiota, Microglia, and Neuroinflammatory Diseases

7.1 Alzheimer's Disease

AD is the most common neurodegenerative disease and causes dementia in millions of people worldwide. Cerebral amyloidosis and abnormal protein deposition are typical pathological features of AD. Microglia in the brain proliferate, activate and concentrate around these amyloid plaques and protein depositions. Genetic studies have shown that most AD risk genes are highly and selectively expressed in microglia, including *TREM2*, *CD33* and *APOE* [67]. In-depth studies of the microbe-gut-brain axis have led to the hypothesis that gut microbes influence the pathological changes observed in AD by affecting microglia. Harach *et al.* [68] found significantly less amyloi-

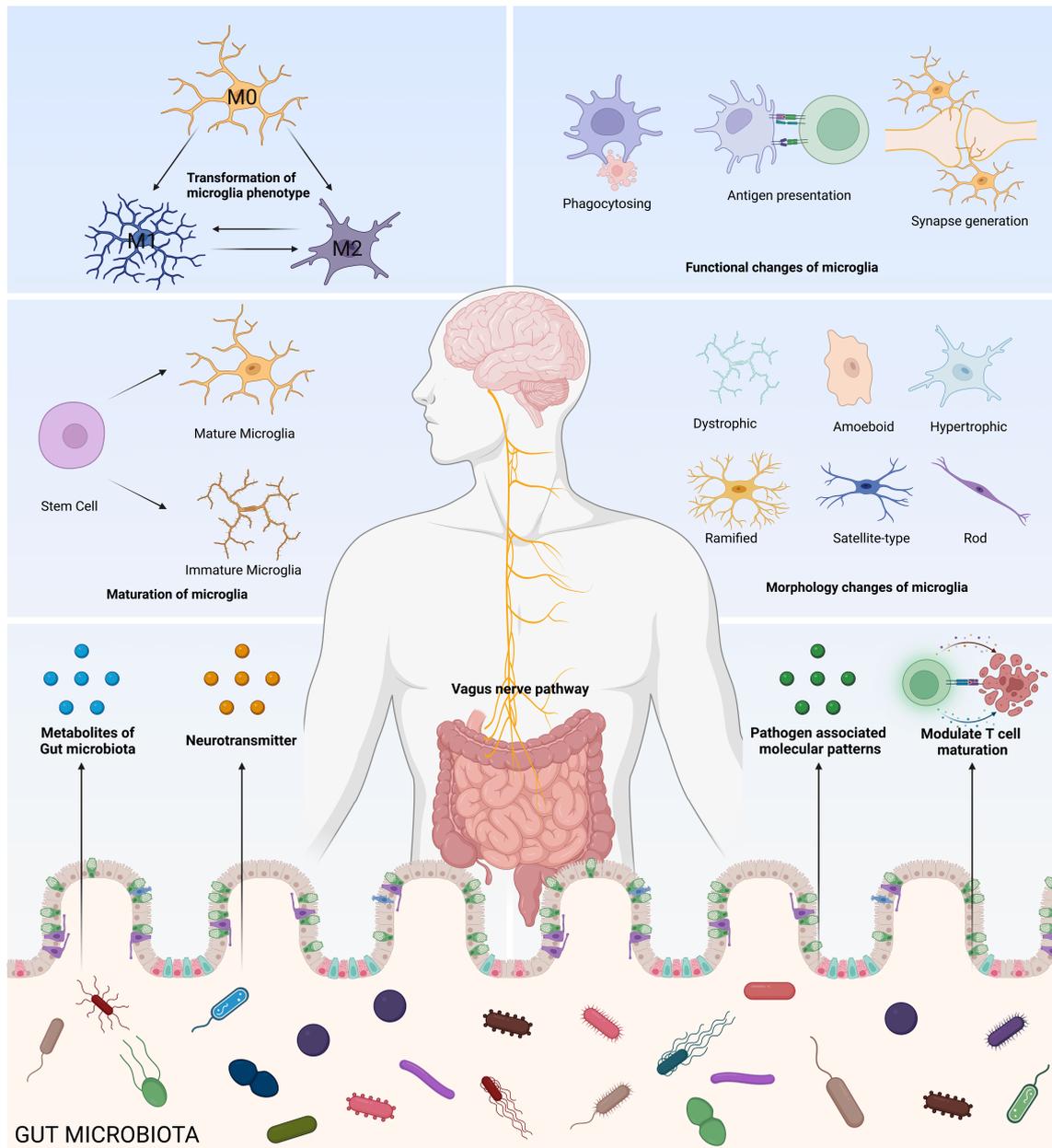


Fig. 1. Gut microbiota-associated substances modulate maturation, morphology, function and phenotype of microglia. Direct pathway includes gut microbiota metabolites, stimulation of the vagus nerve. Indirect pathway includes secretion of neurotransmitters, effect of PAMPs on innate immunity via PRR and regulation of T cell maturation.

dosis in the central nervous system of APPS1 transgenic mice (AD model containing human transgenes for both *APP* bearing the Swedish mutation and *PSEN1* containing an *L166P* mutation, both under the control of the *Thy1*) under GF conditions compared to SPF mice. However, the level of amyloidosis became similar after the transplantation of intestinal microbes from specific pathogen free APPS1 mice. The above studies suggest that the presence or absence of intestinal flora has an important impact on the pathology of AD. Changes in the composition of gut microbiota were also found in the $5 \times$ FAD mouse line, another widely used mouse model for AD [69]. In addition,

the diversity of fecal microbiota was observed to be significantly reduced in a small study of AD patients [70]. Treatment of an AD mouse model with broad-spectrum antibiotics significantly reduced the amyloidosis-related microglia and amyloidosis burden [71], but only in males. The gender difference in response of microglia to AD and intestinal microbiota was later confirmed in another mouse model (APPS1-21) [20]. In summary, the mechanisms by which intestinal microorganisms can affect the function of microglia are only just beginning to be understood and require further in-depth research.

7.2 Parkinson's Disease

PD is the second most common neurodegenerative disease after AD and is characterized by the impairment of movement and cognitive decline. PD is caused by the loss of dopaminergic neurons in the substantia nigra compact area, and accumulation of α -synuclein (α syn) in the endoneurium. Gut microbes have been shown to aggravate the severity of PD in the Thy1- α Syn (alpha-synuclein-overexpressing) mouse model [46]. Treatment of Thy1- α Syn mice with broad-spectrum antibiotics reduced the activation of microglia and the secretion of pro-inflammatory factors such as TNF- α and IL-6. These authors also found transplanted gut microbes from SPF-Thy1- α Syn mice to GF-Thy1- α Syn mice, or increasing the supply of SCFAs could result in more severe pathological manifestations of PD compared to the control group. Other investigators have reported that α -synuclein initially accumulates in the intestine only, and then transfers to the central nervous system through the vagus nerve, leading to PD-like pathological changes [72]. In support of this, vagus nerve amputation has been observed to delay the pathological process of PD in some sporadic cases [73,74]. In summary, there is now sufficient evidence to suggest a close regulatory relationship between intestinal microbes, microglia and PD. However, the specific mechanisms and functional changes require further clarification.

7.3 Multiple Sclerosis

MS is a chronic neurodegenerative disease of the central nervous system mediated by T cells. Immune cells from the central nervous system and from the peripheral immune system are involved in the pathological process of MS. The important role of gut microbiota in this process was demonstrated by Berer *et al.* [75]. These workers used a self-developed mouse model of MS to show that intestinal microbiota and myelin autoantigens jointly induced an autoimmune demyelination response. In follow-up work they found that *Akkermansia* may cause aggravation of MS symptoms. Using Experimental Autoimmune Encephalomyelitis (EAE) mice model (a model widely used to simulate MS), Haghika *et al.* [76] showed that long-chain fatty acids can sustainably reduce the concentration of SCFAs in the intestine, thereby increasing Th1 and Th17 cells in the small intestine and aggravating the disease symptoms. Meanwhile, in the EAE mouse model, treatment with a mixture of *Clostridium* strains reduced microglia activation rate and improved axonal damage. In addition, *Clostridium* treated mice were found to have elevated plasma butyrate levels, but treatment with butyrate alone did not produce a therapeutic effect on EAE mouse model [77]. Further, transplantation of fecal from healthy mice into EAE mice modulates the gut microbiota of EAE mouse model, improves the severity of EAE by alleviating blood-brain barrier (BBB) leakage and decreasing microglia activation [78]. It is not yet known how the gut mi-

crobiota regulates immune cells in the peripheral and central nervous system during the pathological process of MS, although antibiotic treatment of lecithin-induced MS mouse models was shown to alter the central nervous system. Inflammatory conditions such as the functional impairment of myelin fragments and the differentiation of oligodendrocyte progenitor cells were also shown to be affected by the activation of microglia. Recent studies have found that the Aryl Hydrocarbon Receptor (AHR) ligand derived from gut microbiota not only controls astrocytes but also binds to microglia that express AHR. This activates the microglia and increases their secretion of TGF- β and VEGF- β , thereby regulating the inflammatory response of astrocytes and microglia during the pathological process of EAE. Other studies have reported that AHR receptor-deficient microglia can aggravate EAE symptoms. The peripheral immune system was not affected, suggesting that AHR activation can alleviate symptoms of the central nervous system. The above studies indicate that microbial metabolism of tryptophan can affect the activation of astrocytes and microglia, thus providing new treatment options in the future.

7.4 Autism Spectrum Disorder (ASD)

ASD is a severe neurodevelopmental disorder characterized by age inappropriate, impaired social communication, and the presence of stereotypic behavior. During neurodevelopment, microglia are involved in regulating the number and strategic positioning of neurons and shaping neuronal connectivity [79,80], and also support gliogenesis and myelination [79,81]. Due to their important functions, microglia are thought to play an important role in ASD. In a study of pathological samples, Vargas *et al.* [82] found increased microglia activation throughout the cerebrum and cerebellar cortex of ASD patients, as evidenced by increased MHCII expression. In another study, it was also found that microglia density was increased in the brain and cerebellar cortex of patients with ASD and exhibited enlarged cell bodies, synaptic constriction and thickening [83,84]. Also, it was found that overproduction of proteins, such as overexpression of the transcription initiation factor EIF4e, in microglia alone was sufficient to cause mice to exhibit impaired synapse formation and ASD-like behavior [85]. Several studies have found alterations in the composition and abundance of the intestinal flora of children with ASD, with increased proportions of *Clostridium*, *Suterella*, *Ruminococcus*, *Lactobacillus*, and decreased proportions of *Bifidobacterium*, *Akkermansia*, *Blautia*, and *Prevotella* [86–91]. Transplantation of fecal microbiota from the neurotypical control donors group into ASD patients significantly improved irritability, communication skills and sociability [92,93]. Propionate in SCFAs increases inflammatory cytokine production and ASD-like behavior in mice by increasing the activation of microglia [94,95]. While butyrate was found to promote the transcription of genes related to [96] neuronal inhibitor pathways, thereby improv-

ing social behavior in BTBR mouse strains, an idiopathic ASD model. There is a significant relationship between microglia function, gut microbiota and ASD, which may provide us with new therapeutic targets, but the exact mechanisms remain to be fully established.

7.5 Major Depressive Disorder (MDD)

Depression is a major mental disorder, estimated to reach approximately 4.4% of the world population [97]. The main symptoms of Major Depressive Disorder include depressed mood, anhedonia, irritability, difficulty concentrating, changes in appetite and sleep, and others [98]. The main hypothetical theories about MDD include monoamine neurotransmitter depletion hypothesis, neuroplasticity hypothesis, hypothalamus–pituitary–adrenal [99]. Several studies have shown that microglia are closely related to the three hypotheses mentioned above [100,101]. Microglia play an important role in neuroinflammatory diseases because of their functions such as activation of inflammation, synaptic refinement, synaptic pruning, and neuronal connectivity [102]. Thus, MDD is also considered to be a microglia-related disease. Probiotic targeted regulation of gut microbiota has been found to be a new strategy for improving MDD. *Lactobacillus reuteri* NK33 and *Bifidobacterium adolescentis* NK98 were found to ameliorate restraint-induced stress anxiety/depression in mice [103]. A study by Zhang *et al.* [104] showed that administration of an anti-mouse IL-6 receptor antibody (MR16-1) led to antidepressant effects in a model of socially defeat stress and an improvement in *Firmicutes/Bacteroidetes* ratio in susceptible mice, suggesting that IL-6 blockade induces antidepressant effects by normalizing the composition of the altered gut microbiota. The kynurenine pathway (KP) plays an important role in the development of psychiatric disorders, including schizophrenia [105] and MDD [106]. Gut microbiota metabolites such as short-chain fatty acids can promote 5-hydroxytryptamine production via tryptophan, thereby modulating this pathway and preventing the conversion of tryptophan to kynurenine [107]. A small fraction of tryptophan is metabolized to 5-hydroxytryptamine through indoleamine-2,3-dioxygenase (IDO) enzymes, which are contained in all tissues, while another fraction enters the kynurenine pathway [108]. During intestinal inflammation, stimulation of the inflammatory cytokines, such as interferon γ (IFN- γ) and IL-6, induces the production of IDO and leads to changes in tryptophan metabolism, resulting in a shift in 5-hydroxytryptamine synthesis to the kynurenine metabolic pathway [109,110]. KP are divided into two pathways: (a) production of kynurenic acid (KYNA), and (b) production of quinolinic acid (QUIN) [111]. MDD is associated with overproduction of QUIN and a decrease in KYNA [112]. Investigators hypothesize that MDD is associated with pathological pathways that produce inflammatory KP, maintaining a persistent microglia imbalance and the per-

sistence of chronic inflammation [106,113].

8. Gut Microbiota, Microglia, and Tumors

Glioma

Gliomas are the most common and lethal tumor of the central nervous system in adults. About 50% of glioma patients have the most aggressive phenotype. The five-year survival rate of these patients is just 3.3%, with a median survival time of only 14.6 months [114]. Like other solid tumors, gliomas have a complex immune microenvironment that has important effects on tumor proliferation, invasion and metastasis [15]. Microglia are believed to play a role in the pathogenesis of gliomas by promoting the migration and proliferation of glial cells. The relative abundance of *Akkermansia* and *Bacteroides* increased after tumor growth in the syngeneic mouse model of glioma (GL261), but no obvious difference was found in 6 glioma patients [115]. Moreover, the concentrations of fecal microbiota metabolites such as SCFAs, 5-hydroxyindole acetic acid and norepinephrine were reduced. In another study using the GL261 mouse model, treatment with broad-spectrum antibiotics promoted the growth of mouse gliomas and altered the subpopulation and function of NK cells in the brain, bone marrow, and spleen [116]. Although they did not find a significant change in the density of microglia, these authors found that antibiotic treatment increased the expression of *Arg1*, *P2ry12* and *INOS* in the brain. Some researchers have speculated that the observed phenomenon may be due to the lack of signals from *Prevotellaceae*, *Rikenellaceae* and *Helicobacteraceae*, or to the up-regulation of signals from *Burkholderiales*. In summary, the research to date on regulation of microglia by intestinal microbes and the possible effects on glioma development is still preliminary. Further research is needed to clarify these relationships, including identification of the specific mechanisms or signaling pathways.

Multiple experiments have shown that the intestinal microbiota regulates peripheral immune cells, thereby affecting malignant tumors of the digestive system. For example, it was recently reported that the intestinal microbiota of colorectal cancer patients can stimulate cathepsin K secretion, mediate the polarization of TLR4 receptor-dependent M2 microglia, and promote tumor metastasis [117]. Microglia are considered to be macrophages in the central nervous system. Intestinal microbiota could in theory also change the polarization state of microglia, thereby affecting the microenvironment of central nervous system tumors and of brain metastases.

Recent studies have shown that the intestinal microbiota can affect not only the occurrence, invasion and metastasis of tumors, but may also impact the efficacy of tumor treatment. The efficacy of cyclophosphamide, a traditional and widely used cancer drug, in a mouse tumor model has been shown to depend on the intestinal bacteria. Mice with solid tumors showed a reduced Th17 response under

sterile conditions or with the use of broad-spectrum antibiotics. The tumors were also resistant to cyclophosphamide. Specific strains (e.g., *B. fragilis*) in GF mice may overcome tumor resistance to CTLA-4 immunotherapy [118]. In light of the above findings it is likely that microglia, as the only resident immune cell in the brain, are also regulated by the intestinal microbiota. Therefore, microglia occupy an important position in the development of brain tumors (primary or metastatic) and in treatment response. The intestinal microbiota can affect brain tumors by regulating microglia, suggesting this axis may provide a novel therapeutic target as well as new directions for future studies of brain diseases.

9. Discussion

Perhaps five years ago, it would have been difficult to imagine such a broad and tight connection between the intestinal microbiota and microglia, which could serve as an axis through a wide variety of disease pathologies and as potential therapeutic targets. Because of the importance of these connections, we need to go back to the roots of these connections and dig deeper into the underlying mechanisms. First, the pathways through which the gut microbiota affect microglia are diverse and can be functionally oriented to investigate the specific mechanisms of each regulatory pathway and possible synergistic effects in disease models. Second, the regulation of metabolism on the polarization state of microglia can be studied from a metabolomics perspective, similar to the regulation of peripheral macrophage metabolism. Third, the existence of the concept of M1/M2 type polarization of microglia is still controversial and can be studied from the perspective of individual cell genomics, proteomics and metabolomics to expand the understanding of the microglia phenotype.

Author Contributions

XD and FT designed the study. LL, HL, YB, PD, LP and ZL conducted data collection. LL wrote the first draft of the manuscript. All authors contributed to the manuscript revision and read and approved the final version. All authors have participated sufficiently in the work to take public responsibility for appropriate portions of the content and agreed to be accountable for all aspects of the work in ensuring that questions related to its accuracy or integrity.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

This work was supported by the National Natural Science Foundation of China (81573090, 81172595,

81703165).

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Sgritta M, Dooling SW, Buffington SA, Momin EN, Francis MB, Britton RA, *et al.* Mechanisms Underlying Microbial-Mediated Changes in Social Behavior in Mouse Models of Autism Spectrum Disorder. *Neuron*. 2019; 101: 246–259.e6.
- [2] Li N, Wang Q, Wang Y, Sun A, Lin Y, Jin Y, *et al.* Fecal microbiota transplantation from chronic unpredictable mild stress mice donors affects anxiety-like and depression-like behavior in recipient mice via the gut microbiota-inflammation-brain axis. *Stress (Amsterdam, Netherlands)*. 2019; 22: 592–602.
- [3] Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, *et al.* Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proceedings of the National Academy of Sciences of the United States of America*. 2011; 108: 16050–16055.
- [4] Ridaura V, Belkaid Y. Gut microbiota: the link to your second brain. *Cell*. 2015; 161: 193–194.
- [5] Musso G, Gambino R, Cassader M. Obesity, diabetes, and gut microbiota: the hygiene hypothesis expanded? *Diabetes Care*. 2010; 33: 2277–2284.
- [6] Banks WA. The blood-brain barrier in psychoneuroimmunology. *Neurologic Clinics*. 2006; 24: 413–419.
- [7] Liu S, da Cunha AP, Rezende RM, Cialic R, Wei Z, Bry L, *et al.* The Host Shapes the Gut Microbiota via Fecal MicroRNA. *Cell Host & Microbe*. 2016; 19: 32–43.
- [8] Wakselman S, Béchade C, Roumier A, Bernard D, Triller A, Bessis A. Developmental neuronal death in hippocampus requires the microglial CD11b integrin and DAPI2 immunoreceptor. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2008; 28: 8138–8143.
- [9] Roumier A, Béchade C, Poncer J, Smalla K, Tomasello E, Vivier E, *et al.* Impaired synaptic function in the microglial KARAP/DAPI2-deficient mouse. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2004; 24: 11421–11428.
- [10] Ji K, Akgul G, Wollmuth LP, Tsirka SE. Microglia actively regulate the number of functional synapses. *PLoS ONE*. 2013; 8: e56293.
- [11] Colonna M, Butovsky O. Microglia Function in the Central Nervous System During Health and Neurodegeneration. *Annual Review of Immunology*. 2017; 35: 441–468.
- [12] Hansen DV, Hanson JE, Sheng M. Microglia in Alzheimer's disease. *The Journal of Cell Biology*. 2018; 217: 459–472.
- [13] Ho MS. Microglia in Parkinson's Disease. *Advances in Experimental Medicine and Biology*. 2019; 1175: 335–353.
- [14] Voet S, Prinz M, van Loo G. Microglia in Central Nervous System Inflammation and Multiple Sclerosis Pathology. *Trends in Molecular Medicine*. 2019; 25: 112–123.
- [15] Hambardzumyan D, Gutmann DH, Kettenmann H. The role of microglia and macrophages in glioma maintenance and progression. *Nature Neuroscience*. 2016; 19: 20–27.
- [16] Savage JC, Carrier M, Tremblay M. Morphology of Microglia Across Contexts of Health and Disease. *Methods in Molecular Biology (Clifton, N.J.)*. 2019; 2034: 13–26.
- [17] Graeber MB. Changing face of microglia. *Science (New York, N.Y.)*. 2010; 330: 783–788.
- [18] Boche D, Perry VH, Nicoll JAR. Review: activation patterns of microglia and their identification in the human brain. *Neu-*

- ropathology and Applied Neurobiology. 2013; 39: 3–18.
- [19] Erny D, Hrabě de Angelis AL, Jaitin D, Wieghofer P, Staszewski O, David E, *et al.* Host microbiota constantly control maturation and function of microglia in the CNS. *Nature Neuroscience*. 2015; 18: 965–977.
- [20] Dodiya HB, Kuntz T, Shaik SM, Baufeld C, Leibowitz J, Zhang X, *et al.* Sex-specific effects of microbiome perturbations on cerebral A β amyloidosis and microglia phenotypes. *The Journal of Experimental Medicine*. 2019; 216: 1542–1560.
- [21] Bauer KC, York EM, Cirstea MS, Radisavljevic N, Petersen C, Huus KE, *et al.* Gut microbes shape microglia and cognitive function during malnutrition. *Glia*. 2022; 70: 820–841.
- [22] Thion MS, Low D, Silvin A, Chen J, Grisel P, Schulte-Schrepping J, *et al.* Microbiome Influences Prenatal and Adult Microglia in a Sex-Specific Manner. *Cell*. 2018; 172: 500–516.e16.
- [23] Orihuela R, McPherson CA, Harry GJ. Microglial M1/M2 polarization and metabolic states. *British Journal of Pharmacology*. 2016; 173: 649–665.
- [24] Prinz M, Priller J. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. *Nature Reviews. Neuroscience*. 2014; 15: 300–312.
- [25] Durafourt BA, Moore CS, Zammit DA, Johnson TA, Zaguia F, Guiot M, *et al.* Comparison of polarization properties of human adult microglia and blood-derived macrophages. *Glia*. 2012; 60: 717–727.
- [26] Yang X, Wang L, Wu H, Jiao L. Effects of prebiotic galactooligosaccharide on postoperative cognitive dysfunction and neuroinflammation through targeting of the gut-brain axis. *BMC Anesthesiology*. 2018; 18: 177.
- [27] Yu X, Fu X, Wu X, Tang W, Xu L, Hu L, *et al.* Metformin Alleviates Neuroinflammation Following Intracerebral Hemorrhage in Mice by Regulating Microglia/Macrophage Phenotype in a Gut Microbiota-Dependent Manner. *Frontiers in Cellular Neuroscience*. 2022; 15: 789471.
- [28] Soriano S, Curry K, Wang Q, Chow E, Treangen TJ, Villapol S. Fecal Microbiota Transplantation Derived from Alzheimer's Disease Mice Worsens Brain Trauma Outcomes in Wild-Type Controls. *International Journal of Molecular Sciences*. 2022; 23: 4476.
- [29] Chen C, Liao J, Xia Y, Liu X, Jones R, Haran J, *et al.* Gut microbiota regulate Alzheimer's disease pathologies and cognitive disorders via PUFA-associated neuroinflammation. *Gut*. 2022; 71: 2233–2252.
- [30] Zhou X, Lu J, Wei K, Wei J, Tian P, Yue M, *et al.* Neuroprotective Effect of Ceftriaxone on MPTP-Induced Parkinson's Disease Mouse Model by Regulating Inflammation and Intestinal Microbiota. *Oxidative Medicine and Cellular Longevity*. 2021; 2021: 9424582.
- [31] Sun J, Li H, Jin Y, Yu J, Mao S, Su K, *et al.* Probiotic *Clostridium butyricum* ameliorated motor deficits in a mouse model of Parkinson's disease via gut microbiota-GLP-1 pathway. *Brain, Behavior, and Immunity*. 2021; 91: 703–715.
- [32] Ransohoff RM. A polarizing question: do M1 and M2 microglia exist? *Nature Neuroscience*. 2016; 19: 987–991.
- [33] Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nature Immunology*. 2010; 11: 889–896.
- [34] David S, Kroner A. Repertoire of microglial and macrophage responses after spinal cord injury. *Nature Reviews. Neuroscience*. 2011; 12: 388–399.
- [35] Wang Y, Wang D, Lv H, Dong Q, Li J, Geng W, *et al.* Modulation of the Gut Microbiota and Glycometabolism by a Probiotic to Alleviate Amyloid Accumulation and Cognitive Impairments in AD Rats. *Molecular Nutrition & Food Research*. 2022; 66: e2200265.
- [36] Lei L, Ji M, Yang J, Chen S, Gu H, Yang J. Gut microbiota-mediated metabolic restructuring aggravates emotional deficits after anesthesia/surgery in rats with preoperative stress. *Frontiers in Immunology*. 2022; 13: 819289.
- [37] Brown DG, Soto R, Yandamuri S, Stone C, Dickey L, Gomes-Neto JC, *et al.* The microbiota protects from viral-induced neurologic damage through microglia-intrinsic TLR signaling. *ELife*. 2019; 8: e47117.
- [38] Shi H, Ge X, Ma X, Zheng M, Cui X, Pan W, *et al.* A fiber-deprived diet causes cognitive impairment and hippocampal microglia-mediated synaptic loss through the gut microbiota and metabolites. *Microbiome*. 2021; 9: 223.
- [39] Li F, Wang Y, Song X, Wang Z, Jia J, Qing S, *et al.* The intestinal microbial metabolite nicotinamide n-oxide prevents herpes simplex encephalitis via activating mitophagy in microglia. *Gut Microbes*. 2022; 14: 2096989.
- [40] Zhang L, Liu Y, Wang S, Long L, Zang Q, Ma J, *et al.* Vagus nerve stimulation mediates microglia M1/2 polarization via inhibition of TLR4 pathway after ischemic stroke. *Biochemical and Biophysical Research Communications*. 2021; 577: 71–79.
- [41] Lee K, Kim J, Han S, Lee DY, Lee H, Yim S, *et al.* The extracellular vesicle of gut microbial *Paenalcaldigenes hominis* is a risk factor for vagus nerve-mediated cognitive impairment. *Microbiome*. 2020; 8: 107.
- [42] Colombo AV, Sadler RK, Llovera G, Singh V, Roth S, Heindl S, *et al.* Microbiota-derived short chain fatty acids modulate microglia and promote A β plaque deposition. *ELife*. 2021; 10: e59826.
- [43] Huang R, Gao Y, Chen J, Duan Q, He P, Zhang J, *et al.* TGR5 Agonist INT-777 Alleviates Inflammatory Neurodegeneration in Parkinson's Disease Mouse Model by Modulating Mitochondrial Dynamics in Microglia. *Neuroscience*. 2022; 490: 100–119.
- [44] Zarbock KR, Han JH, Singh AP, Thomas SP, Bendlin BB, Denu JM, *et al.* Trimethylamine N-Oxide Reduces Neurite Density and Plaque Intensity in a Murine Model of Alzheimer's Disease. *Journal of Alzheimer's Disease: JAD*. 2022; 90: 585–597.
- [45] Dantzer R, Konsman JP, Bluthé RM, Kelley KW. Neural and humoral pathways of communication from the immune system to the brain: parallel or convergent? *Autonomic Neuroscience: Basic & Clinical*. 2000; 85: 60–65.
- [46] Maynard CL, Elson CO, Hatton RD, Weaver CT. Reciprocal interactions of the intestinal microbiota and immune system. *Nature*. 2012; 489: 231–241.
- [47] Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y, *et al.* Induction of colonic regulatory T cells by indigenous *Clostridium* species. *Science (New York, N.Y.)*. 2011; 331: 337–341.
- [48] Lee H, Lee S, Cho I, Lee SJ. Toll-like receptors: sensor molecules for detecting damage to the nervous system. *Current Protein & Peptide Science*. 2013; 14: 33–42.
- [49] Laflamme N, Rivest S. Toll-like receptor 4: the missing link of the cerebral innate immune response triggered by circulating gram-negative bacterial cell wall components. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2001; 15: 155–163.
- [50] Kielian T, Esen N, Bearden ED. Toll-like receptor 2 (TLR2) is pivotal for recognition of *S. aureus* peptidoglycan but not intact bacteria by microglia. *Glia*. 2005; 49: 567–576.
- [51] Esen N, Tanga FY, DeLeo JA, Kielian T. Toll-like receptor 2 (TLR2) mediates astrocyte activation in response to the Gram-positive bacterium *Staphylococcus aureus*. *Journal of Neurochemistry*. 2004; 88: 746–758.
- [52] Gordon S, Taylor PR. Monocyte and macrophage heterogeneity. *Nature Reviews. Immunology*. 2005; 5: 953–964.
- [53] Villalta SA, Nguyen HX, Deng B, Gotoh T, Tidball JG. Shifts in

- macrophage phenotypes and macrophage competition for arginine metabolism affect the severity of muscle pathology in muscular dystrophy. *Human Molecular Genetics*. 2009; 18: 482–496.
- [54] Hanamsagar R, Torres V, Kielian T. Inflammasome activation and IL-1 β /IL-18 processing are influenced by distinct pathways in microglia. *Journal of Neurochemistry*. 2011; 119: 736–748.
- [55] Shi F, Yang Y, Kouadir M, Fu Y, Yang L, Zhou X, *et al.* Inhibition of phagocytosis and lysosomal acidification suppresses neurotoxic prion peptide-induced NALP3 inflammasome activation in BV2 microglia. *Journal of Neuroimmunology*. 2013; 260: 121–125.
- [56] Rooks MG, Garrett WS. Gut microbiota, metabolites and host immunity. *Nature Reviews. Immunology*. 2016; 16: 341–352.
- [57] Song L, Sun Q, Zheng H, Zhang Y, Wang Y, Liu S, *et al.* Roseburia hominis Alleviates Neuroinflammation via Short-Chain Fatty Acids through Histone Deacetylase Inhibition. *Molecular Nutrition & Food Research*. 2022; 66: e2200164.
- [58] Luo P, Lednovich K, Xu K, Nnyamah C, Layden BT, Xu P. Central and peripheral regulations mediated by short-chain fatty acids on energy homeostasis. *Translational Research: the Journal of Laboratory and Clinical Medicine*. 2022; 248: 128–150.
- [59] Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, *et al.* The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science (New York, N.Y.)*. 2013; 341: 569–573.
- [60] Erny D, Dokalis N, Mezö C, Castoldi A, Mossad O, Staszewski O, *et al.* Microbiota-derived acetate enables the metabolic fitness of the brain innate immune system during health and disease. *Cell Metabolism*. 2021; 33: 2260–2276.e7.
- [61] Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, *et al.* Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson's Disease. *Cell*. 2016; 167: 1469–1480.e12.
- [62] Qiao C, Sun M, Jia X, Shi Y, Zhang B, Zhou Z, *et al.* Sodium butyrate causes α -synuclein degradation by an Atg5-dependent and PI3K/Akt/mTOR-related autophagy pathway. *Experimental Cell Research*. 2020; 387: 111772.
- [63] Frost G, Sleeth ML, Sahuri-Arisoylu M, Lizarbe B, Cerdan S, Brody L, *et al.* The short-chain fatty acid acetate reduces appetite via a central homeostatic mechanism. *Nature Communications*. 2014; 5: 3611.
- [64] Huuskonen J, Suuronen T, Nuutinen T, Kyrölenko S, Salminen A. Regulation of microglial inflammatory response by sodium butyrate and short-chain fatty acids. *British Journal of Pharmacology*. 2004; 141: 874–880.
- [65] Layden BT, Angueira AR, Brodsky M, Durai V, Lowe WL. Short chain fatty acids and their receptors: new metabolic targets. *Translational Research: the Journal of Laboratory and Clinical Medicine*. 2013; 161: 131–140.
- [66] Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, *et al.* The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *The Journal of Biological Chemistry*. 2003; 278: 11312–11319.
- [67] Prinz M, Priller J. The role of peripheral immune cells in the CNS in steady state and disease. *Nature Neuroscience*. 2017; 20: 136–144.
- [68] Harach T, Marungruang N, Duthilleul N, Cheatham V, Mc Coy KD, Frisoni G, *et al.* Reduction of A β amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. *Scientific Reports*. 2017; 7: 41802.
- [69] Brandscheid C, Schuck F, Reinhardt S, Schäfer K, Pietrzik CU, Grimm M, *et al.* Altered Gut Microbiome Composition and Tryptic Activity of the 5xFAD Alzheimer's Mouse Model. *Journal of Alzheimer's Disease: JAD*. 2017; 56: 775–788.
- [70] Vogt NM, Kerby RL, Dill-McFarland KA, Harding SJ, Merluzzi AP, Johnson SC, *et al.* Gut microbiome alterations in Alzheimer's disease. *Scientific Reports*. 2017; 7: 13537.
- [71] Minter MR, Zhang C, Leone V, Ringus DL, Zhang X, Oyler-Castrillo P, *et al.* Antibiotic-induced perturbations in gut microbial diversity influences neuro-inflammation and amyloidosis in a murine model of Alzheimer's disease. *Scientific Reports*. 2016; 6: 30028.
- [72] Kim S, Kwon S, Kam T, Panicker N, Karuppagounder SS, Lee S, *et al.* Transneuronal Propagation of Pathologic α -Synuclein from the Gut to the Brain Models Parkinson's Disease. *Neuron*. 2019; 103: 627–641.e7.
- [73] Bencsik A, Muselli L, Leboindre M, Lakhdar L, Baron T. Early and persistent expression of phosphorylated α -synuclein in the enteric nervous system of A53T mutant human α -synuclein transgenic mice. *Journal of Neuropathology and Experimental Neurology*. 2014; 73: 1144–1151.
- [74] Holmqvist S, Chutna O, Bousset L, Aldrin-Kirk P, Li W, Björklund T, *et al.* Direct evidence of Parkinson pathology spread from the gastrointestinal tract to the brain in rats. *Acta Neuropathologica*. 2014; 128: 805–820.
- [75] Berer K, Gerdes LA, Cekanaviciute E, Jia X, Xiao L, Xia Z, *et al.* Gut microbiota from multiple sclerosis patients enables spontaneous autoimmune encephalomyelitis in mice. *Proceedings of the National Academy of Sciences of the United States of America*. 2017; 114: 10719–10724.
- [76] Haghikia A, Jörg S, Duscha A, Berg J, Manzel A, Waschbisch A, *et al.* Dietary Fatty Acids Directly Impact Central Nervous System Autoimmunity via the Small Intestine. *Immunity*. 2015; 43: 817–829.
- [77] Calvo-Barreiro L, Eixarch H, Cornejo T, Costa C, Castillo M, Mestre L, *et al.* Selected Clostridia Strains from The Human Microbiota and their Metabolite, Butyrate, Improve Experimental Autoimmune Encephalomyelitis. *Neurotherapeutics: the Journal of the American Society for Experimental NeuroTherapeutics*. 2021; 18: 920–937.
- [78] Li K, Wei S, Hu L, Yin X, Mai Y, Jiang C, *et al.* Protection of Fecal Microbiota Transplantation in a Mouse Model of Multiple Sclerosis. *Mediators of Inflammation*. 2020; 2020: 2058272.
- [79] Zhan Y, Paolicelli RC, Sforzini F, Weinhard L, Bolasco G, Pagani F, *et al.* Deficient neuron-microglia signaling results in impaired functional brain connectivity and social behavior. *Nature Neuroscience*. 2014; 17: 400–406.
- [80] Cheadle L, Rivera SA, Phelps JS, Ennis KA, Stevens B, Burkly LC, *et al.* Sensory Experience Engages Microglia to Shape Neural Connectivity through a Non-Phagocytic Mechanism. *Neuron*. 2020; 108: 451–468.e9.
- [81] Miron VE, Boyd A, Zhao J, Yuen TJ, Ruckh JM, Shadrach JL, *et al.* M2 microglia and macrophages drive oligodendrocyte differentiation during CNS remyelination. *Nature Neuroscience*. 2013; 16: 1211–1218.
- [82] Vargas DL, Nascimbene C, Krishnan C, Zimmerman AW, Pardo CA. Neuroglial activation and neuroinflammation in the brain of patients with autism. *Annals of Neurology*. 2005; 57: 67–81.
- [83] Morgan JT, Chana G, Pardo CA, Achim C, Semendeferi K, Buckwalter J, *et al.* Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biological Psychiatry*. 2010; 68: 368–376.
- [84] Tetreault NA, Hakeem AY, Jiang S, Williams BA, Allman E, Wold BJ, *et al.* Microglia in the cerebral cortex in autism. *Journal of Autism and Developmental Disorders*. 2012; 42: 2569–2584.
- [85] Xu Z, Kim GH, Tan J, Riso AE, Sun Y, Xu EY, *et al.* Elevated protein synthesis in microglia causes autism-like synaptic and behavioral aberrations. *Nature Communications*. 2020; 11: 1797.
- [86] Wang L, Christophersen CT, Soricich MJ, Gerber JP, Angley MT,

- Conlon MA. Increased abundance of *Sutterella* spp. and *Ruminococcus torques* in feces of children with autism spectrum disorder. *Molecular Autism*. 2013; 4: 42.
- [87] Strati F, Cavalieri D, Albanese D, De Felice C, Donati C, Hayek J, *et al.* New evidences on the altered gut microbiota in autism spectrum disorders. *Microbiome*. 2017; 5: 24.
- [88] Liu F, Li J, Wu F, Zheng H, Peng Q, Zhou H. Altered composition and function of intestinal microbiota in autism spectrum disorders: a systematic review. *Translational Psychiatry*. 2019; 9: 43.
- [89] Williams BL, Hornig M, Parekh T, Lipkin WI. Application of novel PCR-based methods for detection, quantitation, and phylogenetic characterization of *Sutterella* species in intestinal biopsy samples from children with autism and gastrointestinal disturbances. *MBio*. 2012; 3: e00261–11.
- [90] Wang L, Christophersen CT, Sorich MJ, Gerber JP, Angley MT, Conlon MA. Low relative abundances of the mucolytic bacterium *Akkermansia muciniphila* and *Bifidobacterium* spp. in feces of children with autism. *Applied and Environmental Microbiology*. 2011; 77: 6718–6721.
- [91] Finegold SM, Molitoris D, Song Y, Liu C, Vaisanen M, Bolte E, *et al.* Gastrointestinal microflora studies in late-onset autism. *Clinical Infectious Diseases: an Official Publication of the Infectious Diseases Society of America*. 2002; 35: S6–S16.
- [92] Kang D, Adams JB, Coleman DM, Pollard EL, Maldonado J, McDonough-Means S, *et al.* Long-term benefit of Microbiota Transfer Therapy on autism symptoms and gut microbiota. *Scientific Reports*. 2019; 9: 5821.
- [93] Kang D, Adams JB, Gregory AC, Borody T, Chittick L, Fasano A, *et al.* Microbiota Transfer Therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: an open-label study. *Microbiome*. 2017; 5: 10.
- [94] Choi J, Lee S, Won J, Jin Y, Hong Y, Hur T, *et al.* Pathophysiological and neurobehavioral characteristics of a propionic acid-mediated autism-like rat model. *PLoS ONE*. 2018; 13: e0192925.
- [95] Silva YP, Bernardi A, Frozza RL. The Role of Short-Chain Fatty Acids From Gut Microbiota in Gut-Brain Communication. *Frontiers in Endocrinology*. 2020; 11: 25.
- [96] Kratsman N, Getselter D, Elliott E. Sodium butyrate attenuates social behavior deficits and modifies the transcription of inhibitory/excitatory genes in the frontal cortex of an autism model. *Neuropharmacology*. 2016; 102: 136–145.
- [97] Smith K. Mental health: a world of depression. *Nature*. 2014; 515: 181.
- [98] Nestler EJ, Barrot M, DiLeone RJ, Eisch AJ, Gold SJ, Monteggia LM. Neurobiology of depression. *Neuron*. 2002; 34: 13–25.
- [99] Jesulola E, Micalos P, Baguley JJ. Understanding the pathophysiology of depression: From monoamines to the neurogenesis hypothesis model - are we there yet? *Behavioural Brain Research*. 2018; 341: 79–90.
- [100] Troubat R, Barone P, Leman S, Desmidt T, Cressant A, Atanasova B, *et al.* Neuroinflammation and depression: A review. *The European Journal of Neuroscience*. 2021; 53: 151–171.
- [101] Miller AH, Raison CL. The role of inflammation in depression: from evolutionary imperative to modern treatment target. *Nature Reviews. Immunology*. 2016; 16: 22–34.
- [102] Deng S, Chen J, Wang F. Microglia: A Central Player in Depression. *Current Medical Science*. 2020; 40: 391–400.
- [103] Jang H, Lee K, Kim D. The Preventive and Curative Effects of *Lactobacillus reuteri* NK33 and *Bifidobacterium adolescentis* NK98 on Immobilization Stress-Induced Anxiety/Depression and Colitis in Mice. *Nutrients*. 2019; 11: 819.
- [104] Zhang J, Yao W, Dong C, Yang C, Ren Q, Ma M, *et al.* Blockade of interleukin-6 receptor in the periphery promotes rapid and sustained antidepressant actions: a possible role of gut-microbiota-brain axis. *Translational Psychiatry*. 2017; 7: e1138.
- [105] Zhu F, Guo R, Wang W, Ju Y, Wang Q, Ma Q, *et al.* Transplantation of microbiota from drug-free patients with schizophrenia causes schizophrenia-like abnormal behaviors and dysregulated kynurenine metabolism in mice. *Molecular Psychiatry*. 2020; 25: 2905–2918.
- [106] Réus GZ, Jansen K, Titus S, Carvalho AF, Gabbay V, Quevedo J. Kynurenine pathway dysfunction in the pathophysiology and treatment of depression: Evidences from animal and human studies. *Journal of Psychiatric Research*. 2015; 68: 316–328.
- [107] Reigstad CS, Salmons CE, Rainey JF, Szurszewski JH, Linden DR, Sonnenburg JL, *et al.* Gut microbes promote colonic serotonin production through an effect of short-chain fatty acids on enterochromaffin cells. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2015; 29: 1395–1403.
- [108] Salter M, Pogson CI. The role of tryptophan 2,3-dioxygenase in the hormonal control of tryptophan metabolism in isolated rat liver cells. Effects of glucocorticoids and experimental diabetes. *The Biochemical Journal*. 1985; 229: 499–504.
- [109] Jürgens B, Hainz U, Fuchs D, Felzmann T, Heitger A. Interferon-gamma-triggered indoleamine 2,3-dioxygenase competence in human monocyte-derived dendritic cells induces regulatory activity in allogeneic T cells. *Blood*. 2009; 114: 3235–3243.
- [110] Yeung AWS, Terentis AC, King NJC, Thomas SR. Role of indoleamine 2,3-dioxygenase in health and disease. *Clinical Science (London, England: 1979)*. 2015; 129: 601–672.
- [111] Savitz J, Drevets WC, Wurfel BE, Ford BN, Bellgowan PSF, Victor TA, *et al.* Reduction of kynurenic acid to quinolinic acid ratio in both the depressed and remitted phases of major depressive disorder. *Brain, Behavior, and Immunity*. 2015; 46: 55–59.
- [112] Grant RS, Naif H, Espinosa M, Kapoor V. IDO induction in IFN-gamma activated astroglia: a role in improving cell viability during oxidative stress. *Redox Report: Communications in Free Radical Research*. 2000; 5: 101–104.
- [113] Myint AM, Kim YK. Cytokine-serotonin interaction through IDO: a neurodegeneration hypothesis of depression. *Medical Hypotheses*. 2003; 61: 519–525.
- [114] Thakkar JP, Dolecek TA, Horbinski C, Ostrom QT, Lightner DD, Barnholtz-Sloan JS, *et al.* Epidemiologic and molecular prognostic review of glioblastoma. *Cancer Epidemiology, Biomarkers & Prevention: a Publication of the American Association for Cancer Research, Cosponsored by the American Society of Preventive Oncology*. 2014; 23: 1985–1996.
- [115] Dono A, Patrizz A, McCormack RM, Putluri N, Ganesh BP, Kaur B, *et al.* Glioma induced alterations in fecal short-chain fatty acids and neurotransmitters. *CNS Oncology*. 2020; 9: CNS57.
- [116] D’Alessandro G, Antonangeli F, Marrocco F, Porzia A, Lauro C, Santoni A, *et al.* Gut microbiota alterations affect glioma growth and innate immune cells involved in tumor immunosurveillance in mice. *European Journal of Immunology*. 2020; 50: 705–711.
- [117] Li R, Zhou R, Wang H, Li W, Pan M, Yao X, *et al.* Gut microbiota-stimulated cathepsin K secretion mediates TLR4-dependent M2 macrophage polarization and promotes tumor metastasis in colorectal cancer. *Cell Death and Differentiation*. 2019; 26: 2447–2463.
- [118] Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillère R, Han-nani D, *et al.* The intestinal microbiota modulates the anti-cancer immune effects of cyclophosphamide. *Science (New York, N.Y.)*. 2013; 342: 971–976.